



**Short note**

<https://doi.org/10.26496/bjz.2017.12>

## Two marine parasitic crustaceans new to the Belgian fauna

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Mortelmans J., Debusschere E., Vranken S., Deneudt K., Hernandez F. & Mees J. 2017. Two marine parasitic crustaceans new to the Belgian fauna. *Belgian Journal of Zoology* 147 (2): 143–153. <https://doi.org/10.26496/bjz.2017.12>

**KEYWORDS.** Marine parasites, *Progebiophilus*, *Caligus*, North Sea.

Marine parasites comprise various groups of animals, including crustaceans (copepods, isopods), ecto- and endoparasitic flukes (platyhelminthes), round worms (nematodes), Bryozoa, bivalves and even Cnidaria [1].

Since many commercial fish species and populations are parasitized (e.g., [2]), with often a severe impact on fitness and health, knowledge on parasites is of considerable ecological and economic importance [3]. In addition, organisms from lower trophic levels that form the base of the food chain, can be subject to parasites (e.g., [4]). In Belgium efforts have been taken to assess patterns and prevalence of parasites on whiting and dab during the years 1996–2011 [5], and to quantify parasites originating from marine dredge spoil disposal sites [6]. Otherwise, parasites have only been reported occasionally or as a side-product of a survey (e.g., [7]).

Two marine parasites; *Progebiophilus euxinicus* (Popov, 1929) (isopoda: Bopyridae) and *Caligus brevicaudatus* A. Scott, 1901 (Siphonostomatoida: Caligidae) are here recorded for the first time in Belgian waters [8]. On a global scale, both species are reported infrequently and often without details on collection date and location. Moreover, the environmental parameters of these species remain hitherto unknown (i.e., depth, temperature, salinity). In this paper, we link the collected specimens to a wide range of parameters (Appendix 1), including pigment and nutrient concentrations of water, and measurements on conductivity, temperature, and depth. These new species records bring the list of known occurrences of parasitic crustaceans for Belgium to 11 species of parasitic Isopoda and 14 species of parasitic Copepoda [9] (Table 1).

The Belgian part of the North Sea (BPNS) covers an area of approximately 3447 km<sup>2</sup> and is part of the Southern Bight of the North Sea. The BPNS is unique in the way it is characterized by sand banks: Flemish banks, Zealand banks, Hinder banks and Coastal banks. Currents in the BPNS are dominated by semi-diurnal tides ranging from 3 m (neap tide) to 4.5m (spring tide) [10]. River plumes of the Scheldt, Rijn, Seine and Maas reduce the salinity in the BPNS [11].

TABLE 1

Species of parasitic Isopoda and Caligidae known from Belgium (based on [7, 9, 31]) with reference to the aphiaID in the World Register of Marine Species [32]. *Caligus rapax* Milne Edwards, 1840 was recorded from the BPNS by [31], but is here omitted based on [33].

Species	AphiaID
<b>Isopoda</b>	
<i>Progebiophilus euxinicus</i> (Popov, 1929)	118233
<i>Pseudione hyndmanni</i> (Bate & Westwood, 1868)	118240
<i>Prodajus ostendensis</i> Gilson, 1909	148638
<i>Pleurocrypta porcellanaelongicornis</i> Hesse, 1876	593521
<i>Nerocila orbignyi</i> (Guérin-Méneville, 1832)	118911
<i>Hemiarthrus abdominalis</i> (Krøyer, 1840)	118217
<i>Gnathia oxyuraea</i> (Lilljeborg, 1855)	118995
<i>Ceratothoa oestroides</i> (Risso, 1816)	118871
<i>Athelges paguri</i> (Rathke, 1843)	118196
<i>Portunion kossmanni</i> (Giard & Bonnier, 1886)	148668
<i>Ione thoracica</i> (Montagu, 1808)	118218
<b>Siphonostomatoida</b>	
<i>Caligus brevicaudatus</i> A. Scott, 1901	135746
<i>Caligus curtus</i> O.F. Müller, 1785	135749
<i>Caligus elongatus</i> von Nordmann, 1832	135754
<i>Caligus diaphanus</i> von Nordmann, 1832	135751
<i>Eudactylina acuta</i> Van Beneden, 1853	135806
<i>Haemobaphes ambiguus</i> Scott T., 1900	135987
<i>Kroyeria lineata</i> Van Beneden, 1853	135834
<i>Tripaphylus musteli</i> (Van Beneden, 1851)	136011
<i>Pseudocharopinus malleus</i> (Rudolphi in Nordmann, 1832)	135876
<i>Lernaeocera minuta</i> (Scott T., 1900)	135995
<i>Lernaeocera lusci</i> (Bassett-Smith, 1896)	135994
<i>Lernaeocera branchialis</i> (Linnaeus, 1767)	135993
<i>Lernaeenicus sprattae</i> (Sowerby, 1806)	135991
<i>Lepeophtheirus pectoralis</i> (Müller O.F., 1776)	135779

### First record of *Progebiophilus euxinicus*

On 26<sup>th</sup> October 2016, at 51.30934 N, 2.852588 E, one fished specimen of the mud-shrimp *Upogebia deltaura* was infested with the parasite *Progebiophilus euxinicus* [8] (Fig. 1). Two individual parasites, an adult male and an adult female with full brood sac, were recovered from the branchial cavity. These specimens are stored in 70% ethanol in the Marine Station Ostend (MSO), Flanders Marine Institute, Ostend, Belgium. All details on this record are available online [8], and accessible through EurOBIS (<http://www.iobis.org/mapper/>) and EMODnet biology portal (<http://www.emodnet-biology.eu/portal/#>).

*Progebiophilus euxinicus* is a highly modified epicaridean isopod, parasitic in the branchial cavity of mud-shrimps *Upogebia stellata* (Montagu, 1808), *U. deltaura* (Leach, 1815) and *U. pusilla* (Petagna, 1792) (all Decapoda: Thalassinidea) (Appendix 2). From the coast of Denmark and the Netherlands, *Upogebia* species are considered amongst the most important food sources for plaice *Pleuronectes*

*platessa* Linnaeus, 1758 [12]. The parasite has been previously found in the branchial cavity of *U. stellata*, taken from the gut of plaice *P. platessa* [13]. Occasionally, *Upogebia* can be seen in large numbers (Ranson, ILVO, pers. comm.).

*Upogebia* has a specific ecology, living permanently in a relatively easy system of passages in the sediment [14]. In this stable environment, *Upogebia* serves as an important host for a variety of parasites, including Bryozoa (*Triticella flava* Dalzell, 1848), Bivalvia (*Lepton squamosum* (Montagu, 1803), *Hemilepton nitidum* (Turton, 1822)) and other isopods (*Gyge branchialis* Cornalia & Panceri, 1861). *Gyge branchialis*, is known to have a strong negative impact on the fitness of mud-shrimps, including their reproductive success [4]. Since *Gyge* and *Progebiophilus* can be found simultaneously on the same mud-shrimp *Upogebia* and both are morphologically very similar [15], it is quite possible that *Progebiophilus* may have similar effects on its host but this has not yet been determined. Larval *Progebiophilus* enter the host at an early life stage of the host, and both develop together. So far, no adult individual parasites, have been found outside of the gill chamber [16, 17].



Figure 1. – Infested *Upogebia deltaura*, the left gill chamber conspicuously swollen by the two parasites inside.

BOURDON [15] distinguishes *Progebiophilus* and *Gyge* as follows: females of *Progebiophilus* have biramous and elongated, pointed pleopods (Fig. 2A–B), while pleopods are uniramous and rounded in females of *Gyge*; the males of *Progebiophilus* have large and distinctive pleopods (Fig. 2A, C), while they are small and indistinct in the males of *Gyge*. All life stages of both *Progebiophilus* and *Gyge* are depicted and described in detail by BOURDON [15]. Males and females of *P. euxinicus* differ markedly in size: the female reaches lengths up to 10–12 mm whilst the male reaches lengths up to 2.75–4.5 mm [15] (Fig. 2A). The female is asymmetrically rounded; with one flat and one markedly protruding body side. The male is elongated and segmented, resembling typical isopods, and often resides within a cavity of the female body.

Both *G. branchialis* and *P. euxinicus* have been reported from the Friese Front, the Netherlands [18] and were to be expected in the BPNS. No other observations in the North Sea have been published or are known to be communicated, so the species is considered a rare occurrence in the North Sea (Fig. 3). Eight of the twelve known records originate from Mediterranean and Atlantic waters (Fig. 3). Due to the limited records in literature it is difficult to assess the temporal distribution. Three records ([13, 18, 19]) include collection dates and indicate *Progebiophilus* can be collected all year round.

### First record of *Caligus brevicaudatus*

On 29<sup>th</sup> August 2013, near 51.43319N, 2.80777E, a tub gurnard (*Chelidonichthys lucerna*) was fished that hosted four female *Caligus brevicaudatus* (Fig. 4) [8]. Two specimens were dissected for species identification: all legs (Fig. 5A–D), mouthparts, antennae and antennules were dissected and studied. Two specimens remain intact and stained. Three specimens are stored in 70% ethanol as part of the LifeWatch sample library in the MSO, Flanders Marine Institute, Ostend, Belgium. All details on this

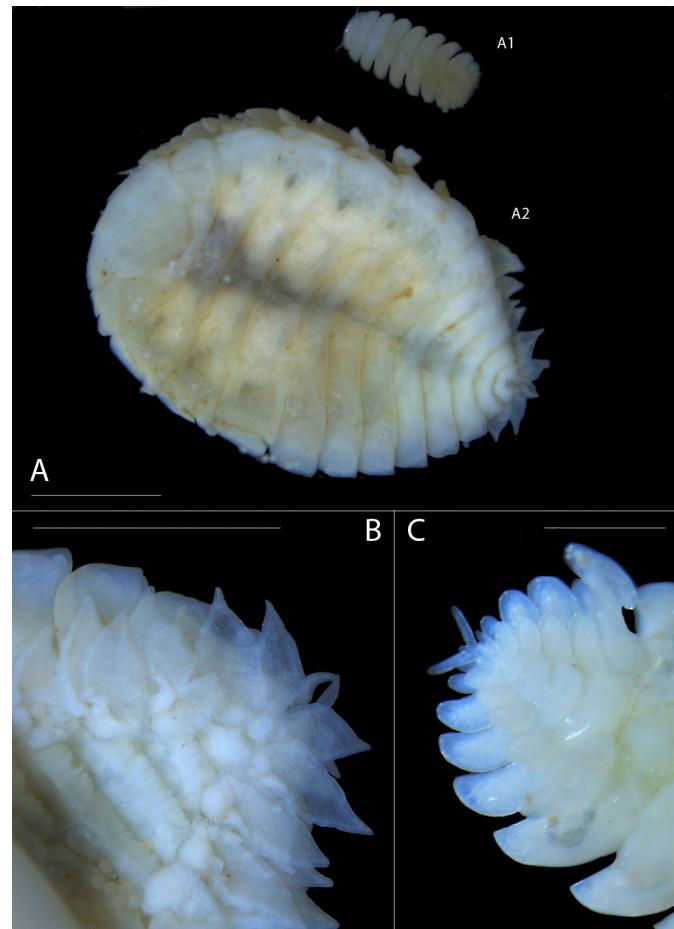


Figure 2. – Male (A1) and female (A2) of *Progebiophilus euxinicus* (Popov, 1927). Female, ventral view of pleopodes and uropods (B). Male, ventral view of pleopodes and uropods (C). Scale bars: A–B = 0.25 mm; C = 0.05 mm.

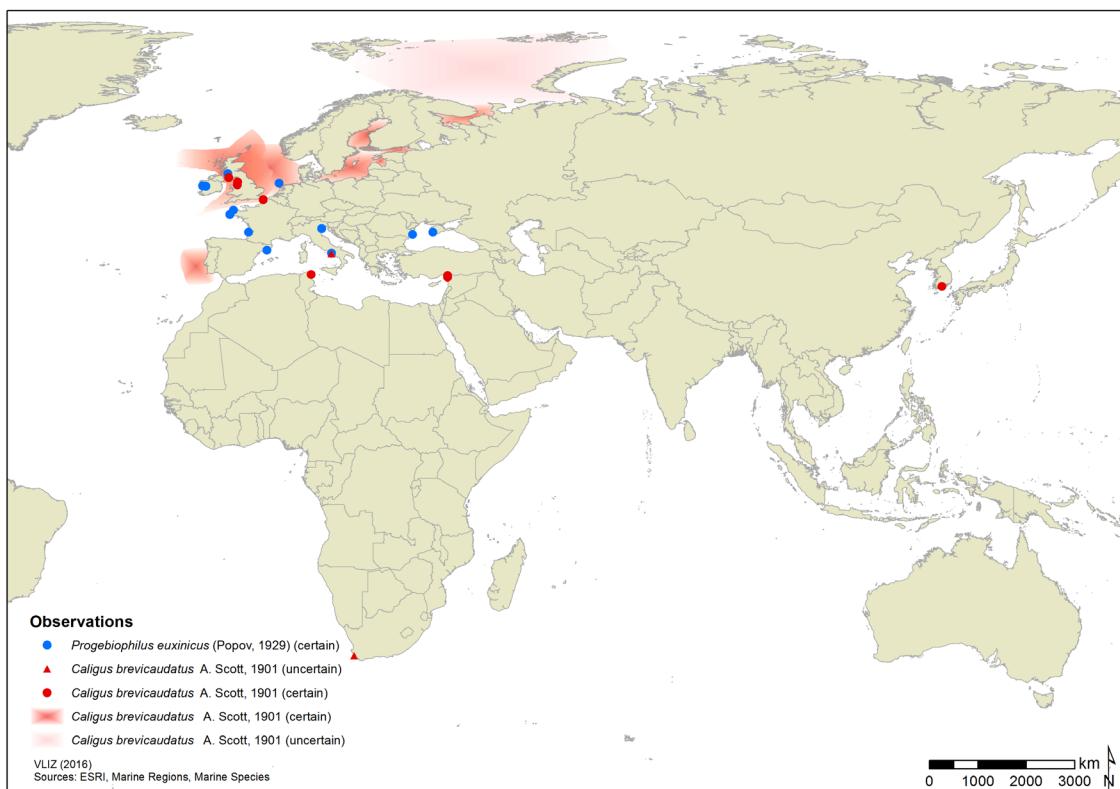


Figure 3. – Known distribution of *Caligus brevicaudatus* (red) and *Progebiophilus euxinicus* (blue), based on [13, 15, 18-20, 24-29]. If no point location is known, the wider sea region is coloured according to MarineRegions [30].

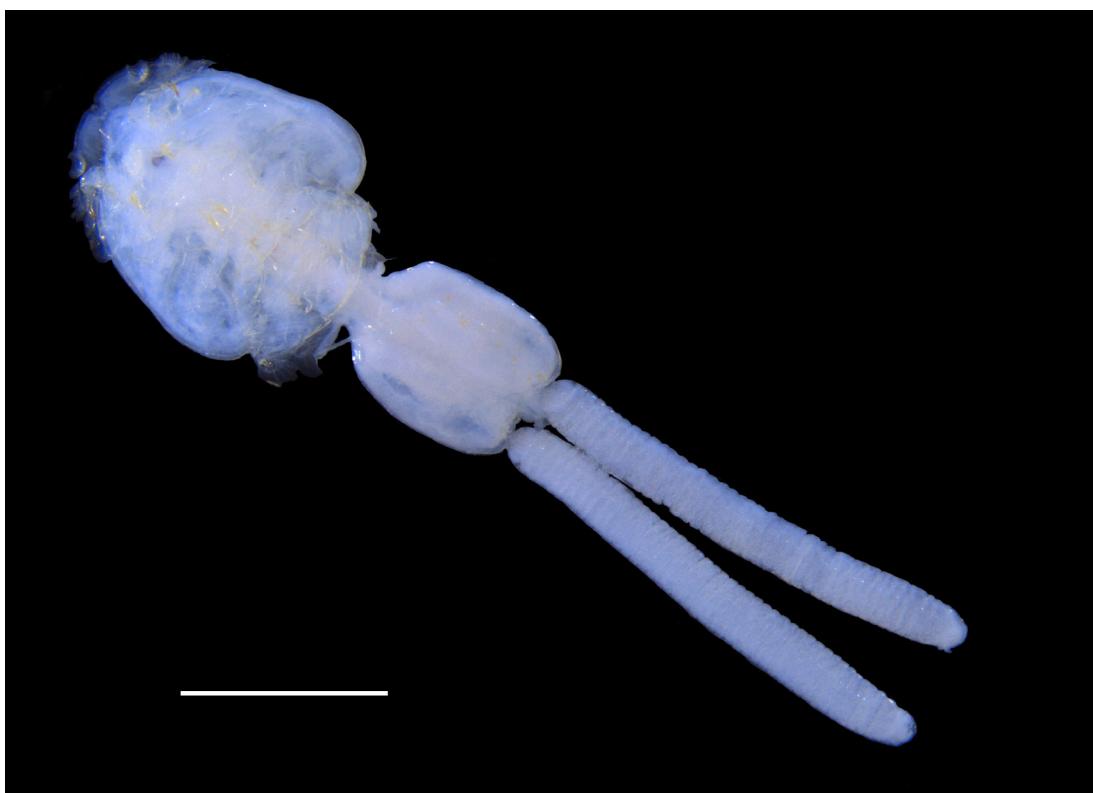


Figure 4. – Female of *Caligus brevicaudatus*, dorsal. Scale bar: 1cm

record are available online [8], and accessible through EurOBIS (<http://www.iobis.org/mapper/>) and EMODnet biology portal (<http://www.emodnet-biology.eu/portal/#>).

*Caligus brevicaudatus* is an ectoparasitic copepod, known to parasitize several species of *Chelidonichthys* and *Eutrigla* (gurnard), including *C. capensis* (Cuvier, 1829), *C. lucerna* (Linnaeus, 1758) and *E. gurnardus* (Linnaeus, 1758) [2, 20]. Besides gurnards, also several flatfish species have been reported as hosts: *Paralichthys olivaceus* (Temminck & Schlegel, 1846), *Pegusa lascaris* (Risso, 1810), *Solea senegalensis* Kaup, 1858, *Solea solea* (Linnaeus, 1758) [21, 22]. An overview on host-parasite relationships is given in Appendix 2. Similar species of ectoparasitic copepods, e.g., *Lepeophtheirus salmonis* (Krøyer, 1837) are known to be able to kill the host [23]. However, no such reports are known for *Caligus brevicaudatus*.

Identification of parasitic copepods is challenging and often involves dissection of legs, antennae and mouthparts. KABATA [2] provides a recent synopsis covering all British species of parasitic copepods on fish enabling researchers to key out the several genera and species around the British Isles. For the BPNS and the wider region of the North Sea, additional literature should be used.

Remarkably, males of *C. brevicaudatus* were only described in 2016 [20]. Females are scarce [2, 20] and were thoroughly redescribed in 2016 [20]. Records are known from Wimereux, France [24] and from the Western Wadden Sea and the Netherlands [25], and therefore, the species was expected to occur in the BPNS. The current distribution of *C. brevicaudatus* is enigmatic and few reliable records exist (Fig. 3). No exact collection dates were found in literature.

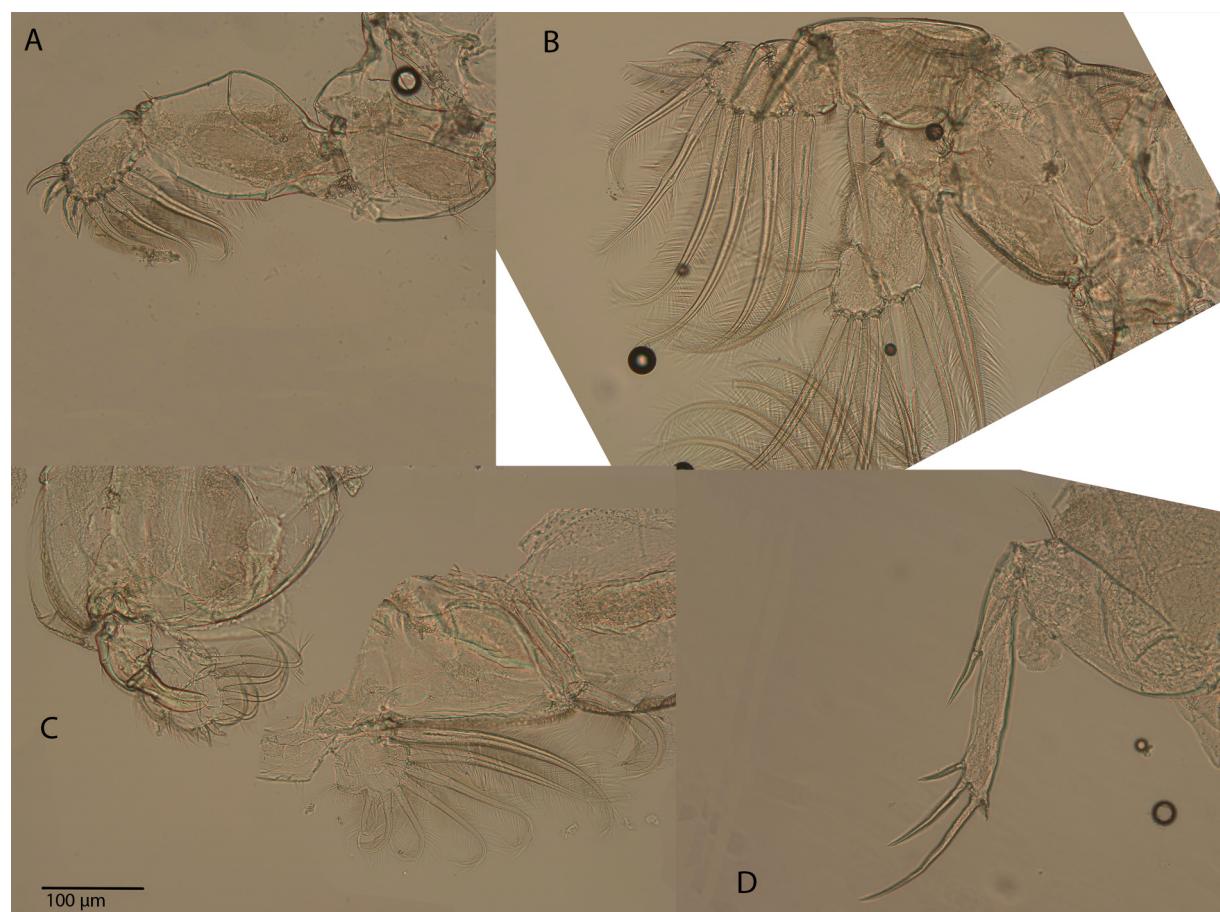


Figure 5. – Female of *Caligus brevicaudatus*. Legs 1 (A), 2 (B), 3 (dissected, C) and 4 (D).

## Acknowledgements

The authors wish to acknowledge Nathalie de Hauwere (VLIZ) for creating the map. Everybody who joined the sampling campaigns is thanked, especially Wim Decock and Dre Cattrijssse. The crew of the RV Simon Stevin and DAB VLOOT are thanked for facilitating campaigns. The research leading to results presented in this publication was carried out with infrastructure funded by LifeWatch (FWO project GOH3417N).

## References

1. HAYWARD P.J. & RYLAND J.S. (1995). *Handbook of the Marine Fauna of North-West Europe*. Oxford University Press, Oxford.
2. KABATA Z. (2003). *Copepods Parasitic on Fishes*. Synopses of the British Fauna 47. Backhuys, Oegstgeest-Leiden.
3. BARBER I., HOARE D. & KRAUSE J. (2000). Effects of parasites on fish behaviour: A review and evolutionary perspective. *Reviews in Fish Biology and Fisheries* 10: 131–165. <https://doi.org/10.1023/A:1016658224470>
4. PASCAL L., DE MONTAUDOUIN X., GRÉMARE A. & MAIRE O. (2016). Dynamics of the *Upogebia pusilla-Gyge branchialis* marine host-parasite system. *Marine Biology* 163 (9): 163–195. <https://doi.org/10.1007/s00227-016-2969-9>
5. DEVRIESE L., BEKAERT K., BOSSAER M. & ROBBENS J. (2012). Patterns and prevalence of marine fish diseases and parasites. In: MEES J. *et al.* (ed.) *Book of Abstracts - VLIZ Young Marine Scientists' Day*: 30. Brugge, Belgium, 24 February 2012.
6. BEKAERT K., HOFFMAN S., PARMENTIER K. (2007). Fish diseases and parasites on the Belgian Continental Shelf. In: MEES J. *et al.* (ed.) *Book of Abstracts - VLIZ Young Marine Scientists' Day*: 20. Brugge, Belgium, 2 March 2007.
7. BIO-ENVIRONMENTAL RESEARCH GROUP, INSTITUTE OF AGRICULTURAL AND FISHERIES RESEARCH (ILVO) (2015). *Zooplankton Monitoring in the Belgian Part of the North Sea Between 2009 and 2010*. <https://doi.org/10.14284/55>
8. FLANDERS MARINE INSTITUTE (VLIZ) (2016). *LifeWatch Observatory Data: Reference Collection of Unique Observations in the Belgian Part of the North Sea*. Vlaams Instituut voor de Zee, Oostende. <https://doi.org/10.14284/267>
9. VANDEPITTE L., DECOCK W. & MEES J. (eds) (2010). *The Belgian Register of Marine Species, Compiled and Validated by the VLIZ Belgian Marine Species Consortium*. VLIZ Special Publication 46. Vlaams Instituut voor de Zee (VLIZ), Oostende, 78 pp.
10. LESCRAUWAET A.-K., PIRLET H., VERLEYE T., MEES J. & HERMAN R. (2013). *Compendium voor Kust en Zee 2013: een geïntegreerd kennisdocument over de socio-economische, ecologische en institutionele aspecten van de kust en zee in Vlaanderen en België*. Vlaams Instituut voor de Zee (VLIZ), Oostende.
11. LACROIX G., RUDDICK K., OZER J. & LANCELOT C. (2004). Modelling the impact of the Scheldt and Rhine/Meuse plumes on the salinity distribution in Belgian waters (southern North Sea). *Journal of Sea Research* 3: 149–163. <https://doi.org/10.1016/j.seares.2004.01.003>
12. RIJNSDORP A.D. & VINGERHOED B. (2001). Feeding of plaice *Pleuronectes platessa* L. and sole *Solea solea* (L.) in relation to the effects of bottom trawling. *Journal of Sea Research* 45: 219–229. [https://doi.org/10.1016/S1385-1101\(01\)00047-8](https://doi.org/10.1016/S1385-1101(01)00047-8)
13. MCGRATH D. & ATKINS P. (1979). Some parasitic Isopoda (Epicaridea) from the Galway Bay area, west coast of Ireland. *Irish Naturalists Journal* 19: 437–439.

14. TEMPELMAN D., VAN MOORSEL G. & FAASSE M. (2013). De molkreeften *Upogebia deltaura* en *U. stellata* en geassocieerde soorten in de Noordzee (Decapoda: Upogebiidae). *Nederlandse Faunistische Mededelingen* 41: 15–30.
15. BOURDON R. (1968). *Les Bopyridae des mers européennes*. Mémoires du Muséum national d'Histoire naturelle 50: 75–424. Muséum national d'Histoire naturelle, Paris.
16. TUCKER B. (1930). On the effects of an Epicaridan Parasite, *Gyge branchialis*, on *Upogebia littoralis*. *Journal of Cell Science* 74: 1–118.
17. PIKE R. (1953). The bopyrid parasites of the anomura from British and Irish waters. *Zoological Journal of the Linnean Society* 42: 219–237. <https://doi.org/10.1111/j.1096-3642.1953.tb02540.x>
18. HUWAE P. (2001). Eerste vondsten van drie parasitaire piszebedden (Isopoda: Epicaridea) en een krabbenzakje (Cirripedia: Rhizocephala) voor het Nederlandse deel van de Noordzee. *Het Zeepaard* 61 (6): 191–203.
19. ASTALL C.M., TAYLOR A.C. & ATKINSON R.J.A. (1996). Notes on some branchial isopods parasitic on upogebiid mud-shrimps (Decapoda: Thalassinidea). *Journal of the Marine Biological Association of the United Kingdom* 76: 821–824. <https://doi.org/10.1017/S0025315400031489>
20. DEMIRKALE I., ÖZAK A.A. & BOXSHALL G.A. (2015). The discovery of the male *Caligus brevicaudatus* Scott, 1901 (Copepoda: Caligidae) parasitic on tub gurnard, *Chelidonichthys lucerna* from the eastern Mediterranean. *Folia Parasitologica* 62 (54): 1–9. <https://doi.org/10.14411/fp.2015.054>
21. ÖZAK A.A., DEMIRKALE I., BOXSHALL G.A. & ETYEMEZ M. (2013). Parasitic copepods of the common sole, *Solea solea* (L.), from the Eastern Mediterranean coast of Turkey. *Systematic Parasitology* 86: 173–185. <https://doi.org/10.1007/s11230-013-9441-8>
22. FERREIRA MARQUES J., SANTOS M.J. & NOGUEIRA CABRAL H. (2009). Zoogeographical patterns of flatfish (Pleuronectiformes) parasites in the Northeast Atlantic and the importance of the Portuguese coast as a transitional area. *Scientia Marina* 73: 461–471.
23. TULLY O. & NOLAN D.T. (2002). A review of the population biology and host-parasite interactions of the sea louse *Lepeophtheirus salmonis* (Copepoda: Caligidae). *Parasitology* 124 (7): 165–182. <https://doi.org/10.1017/S0031182002001889>
24. FLANDERS MARINE INSTITUTE (VLIZ) (2004). *Taxonomic Information System for the Belgian Coastal Area*. Available from <http://www.vliz.be/en/imis?module=dataset&dasid=82> [accessed 27 October 2017].
25. KOCH W., BOER P., WITTE J.I.J., VAN DER VEER H.W. & THIELTGES D.W. (2014). Inventory and comparison of abundance of parasitic copepods on fish hosts in the western Wadden Sea (North Sea) between 1968 and 2010. *Journal of the Marine Biological Association of the United Kingdom* 94: 547–555. <https://doi.org/10.1017/S0025315413001677>
26. CORNALIA E. & PANCERI P. (1858). Osservazioni zoologico-anatomiche sopra un nuovo genere di Crostacei Isopodi sedentarii (*Gyge branchialis*). *Memorie della Reale accademia delle scienze di Torino* 2 19: 85–118.
27. SCOTT A. (1901). Some additions to the fauna of Liverpool Bay, collected May 1<sup>th</sup> 1900 to April 30<sup>th</sup> 1901. *Proceedings and Transactions of the Liverpool Biological Society* 15: 342–353.
28. SCOTT T. & SCOTT A. (1913). *The British Parasitic Copepoda. Volumes I and II*. The Ray Society. London.
29. POPOV V.K. (1929). Rhizocephala and Bopyridae of the bay of Sevastopol. *Trudy Sevastopol'skoi Biologicheskoi Stantsii* 1: 1–26.

30. CLAUS S., DE HAUWERE N., VANHOORNE B., SOUZA DIAS F., OSET GARCÍA P., HERNANDEZ F. & MEES J. (2017). MarineRegions.org. Available from <http://www.marineregions.org> [accessed 27 October 2017].
31. SCHUURMANS STEKHOVEN J.H. (1935). Copepoda Parasitica from the Belgian coast. *Bulletin du Musée royal d'Histoire naturelle de Belgique/Mededelingen van het Koninklijk Natuurhistorisch Museum van België* 11 (7): 1–20.
32. WoRMS Editorial Board (2017). World Register of Marine Species. Vlaams Instituut voor de Zee (VLIZ), Oostende. <https://doi.org/10.14284/170>
33. PARKER R.R. & MARGOLIS L. (1967). A redescription of the syntypes of *Caligus rapax* Milne Edwards, 1840 (Copepoda, Caligidae) and the misuse of this name since 1850. *Crustaceana* 12 (1): 87–101.
34. CARAYON J. (1943). Sur les Epicarides du bassin d'Arcachon (2ième Note). *Bulletin de la Société zoologique de France* 68: 43–48
35. CODREANU R. & CODREANU F. (1963). Sur plusieurs Bopyriens parasites branchiaux des Anomoures de la Mer Noire, de la Méditerranée et du Viêt-Nam. *Rapport de la Commission Internationale pour l'Exploration Scientifique de la Méditerranée* 17: 283–285.
36. BOURDON R. (1965). *Inventaire de la Fauna Marine de Roscoff; décapodes-stomatopodes*. Editions de la station biologique de Roscoff, Roscoff.
37. FLANDERS MARINE INSTITUTE (VLIZ) (2017). *LifeWatch observatory data: nutrient, pigment, suspended matter and secchi measurements in the Belgian Part of the North Sea*. Vlaams Instituut voor de Zee, Oostende. <https://doi.org/10.14284/301>

*Manuscript received: 1 February 2017*

*Manuscript accepted: 11 September 2017*

*Published on: 20 November 2017*

*Branch editor: Steven Degraer*

## Appendix 1

Metadata associated with the sample [37].

<b>Parameter</b>	<b>Value (26/10/2016)</b>	<b>Value (29/8/2013)</b>	<b>Unit</b>
Secchi > Secchi Depth	80	440	cm
Pigments > Chlorophyll c3	0	0	µg/l
Pigments > Chlorophyll c2	0.226	0.500	µg/l
Pigments > Chlorophyllide a	0	0	µg/l
Pigments > Pheophorbide a	0	0	µg/l
Pigments > Peridinin	0	0.196	µg/l
Pigments > 19But-Fucoxanthin	0	0	µg/l
Pigments > Fucoxanthin	1.151	2.286	µg/l
Pigments > Neoxanthin	0	0	µg/l
Pigments > Prasinoxanthin	0	0	µg/l
Pigments > 19Hex-Fucoxanthin	0	0	µg/l
Pigments > Diadinoxanthin	0.099	0.285	µg/l
Pigments > Antheraxanthin	0	0	µg/l
Pigments > Alloxanthin	0.077	0.148	µg/l
Pigments > Diatoxanthin	0	0.018	µg/l
Pigments > Zeaxanthin	0.013	0	µg/l
Pigments > Chlorophyll b	0.057	0	µg/l
Pigments > Chlorophyll a	2.841	5.468	µg/l
Pigments > Pheophytin a	0	0.040	µg/l
Pigments > Beta_Carotene	0.065	0.135	µg/l
Nutrients > Ammonium NH4	3.750	24.478	µmol N_NH4/L
Nutrients > Nitrite NO2	1.340	0.143	µmol N_NO2/L
Nutrients > Nitrate NO3	13.170	?	µmol N_NO3/L
Nutrients > Nitrate Nitrite	14.510	3.242	µmol N_NO3-NO2/L
Nutrients > Phosphate PO4	1.040	0.404	µmol P_PO4/L
Nutrients > Silicate SiO4	13.950	2.793	µmol Si_SiO4/L
SPM > SPM	30.000	2.100	mg/l
CTD > Pressure	3.106	3.026	db
CTD > Temperature	13.080	19.677	°C
CTD > Conductivity	38217.000	45591.000	mS/cm
OBS 3+	8.945	3.292	NTU
Oxygen	7.660	6.239	mg/L
CTD > Density	1024.318	1023.605	kg/m³
CTD > Salinity	32.322	33.355	PSU
CTD > Sound Velocity	1497.340	1518.780	m/s
MaximumDepth			m

## Appendix 2

Host relationships of *P. euxinicus* and *C. brevicaudatus*.

	<b>Host</b>	<b>Reference</b>
<i>P. euxinicus</i>	<i>Upogebia stellata</i> (Montagu, 1808)	[19] (from left branchial chamber), [13, 34]
	<i>Upogebia pusilla</i> (Petagna, 1792)	[19] (from left branchial chamber), [13, 34, 35]
	<i>Upogebia deltaura</i> (Leach, 1816)	[18] (from right branchial chamber), [15, 36]
<i>C. brevicaudatus</i>	<i>Eutrigla gurnardus</i> (Linnaeus, 1758)	[20, 27]
	<i>Chelidonichthys lucerna</i> (Linnaeus, 1758)	[20, 25]
	<i>Paralichthys olivaceus</i> (Temminck & Schlegel, 1846)	[21]
	<i>Pegusa lascaris</i> (Risso, 1810)	[23]
	<i>Solea senegalensis</i> Kaup, 1858	[22]
	<i>Solea solea</i> (Linnaeus, 1758)	[22]
	<i>Chelidonichthys capensis</i> (Cuvier, 1829)	[20]
	<i>Chelidonichthys obscurus</i> (Walbaum, 1792)	[20]