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Research Article

Prevalence of Human T-Lymphotropic Virus Types I and II in Patients With Hematological Disorders in Isfahan, Iran

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Background: Human T-cell lymphotropic virus types I and I I (HTLV-I and HTLV-II) are deltaretroviruses which may cause leukemia, lymphoma and HTLV-associated myelopathy/tropical spastic paraparesis (HAM/TSP). In addition, HTLV-1 may be related to thalassemia and hemophilia cases after blood transfusion.

Objectives: The aim of this study was evaluation of the prevalence of HTLVs in patients with hematological disorders (leukemia, thalassemia, lymphoma and hemophilia).

Patients and Methods: This cross-sectional study was conducted during April to October 2012. A total of 101 serum samples were collected from patients and were stored at -20°C. DNA was extracted from serum by an extraction kit. The extracted DNA was tested by polymerase chain reaction (PCR) for detection of HTLV-I and HTLV-II pol and tax gene sequences, respectively. Samples were collected from 67(66.33%), 20 (19.80%), 4 (3.96%), and 10 (9.90%) patients with thalassemia, leukemia, lymphoma and hemophilia, respectively.

Results: One thalassemia sample was HTLV-I positive, but none of the samples contained the genome of HTLV-II. The prevalence of HTLV- I in this study in patients with hematological disorders was 0.99%.

Conclusions: The prevalence of HTLV- I in hematological disorders was similar to that of other parts of Iran. The present study revealed that HTLV-I screening should be performed before blood transfusion to reduce the risk of virus transmission in patients with hematological disorders. More study should be performed to detect these viruses in blood donors.

Keywords: Human T-lymphotropic Virus; Hematological Diseases; Iran

1. Background

Human T-lymphotropic virus type I (HTLV-I) was first identified in humans in 1980 (1) and 1982 (2). A type C retrovirus, HTLV- I, is the causative agent of two distinct human diseases, adult T-cell leukemia (ATL) or lymphoma, and a chronic progressive demyelinating disorder known as HTLV- I -associated myelopathy/tropical spastic paraparesis (HAM/TSP) (3). It is estimated that 10 to 20 million people worldwide are infected with HTLV- I (4). HTLV I and II are transmitted through breastfeeding, sexual contact and blood transfusion. Transplacental transmission is also suspected (5, 6). Whole blood components, platelets and packed red blood cells, but not fresh frozen plasma, are sources of virus transmission (7). The probability of seroconversion in a recipient of contaminated blood is about 44% (8).

Patients with thalassemia need infusion of four to six blood units per month and therefore the risk of bloodborne diseases caused by viruses such as HTLV-I, hepatitis B virus (HBV), hepatitis C virus (HCV), and human immu-

nodeficiency virus (HIV) increase in these patients (9). The association between retroviruses and hematologic malignancies has also been described previously. However. few studies on the association between HTLV- I infection and malignancy risk have been published (10). There are some case reports regarding finding HTLV- I in lymphoid malignancies except ATL (11). It is still controversial whether or not HTLV- I infection affects the incidence of several malignancies. Adedayo et al. found an association between HTLV- I and lymphoid malignancies in the Dominican population (12). Most of HTLV-I-infected individuals remain asymptomatic throughout their lives and in a few subjects HTLV-I-associated diseases will appear.

This infection is endemic in southern Japan, the Caribbean Basin, central Africa, central and south America, the Melanesian Islands in the Pacific Basin, and in the aboriginal population in Australia (13). In Iran, this virus has been found in isolated pockets in which HTLV- I infection is endemic (Khorasan, the northeastern province of Iran).

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The prevalence of HTLV- I infection in Mashhad, Khorasan was 0.77% among seeming healthy blood donors (14). However, we have few data about the prevalence of HTLV- I and HTLV-II in patients with hematological disorders and malignancies in Iran. According to another reports, the prevalence of HTLV-I infection in Mashhad, Neishbour, and Sabzevar was 2.1%, 3% and 1.6%, respectively (15, 16). However, the virus is less frequent in other parts of Iran including Urmia (0.34%) in northwest and Chaharmahal and Bakhtiari (0.62%) in southwest of Iran (17, 18).

2. Objectives

The aim of this study was to determine the prevalence of HTLVs among patients with chronic myelogenous leukemia (CML), acute lymphoblastic leukemia (ALL), acute myelogenous leukemia (AML), lymphoma, hemophilia and thalassemia, and hemophilia. Therefore, this study was conducted to detect HTLV- I and HTLV-II in patients with hematological disorders from Isfahan province, center of Iran.

3. Patients and Methods

3.1. Population Study

In this cross-sectional study, 101 patients with confirmed hematological disorders admitted to the oncology and thalassemia unit of Seyed Al-Shohada hospital, Isfahan, Iran, were enrolled during April to October 2012. Diagnoses of the diseases were confirmed based on previous clinical and laboratory document findings.

3.2. Sampling

Hematological disorders were as follows; leukemia (20 cases), lymphoma (four cases), thalassemia (67 cases) and hemophilia (10 cases). About 5 mL blood was collected from each patient without adding any anticoagulant. Serum was stored at -20°C before testing.

3.3. DNA Extraction

DNA was extracted from each 100 μ L blood serum using high pure extraction kit (CinnaPure cell DNA kit, Cat No. PR881613-50, Iran).

3.4. Polymerase Chain Reaction

PCR amplification was performed with two sets of prim-

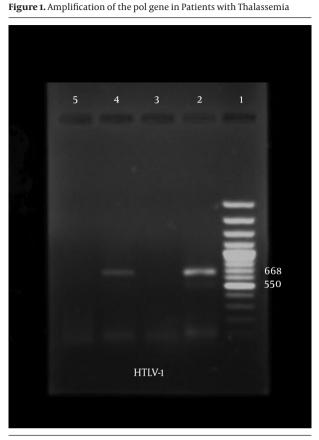
ers, pol (forward, *CCCTACAATCCAACCAGCTCAG*; reverse, *TG-GAGTAACTTACTAGGTTAG*) for HTLV- I (GenBank accession No. J02029); and tax (forward, *CGATTGTGTACAGGCCGATTG*; reverse, *CCTGTACACCAGGCAGTCTGGA*) for HTLV- I I (GenBank accession No. M10060); resulting in amplicons of 668 bp and 628 bp, respectively, according to Dehee et al. (19) guidelines.

The total PCR reaction volume was 25 µL. It contained 5 uL DNA sample, 1 uL of each primer (10 pmol, TAG Copenhagen, Denmark), 0.5 µL Taq DNA polymerase (500 U·TA7506C CinnaGen, Iran), 2.5 µL PCR buffer 10x, 1 µL dNTP mixture (10 mM, DN7603C, CinnaGen, Iran), 1 µL MgCl₂ (50 mM, TP7506C, CinnaGen, Iran) and 13 µL distilled water. For HTLV- I, the reaction mixture was incubated for five minutes at 95°C and then subjected to 40 cycles consisting of 30 seconds at 94°C, 30 seconds at 57°C, and one minute at 72°C, and a final extension for seven minutes at 72°C in a DNA thermal cycler (Corbett, Research, CGI-960, Australia). For HTLV- I I, the reaction mixture was incubated for five minutes at 95°C and then subjected to 40 cycles consisting of 30 seconds at 94°C, 30 seconds at 62°C, and one minute at 72°C, and a final extension for seven minutes at 72°C. Genomic DNA of HTLV- I -infected cell line, TL-Om1, was used as positive control. Distilled water was used as negative control. The PCR products were analyzed by electrophoresis on agarose gel (1%) and stained with ethidium bromide.

4. Results

One hundred and one patients with confirmed hematological disorders were recruited in this study. The mean age of patients was 44.25 years, ranging from 1.5 to 87. Of 101 patients, 67 (66.33%) and 34 (33.66%) were female and male, respectively. According to the type of hematological disorders, the most cases with thalassemia were in the age group of 20 - 40 years (Table 1). Most of the patients with leukemia (20) were in the age group of 1-6 years and in hemophilic cases (10), most of the patients were in the age group of 40 - 60 years. In the lymphoma group (4), all of the four cases were over 12 years. The results showed that only one male (0.99%) was positive for HTLV-I in patients with thalassemia. According to the results of PCR, the overall prevalence of HTLV-I infection in patients with thalassemia in Isfahan was 1.49%. All the samples were negative for HTLV-II (Figure 1).

Age, y	Lymphoma	Hemophilia	Thalassemia	Leukemia
1-6	0	0	3	9
6 - 12	0	0	3	2
12 - 20	1	0	4	0
20-40	1	4	35	3
40-60	1	5	14	3
>60	1	1	8	3
Total	4	10	67	20



A 668 bp fragment was amplified from the pol region of HTLV- I genome. Lane 1, 100 bp marker; lane 2, positive control (TL-Oml); lane 3, negative sample; lane 4, positive sample; lane 5, negative control.

5. Discussion

Approximately 5 - 10% of HTLV-I-infected individuals develop either ATL or HAM/TSP. Some evidences have shown that it has also been associated with other diseases such as cutaneous T cell lymphoma (CTCL), HTLV-I-associated arthropathy (HAAP), Graves' disease, uveitis, polymyositis, chronic respiratory diseases, lymphadenitis, and dermatitis (20). On the other hand, HTLV- I and HTLV-II infections are usually chronic and untreatable diseases; so, adequate standards of diagnosis, prevention, care and support, as well as surveillance should be provided (21). HTLV- I infection is endemic in certain parts of the world (22) as well as in a northern city of Iran, Mashhad (14, 16). In the present study, we tried to determine the prevalence of HTLV-I and HTLV-II infections in patients with hematological disorders in Isfahan province, Iran. Although there is no defined treatment for patients infected with HTLV- I and HTLV-II, the accurate knowledge of the prevalence rates in different populations may be helpful in establishing prophylactic measures to reduce the rates of viral transmission from infected individuals. HTLV- I and HTLV-II are cell associated and spread in cells after blood transfusion, sexual intercourse, or breastfeeding.

Patients with hematological disorders who need repeated blood transfusion are among the most high-risk groups for this infection (23, 24). HTLV- I and HTLV-II, related to thalassemia and hemophilia by blood transfusion, have been reported in previous publications. In Iran, also, there is some evidence suggesting the relatively high prevalence of the virus in these patients. Farias de Carvalho et al. found a seroprevalence of 28.9% among patients with T-cell lymphoid malignancies in Brazil (25). Adedayo and Shehu found a 38.6% HTLV- I seropositivity in all hematological malignancies in India (12). Miyagi et al. found an HTLV-1 prevalence of 26.1% in 88 cases of non-Hodgkin's lymphoma in Japan (26). The prevalence rate of HTLV- I and HTLV-II infections was 18% in southern Chile in patients with malignant hematological diseases, and 27% in patients with chronic lymphoproliferative disorders, as Barrientos et al. reported (27). Monavari et al. reported that the prevalence of HTLV- I among patients with malignant hematological diseases in Tehran was 12% (28).

The information about HTLV- I and HTLV-II prevalence in different populations and patients is crucial, because it may be useful in establishing prophylactic measures to decrease the rates of viral transmissions from infected individuals. Whereas the gold standard method for the diagnosis of HTLV infection is the detection of HTLV genome in specimens of patients, it seems that the prevalence of HTLV- I infection in our study population was 0.99%. PCR is an extremely sensitive technique, but sometimes has some limitations which lead to unexpected results. On the other hand, the presence of antibodies does not affect the results and the few copy number of the target sequence will appear in PCR product. Cross-contamination is the main limitation in PCR. Negative control was the indicator of cross-contamination and the PCR product of negative control did not show any expected amplicon length.

In the present study, we could not find any association between gender, age and occupation and HTLV-I. In conclusion, our study showed that the prevalence of HTLV-I infection in patients with thalassemia in Isfahan was 1.49%, but none of the samples contained the genome of HTLV-II. Therefore, it is necessary to carry out a large epidemiological study in this part of Iran. Although the prevalence of HTLV-I and HTLV-II infection among patients with hematological disorders in Isfahan province compared to other regions of Iran is not too high, HTLV screening should be performed prior to blood transfusion to decline the risk of virus transmission in these patients. Patients with HTLV- I and HTLV-II infection may acquire this infection via blood transfusion, despite all of the precautions for screening the blood supply. Therefore, the present study suggests that serious consideration must be given to prevent HTLV-I and HTLV-II infection via transfusion in hematological disorders. Routine serological screening for HTLV-I and HTLV-II antibody and detection of HTLV- I and HTLV-II genome in blood donors is indicated to permit the deferral of blood product donations by asymptomatic HTLV- I and HTLV-II carriers.

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Authors' Contributions

Mohammadreza Mahzounieh was the supervisor of this research. Mohammadreza Ghorani and Razieh Nikoozad carried out the molecular process. Ali Karimi and Batoul Pourgheysari were the advisors of this project. All the authors read and approved the final manuscript.

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References

- Poiesz BJ, Ruscetti FW, Gazdar AF, Bunn PA, Minna JD, Gallo RC. Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. *Proc Natl Acad Sci U S A*. 1980;77(12):7415–9.
- Kalyanaraman VS, Sarngadharan MG, Poiesz B, Ruscetti FW, Gallo RC. Immunological properties of a type C retrovirus isolated from cultured human T-lymphoma cells and comparison to other mammalian retroviruses. J Virol. 1981;38(3):906–15.
- Matsuoka M, Jeang KT. Human T-cell leukaemia virus type 1 (HTLV-1) infectivity and cellular transformation. *Nat Rev Cancer*. 2007;7(4):270–80.
- Johnson JM, Harrod R, Franchini G. Molecular biology and pathogenesis of the human T-cell leukaemia/lymphotropic virus Type-1 (HTLV-1). Int J Exp Pathol. 2001;82(3):135–47.
- Monplaisir N, Neisson-Vernant C, Bouillot M, Duc-Dodon M, Ugarte E, Valette I, et al. HTLV-I maternal transmission in Martinique, using serology and polymerase chain reaction. *AIDS Res Hum Retroviruses*. 1993;9(9):869–74.
- Murphy EL, Figueroa JP, Gibbs WN, Brathwaite A, Holding-Cobham M, Waters D, et al. Sexual transmission of human T-lymphotropic virus type I (HTLV-I). Ann Intern Med. 1989;111(7):555–60.
- Rafatpanah H, Farid R, Golanbar G, Azad FJ. HTLV-I Infection: virus structure, immune response to the virus and genetic association studies in HTLV-I-infected individuals. *Iran J Allergy Asthma Immunol.* 2006;5(4):153–66.
- Manns A, Wilks RJ, Murphy EL, Haynes G, Figueroa JP, Barnett M, et al. A prospective study of transmission by transfusion of HTLV-I and risk factors associated with seroconversion. *Int J Cancer.* 1992;51(6):886–91.
- Ghaffari J, Kowsarian M, Mahdavi M, Vahid Shahi K, Rafatpanah H, Tafreshian A. Prevalence of HTLV-I Infection in Patients with Thalassemia Major in Mazandaran, North of Iran. Jundishapur J Microbiol. 2012;6(1):57-60.
- 10. Inoue H, Matsushita K, Arima N, Hamada H, Uozumi K, Ozaki A, et al. High prevalence of human T-lymphotropic virus type I car-

riers among patients with myelodysplastic syndrome refractory anemia with excess of blasts (RAEB), RAEB in transformation and acute promyelocytic leukemia. *Leuk Lymphoma*. 2008;**49**(2):315–21.

- Starkebaum G, Loughran TJ, Kalyanaraman VS, Kadin ME, Kidd PG, Singer JW, et al. Serum reactivity to human T-cell leukaemia/ lymphoma virus type I proteins in patients with large granular lymphocytic leukaemia. *Lancet.* 1987;1(8533):596–9.
- Adedayo OA, Shehu SM. Human T-cell lymphotropic virus type 1 (HTLV-1) and lymphoid malignancies in Dominica: a seroprevalence study. *Am J Hematol.* 2004;77(4):336–9.
- Proietti FA, Carneiro-Proietti AB, Catalan-Soares BC, Murphy EL. Global epidemiology of HTLV-I infection and associated diseases. Oncogene. 2005;24(39):6058–68.
- Tarhini M, Kchour G, Zanjani DS, Rafatpanah H, Otrock ZK, Bazarbachi A, et al. Declining tendency of human T-cell leukaemia virus type I carrier rates among blood donors in Mashhad, Iran. *Pathology*. 2009;41(5):498–9.
- Azarpazjhoo M, Hasanpoor K, Ghanbari M, Rezaee SA, Mashkani B, Hedayati Moghaddam M. HTLV-I prevalence in Northeastern Iran, Sabzevar; An epidemiologic based study and phylogenetic analysis. *AIDS Res Hum Retrovir.* 2012;28(9):1095–101.
- Rafarpanah H, Hedayati-Moghaddam MR, Fathimoghadam F, Bidkhori HR, Shamsian SK, Ahmadi S, et al. High prevalence of HTLV-I infection in Mashhad, Northeast Iran: a population-based seroepidemiology survey. J Clin Virol. 2011;52(3):172–6.
- Karimi A, Nafici M, Imani R. Comparison of human T-cell leukemia virus type-1 (HTLV-1) seroprevalence in high risk patients (thalassemia and hemodialysis) and healthy individuals from Charmahal-Bakhtiari Province, Iran. *Kuwait Med J.* 2007;**39**(3):259–61.
- Khameneh ZR, Baradaran M, Sepehrvand N. Survey of the seroprovalence of HTLV I/II in hemodialysis patients and blood donors in Urmia. Saudi J Kidney Dis Transpl. 2008;19(5):838-41.
- Dehée A, Césaire R, Désiré N, Lézin A, Bourdonné O, Béra O, et al. Quantitation of HTLV-I proviral load by a TaqMan real-time PCR assay. J Virol Methods. 2002;102(1):37–51.
- Uchiyama T. Human T cell leukemia virus type I (HTLV-I) and human diseases. Annu Rev Immunol. 1997;15:15–37.
- 21. Dougan S, Payne LJ, Tosswill JH, Davison K, Evans BG. HTLV infection in England and Wales in 2002–results from an enhanced national surveillance system. *Commun Dis Public Health*. 2004;7(3):207-11.
- 22. Payne LJ, Tosswill JH, Taylor GP, Zuckerman M, Simms I. In the shadow of HIV-HTLV infection in England and Wales, 1987-2001. *Commun Dis Public Health.* 2004;7(3):200–6.
- Chiewsilp P, Iamsilp W, Hathirat P. HTLV-I antibody screening in donated blood and thalassemic patients. *J Med Assoc Thai*. 1993;**76** Suppl 2:103–5.
- Covas DT, Boturao Neto E, Zago MA. The frequency of blood-born viral infections in a population of multitransfused Brazilian patients. *Rev Inst Med Trop Sao Paulo*. 1993;35(3):271–3.
- 25. Farias de Carvalho SM, Pombo de Oliveira MS, Thuler LC, Rios M, Coelho RC, Rubim LC, et al. HTLV-I and HTLV-II infections in hematologic disorder patients, cancer patients, and healthy individuals from Rio de Janeiro, Brazil. J Acquir Immune Defic Syndr Hum Retrovirol. 1997;15(3):238–42.
- Miyagi J, Toda T, Uezato H, Ohshima K, Miyakuni T, Takasu N, et al. Detection of Epstein-Barr virus and human T-cell lymphotropic virus type 1 in malignant nodal lymphoma, studied in Okinawa, a subtropical area in Japan. Int J Hematol. 2002;75(1):78–84.
- 27. Barrientos A, Lopez M, Sotomayor C, Pilleux L, Calderon S, Navarrete M, et al. Prevalence of human T-cell lymphotropic virus type 1 and 2 among patients with malignant hematological diseases in south Chile. *J Med Virol.* 2011;**83**(4):745–8.
- 28. Monavari SH, Keyvani H, Salehi Vaziri M, Bokharaei Salim F. Detection of human T-cell lymphotropic virus Type-1 among patients with malignant hematological diseases in Capital of Iran, Tehran. J General Mol Virol. 2011;3(5):67–70.