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Assays to control the development of the green staining alteration in Spanish-style green olives of the Gordal variety

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Running title: Green staining alteration in Gordal olives

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2
3 19 **Abstract**
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5 20 BACKGROUND: Olives of the Gordal variety processed according to the Spanish-style
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7 21 sometimes develop an alteration in color known as green staining (GS) due to the
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9 22 formation of harmless copper-chlorophyll complexes that make the product less
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11 23 valuable. The aim of this study was to investigate methods to minimize the impact that
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13 24 this alteration supposes for the table olive industry.
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17 26 RESULTS: Calcium chloride, sorbic, benzoic and ascorbic acids and SO₂ did not inhibit
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19 27 the development of the alteration in olives packed under their own fermentation brine or
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21 28 new fresh brine. It was also discovered that the incubation of olive samples at 45 °C for
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23 29 20 days accelerates the formation of GS, and it can be a very useful tool to predict the
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25 30 incidence of the alteration in advance. By applying this test to numerous industrial tanks
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27 31 for four consecutive seasons, it was found that GS was mainly present in olives
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29 32 harvested at the beginning of the season.
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33 34 CONCLUSION: The formation of GS in olives of the Gordal variety is time and
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35 35 temperature dependent, and none of the additives tested avoided or retarded the
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37 36 development of the alteration. However, an accelerated test to predict the development
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39 37 of the GS formation has been proposed that could contribute to minimize the effects of
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41 38 the alteration.
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45 40 **Keywords:** table olives; copper-chlorophyll; color; alteration; Gordal
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41 INTRODUCTION

42 Among the olive varieties intended for table olives, Gordal is one of the most
43 appreciated by consumers due to its large size and high flesh/pit.¹ This variety is also
44 known as Sevillana, and is usually processed according to the Spanish-style. The olives
45 are picked during September and October in Spain when they have a green-yellow color
46 on their surface. Upon arrival to the olive factories, the fruit is treated first with a dilute
47 NaOH solution to hydrolyze the bitter glucoside oleuropein. Subsequently, the olives
48 are washed with tap water to remove excess alkali and covered with a NaCl solution in
49 which spontaneous lactic acid fermentation takes place for months.² At the end of the
50 process, the olives have a yellow color that makes them very attractive for consumers.
51 However, the Gordal olives sometimes develop an alteration in color known as “green
52 staining” (GS). This is seen as bluish-green zones distributed over the skin that are
53 initially small spots but with time can cover a great part of the fruit surface (Figure 1).

54 The formation of Cu-chlorophyll complexes with the endogenous copper of the
55 fruit has been associated with the appearance of GS in Gordal olives fermented
56 following the Spanish-style.³ Nevertheless, these metal-chlorophyll complexes have
57 also been detected in olives elaborated following the Castelvetro method,⁴ and in
58 several samples of commercial table olives.⁵

59 It has been proposed that the pectin chain acts as a reservoir of the Cu which
60 forms a complex with chlorophyll derivatives,⁶ but the involvement of oxidative
61 disintegration of the fruit chloroplasts is also necessary to allow contact between the
62 chlorophyll pigments and the copper.⁷ However, the fact that this alteration is mainly
63 expressed in fruit of the Gordal variety raises the question of specific characteristics of
64 this variety.

65 Gordal olives are amongst those of the biggest size employed for table olive
66 processing,¹ and they show a high respiration rate during the post-harvest period.⁸ It is
67 considered a sweet variety due to its low content in phenolic compounds, particularly
68 the bitter glucoside oleuropein,⁹ and it favors lactic acid fermentation.¹⁰ Gordal olives
69 also possess higher polyphenoloxidase activity than Hojiblanca but similar to
70 Manzanilla, which are the two most employed Spanish varieties for processing as table
71 olives.⁹ PPO is a metallo enzyme with Cu in its prosthetic group that could be involved

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3 72 in the formation of the Cu-chlorophyll complexes although this relationship has not
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5 73 been found.¹¹ Besides, none of these two varieties usually develops GS.

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7 74 Although the Cu-chlorophyll complexes possess inhibitory activity against
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9 75 mutagenesis and high antioxidant activity,^{12,13} color is important for consumer
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11 76 acceptance and GS alteration is of great concern for olive factories. Hence, methods to
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13 77 inhibit or retard the development of GS in fermented Gordal olives are required. The
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15 78 aim of this work was to gain knowledge about the variables that affect the formation of
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17 79 GS in this particular olive variety, ways and methods to prevent the formation of the
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19 80 alteration, and to develop an accelerated test to predict the formation of GS.

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22 82 **MATERIALS AND METHODS**

23 24 25 83 **Assessment of GS**

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27 84 An accelerated test was developed to predict the formation of GS with time. To this
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29 85 aim, around 580 g of olives obtained from industrial tanks were packed in quarter-liter
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31 86 jars and covered with 350 mL of fermentation brine. Subsequently, they were sealed
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33 87 and incubated in a thermostatic chamber at 45 °C for 20 days. Finally, five trained
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35 88 experts visually assessed the appearance of bluish-green zones on the surface of every
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37 89 olive. The effectiveness of the method was checked during the season 2006/2007 by
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39 90 comparing the percentage of olives with GS predicted by the accelerated test in January,
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41 91 2007 and the presence of GS in fruit of the same batch stored at ambient temperature
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43 92 until September, 2007.

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45 93 The GS test was carried out on olives of the Gordal variety processed according
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47 94 to the Spanish-style from the 2006/2007 to 2009/2010 season. The fruits were
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49 95 elaborated in three olive factories located in the province of Seville (Spain), and the
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51 96 method consisted of placing the fruits in a NaOH solution of 15-20 g L⁻¹ for 8-10 h until
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53 97 the alkaline solution reached two-thirds of the way to the pit of the olives.
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55 98 Subsequently, the fruit were washed with tap water for 5-10 h and then covered with a
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57 99 100 g L⁻¹ NaCl solution where spontaneous lactic acid fermentation took place for
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59 100 months. The fermentation step was carried out in tanks containing around 10000 kg of
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101 olives and 5000 L of brine. The number of tanks analyzed was 146, 223, 364 and 233
102 during the 2006/2007, 2007/2008, 2008/2009 and 2009/2010 seasons, respectively.

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104 Packing experiments to prevent the development of GS

105 Gordal olives processed according to the Spanish-style were obtained from 10 industrial
106 tanks in June 2006 during the 2005/2006 season (samples A-J). Subsequently, the fruits
107 were packed in the jars, and covered with i) the same fermentation brine, ii) a new
108 brine of 60 g L⁻¹ NaCl, iii) this new brine spiked with 3 g L⁻¹ calcium chloride. The jars
109 were incubated at 45 °C until September, except a lot with fermentation brine that was
110 stored at 7 °C. Finally, the presence of GS was assessed by the trained olive experts.

111 Olives from only one tank of the 2005/2006 season with symptoms of GS were
112 packed in June in the jars, and covered with the fermentation brine alone or the brine
113 spiked with the preservatives potassium sorbate (1 g L⁻¹ sorbic acid), sodium benzoate
114 (2 m L⁻¹ benzoic acid) or citric acid (6 g L⁻¹). The jars were incubated until September at
115 45 °C. Olives packed with the fermentation brine alone were also incubated at 7 °C, 25
116 °C and 65 °C until September for the assessment of GS.

117 Fruits from the season 2006/2007 with symptoms of GS were obtained in
118 February 2007 from an industrial tank of Gordal olives processed according to the
119 Spanish-style. The olives were packed in the jars and covered with the fermentation
120 brine or this brine spiked with ascorbic acid (1 g L⁻¹) or SO₂ (0.2 g L⁻¹). The jars were
121 incubated at 45 °C until June.

123 Fermentation experiments to prevent the development of GS

124 At the beginning of the 2009/2010 season (second week of September 2009) an
125 experiment was performed to study the influence of the processing conditions on the
126 development of GS in olives of the Gordal variety. The fruits were processed during
127 three consecutive days in 15 industrial tanks made of fiberglass and buried into the
128 ground. Around 10000 kg of olives were covered in each tank with 5000 L of NaOH
129 17 g L⁻¹ until the alkaline solution penetrated two thirds of the way to the pit of the fruit.
130 Subsequently, the NaOH solution was discharged and tap water was added. After 4-8 h,
131 the washing water was eliminated, and the olives were immersed in a 100 g L⁻¹ NaCl
132 solution for fermentation. A batch of 15 kg of debittered olives immersed in their own

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3 133 brine was taken out from each tank and transported to the Instituto de la Grasa facilities.
4 134 Three kilograms of olives from each industrial tank were put into four polyethylene
5 135 recipients, covered with 1.5 L of the brines and left for fermentation at ambient
6 136 temperature (Treatment A). Two of the recipients were inoculated with the strain
7 137 *Lactobacillus pentosus* LP99 from the Instituto de la Grasa's own collection (Treatment
8 138 B). The strain was grown in flasks with MRS broth + 40 g L⁻¹ NaCl and incubated
9 139 overnight at 32 °C. The cultures were centrifuged and the pellets washed with saline.
10 140 The washed cells were centrifuged again, re-suspended in saline and added to the
11 141 fermenters 48 h after brining. In order to overcome the unfavorably high pH values, the
12 142 inocula were calculated to achieve an initial population of ca. of 10⁸ CFU mL⁻¹. The
13 143 population of lactic acid bacteria in each fermenter before and after inoculation was
14 144 checked by seeding their brines onto MRS agar plates with a spiral plater, and counting
15 145 the colonies after 72 h of anaerobic incubation at 32 °C.

16 146 At the same time, a batch of 12 kg of raw olives was obtained from each
17 147 industrial tank before processing and transported to the Instituto de la Grasa facilities.
18 148 Three kilograms of fruit from each lot were put into four polyethylene recipients,
19 149 covered with a NaOH solution, washed with tap water and put into brine in a similar
20 150 manner as that done at industrial scale. Two of the recipients were left at ambient
21 151 temperature in the Instituto de la Grasa (Treatment C), and the olives from the other two
22 152 recipients were transported to the olive factory and immersed inside a plastic net into
23 153 each industrial tank for fermentation (Treatment D).

24 154 The accelerated test was performed on olives of all treatment in March, 2010.
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26 156 **Chemical analyses**

27 157 The titratable acidity, pH and combined acidity of the olive solutions were measured
28 158 using a Metrohm 670 Titro-processor (Herisau, Switzerland). Titratable acidity was
29 159 determined by titrating up to pH 8.3 with NaOH 0.2 M and expressed as percent (w/v)
30 160 lactic acid.

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32 162 **RESULTS AND DISCUSSION**

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3 163 The results from the packing experiments carried out to assess the influence of calcium,
4 164 temperature and type of brine on GS development are presented in Table 1. Packing
5 165 with fresh brine did not show a lower incidence of GS that using the same fermentation
6 166 brine, particularly when focusing on the three samples with higher percentages of
7 167 altered fruit (E, H and J). It must be noted that the pH was rather similar for olives
8 168 packed in both types of brine after equilibrium. Moreover, the addition of calcium into
9 169 the brines did not affect the percentage of the alteration despite the fact that a higher Ca
10 170 content has been reported in altered fruit than unaltered, and higher in the altered zones
11 171 than in the unaltered zones of the same fruit.⁶ Pectin chains have been proposed as a
12 172 storage place for Cu, and displacement of this cation by calcium from the galacturonic
13 173 units could occur. Besides, the addition of calcium to olives preserved under acidic
14 174 conditions inhibits cell wall degradation thereby lessening the softening rate of the fruit
15 175 with time.¹⁴ In fact, the formation of Cu-complexes has been associated with cell
16 176 degradation, particularly chloroplasts.⁷ Likewise, this phenomenon seems to be
17 177 temperature dependent, which can be observed in Table 1. Olives preserved at 7 °C had
18 178 a lower incidence of GS than those at 45 °C regardless of the type of brine or calcium
19 179 addition.

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32 180 A new packing assay was performed with olives obtained from industrial tanks
33 181 in June. They were packed in their own fermentation brine, and stored until September.
34 182 Again, GS percentage was higher with increasing storage temperature from 7 °C to 45
35 183 °C (Table 2). Surprisingly, the alteration was lower when olives were kept at 65 °C,
36 184 which could suggest the involvement of enzymatic reactions during the formation of
37 185 GS. However, it must be noted that the olives suffered a severe alkaline treatment at the
38 186 beginning of the Spanish-style processing that probably inactivated most olive
39 187 enzymes.¹⁵ Moreover, the influence of the preservatives sorbic and benzoic acids as
40 188 well as the metal-chelating citric acid was also tested during this season 2005/2206 but
41 189 none of them prevented the formation of GS during summer storage of the fruit at 45
42 190 °C.

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51 191 Because of the suspicion about enzymatic involvement in the formation of GS,
52 192 a new assay was carried out during the next 2006/2007 season with olives that were
53 193 pasteurized after packing in their own fermentation brine with and without the addition
54 194 of the antioxidants ascorbic acid and SO₂. The results presented in Table 3 reveal a
55 195 continuous increase in the GS alteration from February to June regardless of the

196 addition of antioxidants. Also, the formation of GS occurred in these pasteurized olives
197 and therefore the involvement of enzymatic reactions must be ruled out.

198 From the foregoing it is deduced that none of the additives studied and with
199 permitted use in table olives inhibited or retarded the formation of GS in Gordal
200 olives.¹⁶ Hence, an accelerated test was performed to predict the development of the
201 alteration as soon as possible after primary fermentation, which may last from
202 September to December. With this aim, samples of 1-2 kg of olives from 33 industrial
203 tanks from the 2006/2007 season were packed in jars and covered with their
204 fermentation brine in the second fortnight of December. One jar was incubated at 45 °C
205 and another left at ambient temperature. In January, after 20-25 days of incubation at 45
206 °C the jars were opened and the percentage of GS alteration assessed. Likewise, the
207 alteration was checked in the olives left at ambient temperature in September. The
208 results are presented in Table 4. First, it is noteworthy that it was predicted in January
209 that olives from 24 tanks would not develop the alteration, which was confirmed in
210 September with the olives stored at ambient temperature, none of them had GS. There
211 were 9 tanks with symptoms of the alteration, and particularly three of them contained
212 fruit with a high percentage of GS, which was clearly predicted in January. Therefore,
213 the accelerated test is a simple and reliable method to predict the appearance of GS in
214 olives of the Gordal variety as soon as 2-3 months from brining.

215 Once the accelerated test was developed, it was employed for four consecutive
216 seasons during the months of December-January. Figure 2 shows the percentage of
217 tanks liable to be affected by the alteration with olives processed from the beginning to
218 the end of the 2006/2007 and 2007/2008 seasons. It is clear from the data of both
219 seasons that the alteration occurs mainly in olives picked at the beginning of the
220 harvesting period. About half of the tanks containing olives harvested from 13 to 21 of
221 September presented the alteration while olives harvested in October did not develop
222 GS. Changes in sugars, fatty acids and phenolic compounds during the maturation of
223 Gordal olives have been reported,¹ and their behavior was rather similar to that found
224 for other olive varieties non-prone to GS. The evolution of the superoxide dismutase
225 and polyphenoloxidase activities, which are related with the protection of the
226 chloroplast pigments against oxidative species, during the growth of Gordal was similar
227 to that observed for the Manzanilla variety.¹¹ To our knowledge, there is no explanation
228 for the higher formation of GS in olives early harvesting.

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3 229 In addition, the intensity of the alteration depended on the olive season. As can
4 230 be seen in Figure 3, the percentage of tanks with olives showing GS was around 30%
5 231 for the 2006/2007 and 2007/2008 seasons whereas it was lower than 5% for the
6 232 2008/2009 season. In the following 2009/2010 season this percentage raised again to up
7 233 to 23%. These figures are very reliable since a very high number of tanks were analyzed
8 234 each season and it means that just lower than 30% of olive tanks may present olives
9 235 with GS during fermentation. Of course, not all the olives in the tanks possess the
10 236 alteration, though consumers may reject a commercial sample with just a few altered
11 237 olives in it.

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19 238 Gallardo-Guerrero *et al.*,⁷ reported that the alkaline treatment causes chlorophyll
20 239 oxidation as well as cell deterioration, and both phenomena are related with the
21 240 formation of GS. They also suggested that the reduction in pH of the brine below 4.5
22 241 units is also necessary to allow the insertion of Cu into the chlorophyll molecule. We
23 242 carried out an experiment at laboratory and industrial scale to assess the influence of the
24 243 debittering and fermentation steps on the development of the alteration. Table 5 shows
25 244 the results obtained with olives from 5 different tanks although the assay was performed
26 245 with another 10 tanks. Unfortunately, the olives from the latter 10 tanks did not present
27 246 the alteration thereby the data are not shown. The most surprising result from these
28 247 experiments was the absence of GS in the olives which were debittered and fermented at
29 248 the laboratory (treatment C). Likewise, the olives debittered at the laboratory but
30 249 fermented at the olive factory showed a low incidence of the alteration, this was
31 250 particularly observed in the olives of tank n° 5. By contrast, fruit debittered at the olive
32 251 factory developed a high percentage of GS regardless of whether they were fermented at
33 252 the factory or laboratory, even in inoculated batches. As it was explained in the
34 253 materials and methods section, the debittering conditions were rather similar at both
35 254 laboratory and industrial scale. However, the size of the batches was very different,
36 255 around 3 kg in the laboratory fermenters and 10000 kg in the factory tanks. It has been
37 256 indicated that during the alkaline treatment an increase in temperature takes place inside
38 257 the industrial tanks.¹⁷ The intensity of the alkaline treatment has been associated with
39 258 the appearance of GS in Gordal olives due to its effect on chloroplast disintegration and
40 259 could be an explanation for the different results obtained at laboratory and industrial
41 260 scale.⁷

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262 CONCLUSIONS

263 The formation of GS in olives of the Gordal variety is time and temperature dependent,
264 and none of the additives tested avoided or retarded the development of the alteration.
265 Moreover, it has been seen that the elaboration of the fruit at small scale reduced the
266 appearance of the alteration to a large extent, although further assays are necessary to
267 confirm these data.

268 An accelerated test to predict the development of the GS formation has been
269 proposed, which is based on incubation of the olive sample at 45 °C for 20 days. This
270 test was performed on olives from numerous industrial tanks for 4 consecutive seasons
271 and it was found that the incidence of the alteration was higher in fruit elaborated at the
272 beginning of the season. In addition, the percentage of tanks affected by the alteration
273 depended on the season but it was lower than 30 % of the total. Therefore, processors
274 can use this new accelerated test to manage the alteration and minimize the effects of
275 the alteration.

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Table 1

Influence of calcium, type of brine and temperature on the development of green staining alteration. Gordal olives of the 2005/2006 season processed according to the Spanish-style at industrial scale were packed in glass containers in June and stored for analysis. The percentage of fruit with alteration (%) was evaluated in September.

Samples	% olives with green staining				
	Treatment				
	FB-7 ^a	FB-45	FB-45-Ca	NB-45	NB-45-Ca
A	0	2	2	0	2
B	2	0	0	0	0
C	0	0	0	0	0
D	2	0	0	0	0
E	16	21	13	16	31
F	2	2	5	7	2
G	0	0	0	0	5
H	17	35	28	39	31
I	0	5	5	9	7
J	13	28	32	12	28

^aFB-7, fermentation brine and 7° C; FB-45, fermentation brine and 45 °C; FB-45-Ca, fermentation brine, calcium and 45 °C; NB-45, new brine and 45 °C; NB-45-Ca, new brine, calcium and 45 °C.

Table 2

Influence of temperature and several food additives on the development of green staining alteration. Gordal olives of the 2005/2006 season were taken from an industrial tank, put into glass containers, and covered with the fermentation brine and additives in June. The percentage of fruit with alteration (%) was evaluated in September.

Treatment	% olives with green staining
7° C	23
25 °C	28
45 °C	33
65 °C	21
45 °C + sorbic acid (1 g L ⁻¹)	35
45 °C + benzoic acid (2 g L ⁻¹)	25
45 °C + citric acid (6 g L ⁻¹)	34

Table 3

Influence of ascorbic acid and SO₂ on the development of green staining alteration in Gordal olives of the 2006/2007 season. Fruits were put into jars, covered with the fermentation brine and pasteurized in February. The jars were incubated at 45 °C until June and the presence of GS was evaluated.

Treatment	% olives with green staining		
	February	March	June
Control	5.5 (0.7) ^a	18.0 (2.8)	33.0 (1.4)
Ascorbic acid (1 g L ⁻¹)	5.0 (1.4)	18.0 (5.6)	32.0 (5.6)
SO ₂ (0.2 g L ⁻¹)	5.0 (1.4)	18.0 (1.4)	41.0 (2.8)

^aStandard deviation of duplicates

Table 4

Green staining formation predicted with the accelerated test in January and development of the alteration in olives stored at ambient temperature (23 ± 4 °C) up to September. Gordal fruits were obtained from fermentation tanks of the 2006/2007 season.

Percentage of green staining (%)		
Tanks (n°)	Predicted in January	Developed in September
24	0	0
3	1	0
2	2	0
1	2	1
1	36	44
1	52	34
1	81	61

Table 5

Percentage of olives affected with green staining alteration (%) from fruit processed following different processing methods during the season 2009/2010.

Sample	Treatment ^a			
	A	B	C	D
1	5.7	2.8	0	5.1
2	0	1.0	0	3.6
3	0.8	1.8	0	0
4	0	6.6	0	0
5	21.9	32.2	0	6.2

^aA, olives debittered at the olive factory and fermented at the laboratory; B, olives debittered at the olive factory and fermented with inoculation at the laboratory; C, olives debittered and fermented at the laboratory; D, olives debittered at the laboratory and fermented at the olive factory.

Figure legends

Figure 1. Photograph of the green staining alteration.

Figure 2. Percentage of industrial tanks liable to be affected with the green staining alteration during the seasons 2006/2007 and 2007/2008. The number of tanks analyzed with the accelerated test performed in January was 146 and 223 for the seasons 2006/2007 and 2007/2008, respectively.

Figure 3. Percentage of industrial tanks liable to be affected with the green staining alteration from the seasons 2006/2007 to 2009/2010. The number of tanks analyzed is indicated on the top of the bars. The accelerated test was performed in January.

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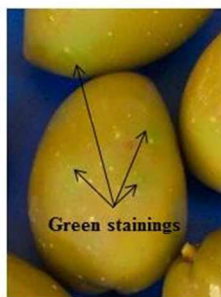


Figure 1
190x254mm (96 x 96 DPI)

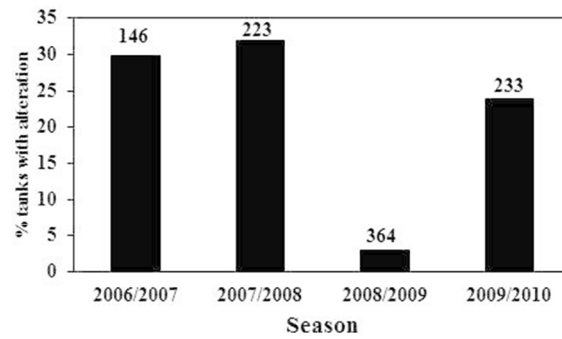


Figure 3
190x254mm (96 x 96 DPI)