## Selective Pd-Catalyzed Oxidative Coupling of Anilides with Olefins through C-H Bond Activation at Room Temperature

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**General Information.** Experiments were carried out under air atmosphere using magnetic stirring unless otherwise noted. Solvents were purchased from commercial suppliers used without further purification. *n*-Butylacrylate was purchased from Aldrich and used as received. Anilides were obtained from commercial suppliers (Acros) or synthesized by reaction of the corresponding aniline with acetic anhydride.  $[(C_6H_4NHC(O)CH_3)Pd(II)(OAc^-)]_2$  was prepared as reported by Fujiwara.<sup>1</sup> <sup>1</sup>H NMR spectra were recorded on a Varian Mercury 300 (300.1 MHz) in CDCl<sub>3</sub> and are reported in ppm using tetramethylsilane as external standard. Data are reported as follows: (b – broad, s – singlet, d – doublet, t – triplet, q – quartet, m – multiplet; integration; coupling constant(s) in Hz; assignment). <sup>13</sup>C NMR spectra were recorded on the same spectrometer (75.5 MHz) in proton decoupled mode. GC measurements were performed on a Shimadzu GC-17A, equipped with a F.I.D. detector and a BPX35 column with an internal diameter of 0.22 mm and a film thickness of 0.25 µm. GC/MS measurements (E.I. detection) were performed on a HP 5890/5971 apparatus, equipped with a ZB-5 column (5% cross-linked phenyl polysiloxane) with an internal diameter of 0.25 µm. Melting points were measured on a Gallenkamp melting point apparatus and are uncorrected. Elemental analyses were performed at the Department of Microanalysis at the Rijksuniversiteit Groningen, The Netherlands.

**Parallel Screening Experiments.** Rapid screening experiments were carried out using a commercially available automated parallel synthesis Chemspeed ASW 2000 apparatus. The reactions were performed under an inert atmosphere of nitrogen. Each reaction vessel was charged with 0.02 mmol (1 mol%) of the desired catalyst, 0.25 mL of dihexylether as internal standard and, if required, 216 mg (2.0 mmol) of benzoquinone (BQ), followed by 2.0 mL of a stock solution of the aniline-substrate (1.0 M) in the desired solvent. Next, the alkene was added (2.0 mmol), the reaction mixture heated to 80 °C and subsequently stirred under vortex agitation for 16 hrs. Samples of the resulting reaction mixtures were diluted with hexanes and analyzed by GC/MS. See the figure on the last page for details on the reactions screened.

General Procedure for the Coupling of Acetanilide Derivatives with *n*-Butylacrylate. In a typical experiment, 3.0 mmol anilide, 13.5 mg (0.06 mmol) of  $Pd(OAc)_2$ , 324 mg (3.0 mmol) of BQ and 286 mg (1.5 mmol) of *p*-toluenesulfonic acid monohydrate are weighed into a one-neck roundbottom flask charged with a stirring bar. Next, 4.5 mL acetic acid is added, followed by a solution of 0.42 mL (3.0 mmol) *n*-butylacrylate in 2.25 mL of toluene. The flask is capped with a rubber septum, and the mixture is stirred overnight. Aliquots of the mixture are taken and diluted in diethyl ether, washed with a saturated NaHCO<sub>3</sub>-solution, dried over MgSO<sub>4</sub>, followed by GC or GC/MS analysis. After 16 hrs., the reaction mixture is diluted with 15 mL of ether, and carefully neutralized with a 2.5 M NaOH solution. After extraction of the aqueous phase with 15 mL ether, the combined organic phases are washed with water (15 mL), dried (MgSO<sub>4</sub>) and evaporated in vacuo. The resulting solids are purified by column chromatography to yield the corresponding product as a white powder. Recrystallization provided analytically pure product (except for **15**).

(*E*)-3-(2-acetylamino-phenyl)-propenoic acid butyl ester (4). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.80 (d, 1H, *J* = 15.7 Hz, olefinic H), 7.73 (d, 1H, *J* = 8.4 Hz, ArH), 7.54 (d, 1H, *J* = 7.8 Hz, ArH), 7.40-7.37 (m, 2H, ArH), 7.20 (m, 1H, ArH), 6.39 (d, 1H, *J* = 15.7 Hz, olefinic H), 4.19 (t, 2H, *J* = 6.6 Hz, C(O)OCH<sub>2</sub>), 2.22 (s, 3H, NHC(O)CH<sub>3</sub>), 1.71-1.62 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.45-1.42 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.95 (t, 3H, *J* = 7.5 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>). (The N-*H* resonance is not observed); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  169.2, 167.2, 139.5, 136.1, 131.0, 127.9, 127.3, 126.2, 125.6, 120.8, 64.9, 30.9, 24.4, 19.4, 14.0. Mp: 86 °C. Calcd. (%) for C<sub>15</sub>H<sub>19</sub>NO<sub>3</sub>: C 68.94, H 7.33, N 5.36; found: C 68.96, H 7.40, N 5.34.

(*E*)-3-(2-acetylamino-5-methylphenyl)-propenoic acid butyl ester (13). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.76 (d, 1H, *J* = 16.0 Hz, olefinic H), 7.65 (bs, 1H, N-*H*), 7.55 (d, 1H, *J* = 8.2 Hz, ArH), 7.36 (s, 1H, ArH), 7.19 (d, 1H, *J* = 8.2 Hz, ArH), 6.38 (d, 1H, *J* = 16.0 Hz, olefinic H), 4.19 (t, 2H, *J* = 6.6 Hz, C(O)OCH<sub>2</sub>), 2.33 (s, 3H, NHC(O)CH<sub>3</sub>), 2.14 (s, 3H, ArCH<sub>3</sub>), 1.70-1.63 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.46-1.38 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.96 (t, 3H, *J* = 7.3 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>):  $\delta$  169.5, 167.3, 139.9, 136.0, 133.7, 131.8, 128.3, 127.5, 126.1, 120.1, 64.8, 30.9, 24.1, 21.2, 19.4, 14.0 Mp: 97 °C. Calcd. (%) for C<sub>16</sub>H<sub>21</sub>NO<sub>3</sub>: C 69.79, H 7.69, N 5.09; found: C 69.65, H 7.82, N 5.08.

(*E*)-3-(2-acetylamino-4-methylphenyl)-propenoic acid butyl ester (14). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.95 (bs, 1H, N-*H*) 7.74 (d, 1H, *J* = 15.9 Hz, olefinic H), 7.40-7.35 (m, 2H, ArH), 6.94 (d, 1H, *J* = 8.0 Hz, ArH), 6.27 (d, 1H, *J* = 15.9 Hz, olefinic H), 4.12 (t, 2H, *J* = 6.6 Hz, C(O)OCH<sub>2</sub>), 2.27 (s, 3H, NHC(O)CH<sub>3</sub>), 2.15 (s, 3H, ArCH<sub>3</sub>), 1.64-1.59 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.40-1.35 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.91 (t, 3H, *J* = 7.3 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.4, 167.4, 141.7, 139.6, 136.0, 127.2, 127.0, 126.2, 125.2, 119.4, 64.8, 30.9, 24.3, 21.7, 19.4, 14.0. Mp: 127-128 °C. Calcd. (%) for C<sub>16</sub>H<sub>21</sub>NO<sub>3</sub>: C 69.79, H 7.69, N 5.09; found: C 69.77, H 7.74, N 5.08.

(*E*)-3-(2-acetylamino-3-methylphenyl)-propenoic acid butyl ester (15). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.79 (d, 1H, *J* = 15.9 Hz, olefinic H), 7.74 (bs, 1H, N-*H*), 7.30-7.22 (m, 3H, ArH), 6.38 (d, 1H, *J* = 15.9 Hz, olefinic H), 4.18 (t, 2H, *J* = 6.5 Hz, C(O)OCH<sub>2</sub>), 2.30 (s, 3H, NHC(O)CH<sub>3</sub>), 2.20 (s, 3H, ArCH<sub>3</sub>), 1.71-1.67 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.46-1.39 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.95 (t, 3H, *J* = 7.4 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>):  $\delta$  168.8, 167.3, 140.7, 136.8, 134.7, 130.7, 129.9, 127.9, 123.9, 120.0, 64.7, 30.9, 23.3, 19.4, 18.6, 14.0. Compound contains a significant amount of starting compound (7), which could not be separated from the product.

(*E*)-3-(2-acetylamino-5-methoxyphenyl)-propenoic acid butyl ester (16). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.75 (d, 1H, *J* = 15.7 Hz, olefinic H), 7.48 (d, 1H, *J* = 8.8 Hz, ArH), 7.08-7.05 (m, 2H, N-*H* and ArH overlapping), 6.95-6.91 (m, 1H, ArH), 6.38 (d, 1H, *J* = 15.7 Hz, olefinic H), 4.19 (t, 2H, *J* = 6.6 Hz, C(O)OCH<sub>2</sub>), 3.80 (s, 3H, OCH<sub>3</sub>), 2.20 (s, 3H, NHC(O)CH<sub>3</sub>), 1.69-1.61 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.46-1.38 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.92 (t, 3H, *J* = 7.4 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  169.8, 167.1, 157.9, 139.7, 130.4, 129.3, 128.2, 120.5, 117.0, 111.3, 64.8, 55.8, 30.9, 23.9, 19.4, 14.0. Mp: 128-129 °C. Calcd. (%) for C<sub>16</sub>H<sub>21</sub>NO<sub>4</sub>: C 65.96, H 7.27, N 4.81; found: C 66.04, H 7.29, N 4.85.

(*E*)-3-(2-benzoylamino-phenyl)-propenoic acid butyl ester (20). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.04 (bs, 1H, N-*H*), 7.88-7.81 (m, 4H, ArH + olefinic H), 7.59-7.38 (m, 5H, ArH), 7.25-7.20 (m, 1H, ArH), 6.40 (d, 1H, *J* = 15.9 Hz, olefinic H), 4.13 (t, 2H, *J* = 6.6 Hz, C(O)OCH<sub>2</sub>), 1.65-1.58 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.41-1.33 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.90 (t, 3H, *J* = 7.2 Hz, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  167.0, 166.3, 139.4, 136.1, 134.4, 132.4, 131.0, 129.1, 128.4, 127.5, 126.3, 125.6, 121.1, 64.8, 30.9, 19.4, 14.0. Mp: 145 °C. Calcd. (%) for C<sub>20</sub>H<sub>21</sub>NO<sub>3</sub>: C 74.28, H 6.55, N 4.33; found: C 73.98, H 6.48, N 4.40.

**Kinetic Competition Experiments.** These were performed as described above for the general catalysis procedures, but with a total Pd-loading of 34 mg (0.15 mmol; 5 mol%), equimolar amounts of competitive substrates (3.00 mmol in total) and at a reaction temperature of 40 °C. The conversion in acetanilide was determined by GC/MS using dihexylether as internal standard.

The kinetic data resulting from the competition experiments using *para*-substituted acetanilides give a Hammett-Brown plot as depicted in figure 1. From this correlation it follows that  $\rho^+ \approx -2.2$ , supporting a reaction pathway which involves attack of the electrophilic Pd<sup>+</sup> species on the arene  $\pi$ -system, resulting in a Wheland-type (arenium ion) intermediate.<sup>2,3</sup>

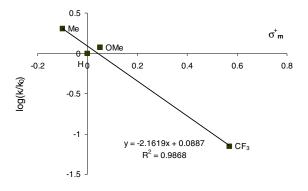


Figure 1. Hammett-Brown plot for the reaction of *para*-substituted acetanilides with *n*-butylacrylate.<sup>4</sup>

The kinetic isotope effect was determined by division of the observed rate-constants ( $k_{obs}$ ) obtained using acetanilide and 2,3,4,5,6-acetanilide- $d_5$  (see figure 2). The reaction follows pseudo first-order kinetics for both acetanilide- $h_5$  and acetanilide- $d_5$  as can be seen from figure 2. The values of  $k_{obs}$  relate to the values of the 'true'  $k'_{obs}$  according to the equation:

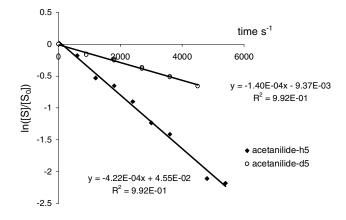
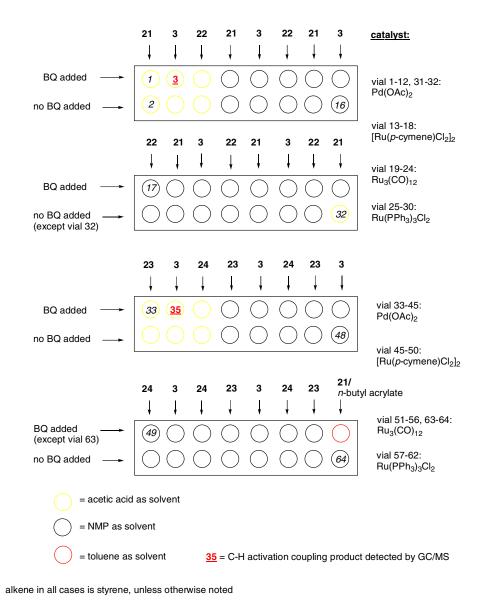


Figure 2. Logarithmic plot of the reaction rate for the reaction of acetanilide- $h_5/d_5$  with *n*-butylacrylate.

The observed kinetic isotope effect has a value  $(k_{\rm H}/k_{\rm D} = 3)$  similar to values reported in other systems.<sup>5</sup> The fact that a small isotope effect is present (instead of having a value of zero) can arise from the partitioning effect.<sup>6</sup>



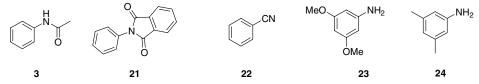


Figure 3. Schematic representation of the reactions performed using the rapid screening setup.

<sup>&</sup>lt;sup>1</sup> Fujiwara, Y.; Moritani, I.; Danno, S.; Asano, R.; Teranishi, S. J. Am. Chem. Soc. **1969**, *91*, 7166-7169.

similar of values have been reported for other aromatic substitution reactions by electrophilic metal species, see for example (a) Fujiwara, Y.; Asano, R.; Moritani, I.; Teranishi, S. J. Org. Chem. **1976**, 41, 1681-1683.

 <sup>&</sup>lt;sup>3</sup> However, Milstein proposed a mechanism without direct involvement of the π system, see: Weissman, H.; Song, X.; Milstein, D. J. Am. Chem. Soc. 2001, 123, 337-338 and references therein.
<sup>4</sup> σ<sub>m</sub><sup>+</sup> values were taken from March, J. Advanced Organic Chemistry, 3rd ed.; John Wiley & Sons: New York, 1985; p 280.
<sup>5</sup> see for example: (a) Shul'pin, G. B.; Nizova, G. V.; Nikitaev, A. T. J. Organomet. Chem. 1984, 276, 115-153; (b) reference 2.
<sup>6</sup> March, J. Advanced Organic Chemistry, 3rd ed.; John Wiley & Sons: New York, 1985; pp. 502-504.