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Foraging by the stoplight parrotfish *Sparisoma viride*. II. Intake and assimilation of food, protein and energy

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ABSTRACT: Daily food intake by the herbivorous parrotfish *Sparisoma viride*, as well as assimilation efficiencies of algal food, protein and energy, were quantified through a combination of laboratory feeding trials and field observations. The intake of algal ash-free dry wt (AFDW) per bite increases linearly with fish wet wt (FWW, g) and algal biomass (mg AFDW cm⁻²), and is further determined by the skeletal density of the limestone substrate. Low-density substrates yield higher amounts of AFDW per bite than do high-density substrates. The percentage of the total food intake that is derived from endolithic and crustose coralline algae increases with the size of the fish, and can be >70% depending on the biomass of epilithic algae. The daily pattern of foraging activity is positively correlated with diurnal changes in food quality, while seasonal daylength variations result in 13% variation in total daily bites taken. Daily number of bites of *S. viride* in the field decreases with fish size, and is further dependent on life phase and foraging depth. In experiments, fish attained an assimilation efficiency of ca 20% from a natural diet of low algal biomass and high-density dead coral substrates that predominates in the shallow reef. Assimilation efficiency was ca 70% from a diet of high algal biomass and low-density substrates that predominates on the deeper reef parts. In spite of lower daily foraging effort, territorial fish, living in deeper parts of the reef, ingest and assimilate higher amounts of AFDW, protein and energy per day than non-territorial fish foraging on the shallow reef. The difference is caused by increased availability of high-yield food and substrate types inside territories compared to the situation on the shallow reef. Daily assimilated energy (kJ d⁻¹) is $0.85 \times \text{FWW}^{0.773}$ for fish foraging in the shallow reef zone, and $1.22 \times \text{FWW}^{0.854}$ for *S. viride* foraging inside territories on the deeper reef.

KEY WORDS: Herbivorous fish · Scaridae · Daily food intake · Assimilation efficiency · Protein · Energy · Social organization

INTRODUCTION

Herbivory on some coral reefs is probably more intense than in any other habitat, either terrestrial or marine (Hatcher & Larkum 1983, Carpenter 1986, Lewis 1986, Choat 1991). On reefs subject to minimal human disturbance, fish, as the main consumers of the primary production, are often estimated to consume between 50 and 100% of total algal production (Carpenter 1986). Parrotfish (family Scaridae) have been widely recognized as a major component of the herbivorous fish community (Williams 1982, Russ 1984a, b, Choat & Bellwood 1985), and can be expected to play an important role in the transfer of food materials and energy from primary producers to the remaining members of the food chain.

Being large and mobile, parrotfish are intense feeders which can profoundly affect the diversity and biomass of reef-inhabiting algae (Brock 1979). In terms of feeding biology, and in analysis of the fish community structure, it has been convenient to view scarids as a relatively uniform group of grazing herbivores that scrape filamentous algae from the reef matrix (Choat 1991). Bellwood & Choat (1990), however, have recently questioned this assumption, and concluded that parrotfish are a diverse group with marked ecological and behavioural differences within the family, occupying a wide range of habitats with different substrata. As a consequence, the impact of these herbivores on the algal communities will vary with species composition and reef habitat.

Scarids are notable in their ability to sustain large population sizes and high growth rates (Russ 1984a, b,

Russ & St. John 1988, J. van Rooij, J. H. Bruggemann & J. J. Videler unpubl.) on food sources which have small standing crop (Hatcher 1983, Russ 1987, Steneck 1988) and which are low in protein (Bowen 1987). However, little is known quantitatively about food intake and assimilation efficiency in scarids (Horn 1989, Choat 1991). The few experimental studies of foraging in herbivorous reef fish tended to concentrate on small territorial species, such as members of the family Pomacentridae (Polunin 1988, Klumpp & Polunin 1989). The paucity of quantitative studies of food intake in larger herbivorous reef fish is probably associated with their mobility and with the difficulty of making direct observations in the field. Moreover, due to their large size and the nature of their food (algae associated with the reef matrix), manipulative studies in the laboratory are difficult and laborious.

The daily intake, assimilation and allocation of energy by a common scarid were investigated in a larger project, aiming to evaluate the role of parrotfish in the trophodynamics of coral reefs. The stoplight parrotfish *Sparisoma viride* (Bonnaterre) was chosen as study animal. This species is a large generalist herbivore that is common on Caribbean reefs. Food selection by *S. viride* has been described by Bruggemann et al. (1994, this issue), while a description of fish growth is provided by van Rooij et al. (unpubl.). Here, daily intake and assimilation of food, protein and energy by *S. viride* are described. More specifically the objectives of this study were to: (1) quantify the daily intake and assimilation of food and nutrients by different life phases and size classes of *S. viride*; and (2) quantify the effect of foraging depth on the acquisition and assimilation of algal food.

This study represents the first published quantitative description of food intake by a larger herbivorous reef fish.

MATERIAL AND METHODS

Study site and study animals. Experimental work was carried out at Karpata Ecological Centre, situated on the leeward coast of Bonaire, Netherlands Antilles, while field observations were made on the fringing reef in front of the field station. For a detailed description of the reef at the study site see Bruggemann et al. (1994). The parrotfish *Sparisoma viride* is an important herbivore on Caribbean reefs, both in terms of population density and algal biomass consumed. It forages only by day, grazing on epilithic, crustose and endolithic algae growing on and in dead coral substrates. *S. viride* employs an excavating grazing mode that leaves deep scars in the limestone (Gygi 1975).

The species is a protogynous hermaphrodite and 3 life phases are distinguished: juveniles (JU: < 15 cm fork length, FL, measured from the tip of the upper jaw to the end of the middle caudal rays); initial phase (IP: 15 to 35 cm FL), mostly female; and terminal phase (TP: 18 to 43 cm FL). Its social organization confines individuals to limited parts of the reef. On the shallow reef (0 to 3.5 m depth) loosely associated groups of IP and TP fish share a common home range. Territories, shared by 3 to 7 harem females, are maintained and fiercely defended against conspecifics by single TP males on the deeper parts of the reef (4 to 25 m depth). JU fish are found in highest densities on the drop-off reef zone (6 to 12 m depth).

Fish used in the laboratory for feeding trials were captured near the study site either at night using hand nets and an anaesthetic (quinaldine, 'Aquavet Seaquin'), or by day using a seine net (84 m long × 10 m deep, 5 cm stretched mesh). Fish were transferred to the laboratory where they were kept in tanks with a continuous supply of seawater that was pumped from the reef. Tanks with a capacity of 115 l were used for juveniles, while 1000 l or 3500 l tanks were used for adults. Shade nets were used to protect the tanks from direct incident light, minimizing temperature increase in the daytime. Fish wet weight (FWW) and fork length were determined for fish with empty guts, anaesthetized in a 10 ppm solution of quinaldine to facilitate handling. FWW was determined to the nearest 0.1 g on an electronic scale (Sartorius PT1200), or to the nearest g using a triple-beam balance (Ohaus), and fork length was determined to the nearest mm with the aid of a measuring board. Between experiments, fish were fed ad libitum with algae growing on coral rubble that was collected from the nearby reef. Before the onset of each experiment, all coral rubble was removed, and all debris was siphoned out of the tanks. JU and IP fish were kept in pairs of approximately equal weight, while TP males were held individually.

Food intake experiments. The amount of algal food ingested per bite was calculated from the decrease in algal biomass after a known number of bites on blocks of dead coral substrate covered with a natural vegetation. Parts of dead coral bearing a homogeneous algal vegetation, representing characteristic food and substrate types, were collected from the main feeding areas of *Sparisoma viride* (see Bruggemann et al. 1994). Substrates originating from *Acropora cervicornis* and *A. palmata* colonies were collected from the shallow reef zone (< 3 m depth) where non-territorial fish feed. *Montastrea annularis* substrates were collected in the deeper reef (> 4 m depth) from the feeding areas of territorial fish. The substrate blocks were trimmed, and subsequently cut into halves using a diamond gem-saw (Raytech Industries) To

verify uniformity of algal growth on either half, the mean percentage cover of filamentous, crustose and endolithic algae was determined from 20 randomly chosen quadrats of 6.25 mm², using a dissecting microscope at 60× magnification. The 2 halves were considered to have a uniform algal cover if the means of either half did not differ by more than 10%. A random sample of 20 quadrats was sufficient to obtain a reproducible estimate of the mean cover by different vegetation components. Designation of experimental and control halves (blocks) was decided by the flip of a coin. To allow later calculation of the no. of bites cm⁻², the total surface area of experimental blocks was determined by modelling heavy-duty aluminum foil of known weight tightly over the algae-covered surface of the block, and then weighing the foil.

The experimental blocks were mounted on heavy concrete bricks with iron wire and placed in the tanks with fish, and the number of bites taken on each block was counted. A block was removed when a decrease in algal cover could be detected with the naked eye. During the experiment, the control blocks were kept under identical conditions as experimental blocks, but without fish being present. Experimental and control blocks were cut in 3 subsamples. From each subsample, the surface area was determined as above, and algal biomass (mg AFDW cm⁻²) was determined using the method described by Bruggemann et al. (1994). Food intake per bite (mg algal AFDW) is calculated as:

$$\frac{\text{mean biomass}_{\text{control blocks}} - \text{mean biomass}_{\text{experimental blocks}}}{\text{no. of bites cm}^{-2}} \quad (1)$$

Each experiment consisted of a series of experimental blocks offered to a fish (or pair of fish) of known weight. The coral origin of the substrate, as well as algal biomass, height and cover of filamentous algal fronds, cover of crustose corallines, and thickness of the endolithic layer on experimental and control blocks were noted. We tested whether the relation between fish weight and food intake per bite differed between substrate types, regardless of differences in algal vegetation on these substrates, using ANCOVA (Norusis 1990) in which fish weight was taken as covariate. If not different, substrate types were pooled, and the effect of algal vegetation parameters on food intake per bite was analyzed for each substrate type/group (ANCOVA and linear regression). In some experiments, differences in the ratio of surface area to perimeter and/or the no. of bites cm⁻² between experimental blocks significantly affected the food intake per bite. This experimental bias was analyzed with multiple regression techniques (Norusis 1990), and corrected for using the corresponding regression equations.

Grazing fish ingest varying amounts of the endolithic algal layer, depending on their size, but they do not excavate this layer completely, as witnessed by the blue-green colour of grazing scars. Epi- and endolithic algal fractions differ in chemical composition (Bruggemann et al. 1994). In order to enable estimation of protein and energy intake per bite, the relative proportion of epilithic and 'substrate-bound' algae (endolithic algae and crustose corallines) was assessed. The percentage of the total food intake per bite that was derived from substrate-bound fractions was calculated from the total food intake per bite minus the intake derived from epilithic algae. Ingestion of epilithic algae was calculated from the surface area of grazing scars (see Bruggemann et al. 1994) and the epilithic biomass. Biomass of epilithic algae on *Acropora* spp. substrates was determined by Bruggemann et al. (1994: Table 5, sparse turfs on endolithic algae = 1.3 ± 0.1 mg AFDW cm⁻²). Experimental *Montastrea annularis* substrates supported higher biomass of epilithic algae, as indicated by increased canopy height and higher percentage cover of epilithic algae (see Table 1). For these substrates, epilithic biomass was estimated by subtracting biomass of endolithic algae as determined by Bruggemann et al. (1994: Table 5, endolithic algae = 17.7 ± 6.0 mg AFDW cm⁻²) from the total algal biomass determined for the experimental substrates. Bite scars left by experimental fish were smaller than those left by free-living fish on the reef (see below), and the relationship between fish fork length and surface area removed per bite was corrected for accordingly.

Food assimilation experiments. Assimilation efficiencies of algal food were determined by direct methods, which involved quantitative measurement of food consumed and faeces produced, and indirect methods, in which the nutrient content of the food was compared with that of the faeces using ash as an assumed non-absorbed marker. The latter method was applicable only to juvenile fish feeding exclusively on epilithic algae that have a relatively constant ash fraction. Larger fish scrape off and ingest large but unknown quantities of carbonates, varying with fish size, food type and skeletal density of substrates (Bruggemann et al. 1994), thus preventing estimates of ash fractions in the food.

Fish were kept over a mesh-wire grate, and fed ad libitum on a diet consisting of coral rubble with associated algae, which was collected from either of the 2 main feeding areas for non-territorial and territorial *Sparisoma viride* respectively (see above). In the assimilation trials we used coral rubble from the same locality, originating from the same coral species and supporting similar algal biomass, as was used in the food intake experiments. Before the onset of each experiment the fish were allowed to get accustomed to

their diet for at least 1 d. Feeding rates (FR, no. of bites h^{-1}) were determined 8 times d^{-1} at regular intervals from 07:30 to 18:30 h. The daily number of bites taken by experimental fish was estimated from extrapolation of the feeding rates to the total daily feeding period from dawn to dusk (see 'Foraging effort'). Daily intake of algal food (AFDW_{in} , mg) was calculated as the product of the daily total number of bites and the experimentally determined food intake per bite. In juveniles that feed only on the epilithic algal fronds, the food intake per bite is not dependent on substrate type. Their food intake per bite was calculated using the regression coefficient of food intake per bite from *Montastrea annularis*, as this substrate type supported highest percentage cover and biomass of epilithic algae (Table 1). At the end of the day all faeces were siphoned from under the grate, collected in buckets and left overnight to allow particulate matter to settle on the bottom. The supernatant was decanted, and the faeces filtered over 5 μm filters (Schleicher & Schuell, no. AE98) under vacuum. The faeces was dried to constant weight at 60 °C, after which the organic fraction was determined (see below). Biochemical composition of faeces was corrected for the loss of organic matter and nutrients between egestion and analysis of faeces. Fresh faeces was collected by massaging the faeces out of the caecum of the fish, and homogenized by stirring. Part of this faeces was dried immediately and served as control, while the rest was divided over a number of Petri dishes, and placed in the tanks under conditions identical to those during experiments. After variable test periods from 3 to 24 h, duplicate faecal samples were siphoned from randomly selected Petri dishes, and the organic fraction and protein contents of control and treated samples were determined following the procedures described below. Faecal nutrient loss was independent of time (least-squares regression of % nutrient loss to time; organic matter: $r = -0.302$, $p = 0.510$, $n = 7$; protein: $r = +0.378$, $p = 0.403$, $n = 7$), indicating that loss of nutrients is caused by collecting and filtering of faeces prior to analysis rather than slow dissolution in the tanks. Loss of organic matter averaged $14 \pm 2\%$ ($n = 7$), while on average $16 \pm 4\%$ ($n = 7$) of faecal protein disappeared. It was assumed that the energy contents decreased to the same extent as that of protein (16% correction). The biochemical composition of daytime faeces was corrected for accordingly. The first defecation of the day, which could easily be distinguished by the mucus film wrapped around it, consists of undigested food remains from the previous day. This 'morning turd' could in most cases immediately be removed whole, and no correction was applied to this portion of the daily defecations. Daily excretion of organic matter (AFDW_{ex} , mg) was calculated as the product of faeces dry wt and the corrected

organic fraction. The proportion of the total food that was absorbed is the 'total assimilation efficiency' (%), which was calculated as:

$$[(\text{AFDW}_{\text{in}} - \text{AFDW}_{\text{ex}}) / \text{AFDW}_{\text{in}}] \times 100, \quad (2)$$

and the proportion of a given nutrient assimilated is the 'nutrient assimilation efficiency' (%), which was calculated as:

$$\frac{\% \text{ nutrient} \times \text{AFDW}_{\text{in}} - \% \text{ nutrient} \times \text{AFDW}_{\text{ex}}}{\% \text{ nutrient} \times \text{AFDW}_{\text{in}}} \times 100. \quad (3)$$

The (indirect) ash-marker method was used to determine assimilation efficiencies in small JU fish (≤ 10 cm FL). They were offered a diet consisting of large algal turfs. To determine the ash fraction of the food, filamentous algae were carefully collected to avoid contamination with carbonate particles or small invertebrates, then dried and ashed (see below). Absorption efficiencies of total algal AFDW and of nutrients were calculated employing the equations of Montgomery & Gerking (1980, p. 146).

Biochemical analysis. The organic fraction (% AFDW) of samples, dried to constant weight at 60 °C, was determined by weighing and reweighing duplicate subsamples after ashing at 500 °C for at least 6 h. Algal biomass (mg algal AFDW cm^{-2}), as well as protein (mg g^{-1} AFDW) and energy (kJ g^{-1} AFDW) contents of food and faeces, were determined following the procedures described by Bruggemann et al. (1994).

To investigate the variation in nutritional quality of algal food over the day, 3 replicate samples of sparse algal turfs were collected at 06:00, 12:00, 18:00 and 00:00 h on 15 January 1991. Since the biomass of sparse algal turfs is very low, each sample was composed of 4 separately collected subsamples. Ash content was determined from replicate subsamples ($n = 3$) as described above. Protein and energy contents were determined from C and N weight fractions using the stoichiometric method (Gnaiger & Bitterlich 1984), following procedures described by Bruggemann et al. (1994). Soluble carbohydrates were extracted by treating replicate ($n = 2$) subsamples of 200 mg with hot (100 °C) 2 M HCl for 2.5 h (Hashimoto et al. 1987). Total soluble carbohydrates were determined by the Anthron method (Morse 1949) using glucose as a standard.

Comparison of bites taken by captive and free-living fish. To verify the validity of the experimentally determined food intake per bite of *Sparisoma viride*, the sizes of grazing scars from captive fish were compared to those from fish foraging on the reef. The bite size, i.e. the surface area removed and the volume of substrate excavated, was determined from the scars left by grazing fish as described by Bruggemann et al. (1994). The mean size of bites recorded from fish kept

in captivity (CB), and that from free-living fish on the reef (FB), were compared pairwise, controlling for fish fork length, substrate type and observer. Differences were tested with the Student's *t*-test (Norusis 1990). Correction factors for surface areas and volumes were calculated as FB/CB for all pairs. The effects of fish fork length and substrate type on the correction factors were tested separately for substrate area and volume of bites with ANOVA (Norusis 1990).

Foraging effort of *Sparisoma viride* in the field.

Field observations of foraging fish were performed using SCUBA, with notes recorded on PVC sheets. Before recording started, the fish were allowed to become accustomed to the presence of the observer for approximately 5 min, during which time the fork length was estimated to the nearest cm. The total number of bites taken during 30 min periods was recorded at regular intervals from dawn to dusk, and the time of day, foraging depth and life phase were noted. Temperature was recorded at 6 m depth at the end of each observation period.

To determine the total daily foraging period, the feeding activity of *Sparisoma viride* was recorded at dawn and dusk relative to the times of sunrise and sunset. Light intensity (photosynthetically active radiation, PAR) was measured simultaneously using a LiCor 192SA quantum sensor which was positioned horizontally on the seabed at 5 m depth, and a LiCor 1000 datalogger. Times of sunrise and sunset were defined as the times of solar elevation of -0.833° (when the upper rim of the sun is level with the horizon), and calculated for the latitude of the study area using the equations described by Dring (1984). Length of the daylight period (DLP) was defined as the time (in h) from sunrise to sunset.

The effects of time of day, fork length, life phase, foraging depth, temperature and season on FR were investigated in 2 series of observations. The first was performed in February, May and August 1989, 3 distinct periods of the year, aiming to encompass a wide range in seasonal temperature and daylength fluctuations. Fish belonging to 4 size classes (i.e. JU fish 10 ± 1 cm FL, IP fish of 20 ± 1 and of 30 ± 1 cm FL and

TP males) were selected in the shallow reef zone and on the deeper reef, and their FRs were recorded (119 30-min protocols). In the second observation series, from May to September 1991, the daily foraging effort of 15 individually recognizable (tagged) TP males of known weight was recorded twice with time intervals ranging from 1 to 4 mo (216 30-min protocols). Feeding activity varies over the day, and the variable 'time of day' can be a confounding factor in the evaluation of the effect of the other variables on FR. When the dataset was restricted to observations made between 09:00 and 17:00 h, no interaction between time of day and the other variables was detected. Therefore, observations made between 09:00 and 17:00 h were used to investigate factors affecting FR (ANCOVA; Norusis 1990).

The total daily number of bites (TDB) was determined from observation series of individual fish, or of well-defined size classes, of which the feeding activity throughout the day was monitored ('daycover'). Complete daycovers were made 28 times at various times of the year for 15 different TP males (territorial and non-territorial individuals), and for fish of 10 ± 1 cm FL (JU), 30 ± 1 cm FL (IP), and 40 ± 2 cm FL (TP) foraging on the deeper reef. Using these daycover observations, the relationship between DLP, TDB and FR between 09:00 and 17:00 h was determined empirically.

RESULTS

Food intake per bite

In Table 1 the characteristics of the experimental substrate types and their associated algal vegetation are summarized. All experimental blocks contained either large or sparse algal turfs; the undergrowth consisted of endolithic algae with a varying proportion of the surface covered with crustose corallines. Substrates bearing endolithic algae and/or crustose corallines, but devoid of epilithic algae, were not used during the experiments.

Table 1. Dead coral substrate types and characteristics of associated algal vegetation used in food intake experiments. Mean \pm SE for each substrate group is shown

Substrate type	Substrate density ^a (g cm ⁻³)	Total algal biomass (mg AFDW cm ⁻²)	Epilithic algal turfs		Crustose corallines (% cover)	Endolithic algal layer (mm)	No. of experimental blocks
			(mm height)	(% cover)			
<i>Acropora cervicornis</i>	2.1 \pm 0.3	14.3 \pm 2.8	2.7 \pm 1.4	49.0 \pm 16.4	35.6 \pm 29.1	1.2 \pm 0.5	17
<i>A. palmata</i>	1.8 \pm 0.2	19.4 \pm 4.5	3.3 \pm 1.2	51.6 \pm 16.5	13.4 \pm 22.1	1.1 \pm 0.4	61
<i>Montastrea annularis</i>	1.3 \pm 0.2	21.5 \pm 5.5	4.5 \pm 1.2	65.4 \pm 18.7	9.1 \pm 18.7	1.4 \pm 0.9	68

^a Values for specific substrate densities from Table 4 in Bruggemann et al. (1994)

Algal biomass differed significantly between substrate types (ANOVA, $F_{(2,144)} = 16.038$, $p < 0.001$). Subsequent comparison of means (Scheffé; Norusis 1990) showed that *Acropora cervicornis* had lowest biomass, while algal biomass on *Montastrea annularis* substrate was highest. Algal biomass on *Acropora* spp. substrates combined was lower than on *M. annularis* substrate (Scheffé, $F_{(1,145)} = 15.810$, $p < 0.001$). Separately and combined, both *Acropora* spp. substrate types had significantly lower turf height (Scheffé, $F_{(1,140)} = 24.467$, $p < 0.001$), lower cover of epilithic algal turfs (Scheffé, $F_{(1,145)} = 25.186$, $p < 0.001$), and higher cover of crustose corallines (Scheffé, $F_{(1,145)} = 6.341$, $p = 0.013$) than *M. annularis* substrates. The thickness of the endolithic algal layer was the only vegetation parameter that did not differ significantly among substrate types (Scheffé, $F_{(1,134)} = 0.931$, $p = 0.336$).

One-way ANCOVA, in which fish weight was taken as covariate, showed no significant difference in food intake per bite between the substrate types *Acropora cervicornis* and *A. palmata* (ANCOVA, $F_{(1,77)} = 0.181$, $p = 0.672$). Food intake per bite differed significantly between *A. cervicornis* and *Montastrea annularis* substrates (ANCOVA, $F_{(1,85)} = 5.648$, $p = 0.020$), and between *A. palmata* and *M. annularis* substrates (ANCOVA, $F_{(1,128)} = 29.76$, $p < 0.001$). In the subsequent analysis the results of experiments with the high-density substrates from the shallow reef (*A. cervicornis* and *A. palmata*) were pooled and treated separately from those obtained with the low-density substrate from the deeper reef areas (*M. annularis*).

Food intake per bite increases linearly with fish weight on all substrate types. In Fig. 1 the least-squares regression of the food intake per bite against fish weight is given for each substrate group. When grazing on dead *Montastrea annularis* substrates, the fish attain a higher food intake per bite than when grazing on dead *Acropora* spp. substrates. For both substrate groups, the deviation from the mean food intake per bite within experiments can be explained by differences in algal biomass on the experimental blocks (ANOVA, $F_{(6,136)} = 4.225$, $p = 0.001$) (Fig. 2). The other vegetation parameters, i.e. percentage cover of epilithic algae and of crustose corallines, and the thickness of the endolithic algal layer did not significantly affect the food intake per bite (ANOVA, epilithic turfs: $F_{(5,141)} = 0.783$, $p = 0.564$; crustose corallines: $F_{(5,141)} = 1.559$, $p = 0.176$; endolithic algae: $F_{(6,129)} = 0.582$, $p = 0.744$). A 1-way ANCOVA, in which algal biomass was taken as a covariate, showed that the effect of algal biomass on food intake per bite was not significantly different for the 2 substrate groups (ANCOVA, $F_{(1,145)} = 1.526$, $p = 0.219$). In Fig. 2 the least-squares regression describing the deviation from the mean food intake per bite (within experiments) as a function of algal biomass is given.

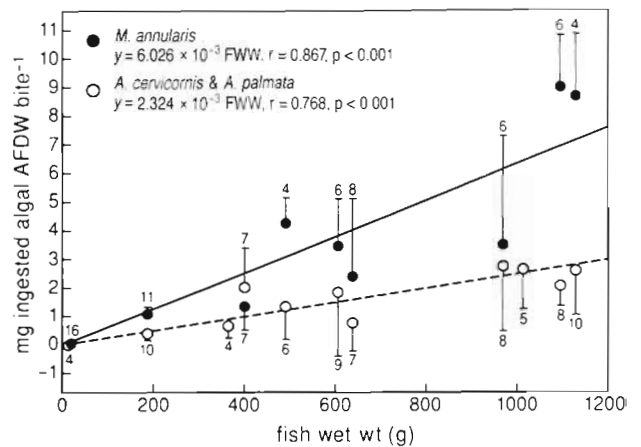


Fig. 1. *Sparisoma viride*. Food intake per bite by stoplight parrotfish as a function of fish wet wt (FWW) and substrate type. Experimental substrates with associated algae were collected from the main feeding areas of the fish: the shallow reef zone (*Acropora cervicornis* and *A. palmata* substrates) and the deeper reef areas (*Montastrea annularis* substrates). The least-squares regression lines are forced through the origin (intercept was not significantly different from zero, $p > 0.05$). Error bar indicates SE of mean food intake per bite for each fish (pair) and substrate type. Only upper or lower error bar is shown for figure clarity. Numbers indicate no. of experimental blocks

The percentage of the total food intake per bite that is derived from crustose and endolithic algae increases with the size of the fish (Fig. 3). Juvenile *Sparisoma viride*, up to 10 cm FL, eat only epilithic algal turfs. The proportion of substrate-bound algal fractions in the diet increases rapidly between 10 and 20 cm FL, and

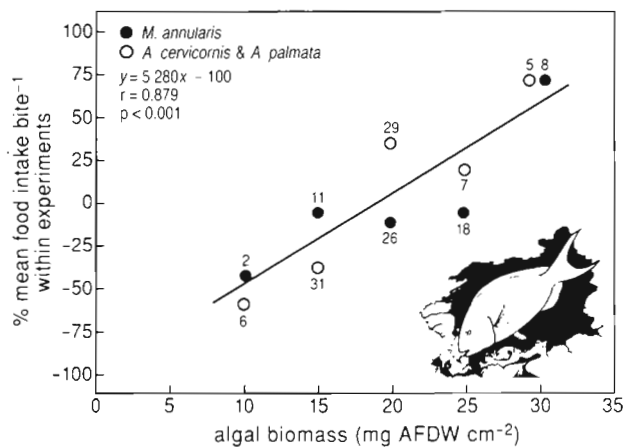


Fig. 2. *Sparisoma viride*. Effect of algal biomass on food intake per bite. The residuals of the mean food intake per bite within experiments are plotted against algal biomass. Mean algal biomass on the different substrate types is given in Table 1. Calculated least-squares regression line through all experimental data is shown. Numbers indicate no. of experimental blocks

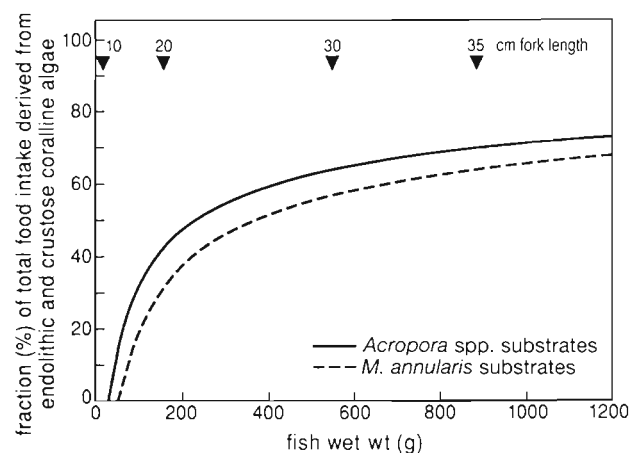


Fig. 3. *Sparisoma viride*. Estimated percentage of the total food intake per bite derived from endolithic and crustose coralline algae as a function of fish size. Percentage intake from substrate-bound algal fractions is calculated as 100% food intake per bite – % food intake from epilithic algae. Intake per bite from epilithic algae is calculated from the surface area of bites in captivity and estimated biomass of epilithic algae on *Acropora* spp. and *Montastrea annularis* substrates respectively

more slowly from 20 cm FL onward. When grazing on *Acropora* spp. substrates, which have a low epilithic biomass, a larger proportion of the food intake per bite is derived from endolithic and crustose coralline algae than when grazing on *Montastrea annularis* substrates, that support a higher epilithic biomass.

Food assimilation experiments

The results of food assimilation experiments are presented in Table 2. Calculated with the direct method, the total assimilation efficiency for adult fish that graze on high-density and low-biomass *Acropora* spp. substrate types averages $20.2 \pm 17.8\%$ ($n = 44$), which is considerably lower than the average assimilation efficiency of 70.5 ± 13.2 ($n = 12$) obtained from a low-density and high-biomass diet on *Montastrea annularis* substrates. The assimilation efficiency attained by juvenile fish feeding on epilithic algal turfs averaged $67.1 \pm 11.1\%$ ($n = 22$). The assimilation efficiency for juveniles obtained from the ash-marker method is much lower: $23.2 \pm 8.6\%$ ($n = 3$).

Table 2. *Sparisoma viride*. Total assimilation efficiency for different diets, calculated using direct and indirect methods. Food intake per bite from each substrate type was calculated using the least-squares regression equation of food intake per bite and fish weight (Fig. 1). Ac+Ap: *Acropora cervicornis* and *A. palmata*; Ma: *Montastrea annularis*. Means \pm SD are calculated over all experimental days

Life phase	Mean fish weight (g)	Substrate type	Duration (d)	Mean intake/bite (mg AFDW _{in})	Mean egestion/bite (mg AFDW _{ex})	Total assimilation efficiency (%) Direct method	Indirect method ^a
JU	5	–	14	0.0404	0.0110	72.8 \pm 8.8	23.9 \pm 8.5
JU	15	–	4	0.0898	0.0391	56.5 \pm 7.9	22.7 \pm 8.6
JU	18	–	4	0.1091	0.0460	57.9 \pm 6.1	23.0 \pm 8.6
						mean: 67.1 \pm 11.1	mean: 23.2 \pm 8.6
IP	183	Ac+Ap	5	0.4253	0.3389	20.3 \pm 12.6	–
IP	362	Ac+Ap	8	0.8401	0.6170	26.6 \pm 4.0	–
IP	397	Ac+Ap	3	0.9233	0.8786	4.8 \pm 8.9	–
IP	561	Ac+Ap	5	1.2964	1.0387	19.9 \pm 4.1	–
IP	571	Ac+Ap	5	1.3277	1.1705	11.8 \pm 6.5	–
IP	640	Ac+Ap	4	1.4874	1.0280	30.9 \pm 23.0	–
TP	958	Ac+Ap	1	2.2264	1.7543	21.1	–
TP	973	Ac+Ap	5	2.2613	2.0516	9.3 \pm 4.9	–
TP	1099	Ac+Ap	5	2.5541	2.0950	18.0 \pm 5.1	–
TP	1130	Ac+Ap	3	2.6245	1.5827	39.7 \pm 40.3	–
						mean: 20.2 \pm 17.8	–
IP	183	Ma	2	1.1028	0.2600	76.4 \pm 12.9	–
IP	397	Ma	3	2.3941	1.0227	57.1 \pm 16.1	–
IP	640	Ma	2	3.8566	1.1499	70.2 \pm 3.9	–
TP	973	Ma	2	5.8633	1.3838	76.4 \pm 0.5	–
TP	1099	Ma	2	6.6226	1.5674	76.3 \pm 0.5	–
						mean: 70.5 \pm 13.2	–

^aAsh fraction of algal turf fronds is $64.1 \pm 11.2\%$ ($n = 59$); ash fraction of faeces of juvenile fish: JU 5 g, $84.2 \pm 0.6\%$ ($n = 4$); JU 15 g, $82.9 \pm 3.5\%$ ($n = 4$); JU 18 g, $83.3 \pm 1.8\%$ ($n = 4$)

Intake of protein and energy was calculated from $AFDW_{in}$ (Table 2), the relative amounts of epilithic and substrate-bound algal fractions in the different diets of experimental fish (Fig. 3), and the biochemical composition of these algal fractions (Table 5 in Bruggemann et al. 1994). Faecal protein content of adult fish was determined for every experimental day; for juveniles the faecal protein content was determined for 2 successive days during each experiment. Faecal caloric content was determined in 22 samples: 15 samples from a wide size-range of fish from different assimilation experiments, and 7 samples collected as fresh 'morning turds' from free-living fish on the reef. Caloric content of experimental and field faeces was not significantly different ($t = -1.36$, $df = 20$, $p = 0.215$), and averaged $17.3 \pm 0.3 \text{ kJ g}^{-1} \text{ AFDW}$. With 1 exception, the average assimilation efficiencies for protein and energy are higher than the average total assimilation efficiencies (Table 3). Only juveniles showed lower assimilation efficiency for protein than for total AFDW, as determined with the direct method. This was due to a slightly higher protein content in their faeces compared to the protein content of their algal food. Protein and energy assimilation efficiencies from the diet on *Montastrea annularis* substrates were higher than from those on the *Acropora* spp. substrate diet.

Table 3. *Sparisoma viride*. Assimilation efficiencies of protein and energy calculated using direct methods. Substrate types: Ac: *Acropora cervicornis*; Ap: *A. palmata*; Ma: *Montastrea annularis*. Means \pm SD are calculated over all experimental days

Life phase	Mean fish weight (g)	Substrate type	Duration (d)	Assimilation efficiency (%)	
				Protein	Energy
JU	5	–	14	69.4 \pm 9.9	78.0 \pm 7.1
JU	15	–	4	47.8 \pm 9.5	64.5 \pm 6.5
JU	18	–	4	51.5 \pm 7.0	65.6 \pm 5.1
				mean: 62.2 \pm 13.4	mean: 73.3 \pm 9.1
IP	183	Ac+Ap	5	69.5 \pm 8.0	33.1 \pm 11.7
IP	362	Ac+Ap	8	67.5 \pm 2.5	42.0 \pm 3.9
IP	397	Ac+Ap	3	70.3 \pm 2.8	26.0 \pm 6.9
IP	561	Ac+Ap	5	62.3 \pm 3.0	38.3 \pm 3.4
IP	571	Ac+Ap	5	56.5 \pm 2.8	31.8 \pm 5.3
IP	640	Ac+Ap	4	65.6 \pm 18.0	46.6 \pm 17.0
TP	958	Ac+Ap	1	56.2	40.9
TP	973	Ac+Ap	5	51.9 \pm 16.7	29.7 \pm 3.4
TP	1099	Ac+Ap	5	67.3 \pm 1.9	37.5 \pm 3.6
TP	1130	Ac+Ap	3	76.2 \pm 22.2	62.7 \pm 31.0
				mean: 64.7 \pm 12.7	mean: 38.8 \pm 15.1
IP	183	Ma	3	88.4 \pm 6.4	81.9 \pm 10.0
IP	397	Ma	3	80.1 \pm 8.0	68.0 \pm 12.0
IP	640	Ma	2	86.1 \pm 7.8	76.6 \pm 3.4
TP	973	Ma	2	88.7 \pm 1.0	82.1 \pm 0.4
TP	1099	Ma	2	88.8 \pm 1.1	82.3 \pm 0.5
				mean: 86.1 \pm 7.0	mean: 77.6 \pm 9.9

Size of grazing scars in captivity and in the field

The surface area and volume of bites was determined from 323 bite scars taken by fish of 27, 30 and 36 cm FL that were kept in the laboratory. Only bite scars from bites taken on the substrate types that were used in the food intake and assimilation experiments were included. On the same substrate types, the size of bites from fish having similar fork length (± 2 cm) was determined from 143 bite scars observed on the reef. The size of bites taken in the field (FB) is usually bigger than of bites taken in captivity (CB), but the difference is not always significant (Table 4). The conversion factor (FB/CB) is not significantly dependent on fish fork length (ANOVA; surface area: $F_{(2,3)} = 0.928$, $p = 0.519$; volume: $F_{(2,3)} = 0.228$, $p = 0.814$) or substrate type (ANOVA; surface area: $F_{(1,4)} = 2.809$, $p = 0.192$; volume: $F_{(1,4)} = 0.318$, $p = 0.612$), and therefore an average conversion factor was calculated for surface area (1.32 ± 0.33) and volume (1.61 ± 0.40).

Foraging effort of *Sparisoma viride*

Total daily foraging period. At first light, the fish leave their nighttime resting places and go to their daytime feeding sites. The onset and cessation of feeding activity is determined by light intensity rather than

the times of sunrise and sunset. For all size classes averaged, the feeding activity starts 53 ± 10 min after sunrise and stops 16 ± 8 min after sunset, with corresponding light intensities of 24 ± 14 and $19 \pm 13 \mu\text{E m}^{-2} \text{ s}^{-1}$. The mean light intensities at the times of the first and last bite of the day are not significantly different ($t = 0.980$, $df = 32$, $p = 0.333$). In the morning, the minimum light intensity for feeding comes late relative to the time of sunrise. This is caused by the location of the study site, being at the base of limestone cliffs that cast a shadow in the morning. Averaged for all size classes of *Sparisoma viride*, the daily foraging period at Karpata equals the daylight period (DLP) – 0.62 h, and ranges from 10.80 h in December to 12.23 h in June.

Initial phase fish of 30 cm FL started feeding at significantly higher light intensities than the other size classes (ANOVA Scheffé, $F_{(3,14)} = 5.368$, $p < 0.05$). The light intensity at the time of the last bite of the day was significantly higher for JU fish than for IP

Table 4. *Sparisoma viride*. Comparison of mean bite size of experimental and field fish. Substrate types: Ac: *Acropora cervicornis*; Ap: *A. palmata*; Ma: *Montastrea annularis*. ns: not significant

Fork length (cm)	Substrate type	Mean size ± SE of n bites of fish in:		p	Correction factor
		Tanks (n)	Field (n)		
Surface area removed (mm ²)					
27	Ac+Ap	36.1 ± 1.5 (102)	34.2 ± 3.8 (20)	ns	0.95
30	Ac+Ap	62.4 ± 9.1 (45)	95.6 ± 12.6 (26)	0.034	1.53
30	Ma	45.4 ± 8.4 (3)	67.6 ± 9.7 (10)	ns	1.49
36	Ac+Ap	71.3 ± 2.1 (168)	70.7 ± 5.7 (61)	ns	0.99
36	Ma	62.6 ± 10.5 (5)	103.6 ± 12.7 (23)	ns	1.66
					mean: 1.32
Substrate volume removed (mm ³)					
27	Ac+Ap	13.0 ± 0.9 (102)	17.6 ± 3.7 (18)	ns	1.35
30	Ac+Ap	43.8 ± 7.8 (45)	81.5 ± 14.0 (26)	0.013	1.86
30	Ma	43.4 ± 6.6 (3)	55.9 ± 9.9 (10)	ns	1.29
36	Ac+Ap	38.5 ± 2.3 (168)	51.9 ± 6.6 (56)	ns	1.34
36	Ma	37.6 ± 9.9 (5)	82.8 ± 17.0 (23)	0.031	2.20
					mean: 1.61

fish of 30 cm FL and TP males, but not significantly different from IP fish of 20 cm FL (ANOVA Scheffé, $F_{(3,14)} = 5.023$, $p < 0.05$). In consequence, IP fish of 30 cm FL start their feeding day on average 15 min later than other size classes, while small fish stop feeding approximately 10 min earlier than larger individuals.

Factors affecting feeding rate. In Fig. 4A all FR observations are plotted against the time of day. FR increases from early morning until noon, followed by a

period with highest FR from noon to approximately 1 h before dusk. This pattern of feeding activity is typical for all observed *Sparisoma viride*, regardless of size, life phase or foraging depth, and does not change qualitatively over the year. Over 70% of the total daily bites are taken after midday.

The feeding rate depends on fish size, life phase and foraging depth. It is inversely linearly related to fork length: smaller fish have higher FR than larger fish. The least-squares regression function of FR to FL is

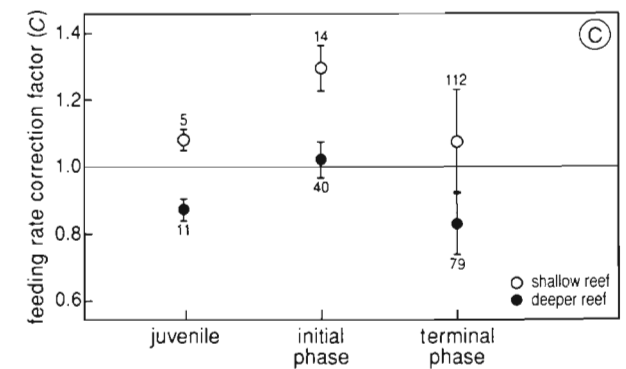
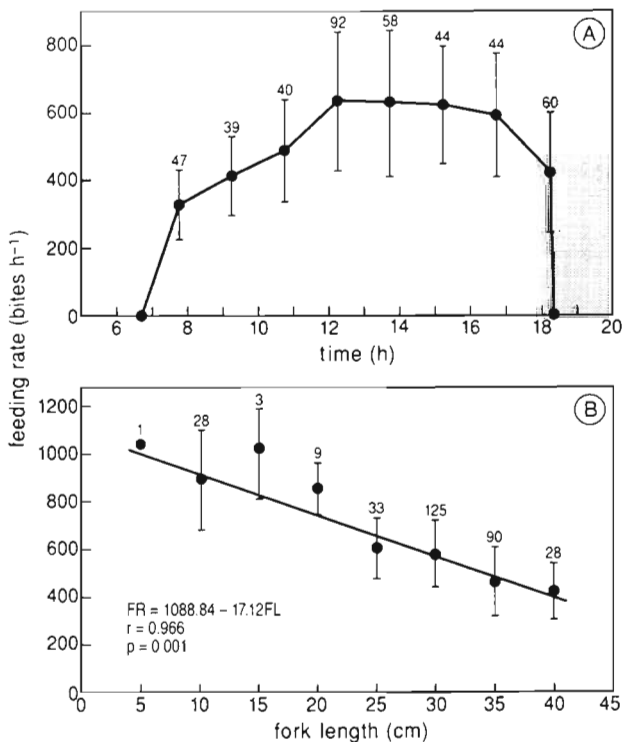


Fig. 4. *Sparisoma viride*. (A) Daily pattern of feeding activity. Feeding rates ± SD of all size classes are plotted against time of day. Value above error bar indicates no. of observations; shaded area indicates time when sun is below the horizon. (B) Effect of fish fork length (FL) on feeding rate (FR). Least-squares regression is fitted through the weighted means ± SD of FR for each 5 cm size class. Dataset is restricted to feeding rate observations made between 09:00 and 17:00 h. Numbers indicate no. of observations for each size class. (C) Effect of life phase and foraging depth on feeding rate. Ratio of group means ± SD to the regression predictions of FR to FL are plotted as feeding rate correction factor. Numbers indicate no. of observations in each group

given in Fig. 4B. Analysis of residuals of the linear regression of FR to FL showed significant effects of life phase and foraging depth (2-way ANCOVA, $F_{(2,258)} = 41.114$, $p = 0.000$). IP *Sparisoma viride* achieve higher feeding rates than both JU and TP fish, and fish foraging on the shallow reef show higher feeding rates than *S. viride* foraging on the deeper reef areas (Fig. 4C). The feeding rate between 09:00 and 17:00 h can be estimated using the formula:

$$FR = C(1088.84 - 17.12 FL), \quad (4)$$

in which *C* represents a correction factor, depending on life phase and foraging depth (Fig. 4C). Seawater temperatures ranged from 26.2 to 29.6°C in the observation period; however, no significant effect of temperature on FR was detected. No effect of season (month of observation) on FR was detected either.

Daily number of bites. From 28 daycover observations, the relation between FR (from 09:00 to 17:00 h), the daylight period, and the total daily bites was established empirically. TDB can be estimated using the equation ($n = 28$, $r = 0.983$, $p < 0.001$):

$$TDB = 284.8 + 0.84 [FR (DLP - 0.62)], \quad (5)$$

Variations in food quality with time of day

The biochemical composition of epilithic sparse algal turfs at different times of day is given in Table 5. Ash contents and soluble carbohydrates showed significant changes over a 24 h period. Ash contents decreased from 06:00 to 18:00 h, and increased during the night. At 18:00 h the ash content was significantly lower compared to the other sampling times ($p = 0.018$). The amount of soluble carbohydrates increased during the day, followed by a decrease during the night. Soluble carbohydrate contents were significantly higher at

Table 5. Biochemical composition of epilithic turf algae at different times within a 24 h period. Means (\pm SE) of replicate samples ($n = 3$) are given. Means were compared using 1-way ANOVA and Scheffé multiple comparison test; $df = 3$ in all cases. Means in horizontal rows with same letter are statistically equivalent

Biochemical composition	Time of day				F	p
	06:00 h	12:00 h	18:00 h	00:00 h		
Ash (% dry wt)	83.1 a (0.9)	81.2 a (1.3)	76.1 b (1.5)	78.5 a (1.2)	6.143	0.018
Soluble carbohydrates (mg g ⁻¹ AFDW)	54.8 a (3.4)	63.5 a (10.2)	89.0 b (0.6)	70.4 a (3.1)	6.725	0.014
Protein (% AFDW)	21.0 a (1.3)	18.8 a (1.9)	17.2 a (0.6)	18.3 a (1.0)	1.555	0.274
Energy (kJ g ⁻¹ AFDW)	17.8 a (0.1)	17.7 a (0.4)	17.6 a (0.1)	17.6 a (0.1)	0.742	0.557

18:00 h compared to other sampling times ($p = 0.014$). Protein and energy contents did not vary significantly with time of day.

DISCUSSION AND CONCLUSIONS

Food intake per bite: effects of food and substrate types

The intake of algal food that is attained from bites taken on dead *Montastrea annularis* substrates is over 2.5 times higher than the food intake attained from bites on *Acropora* spp. substrates. This difference can partly be attributed to differences in vegetation on the experimental substrates. Algal biomass is higher on *M. annularis* substrates than on *Acropora* spp. substrates (21.7 and 18.3 mg AFDW cm⁻² respectively). However, if the effect of algal biomass on the attained food intake per bite is corrected for (using the equation in Fig. 2), the food intake per bite from *M. annularis* is still 2.2 times higher than that from *Acropora* spp. substrates. Not only the biomass, but also the composition of the algal vegetation differed between experimental substrates. *Acropora* spp. substrates had a lower cover of epilithic algae, and a higher cover of crustose corallines than *M. annularis* (Table 1). Crustose corallines yield smaller amounts of algal food per bite, mainly resulting from smaller grazing scars (Bruggemann et al. 1994). Different specific densities of the substrates further contribute to the observed differences in food intake. Bruggemann et al. (1994) demonstrated that the specific density of a substrate affects the size of bite scars left by grazing *Sparisoma viride*: low-density substrates (*M. annularis*) showed bigger grazing scars than high-density substrates (*Acropora* spp.). During the food intake experiments,

we frequently observed that fish were able to denude the surface of *M. annularis* blocks of its algae at a much greater speed than for *Acropora* spp. substrates. In the process, more limestone was also excavated from the former substrate types.

Effect of fish size on composition of ingested algal food

When attaining a larger size, fish not only increase their net food intake per bite, but are also faced with a change in composition of the ingested food. Juvenile *Sparisoma viride* feed almost exclusively on the fronds of epilithic algal

turfs, but as fish grow, an increasing fraction of the ingested food consists of endolithic algae and crustose corallines (Fig. 3). The surface area of bite scars increases linearly with fork length squared (Bruggemann et al. 1994), as does the amount of epilithic algae ingested. The total food intake increases linearly with fish weight, which is a function of fork length cubed. As *S. viride* gains size, the increase in yield from epilithic algae does not match the increase in total AFDW ingested, implying an increasing fraction of the food ingested being harvested from endolithic algal fractions.

When fish are grazing on low-biomass *Acropora* spp. substrates, a larger proportion of the total food intake per bite is derived from endolithic algal fractions than when they graze on high-biomass *Montastrea annularis* substrates. This difference is caused by differences in epilithic algal biomass, biomass being higher on *M. annularis* substrates. A smaller proportion of the total food intake per bite being derived from substrate-bound algal fractions does not imply that the volume of bites on *M. annularis* substrates is smaller. In fact, when the biomass of substrate-bound algae is assumed to be equal for both substrate groups (a reasonable assumption as the thickness of the endolithic layer was not different for the different substrate types, and the biomass of endolithic algae and crustose corallines is not significantly different; see Bruggemann et al. 1994), the higher food intake per bite that *Sparisoma viride* attains from grazing on *M. annularis* is harvested in bigger bites, excavating larger quantities of endolithic and crustose coralline algae.

Factors affecting feeding rate

The feeding rate is defined as the number of bites per hour. Size-related differences in feeding rate may result from differences in bite rate (no. of bites s^{-1}), or from a varying proportion of the time spent on other activities, such as swimming and social interactions with conspecifics. Although there is a negative relationship between bite rate and fish size (slope of linear regression of bite rate to fish fork length: -0.045 ; $r = 0.533$; $p < 0.001$; Bruggemann unpubl. data), this alone cannot account for the reduced feeding rates of larger fish. This implies that smaller fish spend less time on other activities than larger individuals. In a behavioural study of *Sparisoma viride*, Hanley (1984) noted that small individuals were more sedentary and swam less than larger individuals. As an explanation Hanley proposed that small *S. viride* can utilize different food resources, being smaller in size, than large individuals. Small food resources may be uneconomical for large individuals to exploit, as

suggested by the longer swimming times and longer feeding bouts of the latter. Our observations of feeding rates in relation to fish size seem to confirm the hypothesis of different size-related food resources being exploited by *S. viride*.

The effect of life phase on feeding rate can be explained by differences in exploited food resources and by differences in social activity between individuals. JU fish spend less time swimming between food patches than other life phases, but spend more time hovering and looking, presumably for animal prey, before taking a swift bite. The relative feeding rate of JU fish (Fig. 4C) is lower than that of IP fish. TP *Sparisoma viride* also have lower relative feeding rates than IP fish. TP males spend more time on social interactions, related to the defence or potential conquest of territories, than do other life phases (van Rooij pers. comm.). Gladstone (1988) reported reduced feeding rates in territorial males of the sharpnose puffer *Canthigaster valentini* and hypothesized this to be a consequence of the duties associated with defending a territory and maintaining social contact with harem females. IP fish do not partake much in territorial defence; relative to the other life phases, they spend the largest proportion of the day on feeding, presumably to meet the high metabolic demands of daily egg production.

Lower feeding rates on the deeper reef may be related to the spatial distribution of food. The increased structural heterogeneity on the deeper reef (see Table 1 in Bruggemann et al. 1994) causes the food distribution to be more patchy than on the shallow reef. As a consequence, fish on the deeper reef are obliged to swim more between feeding bouts (van Rooij unpubl. data), resulting in lower feeding rates.

Feeding rate and diurnal variation in food quality

Sparisoma viride varies its feeding rate with time of day. The change in feeding rate is correlated with biochemical changes in epilithic algal turfs. Sparse and large turfs constitute important food items for *S. viride*. Both food types are dominated by red algae (Bruggemann et al. 1994). Algal turfs show a decreasing ash content in the course of the day, which means that the potentially digestible organic (AFDW) fraction increases. The soluble carbohydrate content also increases with time of day. The acid-soluble fraction of the total carbohydrates in red algae mainly consists of monosaccharides, such as glucose, fructose and galactose, and of alpha-linked storage polysaccharides, such as Floridean starch (Craigie 1974). As amylase activity is high in the guts of herbivorous fish (Fish 1960, Gohar & Latif 1961), higher fractions of

soluble carbohydrates imply greater digestibility of algal food during the afternoon. *S. viride* seems to exploit these changes in food quality by exerting higher feeding rates during the second half of the day, resulting in >70% of total daily bites being taken after midday. Polunin & Klumpp (1989) also found that the daily pattern of feeding activity of herbivorous fish (pomacentrids and acanthurids) is correlated with diurnal changes in food quality. In a study of 2 different populations of a herbivorous blenny species, Zoufal & Taborsky (1991) concluded that the diurnal foraging periodicity matches the changes in energy content of the algal food. The origins of the diurnal changes in energy content were, however, not explained.

Assimilation efficiency of total organics, protein and energy

Determining the assimilation efficiency in juvenile *Sparisoma viride* using the algal ash fraction as an indigestible marker (Montgomery & Gerking 1980, Horn & Neighbors 1984, Horn et al. 1986) seems an attractive alternative to direct methods which require painstaking quantification of the food consumed. However, the ash-marker method has been criticized (Bjorndal 1985), as absorption of ash has been shown by Buddington (1980). If some of the ash is absorbed, the assimilation efficiency is underestimated. We found the assimilation efficiencies determined by the indirect method to be much lower than those determined by quantification of food consumed and faeces produced. This may well be the result of absorption of some dietary ash. Parrotfish are able to dissolve calcium carbonate in their gut (Smith & Paulson 1974, 1975). From our own measurements, we know the duodenum, i.e. the first quarter of the gut after the pharyngeal mill, to be slightly acidic (pH = 6.5 to 6.7) in both juvenile and adult *S. viride* (pers. obs.). As the ash fraction of epilithic algal turfs consists almost entirely of calcium carbonate, some of this ash fraction probably dissolves during the passage through the gut, resulting in an underestimation of assimilation efficiencies when using the ash-marker method. However, the direct method for estimating the assimilation efficiency also has its drawbacks for JU fish. With the high number of bites taken by JU fish, the daily intake of organic matter can be easily over- or underestimated if a slightly incorrect estimate of the food intake per bite is used.

There was a remarkable difference in assimilation efficiency by fish feeding on a diet that consisted of algae growing on *Acropora* spp. substrates, collected from the shallow reef zone, compared to those feeding

on a diet of algae growing on *Montastrea annularis* substrates that were collected from the deeper reef zones. Two factors may have contributed to the observed difference in assimilation efficiencies. Firstly, the biomass of epilithic algae on *M. annularis* substrates is higher than on the *Acropora* spp. substrates, as indicated by a higher total biomass, and a higher percentage cover of epilithic algal turfs. Epilithic algae are more readily accessible for digestion than endolithic and crustose coralline algae. The latter are embedded in a carbonate matrix that has to be ground in the pharyngeal mill to make them accessible for digestion. This notion is supported by the high assimilation efficiency attained by JU fish from their diet of algal turfs, as determined by direct methods (Table 2). Secondly, the higher skeletal density of *Acropora* spp. substrates will have a negative effect on the pharyngeal grinding efficiency of the carbonate matrix, making the substrate-bound algal fractions less accessible for digestion than those inhabiting lower-density substrates like *M. annularis*. For *Sparisoma viride*, the food scraped off *Acropora* spp. substrates consists to a large extent of these substrate-bound algal fractions (Fig. 3), resulting in a lower assimilation efficiency.

The assimilation efficiencies for total AFDW, protein and energy determined for *Sparisoma viride* fall within the range of values reported by Horn (1989) for other herbivorous fish. In adult *S. viride*, assimilation efficiency was highest for protein, intermediate for energy, and lowest for total AFDW. Similar results were obtained by Montgomery & Gerking (1980), Gerking (1984) and Anderson (1988), using either direct or indirect methods. In JU *S. viride*, protein assimilation efficiency appeared to be lower than the assimilation efficiency of total AFDW. This result is at odds with other values of the protein assimilation efficiency reported here, or by other authors, and is caused by higher protein levels found in faeces than those measured in their algal food (large turfs). JU *S. viride* probably supplemented their algal diet with the small invertebrates that were inadvertently introduced along with the algae, resulting in a higher protein content of the food than that measured for plant material alone.

Food intake per bite: experimental vs field situation

The experimentally determined food intake and assimilation efficiency form the basis for estimating the daily intake and assimilation of food, protein and energy by *Sparisoma viride* foraging on the reef. The size of bites taken by free-living fish was, however, larger than the size of bites taken by fish in experi-

Table 6. *Sparisoma viride*. Conversion factor of experimental to field food intake per bite for 2 substrate types

Fork length (cm)	Conversion factor	
	<i>Acropora</i> spp.	<i>Montastrea annularis</i>
10	1.32	1.32
20	1.44	1.41
30	1.50	1.48
40	1.53	1.52

mental tanks (Table 4). Therefore, conversion factors were determined. One problem here is that the mean conversion factor for the surface area of bite scars was smaller than the conversion factor for bite-scar volume. We therefore calculated a new conversion factor for total food intake per bite, by weighting the conversion factors of surface area and volume with the relative contribution of epilithic and substrate-bound algal fractions in the bite (Fig. 3). This new conversion factor is dependent on fish size and on substrate type (Table 6).

Another problem in translating the experimentally determined food intake to the field situation lies in the variability of algal biomass in the field. With the least-squares regression that describes the effect of algal biomass on the food intake per bite (Fig. 2), differences in algal biomass in the field can be corrected for.

Finally, an assumption has to be made concerning the effect of substrate density on the food intake per bite in the field. Grazing on other high-density substrates (for instance *Agaricia* spp.) is assumed to yield the same amount of food per bite as grazing on the high-density *Acropora* spp. substrates used in the experiments. Similarly, it is assumed that food intake from other low-density substrates (*Madracis mirabilis*, *Diploria* spp. and *Colpophyllia natans*) is comparable to that attained from low-density *Montastrea annularis* substrates used in experiments.

Daily intake and assimilation of food, protein and energy in the field

Total daily bites, food intake, and assimilation of total AFDW, protein and energy d^{-1} were calculated for 4 size classes feeding on the shallow reef and on the deeper reef respectively (Table 7) using the equations presented in this paper and the diet composition of *Sparisoma viride*, as described by Bruggemann et al. (1994: Tables 3 & 4). Daily intake of protein and energy was calculated from the food intake per bite (Fig. 1), the proportion of the food intake derived from epilithic and substrate-bound algal fractions (Fig. 3), and the biochemical composition of these algal fractions (Table 5 in Bruggemann et al. 1994). Daily amounts of food, protein and energy assimilated were calculated using the mean assimilation efficiencies for each diet (Tables 2 & 3). For juvenile parrotfish, the mean of estimates obtained by the direct and indirect method was used.

In spite of taking more bites, *Sparisoma viride* feeding on the shallow reef have lower daily food intake, and lower amounts of assimilated AFDW, protein and energy than similarly sized individuals feeding on the deeper reef (Table 7). This is due to a far greater proportion of bites taken by the latter on low-density substrates, dominating the available grazing surfaces on the deeper reef. These substrates yield a higher food intake per bite, and enable a more efficient assimilation of the ingested food. The increased net yield makes territorial defence economically feasible on the deeper reef.

From the mean C contents of the food (40.5%), we calculated the daily ingestion of organic C by *Sparisoma viride* and some other herbivorous fish. A log-log plot (Fig. 5A) of daily intake of algal C in relation to fish weight shows that intake by *S. viride* falls in the range of values determined in the field for other tropical herbivorous fish, as reported by Chartok

Table 7. *Sparisoma viride*. Effects of size and foraging depth on daily food intake and assimilation of algal AFDW, protein and energy

Foraging depth Life phase	Fork length (cm)	Fish wet wt (g)	Total daily bites	Daily food intake (g AFDW)	Daily assimilation		
					Food (g AFDW)	Protein (g)	Energy (kJ)
Shallow reef							
Juveniles	10	17	9998	0.56	0.25	0.04	7
Initial phase	20	153	9763	5.35	1.25	0.32	44
Initial phase	30	547	7582	15.37	3.59	0.86	130
Terminal phase	40	1350	4528	22.96	5.38	1.24	197
Deeper reef							
Juveniles	10	17	8119	0.94	0.42	0.07	13
Initial phase	20	153	7763	7.35	4.36	0.57	104
Initial phase	30	547	6041	21.36	12.71	1.57	314
Terminal phase	40	1350	3551	31.73	18.91	2.28	474

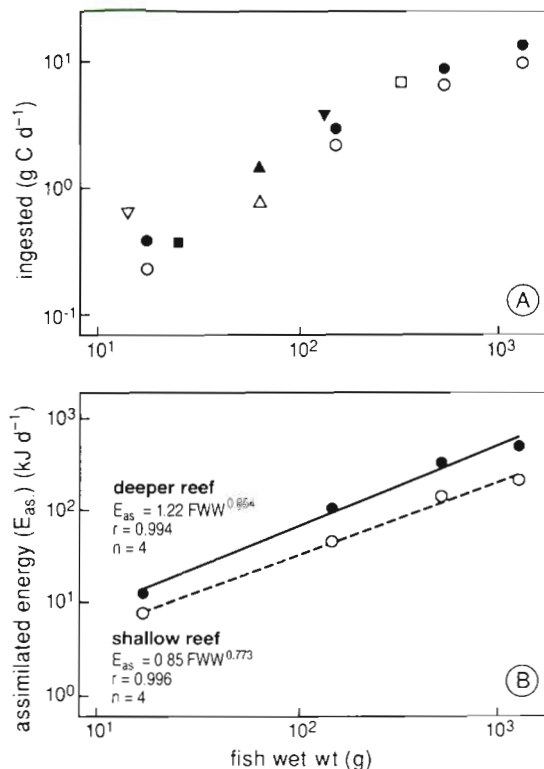


Fig. 5. (A) Daily intake of algal carbon by herbivorous fish versus fish wet wt (FWW). Circles represent values for *Sparisoma viride* foraging on (○) the shallow reef and (●) the deeper reef. Other values were recalculated from Chartok (1983) for *Acanthurus guttatus* (□), Gerking (1984) for *Sarpa salpa* (maintenance ration: ■), Polunin (1988) for *Plectroglyphidodon lacrymatus* (winter: △; summer: ▲), Klumpp & Polunin (1989) for *Stegastes apicalis* (▽), and Montgomery et al. (1989) for *Acanthurus nigrofuscus* (▼). (B) Daily assimilated energy (E_{as}) by various size *S. viride* foraging on the deeper reef and the shallow reef, respectively

(1983), Polunin (1988), Klumpp & Polunin (1989) and Montgomery et al. (1989), and the experimentally determined maintenance ration for the temperate herbivorous fish *Sarpa salpa* (Gerking 1984).

In Fig. 5B the relation of daily assimilated energy to fish weight is shown for *Sparisoma viride* foraging on the deeper reef and the shallow reef, respectively. The weight exponents of the functions fitted through the data points are close to 0.8, and are within the range of values reported for daily food intake by fish as determined in experimental studies (Elliott 1975, Pálsson et al. 1992; see Jobling 1993 for review). Weight exponents <1 for daily assimilated energy indicate lower mass-specific energy requirements in larger fish. With increasing size, an increasing proportion of the food is derived from substrate-bound algal fractions. Exploiting these food resources probably involves additional costs of excavating and grinding these limestone-embedded algae. In this study we quantified the effects

of social organization, life phase and size on the daily amounts of ingested and assimilated algal nutrients and energy. It provides a necessary step for further evaluation of the daily balance of energy intake and allocation by this large herbivorous reef fish. Secondly, it provides a basis for quantifying the impact of scarid grazing on the algal food supply, and for an assessment of their contribution to the energy flow in coral reef ecosystems.

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