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Characterization of Pulmonary and Myocardial Beta-Adrenoceptors with S-1'-[Fluorine-18]Fluorocarazolol

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S-1'-[¹⁸F]fluorocarazolol was administered to healthy volunteers to assess its potential for noninvasive measurement of regional pulmonary and myocardial beta-adrenoceptor densities. Methods: Highspecific activity fluorocarazolol was intravenously injected on two separate occasions within a 1-wk interval. The initial injection was without pretreatment, but before the second injection, the volunteers either inhaled salbutamol (2 \times 200 μ g aerosol) or they ingested pindolol (3 \times 5 mg during a 12-hr interval). Twenty-eight PET time frames of 31 planes were acquired over a period of 60 min after each injection. Blood samples were drawn and analyzed for the presence of fluorocarazolol and radioactive metabolites. Results: Uptake of fluorocarazolol in the target tissues was hardly affected by salbutamol but was strongly depressed by pindolol. Pulmonary and myocardial tissue-to-plasma concentration ratios of fluorocarazolol reached plateau values of 11.6 \pm 0.6 (lungs) and 18.1 \pm 1.0 (heart) at 45–50 min postinjection. These values were reduced to 2.0 \pm 0.4 and 2.0 \pm 0.6 after treatment with pindolol. Conclusion: These data indicate that:

- Pulmonary and myocardial uptake of radioactivity after intravenous administration of S-1'-[¹⁸F]fluorocarazolol represents radioligand binding to beta-adrenoceptors.
- 2. Pulmonary binding occurs mainly in alveoli rather than in airway smooth muscle under these conditions.
- 3. Binding kinetics do not preclude quantification of receptors with compartment models.

Key Words: beta-adrenoceptor density; fluorine-18-fluorocarazolol; PET

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PET with appropriate radioligands offers the possibility of studying receptors noninvasively in man. Carazolol, a lipophilic, nonsubtype-selective beta-adrenoceptor antagonist, can be labeled with the positron emitters $^{11}C(1,2)$ or $^{18}F(3)$. The suitability of carazolol for PET studies of beta-adrenoceptors has been demonstrated in mice, rats and minipigs (1-6). Uptake in the target organs (heart, lungs) is substantial and it can be blocked (1,4) and displaced (2) by propranolol. The in vivo binding is stereoselective as the R-isomer does not accumulate in the target organs, which is in contrast to the S-isomer (2,3). Also, S-propranolol is much more potent than R-propranolol in preventing tissue uptake of S-1'-[¹⁸F]fluorocarazolol (5), which indicates that fluorocarazolol and propranolol compete for binding to the same receptor sites.

In vivo competition studies showed that fluorocarazolol binds to the beta-1 and beta-2-subtypes. Beta-1 subtype-selective antagonists (CGP 20712A and ICI 89,406) inhibited ¹⁸F uptake in rat heart (predominantly beta-1-adrenoceptors) more potently than in rat lungs [predominantly beta-2-adrenoceptors (5)]. In contrast, beta-2 subtype-selective drugs (ICI 118,551 and procaterol) were more potent in the lungs than in the heart (2,5). Radioactive metabolites appeared in rat plasma, but bound radioactivity in heart and lung represented mainly parent compound even at 60 min postinjection (6). In vivo saturation studies indicated receptor densities of 6.0 and 21 pmole/g tissue in rat heart and lung, respectively (7).

Thus, animal experiments indicate that S-(fluoro)carazolol may be a useful ligand for PET evaluation of β -adrenoceptors in patients suffering from asthma, chronic obstructive pulmonary disease, cystic fibrosis, hypertension or heart failure, conditions which are associated with altered receptor densities or an altered coupling of the receptors to distal parts of the transduction chain (8–13). Such studies can also be performed with another radioligand, S-[¹¹C]-CGP 12177 (14–20). However, the synthesis of this compound is difficult and in our hands not sufficiently reliable for routine clinical studies. S-1'-[¹⁸F]Fluorocarazolol is more easily prepared and it may prove a useful alternative to S-[¹¹C]-CGP 12177. Here, we report the results of the first PET studies in healthy volunteers using S-1'-[¹⁸F]fluorocarazolol.

MATERIALS AND METHODS

Subjects

Healthy volunteers were recruited according to the following criteria: age 18-40 yr, prebronchodilator forced expiratory volume in 1 sec (FEV₁) >80% of predicted, nonsmoker or ex-smoker (smoking terminated for more than 1 yr). Excluded were people with a positive history for wheezing and tightness of the chest, upper respiratory tract infections in a period shorter than 4 wk before the study, presence of pulmonary diseases including asthma, presence of airway hyperresponsiveness, atopy (at least one positive reaction to known allergens in intracutaneous skin tests), use of beta-mimetics or theophylline, high blood pressure or heart failure, pregnancy or suspected pregnancy.

All volunteers underwent the following screening: anamnesis, physical examination, routine blood biochemistry to assess kidney and liver function, electrocardiogram, skin tests for allergic reactions and pulmonary function tests (spirometry and determination of airway hyper-responsiveness). Airway responsiveness to methacholine was determined using the 2-min tidal breathing method of Cockroft et al. (21,22). The study was approved by the Medical Ethics Committee of the University Hospital. Each subject gave written, informed consent.

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TABLE 1				
Volunteers and	Study Protocol			

Date	Volunteer no.	Age (yr)	Sex	Weight (kg)	Pretreatment	Injected mass (nmole)	Receptor occupancy lung (%)	Receptor occupancy heart (%)
30-11-94	1	23	M	67	None	0.64	0.76	0.43
07-12- 9 4					None	1.00		
08-03-95	2	21	М	77	None	0.36	1.24	0.52
15-03-95					Salbutamol	2.84		
12-04-95	3	24	м	76	None	0.99	1.51	0.77
19-04-95					Salbutamol	2.40		
24-05-95	4	36	М	67	None	0.88	1.12	0.46
31-05-95					Pindolol	1.01		
02-08-95	5	27	F	56	None	1.75	2.94	1.19
20-09-95					Pindolol	0.59		
29-11-95	6	28	М	76	None	1.33	1.88	1.16
Mean ± s.d.		27 ± 5		70 ± 8		1.3 ± 0.8	1.6 ± 0.8	0.8 ± 0.3
(N)		(6)		(6)		(11)	(6)	(6)

Radioligand

S-Desisopropylcarazolol (enantiomeric excess >98%) was prepared as reported previously (6). S-1'-[18 F]fluorocarazolol was synthesized by reacting the precursor with [18 F]fluoroacetone (3,6) and purified by HPLC. The specific activity was 50 \pm 24 TBq/mmole (1400 \pm 680 Ci/mmole) and the radiochemical purity was >99.8%. The ligand was dissolved in 0.5 ml ethanol/propylene glycol/0.9% NaCl (1/2/2 v/v/v). Before injection this solution was filtered (0.22 μ m) and 7.5 ml 0.9% NaCl was added via the filter. The solution was sterile and apyrogenic. S-1'fluorocarazolol. HCl passed the test on "acute toxicity" (European Pharmacopeia; Dutch Pharmacopeia Ed. IX) at a 10,000-fold higher dose than was administered to humans.

Study Protocol

At the beginning of the study, a cannula was placed in a vein of one of the lower forearms. Another cannula was placed in the radial

artery of the contralateral arm, after patency of the ulnar artery had been proven by the Allen-test. The arterial cannula was inserted under local anesthesia with lidocaine. The venous cannula was used for injection and the arterial line for blood sampling.

The volunteer was then placed in the PET camera (FWHM = 6mm). A rectilinear scan was made for proper positioning (heart and lungs in the field of view). Next, a transmission scan was produced, using the internal ⁶⁸Ge/⁶⁸Ga sources, to correct for attenuation. $S-1'-[^{18}F]$ fluorocarazolol (on average 56 MBq = 1.5 mCi) was injected over a period of 1 min, using a Medrad OP-100 remotecontrolled pump. Lines were carefully flushed with saline to ensure complete delivery of the radioligand.

Data acquisition was started at the onset of injection; 8 frames of 15 sec were followed by 4 frames of 30 sec, 4 frames of 1 min, 4 frames of 2 min, 6 frames of 4 min and 2 frames of 10 min. Total duration of the study was 60 min. Arterial blood samples (2 ml)



FIGURE 1. PET images of a human volunteer acquired with S-1'-[18F]fluorocarazolol. Transaxial cross-sections in the time frame 14-60 min postinjection are displayed. The upper and third rows are from the control study, and the second and bottom rows are from the pindolol-blocked study of the same subject. Subsequent planes are shown from the rostral (left) to the caudal (right) side of the thorax.



FIGURE 2. Myocardial and pulmonary time-activity curves observed after injection of S-1'-[¹⁸F]fluorocarazolol (data plotted as mean \pm s.d.).

were drawn at 0.5-min intervals during the initial 5 min and at 10-min intervals from 10 to 60 min postinjection. Radioactivity in plasma and in a cell pellet (5 min 3000 g) was determined in all samples using a gamma counter that was cross-calibrated with the PET camera. Additional samples (3 ml) drawn at 1, 2, 5, 10, 20, 40 and 60 min were used for metabolite analysis (see below). The volunteer left the camera when data acquisition had ended and the cannulas were removed.

After an interval of at least 1 wk, the volunteer returned for the second part of the study in which the influence of a beta-2 agonist or a beta-adrenoceptor antagonist on tissue uptake of S-1'- $[^{18}F]$ fluorocarazolol was assessed. Two different treatments were compared:

- 1. Some volunteers inhaled salbutamol (Ventolin^R, $2 \times 200 \ \mu g$ aerosol) 30-40 min prior to injection of the radioligand.
- Other volunteers took pindolol (Viskeen^R, orally, 5 mg on the evening before the experiment, 5 mg on the morning before the experiment and 5 mg 30-40 min before injection of the radioligand).

Cannulas were placed in a vein of each of the lower forearms. No arterial catheter was used in the second part of the study to keep inconvenience to the volunteer to a minimum. One cannula was used for injection and the other for blood sampling. Tracer injection, data acquisition and sampling were performed as on day one.

Metabolite Analysis

Plasma was analyzed for the presence of S-1'-[¹⁸F]fluorocarazolol and radioactive metabolites by methods published previously (23). Untreated plasma samples were directly applied to a Chromspher Biomatrix (150 × 4.6 mm i.d.) column with M3 guard column. The mobile phase was 10 mM K₂HPO₄:acetonitrile (90:10 v/v, pH 7.5) and the flow rate 1.5 ml.min⁻¹. Twenty-four fractions of the eluate were collected over a period of 12 min. Radioactivity in the fractions was determined using the gamma counter. An independent estimate of the fraction of unmetabolized ligand was obtained by determining protein-bound radioactivity in human plasma by ultrafiltration (MPS-1 reusable micropartition system with YMT-30 membrane, Amicon, Beverly MA). It has been shown previously that about 73% of native S-1'-[¹⁸F]fluorocarazolol is bound to plasma proteins, but its radioactive metabolites have negligible protein binding (23).

Data Evaluation

Regions of interest (ROIs) were drawn on both lungs, using the transmission scan and avoiding hilar structures. A ROI for the left

 TABLE 2

 Parameters of the 'Slow Kinetic Phase' of Washout in Target Tissues

	Heart			Lungs			
Volunteer no.	Y-intercept (ECAT cts × 10⁴)	Rate constant (min ⁻¹)	Y-intercept (body-weight corrected)	Y-intercept (ECAT cts \times 10 ⁴)	Rate constant (min ⁻¹)	Y-intercept (body-weight corrected)	
1	26.6	0.0106	25.5	13.4	0.0071	12.8	
2	24.1	0.0066	26.5	16.5	0.0089	18.1	
3	27.9	0.0081	30.3	15.6	0.0049	16.9	
4	24.8	0.0089	23.7	17.3	0.0081	16.6	
5	34.1	0.0067	27.3	24.0	0.0080	19.2	
6	24.8	0.0080	26.9	11.5	0.0067	12.5	
Control me	an ± s.d.	0.0082 ± 0.0015	26.7 ± 2.2		0.0073 ± 0.0014	16.0 ± 2.8*	
2	23.4	0.0094	25.7	13.0	0.0055	14.3	
3	22.6	0.0075	24.5	15.0	0.0048	16.3	
Salbutamol	mean ± s.d.	0.0084 ± 0.0010	25.1 ± 0.8		0.0052 ± 0.0004	15.3 ± 1.4	
4	16.2	0.0145	15.5	17.4	0.0218	16.7	
5	13.0	0.0150	10.4	18.5	0.0160	14.8	
Pindolol me	ean ± s.d.	0.0148 ± 0.0003 [†]	$13.0 \pm 3.6^{\dagger}$		0.0189 ± 0.0029 [†]	15.8 ± 1.3	

*Significantly different from the corresponding value for the heart.

[†]Significantly different from the control value.



FIGURE 3. Clearance of radioactivity from human plasma after injection of S-1'- (^{18}F) fluorocarazolol (data plotted as mean \pm s.d.).

ventricle of the heart was drawn using the emission scan and the time frame 14-60 min postinjection of the unblocked study. Time-activity curves for the ROIs were calculated using ECAT software (version 6.5D) running on a SUN/SPARC workstation. Since data from different studies had to be compared, values of tissue uptake were normalized to an injected radioactivity of 74 MBq (2 mCi) and a body weight of 70 kg. The time-activity data were exported to an IBM-compatible PC and characterized using a nonlinear regression data analysis program (EnzFitter, Elsevier Biosoft). Differences between groups were tested using one-way analysis of variance and commercial software (Statistix, NH Analytical Software). A two-tailed probability smaller than 0.05 was considered statistically significant.



FIGURE 4. Time-course of unchanged S-1'-[¹⁸F]fluorocarazolol in human plasma. The fraction of total plasma radioactivity representing unmodified radioligand is plotted as mean \pm s.d.

RESULTS

General

Details regarding the participants and the study protocol are presented in Table 1. All volunteers (5 men, 1 woman; age range 21-36 yr; median age 27 yr) had normal spirometric values: average FEV₁ 103% of predicted, range 92%-114%, average vital capacity 105 (92-123) % of predicted. None of the subjects showed bronchial hyperresponsiveness to methacholine: the provocation concentration for a 20% decrease of FEV₁ (PC₂₀) was >20 mg/ml in all cases.

Injection of S-1'-[¹⁸F]fluorocarazolol by the remote-controlled pump did not cause any change in blood pressure, heart rate or electrocardiogram of the volunteers. Transverse sections of the thorax of a single subject before and after ingestion of pindolol are shown in Figure 1. The volunteer is on his back; the direction of observation is from his feet towards his head. In the control study (without pindolol), the heart and lungs were clearly visible; at planes originating from the lower part of the thorax the right liver lobe came into the field of view. After ingestion of pindolol, heart and lungs were no longer visible, but the hepatic uptake of S-1'-[¹⁸F]fluorocarazolol was unaffected. Inhalation of salbutamol before administration of the radioligand did not alter the PET images in any way. PET images acquired after salbutamol treatment are therefore not shown in Figure 1.

Kinetics of S-1'-[Fluorine-18]Fluorocarazolol Uptake in Heart and Lung

After injection of the radioligand, tissue levels of radioactivity rose to a maximum followed by a rapid decline to a relatively stable plateau (Fig. 2). This 'slow kinetic phase' represented a higher level of radioactivity per volume in the heart than in the lungs (Fig. 2). A biexponential function was fitted to the tissue washout curves; parameters calculated for the slow component of this function (i.e., the slow kinetic phase) are shown in Table 2. Salbutamol did not significantly influence the slow kinetic phase in either heart or lungs, but pretreatment of subjects with pindolol induced a more rapid washout of radioactivity from the target organs (Fig. 2; Table 2). Pindolol reduced cardiac and pulmonary radioactivity to 39% and 56% of the control at 60 min postinjection, whereas salbutamol had no significant effect (average reduction <5% from 30–60 min postinjection, Fig. 2).

Clearance of Radioactivity from Plasma

Injected radioactivity was initially rapidly cleared from plasma (to $6.3\% \pm 0.3\%$ of the peak level within 10 min), but the subsequent clearance was slow (to $3.1\% \pm 0.4\%$ after 60 min, see Fig. 3). Inhaled salbutamol did not affect the plasma clearance of the radioligand, but ingested pindolol retarded it. Circulating levels of radioactivity from 10 to 60 min postinjection were significantly (1.7–1.8 fold) higher in pindolol-treated subjects than in control subjects (Fig. 3).

Appearance of Labeled Metabolites in Plasma

Radioactive metabolites appeared rapidly in human plasma after injection of S-1'-[¹⁸F]fluorocarazolol. The fraction of total plasma radioactivity representing unmodified ligand decreased from >99.8% at t = 0 to ca. 20% after 60 min (Fig. 4). Salbutamol did not affect the ratio of [parent]/[metabolites] in plasma, but pindolol increased it (Fig. 4). The fraction of total plasma radioactivity representing parent compound was significantly higher in pindolol-treated subjects than in control subjects from 5 to 60 min postinjection (Fig. 4).



FIGURE 5. Tissue-to-plasma concentration ratios of S-1'-[¹⁸F]fluorocarazolol in human tissues. Tissue radioactivity was assumed to be 100% unmodified radioligand; plasma radioactivity was corrected for the presence of radioactive metabolites. Data are plotted as mean \pm s.d.

Tissue-to-Plasma Concentration Ratios

If tissue radioactivity is considered to represent bound radioligand only (see Discussion) and the contribution of radioactive metabolites is subtracted from total plasma radioactivity, [tissue]/[plasma] concentration ratios of the radioligand can be calculated. In heart and lung, these ratios slowly rise to a plateau value that is reached after 45–50 min (Fig. 5).

DISCUSSION

Advantages and Disadvantages of Using S-1'-[Fluorine-18-] Fluorocarazolol Rather Than S-[Carbon-11]CGP 12177

We could produce S-[¹⁸F]fluorocarazolol with lower maximum yield than S-[¹¹C]-CGP 12177 but with much higher specific activity (Table 3). The yield could still be optimized and it was already sufficient for human studies. The high specific activity of S-[¹⁸F]fluorocarazolol allowed PET scanning to be performed with low receptor occupancy, i.e., lack of pharmacological effects (see below). In contrast, planned studies with S-[¹¹C]CGP 12177 often had to be canceled for medical-ethical reasons as no low-mass injection was possible.

Both CGP-12177 and fluorocarazolol are potent, nonsubtypeselective beta-adrenoceptor antagonists with subnanomolar affinities to beta-adrenoceptors (Table 3). CGP 12177 is hydrophilic whereas fluorocarazolol is a more lipophilic ligand. The lipophilicity of fluorocarazolol as compared to CGP-12177 resulted in a higher nonspecific binding and a more extensive

		TABLE 3	
Properties	of the	Beta-Adrenoceptor	Ligands

	S-[¹¹ C]-CGP-12177	S-1'-[¹⁸ F]-fluorocarazolol
Maximum yield (GBq) (EOS, not corrected for decay)	1.85	0.37
Specific activity	0.37-18.5	50 ± 24
(TBq/mmole)	Often very low	Usually ca. 74
Physical half-life (min)	20.4	109.8
Lipophilicity (log P, octanol/buffer pH 7.4)	-0.5 (29)	+2.2 (3)
Affinity β_1 -subtype (nM)	0.33 (30)	0.41 (7)
Affinity β_2 -subtype (nM)	0.90 (30)	0.10 (7)
Metabolism in humans (during 60 min scan)	Negligible according to (31, 32) Extensive according to (17)	Fairly extensive (see Fig. 4)

first-pass metabolism in humans (Table 3). However, CGP 12177 did not appreciably cross the blood-brain barrier whereas fluorocarazolol could be used for PET studies of beta-adreno-ceptors in the central nervous system (data reported elsewhere).

Receptor Occupancy in Target Tissues

Tissue concentrations of fluorocarazolol were calculated from PET images using the calibration factor of the camera and the specific activity of the injected radioligand. During the slow kinetic phase, maximal concentrations ranged from 0.03 to 0.085 pmole/ml in the heart and from 0.015 to 0.06 pmole/ml in the lungs (raw data; tissue volume including air).

Washout of ¹⁸F from the target tissues after injection of S-1'-[¹⁸F]fluorocarazolol showed the typical biexponential kinetics that have also been described for S-[¹¹C]CGP 12177, an established beta-adrenoceptor ligand (19). The rapid component of this kinetics represented the vascular phase and nonspecific binding, whereas the slow component mainly represented radioligand binding to beta-adrenoceptors (19). Estimations of beta-adrenoceptor density in the heart of healthy volunteers using S-[¹¹C]-CGP 12177 ranged from 7.0 \pm 1.4 [uncorrected] mean \pm s.d. (24)] to 10.0 \pm 1.7 pmole/ml [corrected for spillover from blood and partial volume effect (25)]. Pulmonary beta-adrenoceptor density in healthy subjects, measured by the same technique, were 2.0 \pm 0.2 pmole/ml tissue (including air) (18). Maximal receptor occupancies in the present study, calculated from tissue concentrations of S-1'-[¹⁸F]fluorocarazolol and in vivo data for tissue B_{max} (18,24,25) ranged from 0.4 to 1.2% in the heart and from 0.75% to 3% in the lungs (see Table 1). At the end of the experiment (50-60 min post)injection), values were ca 0.50% lower. It is therefore not surprising that injection of S-1'-[¹⁸F]fluorocarazolol had no measurable effect on heart rate or blood pressure, although carazolol has no intrinsic sympathomimetic activity.

Mechanisms Underlying Tissue Uptake of Radioactivity

Orally administered pindolol had a strong effect on S-1'-[18 F]fluorocarazolol uptake in the heart and lung but not in the liver of healthy volunteers (Figs. 1, 2, 5; Table 2). The pindolol effect suggests that myocardial and pulmonary radioactivity represents radioligand binding to beta-adrenoceptors, whereas liver uptake is largely determined by other mechanisms (e.g., membrane transport and metabolism). Inhalation of salbutamol had no measurable effect on pulmonary radioactivity at intervals >20 min although salbutamol seemed to induce a more rapid washout of nonspecifically bound radioligand probably due to an increase of pulmonary blood flow (Fig. 2; Table 2). The administered amount of salbutamol (2 × 200 μ g = 1670 nmole) was sufficient to occupy a substantial fraction of receptors, even if only 10% of the dose would enter the lungs [total amount of pulmonary beta-2-adrenoceptors in humans ca. 10 nmole (18); affinity of salbutamol to these receptors 10⁻⁸ M (26)]. Inhaled salbutamol occupies beta-adrenoceptors on smooth muscle of the airways but the β_2 -agonist does not reach the level of the alveoli. The fact that salbutamol did not affect specific binding of S-1'-[¹⁸F]fluorocarazolol in the lung therefore suggests that the radioligand was mainly bound to alveolar beta-adrenoceptors, which constitute >90% of the total betaadrenoceptor pool in human lung (27).

Effect of Pindolol on Radioligand Clearance

In the presence of pindolol, levels of radioactivity in plasma from 10 to 60 min after injection of S-1'-[18 F]fluorocarazolol were almost twice as high as in the control situation (Fig. 3). Moreover, the ratio of the concentrations of parent/metabolites was significantly increased (Fig. 4). The effect of pindolol may be caused by two different (but not mutually exclusive) mechanisms:

- 1. Pindolol blocked beta-adrenoceptors and it caused a more rapid washout of the radioligand from the target organs, resulting in higher concentrations in plasma (see Fig. 2).
- 2. Pindolol competed with S-1'-[¹⁸F]fluorocarazolol for the same metabolic pathway in the liver and thus caused less rapid degradation and excretion of the radioligand.

The former mechanism seemed more likely, as the clearance of a slowly metabolized radiopharmaceutical, $S-1'-[^{11}C]CGP$ 12177, was similarly affected by prior administration of propranolol (16).

CONCLUSION

S-1'-[¹⁸F]fluorocarazolol seemed a useful radiopharmaceutical for PET studies of beta-adrenoceptors in human heart and lungs. The myocardium and the peripheral lung were clearly visualized; uptake in these tissues was strongly inhibited after ingestion of pindolol. Tissue-to-plasma concentration ratios of the radioligand increased to a plateau value which was reached at 45–50 min postinjection. Kinetics of ¹⁸F uptake and release in the target organs were compatible with determination of receptor densities with compartment models. After submission of this article, another article was published in which it was shown that myocardial beta-adrenoceptor density in experimental animals can be accurately determined by a dual-injection protocol (28).

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REFERENCES

- Berridge MS, Cassidy EH, Terris AH, Vesselle JM. Preparation and in vivo binding of [¹¹C]carazolol, a radiotracer for the beta-adrenergic receptor. *Nucl Med Biol* 1992;19: 563-569.
- Berridge MS, Nelson AD, Zheng LB, Leisure GP, Miraldi F. Specific beta-adrenergic receptor binding of carazolol measured with PET. J Nucl Med 1994;35:1665-1676.
- Zheng L, Berridge MS, Ernsberger P. Synthesis, binding properties, and ¹⁸F labeling of fluorocarazolol, a high-affinity beta-adrenergic receptor antagonist. J Med Chem 1994;37:3219-3230.

- Francis BE, Eckelman WC, Grissom MP, Gibson RE, Reba RC. The use of tritium-labeled compounds to develop gamma-emitting receptor-binding radiotracers. *Nucl Med Biol* 1982;9:173-179.
- Van Waarde A, Elsinga PH, Brodde OE, Visser GM, Vaalburg W. Myocardial and pulmonary uptake of S-1'-[¹⁸F]fluorocarazolol in intact rats reflects radioligand binding to β-adrenoceptors. Eur J Pharmacol 1995;272:159-168.
- Elsinga PH, Vos MG, Van Waarde A, et al. (S,S)- and (S,R)-1'-[¹⁸F]fluorocarazolol, ligands for the visualization of pulmonary β-adrenergic receptors with PET. Nucl Med Biol 1996;23:159-167.
- Van Waarde A, Elsinga PH, Visser TJ, et al. In vivo quantification of β-adrenoceptors using S-1'-[¹⁸F]fluorocarazolol. Nucl Med Biol 1996:in press.
- Bai TR, Mak JC, Barnes PJ. A comparison of beta-adrenergic receptors and in vitro relaxant responses to isoproterenol in asthmatic airway smooth muscle. *Am J Respir Cell Mol Biol* 1992;6:647-651.
- Sharma RK, Jeffery PK. Airway beta-adrenoceptor number in cystic fibrosis and asthma. Clin Sci 1990;78:409-417.
- Raaijmakers JA, Beneker C, Dol R, de Ruiter Bootsma AL. Decreased bronchiolar epithelial beta-adrenoceptor densities in patients with chronic obstructive lung disease. Cell Mol Biol 1987;33:515-518.
- Bristow MR, Ginsburg R, Umans V, et al. Beta-1- and beta-2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta-1-receptor downregulation in heart failure. *Circ Res* 1986;59:297-309.
- Brodde OE. Beta1- and beta2-adrenoceptors in the human heart: properties, function and alterations in chronic heart failure. *Pharmacol Rev* 1991;43:203-242.
- Lopes AA, Liberato MH, Brentani MM, Aiello VD, Riso AA, Ebaid M. Lung beta-adrenoceptors in pulmonary hypertension. A study of biopsy specimens in children with congenital heart disease. *Chest* 1991;99:637-641.
- Van Waarde A, Meeder JG, Blanksma PK, et al. Suitability of CGP12177 and CGP26505 for quantitative imaging of beta-adrenoceptors. *Nucl Med Biol* 1992;19: 711-718.
- 15. Van Waarde A, Meeder JG, Blanksma PK, et al. Uptake of radioligands by rat heart and lung in vivo: CGP12177 does and CGP26505 does not reflect binding to β-adrenoceptors. Eur J Pharmacol 1992;222:107-112.
- Law MP. Demonstration of the suitability of CGP 12177 for in vivo studies of β-adrenoceptors. Br J Pharmacol 1993;109:1101-1109.
- Merlet P, Delforge J, Syrota A, et al. Positron emission tomography with ¹¹C CGP-12177 to assess beta-adrenergic receptor concentration in idiopathic dilated cardiomyopathy. *Circulation* 1993;87:1169-1178.
- Ueki J, Rhodes CG, Hughes JMB, et al. In vivo quantification of pulmonary β-adrenoceptor density in humans with (S)-[¹¹C]CGP-12177 and PET. J Appl Physiol 1993;75:559-565.
- Delforge J, Syrota A, Lançon JP, et al. Cardiac beta-adrenergic receptor density measured in vivo using PET, CGP 12177 and a new graphical method. J Nucl Med 1991;32:739-748.
- Valette H, Deleuze P, Syrota A, Delforge J, Crouzel C, Fuseau C, Loisance D. Canine myocardial beta-adrenergic, muscarinic receptor densities after denervation: a PET study. J Nucl Med 1995;36:140-146.
- Cockroft DW, Killian DN, Mellon JJA, Hargreaves FE. Bronchial reactivity to inhaled histamine: a method and clinical survey. *Clin Allergy* 1977;7:235-243.
- Sterk PJ, Fabbri LM, Quanjer PH, et al. Airway responsiveness: Standardized challenge testing with pharmacological, physical and sensitizing stimuli in adults. *Eur Respir J* 1993;6(suppl.16):53-83.
- Van Waarde A, Visser TJ, Posthumus H, et al. Quantification of the β-adrenoceptor ligand, S-1'-[¹⁸F]-fluorocarazolol, in plasma of humans, rats and sheep. J Chromatogr 1996;678:253-260.
- Merlet P, Delforge J, Syrota A, et al. Quantification of beta-adrenergic receptor concentration in human with positron emission tomography [Abstract]. J Nucl Med 1991;32:989.
- Lefroy DC, De Silva R, Choudhury L, et al. Myocardial beta-adrenoceptor density is reduced in hypertrophic cardiomyopathy [Abstract]. Circulation 1992;86:1-246.
- van Koppen CJ, Hermanussen MW, Verrijp KN, et al. Beta-adrenoceptors in human tracheal smooth muscle: characteristics of binding and relaxation. *Life Sci* 1987;40: 2561-2570.
- Carstairs JR, Nimmo AJ, Barnes PJ. Autoradiographic visualization of beta-adrenoceptor subtypes in human lung. Am Rev Respir Dis 1985;132:541-547.
- Muzic RF, Nelson AD, Saidel GM, Miraldi F. Optimal experiment design for PET quantification of receptor concentration. *IEEE Trans Med Imag* 1996;15:2-12.
- Staehelin M, Simons P, Jaeggi K, Wigger N. CGP-12177. A hydrophilic betaadrenergic receptor radioligand reveals high affinity binding of agonists to intact cells. J Biol Chem 1983;258:3496-3502.
- Nanoff C, Freissmuth M, Schutz W. The role of a low beta 1-adrenoceptor selectivity of [³H]CGP-12177 for resolving subtype-selectivity of competitive ligands. *Naunyn* Schmiedebergs Arch Pharmacol 1987;336:519-525.
- Van Waarde A, Anthonio RL, Visser TJ, et al. Quantification of a carbon-11-labeled β-adrenoceptor ligand, S-(-)-CGP 12177, in plasma of humans and rats. J Chromatogr 1995;B 663:361-369.
- Luthra SK, Osman S, Steel CJ, et al. Comparison of S-[¹¹C]-CGP12177 metabolism in rat, dog and man using solid phase extraction and HPLC. J Lab Compd Radiopharm 1992;31:518-520.



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