

## CLONING, OVEREXPRESSION AND BIOCATALYTIC EXPLORATION OF A NOVEL BAEYER-VILLIGER MONOOXYGENASE FROM *ASPERGILLUS FUMIGATUS* AF293

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### Electronic Supplementary Material

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1. PCR protocols: Primers & Vectors

**Table S1**

Cloned genes, expression vectors and primers used for genomic DNA amplification. Restriction sites are underlined

<b>Gene</b>	<b>Size</b>	<b>Vector</b>	<b>Oligonucleotides 5'→3'</b>
<i>Af1</i>	2718	pET200	Fw: CACCATGACCAGAATACGTCCAGAC Rev_EcoRI: <u>CGGAATTCTCAATGCC</u> CATTAGTAGAACGG
<i>Af1 truncated</i>	1614	pCRE2	Fw: ACTCGAGATCTGCAGCTGGTATGACCAGAATACGTCCAG Rev: GTTCGGGCCAAGCTTAACTGTAAAGCTCATA
<i>Af2</i>	1461	pET200	Fw_Nde I: CACCC <u>ATATGG</u> ATTACGATATTATCATTGTTGG Rev_Hind III: GCG <u>AAAGCTT</u> CTATTGCTGCTTCCAGCC
		pCRE2	Fw: ACTCGAGATCTGCAGCTGGTATGGATTACGATATTATCA Rev: GTTCGGGCCAAGCTTATTGCTGCTTCCAG
<i>Af3</i>	1806	pET200	Fw_Nde I: CACCC <u>ATATGC</u> GTTGCATACCCTGCC Rev_Eco RI: GCG <u>GAATTCTT</u> TAGAAGCGGCCGCGGC
<i>Af3 truncated</i>	1632	pCRE2	Fw: ACTCGAGATCTGCAGCTGGTATGTCAGAACACTACCTCG Rev: GTTCGGGCCAAGCTTATAGAAGCGGCCGCGGC

2. Expression conditions

**Table S2**

All the expression conditions assayed are listed below

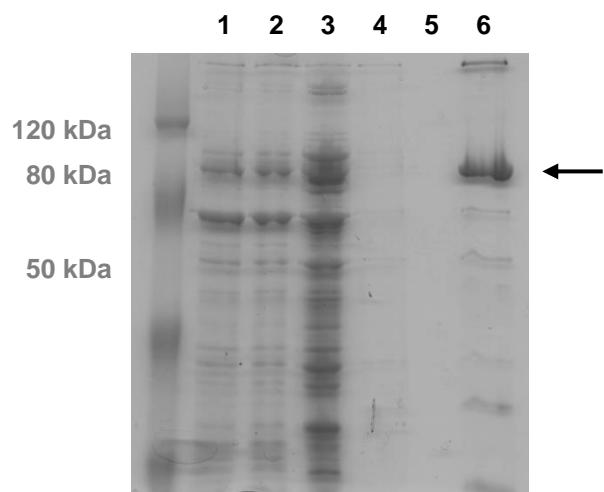
<b>Vector</b>	<b>Cells</b>	<b>Growing T<sup>a</sup></b>	<b>Inductor</b>	<b>Induction T</b>	<b>Induction time</b>
pET-Af1 pET-Af2 pET-Af3	BL21	37 °C (OD <sub>600</sub> =0.5)	IPTG 0.5 mM	17°C	6 h
				24°C	
				30°C	
	BL21	37 °C (OD <sub>600</sub> =0.5)	IPTG 1 mM	37°C	
pET-Af1 pET-Af2 pET-Af3				17°C	6 h
				24°C	
	TOP 10	-	Arabinose 0.002% w/v	30°C	48 h
pCRE-Af1 pCRE-Af2 pCRE-Af3				37°C	
				17°C	36 h
				24°C	
				30°C	
	TOP 10	-	Arabinose 0.02% w/v	37°C	24 h
pCRE-Af1 pCRE-Af2 pCRE-Af3				17°C	
				24°C	
				30°C	
				37°C	12 h
	TOP 10	-	Arabinose 0.2% w/v	17°C	48 h
pCRE-Af1 pCRE-Af2 pCRE-Af3				24°C	
				30°C	
				37°C	
				17°C	12 h

<sup>a</sup> For pCRE2 based constructs, no pre-culture is needed, growing is directly in the presence of inducer

3. Expression of BVMO<sub>Af1</sub>

**Figure S1**

SDS-PAGE analyses for BVMO<sub>Af1</sub> purification fractions. *lane 1*: crude extract, *lane 2*: flow through, *lanes 3, 4, and 5*: sequential washes with imidazole increasing concentrations, *lane 6*: yellow, active fraction. The arrow refers to purified BVMO<sub>Af1</sub>



4. Substrates screening

**Table S3**

BVMO<sub>Af1</sub> substrates profile

<b>Entry</b>	<b>Substrate</b>	<b>BVMO<sub>Af1</sub><sup>a</sup></b>
1	2-propanone	
2	2-butanone	
3	3-buten-2-one	
4	2-octanone	
5	3-octanone	
6	4-octanone	
7	2-decanone	
8	butyl levulinate	
9	3-methyl-2,4-pentanedione	
10	cyclobutanone	
11	cyclopentanone	
12	cyclohexanone	
13	cyclopentadecanone	
14	2-oxocyclohexanecarbonitrile	++
15	4-methyl cyclohexanone	
16	2-propyl cyclohexanone	
17	dehydrocarvone	
18	cyclopropyl methyl ketone	
19	norcamphor	
20	bicyclo[3.2.0]hept-2-en-6-one	+++
21	progesterone	
22	androstenedione	
23	4-dimethylamino benzaldehyde	
24	nicotine	
25	thioanisole	+

26	benzyl ethyl sulfide	+++
27	benzyl phenyl sulfide	+
28	ethionamide	+
29	diphenylmethylthioacetamide	
30	thiacetazone	
31	indole	
32	3-acetyl indole	
33	5-methyl furfural	
34	benzaldehyde	
35	acetophenone	
36	4-hydroxyacetophenone	
37	2,6-dihydroxy acetophenone	
38	3-phenylpentane-2,4-dione	+++
39	phenylacetone	
40	4-(4-hydroxyphenyl)-2-butanone	
41	2-phenyl cyclohexanone	
42	benzoin	
43	phenindione	
44	2-indanone	
45	1-indanone	

<sup>a</sup> Activity was measured employing a colorimetric (phosphate- based detection) screening assay, previously reported for BVMOs substrate screening (Riebel et al. 2012). The activity is indicated as +, ++ or +++ representing 1.2-, 2- or 5-fold phosphate formation (substrate conversion) respectively, when comparing with the blanks

## 5. GC analyses

The following columns were used for the determination of conversions and enantiomeric excesses: Column A: Alltech GT-A (30 m x 0.25 mm x 0.25 µm, 12.2 psi N<sub>2</sub>); column B: Hewlett Packard HP-1 (30m x 0.32 mm x 0.25µm, 12.2 psi N<sub>2</sub>) and C: Chirasil Dex CB (30 m x 0.25 mm x 0.25 µm, 12 psi N<sub>2</sub>). For all the analyses, the injector temperature was 200°C and the FID temperature was 250°C

**Table S4**

GC employed conditions and retention times (*t<sub>R</sub>*) of substrates and products

<b>compound</b>	<b>program<sup>a</sup></b>	<b>column</b>	<b><i>t<sub>R</sub></i> (min)</b>	<b>substrates</b>	<b><i>t<sub>R</sub></i> (min) products</b>
<b>1</b>	130°C isotherm	C			16.5 Abnormal (1 <i>R</i> ,5 <i>S</i> )
				9 (1 <i>S</i> ,5 <i>R</i> )	16.9 Normal (1 <i>R</i> ,5 <i>S</i> )
				9.2 (1 <i>R</i> ,5 <i>S</i> )	17.2 Abnormal (1 <i>S</i> ,5 <i>R</i> )
<b>2</b>	70/0/5/200/0	B	9.63		17.4 Normal (1 <i>S</i> ,5 <i>R</i> )
					13.4 sulfoxide
<b>3</b>	100/0/10/160/8	A	6.9		14.2 sulfone
					13.5 ( <i>R</i> )
<b>3</b>	70/5/5/200/5	B	12.5		14.9 ( <i>S</i> )
					16.5 sulfoxide
					16.9 sulfone
					14.4 ( <i>R</i> )
<b>3</b>	40/0/10/160/8	A	8.8		15.5 ( <i>S</i> )

<sup>a</sup> Program: initial T (°C)/ time (min)/ slope (°C/min)/T (°C)/ time (min)/ slope (°C/min)/T (°C)/ time (min).

Bicyclo[3.2.0]6hepten-2-one (**1**), thioanisol (**2**), benzyl ethyl sulfide (**3**)

## 6. *Aspergillus fumigatus* Af293 BVMO encoding genes

**Figure S2**

Multiple sequence alignment of *A. fumigatus* Af293 BVMOs sequences. Sequences are: BVMO<sub>Af1</sub> (XP\_747160), BVMO<sub>Af2</sub> (XP\_746949), BVMO<sub>Af3</sub> (XP\_755274), XP\_751302, XP\_747774, XP\_754119, XP\_752204, XP\_756084 from *A. fumigatus* Af293, PAMO (YP\_289549) from *Thermobifida fusca*, CHMO (AAG10021) from *Acinetobacter sp.*, and CAMO (AET80001.1) from *Cylindrocarpon radicicola*. The two Rossmann folds (GxGxxG) and the BVMO fingerprint (FxGxxHxxWP/D) are in bolds

	61		120
BVMOAF1_Af293	(5) RPDYDAIVIGAGFSGVRSLWEI R---RLGLTARCFDAGS DVGGTWWNRYPGCRTDGEA		
BVMOAF2_Af293	(1) -MDYDIIIVCAGISGINAAAYRIQS--QLPSHRYAILEARNAIIGGTWDLFKYPGIRSDSDL		
BVMOAF3_Af293	(61) EHYLDALVVCTGFSGLYALQSLL---KLNLKVKAIADAAS DVGGTWWSRYPGAMSDSWS		
PAMO	(14) PEEVDVLVVAGAGFSGLYALYRLR---ELGRSVHVIETAGDVGGVWYWNRTYPGAMSDTET		
CAMO	(7) VLNVDALVVAGAGVAGIYSTYRLS---RAGLNVCICIDTAGDVGGTWWNTYPGAMSDTET		
CHMO	(4) KMDFDAIVIGCGFGGLYAVKKLRD---ELELKVKQAFDKATDVAGTWYWNRTYPGALSDTET		
XP_751302	(51) LPKKRVAVIGAGCLGVSSAAHCIG---HGFDVQLFEARP-----KDKGLGGIWSV		
XP_747774	(32) ARPLRVVIIAGSCISGIIASIRFRQR--IPNVDLCVYEKNEDIGGTLLENRYPGCACDIPA		
XP_754119	(54) VRQIKVGVIGAGCISGITAGVLLPAK--LPGLDLRILYDKNA DVGGTWFENTYPGVRCIPA		
XP_751255	(29) PRKLRVVCIGAGFSGLILAYKLKHERPIDFVDTIYEKNPEVGGTWYENVYPGVGCDIPA		
XP_752204	(19) HTYSPVLVIGAGVSGIATGCRIKEA-LGFDQFRIFERQSGIGGTTWINRYPGAACDVA		
XP_756084	(37) FTRATAVIIAGAGISGICTAIDLKR-SKCHNFIILEKSSGVGGTWNNDNRYPGSCCDVWS		
	<b>GXGXXG</b>		
	121		180
BVMOAF1_Af293	(61) WVYALKELPELLEEWDFTERYPPQEEIQWYLSRVLDYRDIRKDIEFNTEVKSAHYSDHDS		
BVMOAF2_Af293	(58) FTFGFSWNFWN----QDTPIAEGASISKYMRDTAAQYGIDKHIIHFQHRLLAADWSSADN		
BVMOAF3_Af293	(117) HLYRYSFDYE---YPLYRRYVSQPEILAYLIRHVEKYDLRGHMQFNTDMTSAVWDEGTS		
PAMO	(70) IEYCYSFSEEVLQEWNWTERYASQPEILRYINFVADKFDLRSGITFHHTVTAAAFDEATN		
CAMO	(63) YLYRYSWDKEDLRSYPWSNHVYQPEILQYLRHVVERHDLRKYIRFDTEMQKAVWDDQRS		
CHMO	(61) HLYCYSWWDKEILLQSLEIKKKYVQGPDRVRYLQQVAEKHDILKESYQFNTAVQSAHYNEADA		
XP_751302	(98) HSIMYRFHES---VKYTATYPSQOEIRDQIIDVWKRYGLOKRTAFDTPTSVVKQTK-DG		
XP_747774	(90) HTYQATEEPN---KEWSTFYAAAPEIHAZYWKRAEKYGCMLYIKLKQAVVEAVWDDSKS		
XP_754119	(112) HVYQSGEEPN---TQWTEEFAGQGHEIREYWQRVARKYDVYKYLRLPRQKVQKVVWRPEEA		
XP_751255	(89) HSYVFPFEEPN---PNWSKFTVSGPEIQDYIVKTTDKYGLRDKITFNTKLLQVAWDEGDG		
XP_752204	(77) LLYSFSEAPK---KDWTTLHPPGPEIVQYLADEVCEKYQIVDKIQLNTAVKEMRWLEDAE		
XP_756084	(95) SLYSYSFEEQK---ADWTREYPGQEEILDYLGIVAQKYGLYRYIRFNSAVEEARWDEADL		
	241		300
BVMOAF1_Af293	(151) PFPGLQSFKCEVYQTSTWPAHEIEFENKRIGVIGTGSTGIQVITKIALP---VAEQLIVFQ		
BVMOAF2_Af293	(148) DVPGLISQFQGQVIHPQFWPQD-LDYTDKKVVIIGSCATAVTLIPKMAE---KAAKVTMLO		
BVMOAF3_Af293	(203) DLPGIQTFRGEIRHTSAWDTD-LDLKGKRVGLVCGSSGVOLPVAVAD---TVQSLHVFI		
PAMO	(160) NFPGLKDFAGMUYHTGNWPHEPVDFSGQRVGVIIGTGSQGQVQVMTAIAP---QAAELFVFQ		
CAMO	(153) DIKGISSEFAGDIHTAKWDHS-VELQGKTVGIIINGSTGVQVMTAIAP---KVSRLVSFQ		
CHMO	(151) NIKGINQEKGEIHTSRWPDD-VSFEKGKRVGVIGTGSTGVQVITAVAP---LAKHLTVFQ		
XP_751302	(183) PLEDQGRFKGKIFHSSSELDGK--DVEGKKVLIIGGGASAIEALEFAVKS--KARAIDVLS		
XP_747774	(181) DIPGLIHDKEKCKLQHSARWDES-YDWTGKRAAVIGNGSSGQIIVPGMLP---KVAHIDHYI		
XP_754119	(203) DYEGIINDYQCDLFHSSNWHN-VELKGKRVALIINGNGASGLQVLPQIOP---IAAHVHDHYA		
XP_751255	(178) DVEGINLFEQCKLILHSARWDPD-YDWTGKGVIAVIGNGSSALQIVPELQP---KAAKIVNYI		
XP_752204	(193) TVPGIETFECEGVVHTARWKS-DLDCGKDVIIIGSGCSATQVSELTKEPEYKIRSITQLM		
XP_756084	(191) DIPGLDDYKCKLMESARWDS-FDWTGKRVAVIGNGATAAOIIPEIAK---TAAHVSVYQ		
	<b>FXGXXXHXXXW (P/D)</b>	<b>GXGXX (G/A)</b>	