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RESEARCH PAPER

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Anti-platelet aggregation assay and chemical composition of essential oil from *Allium atroviolaceum* Boiss growing in Iran

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Key words: *Allium atroviolaceum*, essential oil, GC-MS, antiplatelet aggregation, steam distillation.

<http://dx.doi.org/10.12692/ijb/5.2.151-156>

Article published on July 28, 2014

Abstract

Plants belonging to genera *Allium* have widely been acquired as food and medicine. Their wide use was mainly due to the medicinal properties attributed to these plants over the centuries, lately supported by epidemiological and research studies. In this study, essential oil constituents of *Allium atroviolaceum* growing in Shahr-e-kord, Iran, were investigated through gas chromatography/mass spectrometry (GC-MS) technique. In this essential oil two major constituents were trisulfide, di-2-propenyl (26.85%) and diallyl disulphide (10.98%) while trans-2-(2-pentenyl) furan (0.02%) and Limonene (0.06%) have been identified in lower amounts. The *in-vitro* antiplatelet activity of essential oil was evaluated, using arachidonic acid (AA) and adenosine diphosphate (ADP) as the platelet aggregation inducers. The results showed that essential oil of *Allium atroviolaceum* with IC₅₀; 0.25 mg/ml and 0.47 mg/ml inhibited *in-vitro* platelet aggregation induced by AA and ADP respectively.

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Introduction

The genus *Allium* is a very diverse and taxonomically complicated group belonging to the family Alliaceae (Fritsch and Maroofi). The genus *Allium* comprises of around 750 species according to Stearn (Hirschegger *et al.*, 2010). It is well known that the *Allium* genus is rich of flavonoids, saponins, saponinins and volatile sulfur compounds. The later compounds are responsible for their characteristic pungent aroma and taste, though they are unstable and easily transform to other compounds (Lanzotti, 2006). However, only a few of them have been used for their pungency and flavoring value and in some parts of the world, for religious connotations. Since ancient times, species in genus *Allium* have been used in folklore of many cultures as food, preventive and therapeutic medicinal agents (Fenwick *et al.*, 1985). There has been an increase in awareness and usage of all forms of alternative medical therapies often mentioned as complementary medicine (CAM) (Rahman, 2007).

Allium species are reported to have several positive health effects on immune functions, antibacterial, antifungal, antiviral, anticancer and practically cardiovascular activities. They are known to have direct effect on vessel wall, hypotensive and cholesterol- and triglyceride-lowering properties (Iciek *et al.*, 2009).

Cardiovascular disease is the main reason of morbidity and mortality in the world, particularly in developing countries (21.9% of total death) (Amidi *et al.*, 2013). Cardiovascular disease is a complex and multifactorial disease. Among these factors increased platelet aggregation and thrombus formation plays a significant role in the etiology of cardiovascular disease (Rahman, 2007). Clot formation, decreased or interrupted blood supply to vital organs such as the heart and brain lead to cardiovascular disorders such as myocardial infarction, unstable angina, stroke, venous thromboembolism (Weller *et al.*, 1994).

Platelets are activated by a variety of metabolic pathways. The mechanism of platelet aggregation pathway is very complex and involves multiple components and it can be controlled by

heterogeneous group of endogenous compounds such as ADP, ATP, collagen, thrombin, tryptophan, epinephrine, thromboxane A₂ and calcium. Each can independently and together begin the process leading to platelet aggregation. These compounds on platelets have specific receptors. Their effect on platelet aggregation is applied through binding to these receptors (Steinhubl *et al.*, 2007).

The aim of this study was to analyze and clarify the medicinal constituents of essential oils of *Allium atrovioleaceum* and to determine its *in-vitro* antiplatelet activity, using AA and ADP as the platelet aggregation inducers.

Materials and methods

Plant material

Aerial parts of *Allium atrovioleaceum* (The local name; Sirdeng) were collected in May 2010 in Rig mountain, Lordegan, Shahr-e-kord province, at 2610 m above sea level, A voucher specimen (SBMU-8013) has been deposited in the Herbarium of Department of Pharmacognosy, Faculty of Pharmacy of Shahid beheshti University of medical science, Tehran, Iran.

Isolation of essential oil

Aerial parts were carefully left to dry in controlled temperature (22°C) without exposure to the light and moisture. They were chopped and then passed from sieve size 60 (25/0 mm) and 142g subjected to hydrodistillation for 4 h, using a Clevenger-type apparatus. The oils were dried over anhydrous sodium sulfate and stored at 4°C.

Gas chromatography-Mass spectrometry

The GC-MS analyses were carried out on a Hewlett Packard GC-MS system, model 5973, fitted with a 30m long, cross-linked 5% phenylmethyl siloxane (HP- 5MS 5% Phenyl Methyl Silox, Agilent 19091S-433) (30 m x 250 µm x 0.25 µm). The source temperature was 230°C, the quadrupole temperature 150°C, the initial oven temperature was 60°C; this was then raised to 260°C at 4°C/min and the final temperature maintained for 20 min. The injector and detector temperatures were 200°C and 250°C,

respectively. The carrier gas was helium at 1.0 mL/min. The sample was injected using a split ratio of 1:100. The carrier gas helium, adjusted to a linear velocity of 34 m/s. the ionization energy was 70 eV, and the scan range 40-650 amu at 3.9 scans/s. The injected volume was 1.0 µl of a 2% dilution of oil in n-heptane. The identification of the oil components was based on calculated relative retention time to those of C8-C24 n-alkanes, and compared with values reported in the literature and Wiley MS data library (6th ed).

Blood collection

Blood was obtained from healthy volunteers who did not take any medication for 14 days and were fasting overnight prior to the study. Blood collected at falcon tube containing 0.1 volume of 2.2% sodium citrate. Platelet rich plasma (PRP) was prepared by the centrifugation of citrated blood at 100g for 10 min. The residual blood was centrifuged at speed of 1500 g for 15 min to give platelet poor plasma (PPP). Platelets were counted under microscope and the platelet count was adjusted to $(250 \pm 25) \times 10^9/L$ by diluting the supernatant PRP with PPP.

Platelet aggregation studies

Platelet aggregation responses were monitored with a turbidmetric method using an optical aggregometer. Aliquots of 200 µl of PRP were distributed in the test cuvettes and placed in incubation chamber of AACT-4004 aggregometer (LABiTec, Ahrensburg, Germany), at 37°C. Platelet aggregation was measured using PRP after activation by the addition of ADP or AA according to Born method. The essential oil was dissolved in DMSO (at 0.05% final

concentration) and added to the PRP, 5 min prior to the activation with ADP or AA. The extent of aggregation was quantified by determining the maximum height of the curve. The platelet aggregation inhibitory activity was expressed as percent inhibition by comparison with that measured for the vehicle (DMSO) alone (Amidi *et al.*, 2013).

Statistical analysis

The anti-aggregation value of each compound was expressed as either % inhibition or IC₅₀ values (the concentration of the compound causing 50% inhibitory effects). The IC₅₀ values were determined from the Graph pad Prism version 3.02.

Result and discussion

Essential oil of aerial part (0.7 mL; 0.49%) of *Allium atroviolaceum* analyzed by GC/MS/MS showed the presence of Forty-two components. A list of the identified compounds, along with their percentages of the total oil, Kovats index and retention time is given in Table 1. Forty-two compounds were identified, representing 84.95% (w/w) of the total oil. The two major constituents of the oil samples were trisulfide, di-2-propenyl (26.85%) and diallyl disulphide (10.98%) while trans-2-(2-pentenyl)furan (0.02%) and Limonene (0.06%) were detected in lower amounts. The presence of compounds showed mono-sulfur (5.15%), disulfide (19.38%), tri-sulfur (36.82%) and tetra-sulfur compounds (7.43%). The results indicated that the highest amount of sulfur compounds is related to tri-sulfur compounds. Differences were observed in the sulfur content of the constituents of this plant with other *Allium* species (Lazarevic *et al.*, 2011).

Table 1. Inhibitory effect of quercetin as positive control on *in-vitro* platelet aggregation induced by arachidonic acid (AA) and ADP.

| Compound | AA (1.35mM) | | ADP (5µM) | |
|------------------------------------|-------------|-----------|-----------|-----------|
| | 0.15mg/ml | 0.07mg/ml | 0.15mg/ml | 0.07mg/ml |
| QUERCETIN (Inhibition%) | 36% | 1% | 2% | - |
| QUERCETIN IC ₅₀ (mg/ml) | 0.1 | | - | |

Also, it could be related to differences in the composition of the essential oil from aerial parts of our study with the bulb in the other studies.

Remarkably, the presence of sulfur compounds in the aerial part such as bulb in a significant amount.

Table 2. Effect of essential oil of *A. atroviolaceum* on *in-vitro* platelet aggregation induced by AA and ADP.

| Concentration of essential oil | AA (1.35mM) | | ADP (5µM) | |
|--------------------------------|-------------|--------------|-------------|--------------|
| | %Inhibition | %Aggregation | %Inhibition | %Aggregation |
| 2 mg/ml | 99.85 | 0.11± 1.31 | 98.74 | 0±2.21 |
| 1 mg/ml | 98.35 | 1.28±2.13 | 91.88 | 7.78±1.45 |
| 0.5mg/ml | 97.80 | 1.71±2.35 | 58.95 | 39.32±3.14 |
| 0.33mg/ml | 97.68 | 1.79±3.57 | 3.26 | 92.69±1.17 |
| 0.28mg/ml | 97.43 | 1.98±3.11 | - | - |
| 0.25mg/ml | 37.41 | 48.32±2.61 | - | - |
| 0.2mg/ml | 9.05 | 70.22±1.78 | - | - |
| Solvent | - | 77.84±4.7 | - | 95.82±4.8 |
| IC ₅₀ (mg/ml) | 0.25 | | 0.47 | |

Table 3. Chemical composition of essential oil of *A. atroviolaceum*.

| Peak No. | RT | KI | Area% | |
|----------|--------|------|-------|-------------------------------------------------------|
| 1 | 4.094 | 808 | 0.21 | Norbornene,5-methylene-2> |
| 2 | 5.898 | 907 | 0.09 | Heptanal |
| 3 | 6.059 | 914 | 0.15 | 3,4-dimethylthiophene |
| 4 | 6.309 | 924 | 2.06 | Isocitronellene |
| 5 | 6.663 | 938 | 1.92 | Disulfide,methyl 1-propenyl |
| 6 | 7.694 | 979 | 5.69 | Dimethyl trisulfide |
| 7 | 8.153 | 997 | 0.33 | Furan,2-pentyl- |
| 8 | 8.354 | 1004 | 0.11 | Pyrazine, 2-ethy-6-methyl- |
| 9 | 8.427 | 1006 | 0.02 | Trans-2-(2-pentenyl)furan |
| 10 | 8.668 | 1014 | 0.17 | Terpinene<alpha> |
| 11 | 8.966 | 1024 | 0.06 | Limonene |
| 12 | 9.474 | 1040 | 0.13 | Ocimene<-beta-> |
| 13 | 9.715 | 1048 | 0.89 | Terpinene<Gamma-> |
| 14 | 10.754 | 1082 | 0.13 | Pyrazine, 2,6-diethyl- |
| 15 | 10.827 | 1084 | 10.98 | Diallyl disulphide |
| 16 | 11.004 | 1090 | 0.29 | Pyrazine,2-ethyl-3,5-dimethyl- |
| 17 | 11.737 | 1113 | 0.24 | 1,3-dithiane, 2,2-dimethyl- |
| 18 | 11.963 | 1119 | 0.37 | 1,2-dithiolane |
| 19 | 12.341 | 1131 | 0.67 | Disulfide,methyl (methylthio)methyl |
| 20 | 13.146 | 1155 | 3.78 | 2-thiatricyclo[3.3.1.1(3,7)]decane |
| 21 | 14.588 | 1198 | 0.32 | Tetradecane |
| 22 | 15.289 | 1219 | 1.51 | Dimethyl,tetrasulfide |
| 23 | 15.74 | 1233 | 2.22 | 4,6-dimethyl-[1,2,3]trithiane |
| 24 | 17.729 | 1292 | 0.92 | Methane,(methylsulfinyl)methylthio)- |
| 25 | 18.019 | 1302 | 26.85 | Trisulfide,di-2-propenyl |
| 26 | 18.857 | 1328 | 5.57 | 1,2,4-trithiolane,3,5-diethyl- |
| 27 | 20.532 | 1382 | 3.46 | Tetrasulfide, di-2-propenyl |
| 28 | 20.943 | 1395 | 1.23 | Bicyclo[3,2,1]oct-2-ene, exo-4-(phenylthio)- |
| 29 | 22.618 | 1450 | 0.22 | 5,9-undecadien-2-one, 6,10-dimethyl-(Z) |
| 30 | 23.351 | 1474 | 1.19 | 1,1'-thiobis(3-(methylthio)-propane |
| 31 | 23.697 | 1486 | 0.97 | 3-buten-2-one, 4-(2,6,6-trimethyl-1-cyclohexen-1-yl)- |
| 32 | 23.749 | 1487 | 0.25 | 5-methyl-2-phenyl-2hexenal |
| 33 | 24.647 | 1518 | 0.51 | Cyclohexanebutanal,2-methyl-3-oxo-cis |
| 34 | 24.849 | 1525 | 1.10 | 1,2,4-cyclopentanetrione,3-(2-pentenyl)- |
| 35 | 25.324 | 1541 | 2.46 | Tetrasulfide, di-2-propenyl |
| 36 | 25.614 | 1551 | 0.47 | Formic acid,2-methyl-[1,3]dithian-2-ylmethyl ester |
| 37 | 27.732 | 1624 | 2.25 | 1-(2-ethyl-[1,3]dithian-2-yl)-3-methyl-butan-1-ol |
| 38 | 27.966 | 1633 | 0.17 | Eudesmol<beta> |
| 39 | 28.441 | 1650 | 0.54 | 1,2-dithiolane, 1,1-dioxide |
| 40 | 31.123 | 1750 | 0.36 | Tetradecanoic acid |
| 41 | 31.654 | 1771 | 1.78 | 1-(2-ethyl-[1,3]dithian-2-yl)-3-methyl-butan-1-ol |
| 42 | 33.329 | 1838 | 2.34 | 6,10,14-trimethylpentadecan-2-one |

Currently, there is a growing attention both in industry and scientific investigation in spices and aromatic herbs because of their strong antioxidant

and antimicrobial properties. These properties are in line for many substances, including some terpenoids, flavonoids, vitamins, carotenoids, phytoestrogens,

etc. (Bareemizadeh *et al.*, 2014). Moreover *Allium* species are reported to have several effects on immune functions and antibacterial, antifungal, antiviral, anticancer and practically effect on cardiovascular diseases. In view of that we examined the anti-platelet aggregation activity of essential oil of *A. atrovioleaceum* (Table 2, 3).

Essential oil of *A. atrovioleaceum* showed a dose-dependent inhibitory effect against AA and ADP-induced aggregation with IC₅₀ values of 0.25 and 0.47 mg/mL, respectively. Platelet aggregation inhibitory effect of essential oil of *A. atrovioleaceum* is about two times weaker than that of quercetin when ADP is used as aggregation inducer and five times weaker when AA is used as the inducer.

The antiplatelet activity of *A. atrovioleaceum* oils may not be due solely to any individual components but could be due to the synergistic effects of a group of constituents. Further studies need to be carried out to identify anti-platelet aggregation activity of major components of this essential oil and comparing the IC₅₀ values with IC₅₀ values obtained for the total essential oil used in this study. Previous studies have reported that the antiplatelet properties of *Lavandula hybrida* and *Goniothalamus* oil could be due to the synergistic effect of their components (Ballabeni *et al.*, 2004, Moharam *et al.*, 2010).

Plants of the genus *Allium* such as onion and garlic are often consumed as a source of compounds which inhibit human platelet activity, with the goal of decreasing vascular diseases. Antiplatelet activity of these plants is in part due to the concentrations of organosulfur compounds. Goldman *et al.* (1996) demonstrated antiplatelet activity is genotype dependent and correlated with bulb sulfur content (Goldman *et al.*, 1996).

Acknowledgments

The authors would like to acknowledge Dr. Keramat Saidi, Faculty of horticulture, University of Shahrekord for identification of *Allium atrovioleaceum*. This study was a part of PhD thesis of

Zahra Lorigooini, proposed and approved in Faculty of Pharmacy, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

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