



# Altered calcium signaling following traumatic brain injury

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Cell death and dysfunction after traumatic brain injury (TBI) is caused by a primary phase, related to direct mechanical disruption of the brain, and a secondary phase which consists of delayed events initiated at the time of the physical insult. Arguably, the calcium ion contributes greatly to the delayed cell damage and death after TBI. A large, sustained influx of calcium into cells can initiate cell death signaling cascades, through activation of several degradative enzymes, such as proteases and endonucleases. However, a sustained level of intracellular free calcium is not necessarily lethal, but the specific route of calcium entry may couple calcium directly to cell death pathways. Other sources of calcium, such as intracellular calcium stores, can also contribute to cell damage. In addition, calcium-mediated signal transduction pathways in neurons may be perturbed following injury. These latter types of alterations may contribute to abnormal physiology in neurons that do not necessarily die after a traumatic episode. This review provides an overview of experimental evidence that has led to our current understanding of the role of calcium signaling in death and dysfunction following TBI.

**Keywords:** calcium stores, excitotoxicity, *in vitro*, trauma, signal transduction

## INTRODUCTION

The calcium ion ( $\text{Ca}^{2+}$ ), and maintenance of cellular  $\text{Ca}^{2+}$  homeostasis, is essential for the proper function of the nervous system. For example,  $\text{Ca}^{2+}$  is critical for growth and development (Ghosh and Greenberg, 1995; Spitzer and Ribera, 1998; Takei et al., 1998), is a necessary component for neurotransmission (Berridge, 1998; Iwasaki et al., 2000), and contributes to distinct patterns of differential gene expression in neurons (Bading et al., 1993; Lerea and McNamara, 1993; Finkbeiner and Greenberg, 1998). Calcium also appears to be required for most forms of activity-dependent synaptic plasticity, generally believed to be the cellular correlate for learning and memory (Bliss and Collingridge, 1993; Malenka and Nicoll, 1999; Lamont and Weber, 2012; Kawamoto et al., 2012).

As the importance of  $\text{Ca}^{2+}$  in a wide range of physiological functions in the nervous system is now well established, it is understandable that changes in the levels of intracellular  $\text{Ca}^{2+}$  or alterations in  $\text{Ca}^{2+}$  signaling could contribute to a variety of pathologies. For example, changes in  $\text{Ca}^{2+}$  metabolism and homeostasis are well noted during the aging process (Kirischuk and Verkhratsky, 1996; Oliveira and Bading, 2011). Altered  $\text{Ca}^{2+}$  homeostasis is postulated to contribute to neurodegenerative diseases such as Huntington's, Parkinson's, and Alzheimer's disease, as well as amyotrophic lateral sclerosis (Mattson et al., 2000; Heath and Shaw, 2002; Wojda et al., 2008; Nakamura and Lipton, 2010; Surmeier et al., 2010). Disrupted signaling through  $\text{Ca}^{2+}$  channels also contributes to epilepsy and migraine (Cain and Snutch, 2011). Perhaps two of the most studied pathologies that are believed to involve disturbed  $\text{Ca}^{2+}$  homeostasis are cerebral ischemia and traumatic brain injury (TBI; Young, 1992; Tymianski and Tator, 1996; Kristián and Siesjö, 1998; Weber, 2004). The purpose of this review is to provide an overview of our current understanding of disrupted  $\text{Ca}^{2+}$  homeostasis following TBI, primarily at a

cellular level with a focus on neurons. Although the role of  $\text{Ca}^{2+}$  in TBI was arguably of considerably more attention to scientists a few decades ago, some important studies have been conducted in recent years that have advanced our understanding in this research field. These studies have provided more data not only on the role of  $\text{Ca}^{2+}$  in neuronal death, but on the role that  $\text{Ca}^{2+}$  could play in altered physiology in cells that do not die, such as the effects of disturbed signal transduction mechanisms or synaptic plasticity. These findings are highlighted in latter sections of the review.

## PATHOPHYSIOLOGY OF TRAUMATIC BRAIN INJURY

Traumatic brain injury is an insult to the brain caused by an external physical force, resulting in functional disability. Falls and motor vehicle accidents are the primary causes of TBI, while sports, assaults, and gunshot wounds also contribute significantly to these types of injuries (Centers for Disease Control and Prevention, 2010). TBI is one of the leading causes of death and disability worldwide, including the developing world (Reilly, 2007). TBI is characterized as mild, moderate or severe. Mild TBI, i.e., concussion, accounts for 70–90% of all TBI cases and 15–20% of individuals with a mild TBI have long-term dysfunction (Ryu et al., 2009). The principle mechanisms of damage from a TBI are due either to contact or acceleration/deceleration types of injury (Gennarelli, 1994). Contact injury results from an object striking the head and is generally associated with focal brain damage, which includes surface contusions, intracranial hematoma, and raised intracranial pressure (McIntosh et al., 1996; Patt and Brod-hun, 1999). Acceleration/deceleration brain injury results from unrestricted head movement in the instant after injury and leads to shear, tensile, and compressive strain. These types of injuries are generally associated with diffuse brain damage which includes

widespread damage to axons, ischemic brain injury, and diffuse brain swelling (Gennarelli, 1994).

The pathophysiology of TBI consists of two main phases, a primary (mechanical) phase of damage and secondary (delayed) damage. Primary damage occurs at the moment of insult and includes contusion and laceration, diffuse axonal injury, and intracranial hemorrhage (McIntosh et al., 1996; Patt and Brod-hun, 1999). Secondary damage includes processes that are initiated at the time of insult, but do not appear clinically for hours or even days after injury. Such processes include brain damage due to ischemia, swelling (edema), altered neurochemical mechanisms, and activation of degradative enzymes. The overall events that constitute secondary or delayed cell damage after TBI are complex, and still only partially understood.

A wide variety of mechanisms are proposed to contribute to secondary damage following trauma (for reviews, see Leker and Shohami, 2002; Weber, 2004; Slemmer et al., 2008; McAllister, 2011). Ischemia, caused by a reduction in cerebral blood flow (CBF), is an important mechanism underlying secondary brain damage, especially in severely injured individuals (Maas et al., 2000; van den Brink et al., 2000). A reduction of CBF damages mitochondria, which is followed by a shift from aerobic to anaerobic metabolism in neurons. In addition, ionic homeostasis becomes compromised and increases in intracellular  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Ca}^{2+}$  can lead to cell swelling and an even greater reduction in CBF. Damage to mitochondria leads to the release of reactive oxygen species (ROS), such as superoxide and hydroxyl radicals. These oxygen free radicals can damage cellular membranes by causing lipid peroxidation, can cause DNA strand breaks and damage purine and pyrimidine bases, and can also feedback and further damage mitochondrial membranes leading to altered function (Novack et al., 1996; Raghupathi and McIntosh, 1998; Slemmer et al., 2008). Inflammation also likely plays an important role in secondary brain damage after TBI. Neurons and glia synthesize and release cytokines such as tumor necrosis factor and interleukins, which may contribute to the immunologic response after TBI (Morganti-Kossmann et al., 1992; Stelmasiak et al., 2000). Enhancement of inflammation due to cytokine release and accumulation of immune cells could result in a further decrease in CBF due to swelling of the brain.

The extracellular level of glutamate, the major excitatory neurotransmitter in the mammalian nervous system, is known to be dramatically elevated after TBI *in vivo* (Faden et al., 1989; Katayama et al., 1990; Palmer et al., 1994). Glutamate activates a variety of receptors on neurons including  $\text{Ca}^{2+}$ -permeable ion channels. Excessive release of glutamate and over-stimulation of excitatory amino acid receptors, a process known as “excitotoxicity” as originally coined by Olney (1969), is believed to play a major role in secondary injury and death of cells of the CNS after traumatic injury (Hayes et al., 1992; Young, 1992; Weber, 2004; Lau and Tymianski, 2010). Furthermore, activation of voltage-gated calcium channels (VGCCs) or leakage through damaged cell membranes, can lead to a large elevation of  $\text{Ca}^{2+}$  within cells after injury (Young, 1992). This influx of  $\text{Ca}^{2+}$  into cells causes a corresponding elevation of intracellular free calcium ( $[\text{Ca}^{2+}]_i$ ), which can activate a variety of enzymes, setting in motion a cascade of events which eventually leads to cell injury and death

(Trump and Berezsky, 1995; Wojda et al., 2008). This alteration in cellular  $\text{Ca}^{2+}$  homeostasis is likely one of the key mechanisms contributing to secondary neuronal damage and altered physiology following TBI.

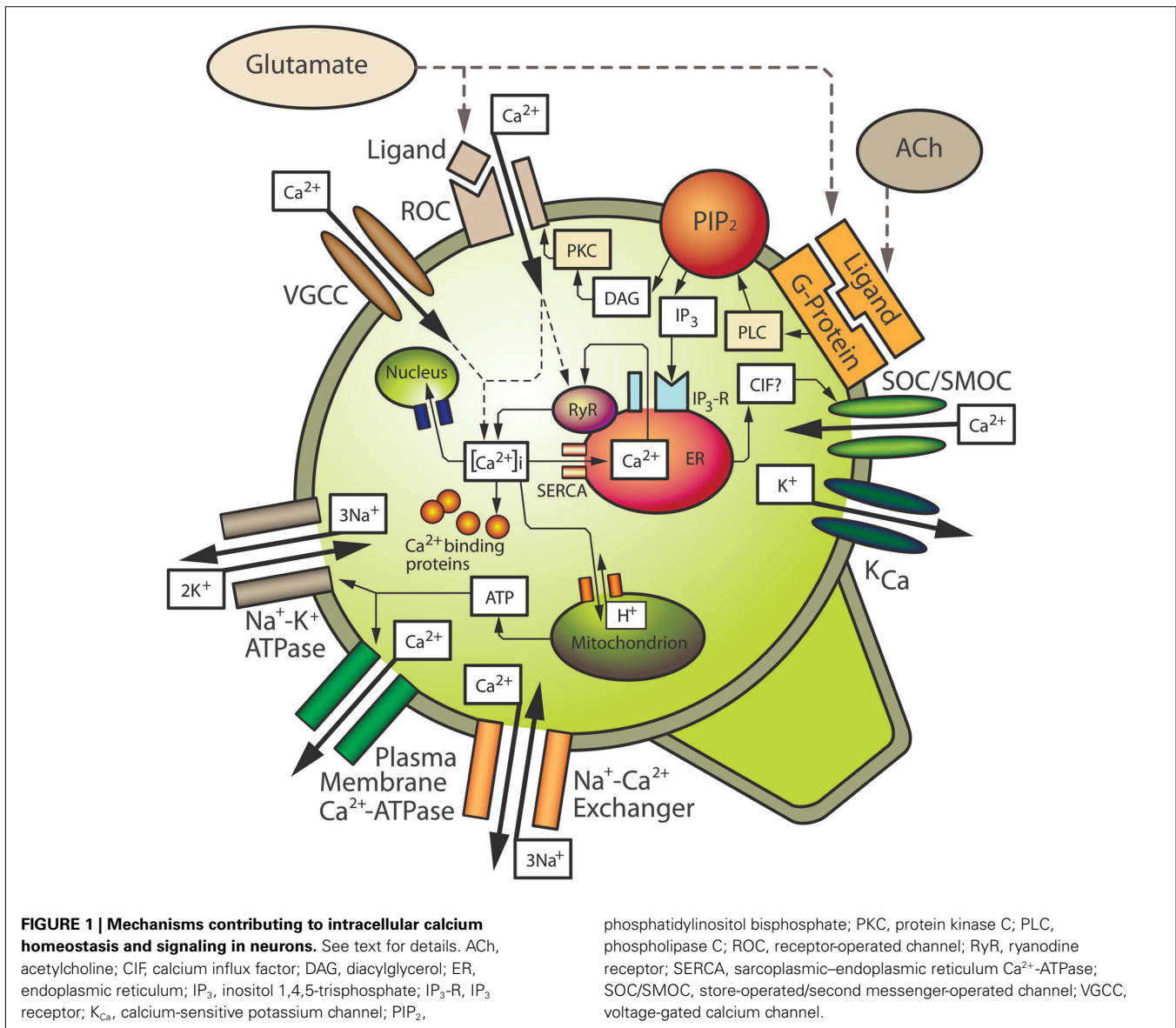
## MECHANISMS OF MAINTAINING CALCIUM HOMEOSTASIS IN NEURONS

### MECHANISMS FOR INCREASING $[\text{Ca}^{2+}]_i$ LEVELS

Arguably,  $\text{Ca}^{2+}$  is the most ubiquitous signal transduction molecule in the body as it acts as an important intracellular second messenger that regulates vital processes in almost all mammalian cells (Petersen et al., 2005). Calcium is so important to normal neuronal function in particular, that these cells have developed several homeostatic mechanisms to control the subcellular location of  $\text{Ca}^{2+}$  and the level of  $[\text{Ca}^{2+}]_i$ . These mechanisms include  $\text{Ca}^{2+}$  influx through voltage-operated and receptor-operated channels (ROCs),  $\text{Ca}^{2+}$  buffering by the plasma membrane and cytosolic proteins,  $\text{Ca}^{2+}$  storage in intracellular organelles and  $\text{Ca}^{2+}$  efflux (Miller, 1991; Kostyuk and Verkhratsky, 1994; Wojda et al., 2008; see **Figure 1**). These homeostatic mechanisms maintain  $[\text{Ca}^{2+}]_i$  at low levels, usually about 100 nM in neurons compared to an extracellular concentration of approximately 1 mM. Maintaining  $[\text{Ca}^{2+}]_i$  at such low levels in the cell allows relatively small or localized increases in  $[\text{Ca}^{2+}]_i$  to be used as a trigger to activate signal transduction pathways which lead to physiological processes such as activation of specific enzymes or modulation of ion channels (Racay and Lehotsky, 1996; Tymianski and Tator, 1996; Barish, 1998; Weber, 2004).

In order to activate various signal transduction mechanisms leading to a physiological response, an elevation in  $[\text{Ca}^{2+}]_i$  is often required. Neurons have various ways of increasing  $[\text{Ca}^{2+}]_i$ . One such way is by ROCs, which are  $\text{Ca}^{2+}$  ion channels activated by ligands (**Figure 1**). An important example of a ROC is the *N*-methyl-D-aspartate (NMDA) receptor. Activation of NMDA receptors by the endogenous ligand, glutamate, leads to the opening of its associated ion channel and an influx of  $\text{Ca}^{2+}$  from the extracellular space into the cytosol. Another major route of  $\text{Ca}^{2+}$  influx into neurons is through several types of VGCCs (Mintz et al., 1992; Snutch and Reiner, 1992; Takahashi and Momiyama, 1993; Westenbroek et al., 1998; Iwasaki et al., 2000; Schmolesky et al., 2002; Cain and Snutch, 2011). The activation of VGCCs is dependent on the membrane voltage of the cell and they are generally in a closed state when the neuronal membrane is polarized. When the membrane becomes depolarized, e.g., due to  $\text{Na}^+$  influx through ion channels or due to  $\text{Ca}^{2+}$  influx through ROCs, these VGCCs allow the influx of  $\text{Ca}^{2+}$  into the cell. An individual neuron may have several different types of VGCCs and ROCs, and  $\text{Ca}^{2+}$  influx through specific channels is quite important. For example, certain classes of VGCCs trigger neurotransmitter release at synaptic terminals (Robitaille et al., 1990; Augustine et al., 1991; Robitaille et al., 1993; Wu et al., 1998), and specific ionotropic glutamate receptors can lead to  $\text{Ca}^{2+}$ -dependent gene induction (Bading et al., 1993; Lerea and McNamara, 1993; Barish, 1998).

Another important mechanism by which neurons increase  $[\text{Ca}^{2+}]_i$  is through the release of  $\text{Ca}^{2+}$  from intracellular stores located on the endoplasmic reticulum (ER). As with  $\text{Ca}^{2+}$  influx, an elevation of  $[\text{Ca}^{2+}]_i$  through the release of stored  $\text{Ca}^{2+}$  is also



believed to play an important role in the normal physiological function of neurons (Kostyuk and Verkhratsky, 1994). The ER consists of inositol 1,4,5-trisphosphate (IP<sub>3</sub>)-sensitive Ca<sup>2+</sup> stores, and ryanodine-sensitive Ca<sup>2+</sup> stores, so named because of their sensitivity to the plant alkaloid ryanodine, which is generally an agonist at low doses and an antagonist at higher doses (Ehrlich et al., 1994). Ryanodine receptors (RyRs) are associated with Ca<sup>2+</sup>-induced Ca<sup>2+</sup> release (CICR) channels. Both IP<sub>3</sub>-sensitive stores and ryanodine-sensitive stores have been identified in neuronal somata and dendrites in various areas of the brain including the hippocampus, cerebellum, and the neocortex (Challiss et al., 1990; Sharp et al., 1993; Simpson et al., 1996; Kurumatani et al., 1997; Nakashima et al., 1997; Oberdorf et al., 1997).

Release of Ca<sup>2+</sup> from IP<sub>3</sub>-sensitive stores occurs upon the activation of G protein-coupled receptors that activate phospholipase C (PLC), e.g., Group I metabotropic glutamate receptors (mGluRs), or the M<sub>1</sub> or M<sub>3</sub> type of acetylcholine (ACh) receptors.

Upon activation, PLC cleaves phosphatidylinositol bisphosphate (PIP<sub>2</sub>) from cell membranes, resulting in the release of diacylglycerol (DAG) and IP<sub>3</sub> (Figure 1). IP<sub>3</sub> binds to receptors (IP<sub>3</sub>R) on IP<sub>3</sub>-sensitive intracellular Ca<sup>2+</sup> stores located on the ER, resulting in release of Ca<sup>2+</sup> (Berridge, 1993; Ozawa et al., 1998). In addition, depletion of Ca<sup>2+</sup> stores stimulates influx of extracellular Ca<sup>2+</sup> through store-operated or second messenger-operated channels (SOCs/SMOCs) in the plasma membrane, a process known as “capacitative calcium influx” (CCI) as originally postulated by Putney (1986). Capacitative Ca<sup>2+</sup> influx is analogous with a capacitor in an electrical circuit in which Ca<sup>2+</sup> stores prevent entry when they are filled but promote entry when they become depleted. Capacitative Ca<sup>2+</sup> entry has been extensively studied in non-excitable cells (Putney and Bird, 1993; Berridge, 1995), and more recently, capacitative Ca<sup>2+</sup> influx has been described in neurons (e.g., Bouron, 2000; Emptage et al., 2001; Weber et al., 2001; Baba et al., 2003; Bouron et al., 2005). Influx likely occurs through

members of the transient receptor potential channel (TRPC) family, which is a group of membrane bound ion channels (see Salido et al., 2011 for review).  $\text{Ca}^{2+}$  influx through TRPCs thereby represents a way in which neurons can further increase  $[\text{Ca}^{2+}]_i$ . CICR can be triggered by  $\text{Ca}^{2+}$  entering through VGCCs or ROCs, however the putative second messenger, cyclic ADP ribose, may also contribute to CICR in neurons (Galione, 1994; Lee, 1994), most likely by increasing the sensitivity of RyR to  $\text{Ca}^{2+}$  (Hua et al., 1994). In addition, both  $\text{IP}_3$ Rs and RyRs are sensitive to  $\text{Ca}^{2+}$  that has entered through membrane channels or has been released from adjacent receptors on the stores (Kano et al., 1995; Verkhratsky and Shmigol, 1996). A buildup of  $\text{Ca}^{2+}$  within the ER also seems to increase the sensitivity of  $\text{IP}_3$ Rs and RyRs (Berridge, 1998). It should also be noted that release of  $\text{Ca}^{2+}$  from intracellular stores, or mGluR activation alone, can alter the sensitivity of membrane ROCs or VGCCs, further contributing to  $\text{Ca}^{2+}$  elevation in the cytosol (Bruno et al., 1995; Choi and Lovinger, 1996; Rahman and Neuman, 1996; Pisani et al., 2001).

### MECHANISMS FOR DECREASING $[\text{Ca}^{2+}]_i$ LEVELS

The maintenance of  $\text{Ca}^{2+}$  homeostasis in neurons requires mechanisms that can compensate for the influx of extracellular  $\text{Ca}^{2+}$  and release of  $\text{Ca}^{2+}$  from intracellular stores, once a target protein or biological process has been activated. These mechanisms of lowering  $[\text{Ca}^{2+}]_i$  include a balance of  $\text{Ca}^{2+}$  extrusion systems in the plasma membrane, buffering by  $\text{Ca}^{2+}$  binding proteins, and uptake and storage by various intracellular organelles (Figure 1). Extrusion of  $\text{Ca}^{2+}$  from the cell to the extracellular space occurs by membrane pumps such as the plasma membrane  $\text{Ca}^{2+}$ -ATPase (PMCA), and a  $\text{Na}^+/\text{Ca}^{2+}$  exchanger which is activated by high levels of  $[\text{Ca}^{2+}]_i$  and is dependent on the  $\text{Na}^+$  gradient, which in turn is dependent on the membrane  $\text{Na}^+/\text{K}^+$  ATPase (Garcia and Strehler, 1999; Wojda et al., 2008; Brittain et al., 2012). In addition to these cell extrusion mechanisms for lowering  $[\text{Ca}^{2+}]_i$ , elevated  $\text{Ca}^{2+}$  is buffered by  $\text{Ca}^{2+}$  binding proteins in the cytosol (Baimbridge et al., 1992; Racay and Lehotsky, 1996), which is believed to be the major way of decreasing elevated  $[\text{Ca}^{2+}]_i$  in neuronal cells (Neher and Augustine, 1992; Zhou and Neher, 1993). Some cytoplasmic proteins, such as calbindin and parvalbumin, act primarily as  $\text{Ca}^{2+}$  acceptors; these proteins however, are thought to deliver  $\text{Ca}^{2+}$  ions to sites of extrusion, such as the PMCA, or to sites of storage such as the ER. The plasma membrane also contains several  $\text{Ca}^{2+}$  binding proteins, which can buffer  $\text{Ca}^{2+}$  but also play a structural role in the cell by contributing to the maintenance of cytoskeletal elements (Baimbridge et al., 1992; Racay and Lehotsky, 1996). Some of these buffering proteins may also play a physiological role in addition to being able to reduce  $[\text{Ca}^{2+}]_i$ . For example, calmodulin, once bound to  $\text{Ca}^{2+}$ , can activate several protein kinases, which can trigger a wide variety of physiological responses in neurons (Braun and Schulman, 1995).

Although  $\text{Ca}^{2+}$ -binding proteins can buffer much of the  $\text{Ca}^{2+}$  in neurons, these proteins can become saturated. Therefore, neurons also sequester  $\text{Ca}^{2+}$  into intracellular organelles such as the ER, mitochondria, and the nucleus in order to maintain homeostatic levels of  $[\text{Ca}^{2+}]_i$  (Miller, 1991; Kostyuk and Verkhratsky, 1994). However, excess  $\text{Ca}^{2+}$  in the mitochondria can lead to altered energy metabolism (Budd and Nicholls, 1996; Schinder

et al., 1996; White and Reynolds, 1996) and elevated  $\text{Ca}^{2+}$  in the nucleus can activate endonucleases, which can cause DNA damage (Trump and Berezsky, 1995; McConkey and Orrenius, 1996). Therefore, these storage methods may only be utilized as secondary mechanisms in cases of excess  $[\text{Ca}^{2+}]_i$  levels not achieved under normal physiological circumstances. In addition to contributing to an elevation of  $[\text{Ca}^{2+}]_i$  for signal transduction events,  $\text{Ca}^{2+}$  stores on the ER also take up  $\text{Ca}^{2+}$  from the cytosol in order to return  $[\text{Ca}^{2+}]_i$  to normal resting levels. This is accomplished by a sarcoplasmic–endoplasmic reticulum  $\text{Ca}^{2+}$ -ATPase (SERCA) on the ER membrane (Figure 1). Once stores of the ER are filled,  $\text{Ca}^{2+}$  is again available for second messenger signaling events mediated by  $\text{IP}_3$  and CICR. Based on the essential role that calcium ions play in normal neuronal function, it is easy to envision the profound effects that an imbalance in  $\text{Ca}^{2+}$  homeostasis would have in these cells, which ultimately could be manifested at a behavioral level. Also, given the complexity of the mechanisms by which neurons maintain  $[\text{Ca}^{2+}]_i$  within a very small range, an alteration to just one component of the homeostatic system could result in altered  $[\text{Ca}^{2+}]_i$  levels and abnormal physiology.

### ROLE OF ELEVATED CALCIUM IN THE PATHOLOGY OF TBI

#### MECHANISMS OF ELEVATED INTRACELLULAR $[\text{Ca}^{2+}]_i$ LEVELS AFTER TBI

One of the major causes of secondary injury following TBI is likely related to an excessive influx of  $\text{Ca}^{2+}$  into cells of the brain, which can cause cell death, especially after moderate or severe TBI (Young, 1992; Farooqui and Horrocks, 1994). This original “calcium hypothesis” for TBI is based in part on findings that  $\text{Ca}^{2+}$  in the cerebrospinal fluid (CSF) decreases from 1 to 0.01 mM following traumatic injury (Young, 1992; Nilsson et al., 1993), and that total tissue  $^{45}\text{Ca}^{2+}$  uptake is increased after injury *in vivo* (Hovda et al., 1992; Fineman et al., 1993). The hypothesis is also based on previous evidence from other studies in the CNS not directly related to TBI. For example, Schlaepfer and Bunge (1973) showed that amputated axons in tissue culture would degenerate only in the presence of extracellular  $\text{Ca}^{2+}$ . Additional studies demonstrated that the toxicity of excitatory amino acids appeared to be  $\text{Ca}^{2+}$ -dependent (Berdichevsky et al., 1983; Choi, 1985; Garthwaite et al., 1986). Therefore, the prevailing hypothesis three decades ago was that  $\text{Ca}^{2+}$  overload following head injury triggered a cascade of events leading to cell damage and eventual cell death.

There are several mechanisms by which neuronal  $\text{Ca}^{2+}$  becomes elevated following TBI (see Figure 1). Because TBI is caused by a physical insult it is widely proposed that a major route of  $\text{Ca}^{2+}$  entry is through plasma membrane pores formed by tear-shearing forces during the primary injury (Maxwell et al., 1993; Gennarelli and Graham, 1998; Büki et al., 1999). This concept is supported by the increased permeability of neuronal membranes to large molecules following experimental TBI *in vivo* (Pettus et al., 1994; Pettus and Povlishock, 1996) and after mechanical stretch *in vitro* (McKinney et al., 1996; Pike et al., 2000; Slemmer et al., 2002). In addition to producing pores in cell membranes, mechanical strain can cause disturbances to existing membrane proteins, such as voltage-gated  $\text{Na}^+$  channels, which are known to be sensitive to mechanical stress (Shcherbatko et al., 1999; Tabarean et al., 1999). An increased influx of  $\text{Na}^+$  into the cell would cause



membrane depolarization, which could then activate VGCCs leading to  $\text{Ca}^{2+}$  influx (Wolf et al., 2001). Also, a high level of  $\text{Na}^+$  in the cell could cause the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger to operate in reverse, further contributing to a buildup of  $\text{Ca}^{2+}$  inside the cell (Stys et al., 1992; Kiedrowski et al., 1994; Stys and Lopachin, 1998; Brittain et al., 2012). Membrane depolarization causes the release of neurotransmitters such as glutamate and ACh from presynaptic terminals. The extracellular concentration of glutamate has long been known to be elevated after TBI (Faden et al., 1989; Katayama et al., 1990; Palmer et al., 1994; Zauner et al., 1996) and levels of ACh are also increased in CSF (Hayes et al., 1992).

Upon release, glutamate can activate a variety of functionally distinct receptors on neurons in the CNS (for reviews, see Ozawa et al., 1998; Coutinho and Knöpfel, 2002; Slemmer et al., 2005). These include ionotropic receptors such as  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionate (AMPA), kainate, and NMDA receptors, as well as mGluRs. NMDA receptors are well-distributed in the brain, and over-activation of this receptor subtype is thought to be a major route of  $\text{Ca}^{2+}$  influx into cells. Under normal resting membrane potential, NMDA receptors are subject to a blockade of its ion pore by  $\text{Mg}^{2+}$ , even in the presence of glutamate and glycine, which is a coagonist of the receptor. When the cell membrane becomes depolarized, the  $\text{Mg}^{2+}$  block is decreased or lost leading to  $\text{Ca}^{2+}$  influx through the ion pore of the receptor. NMDA receptors are also sensitive to mechanical deformation, as membrane stretch causes potentiated responses mediated by these receptors (Paoletti and Ascher, 1994; Casado and Ascher, 1998). Stretch injury also causes a reduction in the voltage-dependent  $\text{Mg}^{2+}$  blockade of NMDA receptors, which leads to elevated  $[\text{Ca}^{2+}]_i$  levels (Zhang et al., 1996). AMPA receptors probably play less of a role in  $\text{Ca}^{2+}$  influx into neurons following injury than do NMDA receptors, because these channels are primarily permeable to  $\text{Na}^+$  (Ozawa et al., 1998). However, some types of neurons do express  $\text{Ca}^{2+}$ -permeable AMPA receptors (Iino et al., 1990; Lu et al., 1996; Carriedo et al., 1998). AMPA receptors are also susceptible to stretch injury, which causes an enhancement in current through these channels (Goforth et al., 1999). Increased  $\text{Na}^+$  flux through AMPA receptors could indirectly lead to further  $\text{Ca}^{2+}$  influx by membrane depolarization, and a corresponding increase in the activation of NMDA receptors and VGCCs. There are also several types of mGluRs, divided into three groups, which are activated directly by binding of glutamate (see Coutinho and Knöpfel, 2002 for a review); however only Group I mGluRs produce a direct increase in  $[\text{Ca}^{2+}]_i$ . Once activated, these G protein-coupled receptors cause an increase in  $\text{IP}_3$  which binds to  $\text{IP}_3$ Rs on the ER, resulting in a release of  $\text{Ca}^{2+}$  from the stores (Figure 1). This depletion of  $\text{Ca}^{2+}$  stores also stimulates influx of extracellular  $\text{Ca}^{2+}$  through channels (SOCs/SMOCs) in the plasma membrane in neurons (Bouron, 2000; Weber et al., 2001; Baba et al., 2003), which are likely members of the TRPC family. In addition, some studies have shown that activation of mGluRs coupled to intracellular  $\text{Ca}^{2+}$  stores can enhance  $[\text{Ca}^{2+}]_i$  elevation mediated by NMDA receptors (Bruno et al., 1995; Rahman and Neuman, 1996; Pisani et al., 2001), and mGluRs in cerebellar Purkinje cells can directly couple to TRPC1, to cause  $\text{Ca}^{2+}$  influx through these channels (Kim et al., 2003). Besides glutamate, ACh can also activate G protein receptors coupled to  $\text{IP}_3$  (muscarinic receptor types 1 and

3) and contribute to intracellular  $\text{Ca}^{2+}$  elevation. ACh also activates nicotinic receptors on some neurons, which are permeable to both  $\text{Ca}^{2+}$  and  $\text{Na}^+$  (Le Novère et al., 2002). As with other types of  $\text{Na}^+$  channels, influx of  $\text{Na}^+$  leads to depolarization and subsequent activation of other types of  $\text{Ca}^{2+}$  channels. Elevated  $\text{Ca}^{2+}$  levels can also cause over-activation of  $\text{Ca}^{2+}$ -sensitive potassium channels ( $\text{K}_{\text{Ca}}$ ), which could alter the membrane potential of cells and alter signaling cascades (Wulff et al., 2007).

Elevated intracellular  $\text{Ca}^{2+}$  after TBI may also be a result of reduced ATP levels. This may be due to a reduction in glucose levels from decreased CBF, or direct mechanical damage to mitochondria (Slemmer et al., 2008). For example, mechanically induced injury can lead to altered mitochondrial membrane potential and decreased ATP levels in neurons in the absence of hypoxia (Tavalin et al., 1995, 1997; Ahmed et al., 2000, 2002). A reduction in ATP is a serious consequence for neurons, i.e., as the supply of ATP to the PMCA and the SERCA of intracellular stores is reduced, the cells are unable to pump  $\text{Ca}^{2+}$  either into the extracellular space or into stores and thus  $\text{Ca}^{2+}$  accumulates in the cytosol (Parsons et al., 1997; Limbrick et al., 2001). Other transmembrane pumps are compromised, such as the  $\text{Na}^+/\text{K}^+$ -ATPase, furthering contributing to a breakdown of the ionic gradient of the membrane (Tavalin et al., 1997). The cell becomes further depolarized, again with subsequent activation of glutamatergic ion channels and VGCCs, leading to elevated  $[\text{Ca}^{2+}]_i$ . This can initiate a cyclical process whereby many depolarized neurons release additional glutamate leading to further over-activation of all classes of glutamate receptors on adjacent cells.

#### MECHANISMS OF CALCIUM-MEDIATED CELL DAMAGE AND DEATH

Elevated neuronal  $[\text{Ca}^{2+}]_i$  leads to damage and death through several mechanisms following TBI; the activation of  $\text{Ca}^{2+}$ -dependent enzymes and  $\text{Ca}^{2+}$  overload of the mitochondria appearing to be major causes of neuronal damage (for reviews, see Trump and Berezsky, 1996; Kermer et al., 1999; Weber, 2004; Slemmer et al., 2008). Depending on the  $\text{Ca}^{2+}$  load and route of  $\text{Ca}^{2+}$  entry, these mechanisms can cause cell death by rapid necrotic means or by a more delayed cell death (apoptosis).

Several cysteine proteases, such as calpains, are activated by high levels of  $[\text{Ca}^{2+}]_i$ . In neurons, specific calpain isoforms are activated in the low micromolar  $[\text{Ca}^{2+}]_i$  range, a level that is achieved by hypoxic or excitotoxic insults, and is believed to be reached after TBI (Wang and Yuen, 1994; Tymianski and Tator, 1996). Several cytoskeletal elements serve as protein substrates for calpain, including spectrin, tubulin, tau, microtubule-associated protein, and neurofilaments (Wang and Yuen, 1994; Kermer et al., 1999). Calpain is also reported to be involved in the degradation of membrane proteins such as glutamate receptors and transporters (Takahashi, 1990). Cell-adhesion molecules and other enzymes such as protein kinases and phosphatases can also serve as substrates for calpain (Takahashi, 1990). Therefore, injury-induced over-activation of calpain can lead to abnormal and possibly irreversible changes to the membrane and cytoskeleton of cells in the CNS (Wang and Yuen, 1994; Bartus, 1997). These changes can compromise the structural integrity of the cell, leading to increased membrane permeability to ions and even macromolecules. More recent data has suggested that calpain activation may not occur due

the rapid, initial increase in  $[Ca^{2+}]_i$  after injury or exposure to high levels of glutamate (Araújo et al., 2010). For example, some studies investigating excitotoxicity have shown that calpain activation occurs only after there is a delayed  $Ca^{2+}$  deregulation in neurons (Gerencser et al., 2009; Brustovetsky et al., 2010). Therefore, calpain activation may occur only after a neuron is physiologically compromised, rather than directly contributing to initial cell damage.

Activation of the caspase gene family is also associated with elevated intracellular  $Ca^{2+}$ . The caspases are a family of cysteine proteases that are distinct from the calpains (Toescu, 1998). These proteins undergo proteolytic processing which leads to the induction of apoptotic cascades. Caspases have been intensely studied following TBI, with enhanced activation being noted following trauma (for reviews, see Büki and Povlishock, 2006; Saatman et al., 2010).  $Ca^{2+}$ -dependent endonucleases can also be activated following TBI, which generate DNA damage in the form of the orderly chromatin cleavage patterns that are typical of apoptosis or delayed cell death (Trump and Berezsky, 1995; McConkey and Orrenius, 1996; Toescu, 1998).

An increase in  $[Ca^{2+}]_i$  after injury can activate phospholipases, most notably PLC and cytosolic phospholipase  $A_2$  (cPLA<sub>2</sub>). Both PLC (Wei et al., 1982; Dhillon et al., 1999) and cPLA<sub>2</sub> (Shohami et al., 1989) are reported to be elevated after TBI. cPLA<sub>2</sub> activation leads to the release and accumulation of free arachidonic acid (AA), which itself may alter membrane permeability and increase ROS (Slemmer et al., 2008). The production of ROS such as the oxygen free radicals, superoxide ( $O_2^{\bullet-}$ ) and hydroxyl radicals ( $OH^{\bullet}$ ), is part of the normal metabolism of the brain. Under normal physiological conditions, oxygen free radicals are kept under strict control by cellular defense mechanisms. Following a traumatic episode, however, these defense mechanisms may be compromised leading to an abundance of free radicals which can cause cellular damage by peroxidation of proteins, DNA, and lipid membranes (Slemmer et al., 2008). As previously described, PLC cleaves PIP<sub>2</sub> from cell membranes, resulting in the release of DAG and IP<sub>3</sub>, which can cause release of  $Ca^{2+}$  from intracellular stores, and contribute further to an increase in  $[Ca^{2+}]_i$ . Injury-induced formation of DAG has been associated with post-traumatic edema (Dhillon et al., 1994, 1995). In addition, DAG activates protein kinase C (PKC), which can modulate other signal transduction pathways. PKC has been shown to be elevated after experimental TBI (Sun and Faden, 1994) and its activation can directly modulate NMDA and VGCCs leading to potentiated activity and an enhanced influx of  $Ca^{2+}$  into neurons (Choi and Lovinger, 1996; Zhang et al., 1996).

Another toxic enzymatic pathway in neurons regulated by  $Ca^{2+}$  is the production of nitric oxide (NO). Upon the activation of NMDA receptors and subsequent  $Ca^{2+}$  influx,  $Ca^{2+}$  binds to the enzyme calmodulin, which then regulates NO synthase (NOS), thereby producing NO from L-arginine (Dawson and Dawson, 1996). Under physiological conditions, the level of NO is low and it contributes to several important biological processes including cerebral vasodilation, neurotransmission, and synaptic plasticity (Dawson and Dawson, 1996). However, excessive NO can combine with superoxide to produce peroxynitrite ( $ONOO^-$ ), which is highly toxic to proteins, membrane lipids, and DNA (Dawson,

1999; Liu et al., 2002; Slemmer et al., 2008). NOS itself produces superoxide, but levels are generally kept in check by endogenous free radical scavengers like superoxide dismutase (SOD). However following trauma, when there may be over-activation of NOS and additional mechanisms producing superoxide (i.e., metabolism of AA and damage to mitochondria), endogenous scavengers like SOD may become saturated, allowing high levels of peroxynitrite to form, ultimately leading to neuronal damage.

## CALCIUM LOAD VERSUS SOURCE SPECIFICITY

The available current evidence strongly supports the fact that intracellular  $Ca^{2+}$  levels are elevated following traumatic neuronal injury. However, the levels of  $Ca^{2+}$  in cells may differ based on the severity of injury and the temporal  $Ca^{2+}$  dynamics may also depend on the level of TBI. The exact mechanisms by which  $Ca^{2+}$  causes neuronal dysfunction are obviously complex, and exactly how  $Ca^{2+}$  causes toxicity to neurons is still not completely understood. One of the original major hypotheses in this field postulated that there is simply a correlation of neuronal death with the amount of  $Ca^{2+}$  that enters the cytosol (Hartley et al., 1993; Eimerl and Schramm, 1994; Lu et al., 1996). This “calcium load” hypothesis is in line with the original calcium hypothesis for TBI, suggesting that after a traumatic insult, neurons are simply overwhelmed with a high  $Ca^{2+}$  load that activates all of the processes discussed previously. However, additional data led to the formation of the refined “source specificity” hypothesis, which suggests that  $Ca^{2+}$ -induced neurotoxicity is caused by specific pathways of influx (Sattler and Tymianski, 2000), or by release of  $Ca^{2+}$  from distinct intracellular sources such as the ER (Paschen, 2000). Much of the data in support of this latter hypothesis has been generated by studies of excitotoxicity or ischemia in neurons, however, because TBI can contain both excitotoxic and ischemic components, this idea can be translated to traumatic neuronal injury.

Determining  $Ca^{2+}$  accumulation in cells or measuring  $[Ca^{2+}]_i$  *in vivo* is technically challenging, therefore most studies aimed at achieving these goals have been conducted *in vitro*. Several reports using cultured neuronal cells have exhibited a strong relationship between the intracellular  $Ca^{2+}$  load and neuronal death following excitotoxic or ischemic challenges (e.g., Manev et al., 1989; Marcoux et al., 1990; Hartley et al., 1993; Eimerl and Schramm, 1994; Lu et al., 1996). However, several other studies have resulted in findings that seem to oppose this correlation between  $Ca^{2+}$  concentration and cell death. For example, Turner et al. (2002) showed that reducing intracellular  $Ca^{2+}$  levels induces injury in developing neurons *in vitro* and *in vivo*. In a model of neurite transection, injury caused cell death but was independent of the level of extracellular  $Ca^{2+}$  in rat sympathetic ganglion cells (Chu et al., 2001). Important studies conducted by Nilsson et al. (1993, 1996) *in vivo* demonstrated a decrease in extracellular  $Ca^{2+}$  after injury and a resulting increase in tissue  $^{45}Ca^{2+}$  uptake, however, neuronal damage did not correlate with the extent of intracellular  $Ca^{2+}$  accumulation (Nilsson et al., 1996).

Due to these conflicting findings, several investigators have suggested that cell death is due to the distinct type of  $Ca^{2+}$  pool that is elevated. Despite the evidence that intracellular  $Ca^{2+}$  is elevated in cells of the CNS after trauma, the relative distribution of  $Ca^{2+}$  throughout the cell after traumatic injury is still unknown.

Much of the  $\text{Ca}^{2+}$  influx after injury may be buffered by the plasma membrane, cytoskeletal elements, and by cytoplasmic proteins such as calmodulin, calbindin, and parvalbumin (Baimbridge et al., 1992), which could provide rapid protection from such dramatic elevations in intracellular  $\text{Ca}^{2+}$  levels. Another major pool that absorbs a high  $\text{Ca}^{2+}$  load under times of stress is the mitochondria (Kristián and Siesjö, 1998), and as discussed earlier, this uptake of  $\text{Ca}^{2+}$  by mitochondria and resulting energy failure may ultimately lead to neuronal degeneration.

An essential form of  $\text{Ca}^{2+}$  within the cell is  $[\text{Ca}^{2+}]_i$ , the type of  $\text{Ca}^{2+}$  necessary to activate the degradative enzymes proposed to lead to neuronal injury and death. Several studies using *in vitro* models of mechanical stretch or shear strain have measured levels of  $[\text{Ca}^{2+}]_i$  after injury in neuronal cell lines (Cargill and Thibault, 1996; LaPlaca et al., 1997; LaPlaca and Thibault, 1998; Geddes and Cargill, 2001; Tárnok and Ulrich, 2001), primary cultures of cortical neurons (Weber et al., 1999, 2001, 2002; Spaethling et al., 2008; Shahlaie et al., 2010; Staal et al., 2010; Goforth et al., 2011), and hippocampal neurons (Geddes et al., 2003; Lusardi et al., 2004). These studies generally show that  $[\text{Ca}^{2+}]_i$  is acutely elevated following a mechanical insult, and although somewhat dependent on the strain level, estimates of  $[\text{Ca}^{2+}]_i$  levels after injury are generally in the nanomolar range. However, most of these studies used the high affinity  $\text{Ca}^{2+}$  indicator fura-2 for  $[\text{Ca}^{2+}]_i$  measurements and fura-2 may underestimate  $[\text{Ca}^{2+}]_i$  levels when reaching the micromolar range, which is considered to be a toxic level (Hyrc et al., 1997; Stout and Reynolds, 1999). Therefore, it is possible that  $[\text{Ca}^{2+}]_i$  does in fact reach micromolar levels following traumatic injury. Despite the absolute levels achieved,  $[\text{Ca}^{2+}]_i$  returns to basal levels within a few hours after injury in cortical neurons and remains normal through 24 h except after very high magnitudes of stretch (Weber et al., 1999). A topic of considerable debate is whether a sustained level of  $[\text{Ca}^{2+}]_i$  is needed to cause neurotoxicity, or if a rapid, transient increase is enough to activate degradative enzymes and other damaging mechanisms, even if  $[\text{Ca}^{2+}]_i$  levels return to normal. A sustained elevation of  $[\text{Ca}^{2+}]_i$  would likely be detrimental to neurons, however it is also possible that an initial transient elevation of  $[\text{Ca}^{2+}]_i$  is sufficient to cause neuronal death. Some cells also exhibit a delayed secondary increase in  $[\text{Ca}^{2+}]_i$  levels after glutamate exposure, which represents a loss of the ability to regulate  $\text{Ca}^{2+}$  levels, leading to cell death (Glaum et al., 1990; DeCoster et al., 1992; Randall and Thayer, 1992; Tymianski et al., 1993a; Brustovetsky et al., 2010; Brittain et al., 2012). In a few studies, removal of extracellular  $\text{Ca}^{2+}$  eliminated the delayed  $\text{Ca}^{2+}$  elevation but neurons still died (Tymianski et al., 1993a,b). Therefore, the secondary  $\text{Ca}^{2+}$  rise does not appear to cause neuronal death, suggesting that neurodegenerative processes may have been initiated with the first  $\text{Ca}^{2+}$  rise. These findings implicate that an initial  $[\text{Ca}^{2+}]_i$  rise may trigger pathways of neuronal death following TBI. However, the effects of  $\text{Ca}^{2+}$  on cell death may also be injury dependent, e.g., Lusardi et al. (2004), found that mechanically injured hippocampal cells exhibited increased death in  $\text{Ca}^{2+}$ -free media at low levels of stretch. In a few recent studies (Deshpande et al., 2008; Sun et al., 2008),  $[\text{Ca}^{2+}]_i$  was measured in hippocampal neurons that were isolated from rats that had received a moderate TBI *in vivo*. When measured at 24 h or 7 days after injury,  $[\text{Ca}^{2+}]_i$  levels were elevated even in the absence of cell death,

which suggests that at least some cells may be able to tolerate long-lasting elevations in  $[\text{Ca}^{2+}]_i$ . Therefore, based on all of the current evidence it is fairly evident that there is no strong correlation with absolute  $[\text{Ca}^{2+}]_i$  levels in neurons and resulting cell death.

Another idea related to  $\text{Ca}^{2+}$ -mediated neurodegeneration is that this process is regulated through specific  $\text{Ca}^{2+}$  influx signaling pathways (Sattler and Tymianski, 2000). This hypothesis is based on the fact that a multitude of data supports the role of specific  $\text{Ca}^{2+}$  influx pathways in regulating normal physiological processes. Therefore, therapeutic strategies aimed at completely blocking  $\text{Ca}^{2+}$  load may cause alterations to essential physiological processes, even in an injured brain. For example, although antagonists of VGCCs and glutamate receptors have shown some efficacy in experimental TBI (Tymianski and Tator, 1996; Obrenovitch and Urenjak, 1997), in the clinic, these drugs have not been successful (Reinert and Bullock, 1999; Ikonomidou and Turski, 2002; Schouten, 2007). Sattler et al. (1998) demonstrated that equivalent  $\text{Ca}^{2+}$  loads through VGCCs and NMDA receptors do not produce equal toxicity, as influx through NMDA receptors caused neurotoxicity while influx through VGCCs was not toxic. In addition, glutamate-mediated neurotoxicity appears to be due to increases in  $[\text{Ca}^{2+}]_i$  in microdomains located near NMDA receptors (Tymianski et al., 1993a, 1994). Further studies were conducted to specifically target  $\text{Ca}^{2+}$  toxicity mediated by NMDA receptors. These receptors are coupled to intracellular signaling molecules through a post-synaptic density protein (PSD-95; Sattler et al., 1999). Specifically, PSD-95 couples NMDA receptors to neuronal NOS (nNOS), which produces NO subsequently leading to neurotoxicity. By uncoupling this pathway, toxicity to cortical neurons triggered through NMDA receptors was significantly attenuated, while normal function of NMDA receptors remained intact (Sattler et al., 1999). When extended to a model of cerebral ischemia, protection from hypoxic damage was elicited when NMDA receptor-PSD-95 interactions were disrupted as late as 1 h after the insult (Aarts et al., 2002). These findings may translate to TBI as well, especially if there is a hypoxic component involved. In another recent study, Shahlaie et al. (2010) showed that blockade of N-type  $\text{Ca}^{2+}$  channels specifically was neuroprotective *in vitro*. Therefore, although VGCC blockade and glutamate receptor antagonism in general has not been useful therapeutically, it is possible that more specific targeting of  $\text{Ca}^{2+}$  channels could prove to be a viable means of clinical neuroprotection after TBI.

Another source specificity hypothesis is that  $\text{Ca}^{2+}$  released specifically from intracellular stores on the ER is toxic to neurons, and a fair amount of data supports this notion. Release of  $\text{Ca}^{2+}$  from internal stores causes apoptosis in developing cerebellar granule neurons (Levick et al., 1995), and depletion of  $\text{Ca}^{2+}$  stores is toxic to neuronal cell lines (Nguyen et al., 2002). The injurious effects of  $\text{Ca}^{2+}$  in traumatic injury of spinal cord white matter are mediated specifically through RyR and  $\text{IP}_3$ -mediated  $\text{Ca}^{2+}$  release (Thorell et al., 2002). Recently, Staal et al. (2010) found that a release of  $\text{Ca}^{2+}$  from intracellular stores was associated with degeneration of mechanically injured axons. Pharmacological studies have indicated that dantrolene, which blocks RyR-mediated  $\text{Ca}^{2+}$  release, is neuroprotective against various insults. For example, dantrolene is protective against neuronal loss in the hippocampus after *in vivo* ischemia (Wei and Perry, 1996;

Nakayama et al., 2002). Blocking the release of  $\text{Ca}^{2+}$  from stores can prevent glutamate and NMDA-induced neurotoxicity even though  $[\text{Ca}^{2+}]_i$  is elevated (Frandsen and Schousboe, 1991; Lei et al., 1992). Traumatically injured hippocampal neurons *in vitro* are also protected by dantrolene, independent of  $\text{Ca}^{2+}$  level (Yoon et al., 1996). In addition to these experimental findings, low molecular weight heparin, an antagonist of  $\text{IP}_3$ -mediated  $\text{Ca}^{2+}$  release, has been demonstrated to be neuroprotective in stroke patients (Jonas et al., 1997). Lastly,  $\text{IP}_3$ -induced  $\text{Ca}^{2+}$  mobilization causes a corresponding increase in mitochondrial  $\text{Ca}^{2+}$  concentration (Rizzuto et al., 1993); therefore, an important interplay seems to exist between these  $\text{Ca}^{2+}$  storage mechanisms. Also, NMDA receptor activation can also lead to a significant amount of  $\text{Ca}^{2+}$  release from intracellular stores (Mody and MacDonald, 1995). These findings suggest that it is unlikely that a single source of  $\text{Ca}^{2+}$  is responsible for neurotoxicity, and that several signaling pathways may contribute to neuronal dysfunction following TBI.

### ALTERATIONS TO $\text{Ca}^{2+}$ -MEDIATED SIGNAL TRANSDUCTION PATHWAYS

There is strong evidence in support of the concept that abnormal neurotransmission and  $\text{Ca}^{2+}$ -mediated signal transduction pathways could be responsible for neuronal dysfunction following trauma. After moderate TBI *in vivo*, extracellular glutamate is cleared within 5 min, but antagonists of glutamate receptors and presynaptic glutamate release inhibitors are efficacious even if administered 30 min after the insult (Obrenovitch and Urenjak, 1997). This finding suggests that immediate release of glutamate and over-activation of glutamatergic receptors alone may not be responsible for cellular damage after TBI, and that alterations in glutamatergic neurotransmission persist after injury. A possible abnormality of neurotransmission at glutamate receptors is a deficient  $\text{Mg}^{2+}$  blockade of  $\text{Ca}^{2+}$ -permeable NMDA receptors. Using an *in vitro* model of stretch-induced injury (Ellis et al., 1995), a reduction in the voltage-dependent  $\text{Mg}^{2+}$  blockade of the NMDA current was discovered in cortical neurons (Zhang et al., 1996). An enhanced elevation of  $[\text{Ca}^{2+}]_i$  in response to NMDA or glutamate is also observed after injury, even if basal  $[\text{Ca}^{2+}]_i$  levels are above normal (Weber et al., 1999). Although the response to these agonists appeared to recover to control levels after injury, 24 h post-injury some of these cells again display enhanced glutamate-mediated responses, suggesting a biphasic enhanced response to glutamate.

Signaling through G protein-coupled receptors that elevate  $\text{Ca}^{2+}$  by  $\text{IP}_3$  production also appears to be perturbed following TBI *in vivo* (Delahunty et al., 1995) and mechanical injury *in vitro* (Rzagalinski et al., 1998; Weber et al., 1999, 2001). However, unlike ionotropic glutamate receptors, immediately after stretch injury  $\text{Ca}^{2+}$  elevation mediated by mGluRs is actually decreased upon agonist stimulation. When measured at later time points (3 or 24 h post-injury), the mGluR-mediated  $[\text{Ca}^{2+}]_i$  response is potentiated. mGluRs appear to contribute to enhanced NMDA current as modulation of these receptors can attenuate enhanced NMDA currents after injury (Lea et al., 2002). In addition, downstream activation of PKC also contributes to the enhanced NMDA response (Zhang et al., 1996). Signaling through muscarinic ACh receptors is also disrupted following mechanical stretch (Weber

et al., 2002), and many neurons that have low  $[\text{Ca}^{2+}]_i$  levels after injury are still unresponsive to  $\text{M}_1$  receptor agonists, suggesting an uncoupling of signal transduction mechanisms, possibly at the receptor or G protein level. Another interesting finding is that differential levels of  $[\text{Ca}^{2+}]_i$  and responses to muscarinic receptor agonists were observed even within the same culture well of neurons after injury. Similar to these findings, Goforth et al. (1999) showed a decreased desensitization of the AMPA receptor subtype of glutamate receptors in a subpopulation of cortical neurons following stretch injury. A subsequent study showed that this reduced desensitization was mediated through NMDA receptors and activation of  $\text{Ca}^{2+}$  calmodulin-dependent protein kinase II (CaMKII; Goforth et al., 2004), which is generally elevated after TBI (Folkerts et al., 2007; Zhang et al., 2012). Interestingly, using another *in vitro* injury model, cortical neurons display an injury-induced increase in  $\text{Ca}^{2+}$ -permeable AMPA receptors, which is mediated by CaMKII $\alpha$  phosphorylation (Spaethling et al., 2008). This form of AMPA receptor upregulation could lead to a greater  $\text{Ca}^{2+}$  influx into cells upon activation by glutamate, and would thus be a form of altered  $\text{Ca}^{2+}$  signaling.

Disrupted mGluR-mediated  $\text{Ca}^{2+}$  signaling also involves alterations in intracellular  $\text{Ca}^{2+}$  stores. The status of intracellular  $\text{Ca}^{2+}$  stores in neurons can be determined by using SERCA inhibitors, such as thapsigargin, which causes a slow leak of  $\text{Ca}^{2+}$  from the stores, resulting in an elevation of  $[\text{Ca}^{2+}]_i$  (Thastrup et al., 1989). Fifteen minutes after stretch-induced injury, thapsigargin-stimulated elevation of  $[\text{Ca}^{2+}]_i$  is abolished in cortical neurons, suggesting that intracellular  $\text{Ca}^{2+}$  stores are depleted (Weber et al., 1999). This finding may be due to increased levels of  $\text{IP}_3$ , as has been demonstrated in astrocytes following stretch injury (Floyd et al., 2001). Also, inhibition of PLC maintains  $\text{Ca}^{2+}$  replete stores after injury (Weber et al., 2001), as does antagonism of Group I mGluRs (Chen et al., 2004). Taken together, these results suggest that elevated levels of  $\text{IP}_3$  may maintain  $\text{Ca}^{2+}$  stores in an empty state in the early period after injury. Also contributing to store depletion is the possibility of enhanced CICR. After stretch injury, the number of neurons that respond to caffeine, which stimulates CICR, is dramatically reduced (Weber et al., 2002).

When measured at later time points following stretch-induced injury (3 and 24 h), mGluR and thapsigargin-stimulated elevation of  $[\text{Ca}^{2+}]_i$  is potentiated, a response which appears to be due to an enhancement of CCI (Weber et al., 2001). As earlier described, depletion of intracellular  $\text{Ca}^{2+}$  stores leads to further influx of extracellular  $\text{Ca}^{2+}$  into cells through SOCs/SMOCs on the plasma membrane. The exact mechanism for the activation of CCI is still a topic of considerable debate. The “conformational coupling” hypothesis as first proposed by Irvine (1990) suggests a direct physical connection between the  $\text{IP}_3$  receptor and plasma membrane channels which controls CCI. When the  $\text{IP}_3$  receptor is activated, it is proposed that an alteration in the physical coupling causes influx of  $\text{Ca}^{2+}$  into the cell via plasma membrane channels. An alternate hypothesis suggests that when  $\text{IP}_3$ -sensitive stores are activated and depleted of  $\text{Ca}^{2+}$ , a diffusible messenger termed “calcium influx factor” (CIF) is released from the stores and binds to SOCs/SMOCs on the plasma membrane and activates CCI (Randriamampita and Tsien, 1993; Rzagalinski et al., 1999). There are several findings in support of a physical coupling and also the possible involvement



of a secretory mechanism in CCI (Kiselyov et al., 1998, 1999; Paterson et al., 1999; Yao et al., 1999), however there is also evidence of a soluble CIF that may be produced from the release of AA from phospholipid stores by cPLA<sub>2</sub> and subsequent cytochrome P450 metabolism of AA (Rzizgalinski et al., 1999). Using the SOC/SMOC antagonist SKF 96365, the potentiation in thapsigargin-stimulated elevation of  $[Ca^{2+}]_i$  at 3 h post-injury in neurons is blocked, a result that mirrored findings in  $Ca^{2+}$ -free conditions (Weber et al., 2001). This delayed enhancement in store-mediated signaling could have detrimental effects on neuronal physiology and may be due to alterations in CIF production, the sensitivity of SOCs/SMOCs to CIF, a damaged physical connection of the stores with the plasma membrane, or to a combination of these factors. As previously mentioned, SOC/SMOCs are likely members of the TRPC family, and they contribute to cell death following excitotoxic insults (Aarts and Tymianski, 2005). These channels therefore represent potential future therapeutic targets.

$Ca^{2+}$ -mediated intercellular signaling may also be perturbed after traumatic neuronal injury. Neurons in pure culture often display spontaneous oscillations in  $[Ca^{2+}]_i$  that are generally considered to be indicative of synaptic connectivity and intercellular communication (Wang and Gruenstein, 1997). Immediately after stretch injury, oscillations in  $[Ca^{2+}]_i$  were inhibited in pure neuronal cultures (unpublished data), therefore injury may inhibit intercellular communication in neurons. This effect was later observed in neurons grown in the presence of glia, where the amplitude of neuronal oscillations was decreased following injury (Goforth et al., 2011). Interestingly, Geddes-Klein et al. (2006) showed that pharmacological induction of neuronal oscillations with bicuculline (a GABA<sub>A</sub> receptor antagonist) resulted in a lower  $[Ca^{2+}]_i$  transient induced by stretch, which conferred protection. It is known that IP<sub>3</sub>-sensitive stores play an important role in intercellular communication between neurons, but also between neurons and glia (Berridge, 1993; Sanderson, 1996; Winder and Conn, 1996). Intercellular waves appear to be mediated by the diffusion of IP<sub>3</sub>, thereby providing a mechanism of multicellular cooperativity (Sanderson, 1996). In relationship to this, inositol polyphosphates are released from injured astrocytes into the extracellular medium, suggesting that release of IP<sub>3</sub> from glial cells could affect neighboring neurons (Floyd et al., 2001). Gap junctions may also play a role in the injured brain, e.g., gap junctions can increase the vulnerability of hippocampal neurons to *in vitro* trauma (Frantseva et al., 2002). In a finding opposite to this, blocking gap junctions can increase glutamate-induced toxicity in glial-neuron co-cultures from mouse cortices (Ozog et al., 2002). Therefore, the exact role of gap junctions in contributing to secondary injury appears to be complex.

### PHYSIOLOGICAL CONSEQUENCES OF ALTERED SIGNAL TRANSDUCTION MECHANISMS

It is not conclusively known whether neurons displaying altered  $Ca^{2+}$  signaling in the early post-traumatic period eventually die after injury, while those that display or have recovered normal signaling survive. It is likely that many cells that survive TBI have a prolonged alteration in their physiology, and this may contribute to functional impairment after a traumatic insult (Cohen et al., 2007). Recent evidence supports this notion, e.g., Witgen et al.

(2005) demonstrated alterations in inhibitory synaptic transmission in the hippocampus after TBI *in vivo*. As previously discussed, Sun et al. (2008) and Deshpande et al. (2008) measured  $[Ca^{2+}]_i$  levels in hippocampal neurons at 1, 7, or 30 days after fluid percussion injury in rats.  $[Ca^{2+}]_i$  levels were elevated at 1 and 7 days post-injury, and these animals also displayed cognitive deficits on these days as measured by Morris water maze performance. At 30 days post-injury  $[Ca^{2+}]_i$  levels were no longer significantly elevated and cognitive function had also returned to normal. This is strong evidence of a correlation between  $Ca^{2+}$  homeostasis and cognitive function. Extended disruption of intracellular signal transduction could alter the normal response to neurotransmitters such as glutamate and ACh, which may have detrimental consequences for cells due to their importance in vital functions such as the control of neuronal membrane excitability, intercellular communication, and further neurotransmitter release. Injury of cortical neurons *in vitro* has shown that synaptic transmission is in fact depressed at 4 h and 2 days post-injury (Goforth et al., 2011), which could have long-term effects on plasticity.  $[Ca^{2+}]_i$  signaling also plays a pivotal role in the maintenance of cytoskeletal integrity in neurons, so a sustained alteration in neuronal physiology may affect spine morphology, which could have profound effects on synaptic communication (Slemmer et al., 2005). Consistent with this idea is the finding that cerebellar cells exhibit synaptic alterations after TBI *in vivo* (Ai et al., 2007).

Specific disruption of  $Ca^{2+}$  signaling through intracellular stores may also contribute to abnormal neuronal function, as depletion of IP<sub>3</sub>-sensitive  $Ca^{2+}$  stores and inhibition of IP<sub>3</sub>-mediated signaling can inhibit neurite extension and axon guidance (Takei et al., 1998). Thus, interference with IP<sub>3</sub>-mediated signaling may inhibit neuronal recovery and plasticity after injury. Group I mGluRs coupled to IP<sub>3</sub>-sensitive stores are reported to modulate VGCCs in cortical neurons, thereby contributing to the control of neuronal membrane excitability (Choi and Lovinger, 1996). IP<sub>3</sub>-sensitive  $Ca^{2+}$  stores also play a role in modulating pre- and post-synaptic neurotransmission. Group I mGluR are reported to both amplify (Bruno et al., 1995; Rahman and Neuman, 1996; Pisani et al., 2001) and attenuate (Yu et al., 1997) elevation of  $[Ca^{2+}]_i$  elicited by NMDA receptors. IP<sub>3</sub>-linked receptors also contribute to the modulation of neurotransmitter release (Ozawa et al., 1998). The physiological functions of CICR in neurons are not as well understood, however, evidence suggests that quantal release of  $Ca^{2+}$  from ryanodine-sensitive stores may contribute to  $Ca^{2+}$  oscillations or waves in neurons (Bootman and Berridge, 1995; Verkhratsky and Shmigol, 1996). These mechanisms may serve as spatial or temporal signals contributing to physiological functions within cells, or they may contribute to intercellular communication. CICR may also facilitate the release of neurotransmitters in hippocampal neurons (Savic and Sciancalepore, 1998; Sharma and Vijayaraghavan, 2003). Therefore, altered  $Ca^{2+}$  store-mediated signaling could have adverse effects on neuronal function.

In addition to altered signal transduction events mediated by release of  $Ca^{2+}$  from internal stores, a low level of  $Ca^{2+}$  in the ER can have further debilitating effects at the subcellular level. Maintenance of  $Ca^{2+}$  homeostasis is important for normal processing of proteins, and  $Ca^{2+}$  store depletion can result in decreased protein

synthesis (Doutheil et al., 1997). A low level of  $\text{Ca}^{2+}$  in the ER lumen can also cause misfolded proteins and disaggregation of ribosomes (Doutheil et al., 1997; Lehotsky et al., 2003). These observations are also documented during ischemia, which causes disrupted ER  $\text{Ca}^{2+}$  homeostasis (Paschen and Doutheil, 1999; Paschen and Frandsen, 2001; DeGracia et al., 2002). Ischemia-induced alterations in  $\text{Ca}^{2+}$  store homeostasis also leads to the induction of a cellular stress response, in which several stress-related genes become activated (Paschen and Doutheil, 1999; DeGracia et al., 2002). It is thus possible that documented changes in intracellular stores after traumatic neuronal injury initiate a similar response, leading to dysfunctional protein synthesis after TBI.

$\text{Ca}^{2+}$  is believed to play a major role in activity-dependent changes in synaptic strength in neurons, thought to represent the underlying cellular mechanisms of learning and memory (Lamont and Weber, 2012; also see Kawamoto et al., 2012). An increase in  $[\text{Ca}^{2+}]_i$  in neuronal dendrites through NMDA channels is the critical trigger in the most intensively studied form of synaptic plasticity, hippocampal long-term potentiation (LTP; Bliss and Collingridge, 1993; Malenka and Nicoll, 1999). A rise in cytoplasmic  $\text{Ca}^{2+}$  is also required for other types of synaptic plasticity, for example long-term depression (LTD) in the hippocampus (Bear and Abraham, 1996; Reyes and Stanton, 1996), and LTD in the cerebellum, at both parallel fiber (PF-LTD) and climbing fiber (CF-LTD) synapses with Purkinje cells (Hansel and Linden, 2000; Hansel et al., 2001). In addition, CF-LTD is associated with a long-term reduction in calcium transients (Weber et al., 2003), and low levels of  $\text{Ca}^{2+}$  influx can result in PF-LTP (Coemans et al., 2004). Given the important role of  $\text{Ca}^{2+}$  in synaptic plasticity, it is logical to assume that altered  $\text{Ca}^{2+}$  homeostasis following traumatic neuronal injury could contribute to changes in the ability to induce LTP or LTD, which could underlie memory impairments following trauma (Cohen et al., 2007).

Because of the importance of the hippocampus in memory formation, the majority of research aimed at evaluating synaptic plasticity after TBI has been conducted in this brain region (Albensi, 2001). Several studies have reported an impairment in the induction of hippocampal LTP following experimental TBI *in vivo* (Miyazaki et al., 1992; Reeves et al., 1995; D'Ambrosio et al., 1998; Sick et al., 1998; Albensi et al., 2000; Sanders et al., 2000; Schwarzbach et al., 2006). However, the effects of TBI on hippocampal LTD are less clear, with some studies showing either no effect (D'Ambrosio et al., 1998) or an enhancement in LTD following injury (Albensi et al., 2000). The underlying cellular mechanisms by which different forms of synaptic plasticity are altered following TBI are not understood. It is possible that perturbed  $\text{Ca}^{2+}$  signaling contributes to these alterations. For example, the release of  $\text{Ca}^{2+}$  from internal stores contributes to

synaptic plasticity in the hippocampus and cerebellum (Svoboda and Mainen, 1999; Mattson et al., 2000; Rose and Konnerth, 2001), thus altered  $\text{Ca}^{2+}$  store-mediated signaling could impair the ability to induce LTP or LTD. Blockade of CCI has been shown to attenuate hippocampal LTP (Baba et al., 2003). In hippocampal neurons, CCI may directly activate CaMKII (Scholz and Palfrey, 1998), which plays a vital role in the induction of LTP. Therefore, altered CCI may also contribute to changes in synaptic plasticity after injury. Alterations in synaptic plasticity following TBI could have long-lasting effects in the brain, which may be manifested at the behavioral level. Accordingly, a further understanding of the underlying mechanisms that contribute to injury-induced changes in hippocampal plasticity is warranted, as well as additional studies on the effects of TBI on synaptic plasticity in other brain areas, such as the cerebellum and cortex.

## CONCLUSION

The relationship of the calcium ion to neurotoxicity has been intensely studied in the past decades. Originally, it appeared that a high  $\text{Ca}^{2+}$  load within neurons was solely responsible for cell damage and death following insults such as ischemia, excitotoxicity, and TBI. Therefore, therapeutic strategies were aimed at inhibiting  $\text{Ca}^{2+}$  influx into cells. However, the failure of VGCC and glutamate receptor antagonists in the clinical setting demonstrates that treatment strategies need to be redesigned. Research in recent years demonstrates more clearly that altered neuronal  $\text{Ca}^{2+}$  homeostasis, rather than absolute intracellular  $\text{Ca}^{2+}$  levels *per se*, may lead to perturbed cellular physiology and neuronal death. Although an elevated level of intracellular  $\text{Ca}^{2+}$  can still initiate destructive pathways, this probably occurs at very defined regions within neurons, and the severity of TBI is also likely important in determining the extent of  $\text{Ca}^{2+}$ -dependent cell death and altered physiology. How do we translate this recently gained knowledge into viable new therapeutic strategies for TBI patients? Drugs aimed at specific intracellular  $\text{Ca}^{2+}$  signaling pathways or subcellular components show some promise for the future treatment of TBI, as well as other disease states of the brain such as neurodegenerative disorders. These agents may be particularly useful for mild TBI, i.e., concussion, where altered physiology in surviving cells may be a more prominent portion of pathology than cell death. The goal of future therapeutic agents should not only be to inhibit cell death, but to establish normal neuronal physiology in cells that do not die following a traumatic episode.

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