

Wainwright, C.L., McCabe, C. and Kane, K.A. (2005) *Endothelin and the ischaemic heart*. Current Vascular Pharmacology, 3 (4). pp. 333-341. ISSN 1570-1611.

http://eprints.gla.ac.uk/16851

Deposited on: 25 July 2011

# Endothelin and the ischaemic heart

<sup>1</sup>Cherry L Wainwright, <sup>3</sup>Christopher McCabe and <sup>2</sup>Kathleen A Kane

<sup>1</sup>School of Pharmacy, The Robert Gordon University, Aberdeen, UK, <sup>2</sup>Department of Physiology & Pharmacology, University of Strathclyde, Glasgow, UK and <sup>3</sup>Division of Clinical Neurosciences, Wellcome Surgical Institute, University of Glasgow, Glasgow, UK.

# **Author for Correspondence:**

Professor Cherry L Wainwright School of Pharmacy The Robert Gordon University Schoolhill Aberdeen AB10 1FR Scotland UK.

Tel: +44 1224 262450

Fax: +44 1224 262555

E-mail: <u>c.wainwright@rgu.ac.uk</u>

#### Introduction

It has been widely documented that plasma ET-1 and big ET-1 levels are increased in patients within a day of onset of acute myocardial infarction [1,2,3,4]. Similar changes in systemic plasma ET-1 are seen in animal models of myocardial ischaemia [5,6]. The source of the released ET-1 appears to be the area subjected to ischaemic damage, since in a study in anaesthetised pigs both short and long periods of left anterior descending (LAD) coronary artery occlusion were found to enhance overflow of ET-1 from the area of myocardium subjected to the ischaemia/reperfusion insult [7]. The myocardial tissue level of ET-1 has been found to be increased by 3 to 7 fold in the ischaemic area compared to the non-ischaemic myocardium, suggesting an increased biosynthesis of ET-1 during ischaemia/reperfusion [5,7]. Indeed, the demonstration of increased ET-1 mRNA levels in cardiomyocytes subjected to ischaemia [8], supports this notion that that the raised circulating levels of endothelin are not simply due to release of stored peptide. The observation that ischaemia and reperfusion increases <sup>125</sup>I-labelled ET-1 binding sites in rat cardiac membranes [9] further supports the view that the endothelin system is up-regulated during these events, although the observation that there is no difference in ET-1 mRNA levels between animals subjected to ischaemia only and hearts subjected to ischaemia followed by reperfusion, implies that it is ischaemia that stimulates enhanced ET-1 production. Taken together, these observations have led to the concept that endothelin may play an important role in determining the outcome of myocardial ischaemia/reperfusion.

Based on our knowledge that ET-1 is a potent vasoconstrictor peptide, early hypotheses focused on the role of ET-1 as a detrimental factor in the

ischaemic/reperfused heart. However, over the last decade or so, as our understanding of the cellular effects of endothelin has extended beyond that of vasoconstriction, evidence is emerging that endothelin may play a very complex role in the setting of the ischaemic heart, with the potential to both contribute to cellular injury and cellular repair. The aim of this review therefore is to provide an overview of the effects of endothelin on the cardiomyocyte, in the setting of myocardial ischaemia, and within this to consider not only its actions mediated through vasoconstriction, but also through its effects on ion channels, cellular integrity and inflammatory cells.

#### Contribution of endothelin to ischaemia/reperfusion-induced myocardial injury.

The possibility that the coronary vasoconstrictor effects of endothelin could play a role in the development of injury resulting from myocardial ischaemia arose following a number of demonstrations that intracoronary infusion of exogenous ET-1 reduces coronary blood flow by approximately 90% [10-12], resulting in marked myocardial ischaemia. Substantive evidence for a contributory role of *endogenous* ET-1 in infarct extension subsequently emanated from studies that demonstrated that a monoclonal antibody to ET-1 can reduce infarct size in rats following ischaemia/reperfusion [5]. The ensuing development of both selective ET<sub>A</sub> receptor and mixed ET<sub>A</sub>/ET<sub>B</sub> receptor antagonists led to the opportunity to demonstrate that blocking the actions of endogenous ET-1 prior to the onset of ischaemia resulted in preservation of the myocardium. Subsequently, there is now a very large literature to demonstrate that both ET<sub>A</sub>-selective (e.g. BQ123; [13]) and mixed ET<sub>A</sub>/ET<sub>B</sub> receptor antagonists (e.g. LU135252 and bosentan; [12, 14]) can reduce myocardial infarct size following ischaemia and reperfusion. Since ET<sub>B</sub> receptor blockade does not appear to be a

prerequisite to observe this effect, this led to the conclusion that  $ET_A$  receptors mediate the actions of endothelin that contribute to tissue injury.

Blockade of the intense coronary vasoconstriction induced by endothelin, which would contribute to the "no-reflow" phenomenon, can be considered a prime candidate for the mechanism underlying the protective effects of endothelin antagonists on cardiomyocyte integrity. However, there are other actions of endothelin that may also contribute to tissue injury. For example, ET-1 is known to activate polymorphonuclear leukocytes to generate reactive oxygen species [15,16], which have been identified as mediators of reperfusion injury, and studies with the ET<sub>A</sub> receptor antagonist LU135252 have shown that this agent reduces both myocardial necrosis and tissue myeloperoxidase activity in the ischaemia/reperfused heart [17]. Furthermore, the protective effects of ET-1 antagonists have been proposed to involve inhibition of ET-1-induced activation of phospholipase C and subsequent release of Ca<sup>2+</sup> [18], superoxide production [19] and suppression of NO production [20,21].

Although the majority of studies assessing the effects of ET-1 antagonists on infarct size demonstrate a cardioprotection, there are also numerous studies that report no beneficial effects in the setting of myocardial ischaemia. Although both bosentan [12] and BQ123 [13,22,23] have shown cardioprotective effects in various animal models, there are several studies using these agents that have failed to demonstrate protection against myocardial infarction [24,25]. Studies with other ET<sub>A</sub> selective (FR-139317 and PD-156707) antagonists have also shown either a reduction in [26,27] or no effect on [28] infarct size. There are several explanations for these conflicting findings.

First, the outcome may be dependent upon the animal model used, although this can not always explain the discrepancy since studies with the same antagonist in the same species do not always agree [29,30]. A second explanation could be the dose employed since there is evidence that high doses of  $ET_A$  receptor antagonists may block the low affinity  $ET_B$  binding site linked to nitric oxide release [31], which would abrogate any potentially protective contribution of nitric oxide production through activation of these receptors by endogenous endothelin, but not the high affinity  $ET_B$  binding site linked to vasoconstriction. A similar argument may apply to mixed  $ET_A/ET_B$  receptor antagonists, since they too appear to possess different affinities for two different  $ET_B$  receptor subtypes [32]. Neither of these explanations can account for all of the incongruities between the large number of studies using ET receptor antagonists against myocardial infarction, giving us an inkling that perhaps, in the setting of the ischaemic heart, the actions of ET-1 in myocardial survival cannot all be labelled as "bad".

# Cardioprotective effects of ET-1 mediated by the ET<sub>A</sub> receptor

Following on from the endothelin antagonist revolution, a small number of studies have emerged to demonstrate that, in the *in vivo* and *in vitro* rabbit heart, exogenous administration of ET-1 as a preconditioning stimulus can reduce myocardial infarct size [33,34], with the subsequent confirmation of this effect in the isolated rat heart [35]. These studies also provided evidence that this effect appeared to be achieved through ET<sub>A</sub> receptor-mediated activation of protein kinase C and K<sub>ATP</sub> channels, as has been suggested for ischaemic preconditioning [36]. This has recently been corroborated in an *in vivo* rat study [37] which provides evidence to support the hypothesis that it is activation of the mitochondrial K<sub>ATP</sub> channel (through the ET<sub>A</sub>

receptor; [38]) that is responsible for ET-1 induced cardioprotection. However, as we begin to understand ET-1 physiology a little more, there are several alternative paradigms that might explain ET<sub>A</sub> receptor mediated cardioprotection.

Inhibition of cardiac myocyte apoptosis.

There is growing evidence that apoptosis is associated with acute myocardial infarction, with apoptotic cardiomyocytes being found predominantly in the hypoperfused border area lying between the "normal" myocardium and the core of the infarct (reviewed in [39]). Moreover, hypoxia and re-introduction of oxygen in isolated cardiomyocytes has been shown to induce apoptosis [40,41]. Evidence for a pathophysiological significance of apoptosis emanates from studies that have demonstrated the ability of caspase inhibitors to reduce infarct size [42]. There is now increasing evidence to show that ET-1 can inhibit apoptosis in cardiomyocytes [43], as well as other cells, through a signalling mechanism that involves induction of GATA-4 [44], which is one of a number of DNA binding proteins responsible for regulating cell lineage differentiation, progenitor cell proliferation and organ morphogenesis [39]. ET-1 has been shown to induce phosphorylation of GATA-4 [44] in an atrial cell line, to increase the interaction between GATA-4 and NFATc in cardiomyocytes [45] and increase GATA-4 DNA-binding activity [44], all of which may contribute to its anti-apoptotic effect. Furthermore, PPARy has been implicated in the perturbation of the anti-apoptotic signalling pathway of ET-1, since activators of PPARγ inhibit the cardiomyocyte protection afforded by ET-1 [46]. Moreover, ET-1 has recently been shown to activate a protective pathway that is unique to cardiomyocytes, in addition to common pathways among other cell types [47], which provides an exciting avenue to explore for selective inhibition of apoptosis within the heart. Interestingly, ET-1 has been shown to act through stabilisation of the mitochondrial membrane to protect the cardiomyocyte from apoptosis, rather than through preservation of the mitochondrial respiratory chain [48], thus implying that its action is required prior to the induction of injury, rather than once injury has begun, Taking this evidence together, it would therefore be of great interest to elucidate whether or not inhibition of apoptosis represents a mechanism by which exogenous ET-1 improves cardiomyocyte survival during ischaemia/reperfusion.

#### Mast cell activation

While the traditional view regarding ET-1 release in the ischaemic heart is that it largely emanates from the vascular endothelium, we now know that it is generated by a range of cells, including granulation tissue and fibroblasts [49,50], at the site of infarction. One very early observation of an alternative source of ET-1 comes from studies by Ehrenreich et al [51], who demonstrated that mast cells synthesize and secrete ET-1. Furthermore, these cells were also seen to express ET<sub>A</sub> receptors. Since then, ET-1 has been shown to induce mast cell degranulation in the gut [52] and, more recently, in the rat heart [53]. Recent work from our own group has similarly demonstrated mast cell degranulation in response to ET-1 in the murine heart, with an associated increase in tryptase activity in the myocardium [54]. Mast cells are recognised as important sentinels of the innate immune response [55] and have long been implicated in the inflammatory events that occur after a myocardial infarction. The effects of ET-1 on mast cell degranulation appear to be dependent upon both the type of mast cell (e.g. bone-marrow-derived versus fetal skin-derived mast cells) [56] and on the microenvironment (reviewed in [57]) and, until very recently, a phenomenon that could not be easily explained. However, the recent study

by Maurer et al [58] has provided unique insight into what now appears to be an ingenious homeostatic mechanism by which ET-1 induced stimulation of mast cells results in protection against ET-1 mediated toxicity. In these studies, mast cell deficient mice administered ET-1 intraperitoneally fared much worse with respect to toxicity to ET-1 than wild type mice. Through an elegant series of experiments, the authors demonstrated that one of the mechanisms by which mast cell-mediated protection against ET-1 associated pathology is achieved is through a reduction in the concentration of ET-1 by release of proteolytic enzymes. This observation implies a "double-edged sword" role for endothelin, whereby not only is it responsible for inducing damage, but also for limiting its own effects. In the setting of myocardial ischaemia/reperfusion, therefore, this may offer a possible explanation for the cardioprotective effects of an agent that, upon its original discovery, was automatically labelled as a "bad guy".

# Cardioprotective effects mediated through the ET<sub>B</sub> receptor.

The above evidence implying a cardioprotective role for ET-1 via an ET<sub>A</sub> receptor mechanism is all well and good, but we should not overlook the potential role of the action of exogenous ET-1 at the ET<sub>B</sub> receptor. Based upon observations that short term ischaemia/reperfusion in the isolated rat heart results in a trend to an increase in ET<sub>B</sub>, but not ET<sub>A</sub> receptor, binding sites [59]) and an increase in mRNA expression for both ET<sub>A</sub> and ET<sub>B</sub> receptors [60], we recently undertook a series of studies to determine the effects of ET<sub>B</sub> receptor activation with the ET<sub>B</sub> receptor agonist sarafotoxin 6c (S6c) on myocardial infarct size and on ischaemia/reperfusion induced changes in ET<sub>A</sub> and ET<sub>B</sub> receptor mRNA levels in the *in vivo* rat heart. We demonstrated that while ischaemia/reperfusion resulted in a marked reduction in

mRNA for both receptor subtypes in the ischaemic (but not the normal) zone of the heart, pre-treatment with S6c significantly reduced myocardial infarct size concomitant with a preservation of ET<sub>B</sub> receptor mRNA [61]. Preservation of ET<sub>B</sub> receptors within the ischaemic zone could conceivably benefit the ischaemic heart in two ways. First, by preferentially preserving the ET<sub>B</sub> receptor; this would allow endogenously generated ET-1 to act at this receptor, which mediates physiological responses (such as vasodilatation) that could be categorised as protective rather than destructive, rather than inducing damaging effects through the ETA receptor. The second benefit could be achieved through increased clearance of ET-1 from the local environment, since this is largely achieved through the ET<sub>B</sub> receptor [62]. In addition to preserving ET<sub>B</sub> receptor mRNA, activation of ET<sub>B</sub> receptors prior to ischaemia could similarly activate a number of mechanisms that could afford protection to the heart, such as direct release of prostacyclin and/or nitric oxide, both of which are cardioprotective [63,64] or induction of a leukocyte-mediated oxidative stress response, which has similarly been linked to cardioprotection [65]. In support of the latter, we have previously demonstrated that S6c can increase the ex vivo generation of reactive oxygen species from leukocytes following in vivo administration [66].

Thus, the role of endothelin in the cellular outcome following myocardial ischaemia is far from straightforward and involves both cytotoxic and cytoprotective effects, providing various paradigms for therapeutic targets (summarised in Figure 1).

# Contribution of endothelin to ventricular arrhythmias.

Cellular injury is not the only adverse outcome in the ischaemic heart to which ET-1 is thought to contribute. The earliest studies aimed at determining the effects of intra-

coronary ET-1 in the normal heart all demonstrated in pigs [67], rats [68] and dogs [69] that there was marked arrhythmic activity associated with a powerful vasoconstriction that induced ischaemia. However, a subsequent study by Yorikane et al [70] provided the first implication that exogenous ET-1 may exert direct electrophysiological effects (prolongation of the action potential and induction of early after-depolarisations) that underlie its ability to induce arrhythmias. However, the effect of exogenous ET-1 on arrhythmias in the setting of myocardial ischaemia was not addressed until, in an early study from our group [71], we demonstrated that exogenous administration of ET-1 during in vivo myocardial ischaemia in rats substantially increased arrhythmia severity and that this could be abrogated by concomitant administration of the ET<sub>A</sub> antagonist BQ123, implying an action of ET-1 through the ET<sub>A</sub> receptor. The importance of the ET<sub>A</sub> receptor in arrhythmogenesis was subsequently corroborated in an in vitro study in rat hearts, whereby BQ-485 prevented ET-1-induced arrhythmias in normal, non-ischaemic hearts [72]. However, our study also demonstrated that BQ123 alone could also reduce ischaemia-induced arrhythmias, providing the first demonstration of a role for endogenous ET-1 in arrhythmogenesis in the ischaemic heart [71]. The potential for ET<sub>A</sub> antagonists to act as anti-arrhythmic agents as well as to reduce myocardial injury has subsequently been well examined and, overall, the evidence points to a potentially beneficial effect of these agents against ischaemic arrhythmias (for a thorough review of this topic see [73]), although this is by no means universal [74]. As with cellular injury, however, there remains some debate as to whether selective ET<sub>A</sub> receptor blockade is preferable to mixed ET<sub>A</sub>/ET<sub>B</sub> receptor inhibition, since some studies show that ET<sub>B</sub> receptor blockade with either of the ET<sub>B</sub> selective antagonists IRL-1038 and IRL-1025 has no effect on arrhythmias [75], thus precluding the need for mixed antagonism in this

setting, whereas others demonstrate an antiarrhythmic effect of ET<sub>B</sub> receptor blockade [76]. The antiarrhythmic effect of ET-1 antagonism has also been observed against high-glucose induced electrical instability (QT prolongation) through an action predominantly at the ET<sub>A</sub> receptor, since both an ET<sub>A</sub> selective (FR139317) and a mixed ET<sub>A</sub>/ET<sub>B</sub> (SB209670) antagonist, but not a selective ET<sub>B</sub> antagonist (BQ788), was able to block its effects [77]. As with all studies on arrhythmias, consideration should always be made of the possible direct effects of agents on cardiac electrophysiology independent of their action at endothelin receptors. Indeed, there is some evidence that, at least in the case of BQ123, such a direct effect on cardiac electrophysiology may, in part, contribute to its antiarrhythmic effects [78]. Notwithstanding this, while there is clear evidence for an antiarrhythmic effect of ET-1 antagonism, what remains highly controversial is whether the pro-arrhythmic effect of ET-1 itself is mediated through a direct electrophysiological effect or occurs as a consequence of its other actions, such as vasoconstriction, and warrants detailed review.

#### Mechanisms underlying the pro-arrhythmic effect of ET-1

Several in vivo studies have attempted to identify the mechanisms by which exogenous ET-1 induces a pro-arrhythmic effect. In view of ET-1's known coronary vasoconstrictor action leading to myocardial ischaemia, the in vivo electrophysiological actions of ET-1 have been compared to those of myocardial ischaemia. Such studies, all carried out in anaesthetised dogs, have concluded that ET-1 has a direct arrhythmogenic effect independent of its ability to cause myocardial ischaemia. The evidence from a study by Szabo et al. [79] was that ET-1, given directly into the coronary artery, induced arrhythmias without electrocardiographic or metabolic signs of ischaemia. Indeed, ET-1 significantly

increased left ventricular epicardial (LV<sub>epi</sub>) and right ventricular endocardial (RV<sub>endo</sub>) monophasic action potential duration (MAPD) with no change in the upstroke velocity. This was in direct contrast to coronary artery occlusion, which decreased MAPD and upstroke velocity in the ischaemic area [79]. In another study from the same group by Becker et al. [80], intra-coronary administration of ET-1 induced ventricular arrhythmias that were focal in nature, without changing local refractory periods, left ventricular conduction time or the overall activation pattern. This was also quite distinct from the changes induced by coronary artery occlusion, which resulted in local conduction delay, prolonged refractoriness in all layers and arrhythmias that were maintained by re-entrant mechanisms [80]. Low dose intracoronary ET-1 (at a concentration that did not cause a significant reduction in coronary flow) was also shown to prolong epicardial and endocardial MAPD [81]. An increase in the spatial dispersion of MAPD was demonstrated with the MAP close to the infusion site showing the most pronounced prolongation. Significant differences were observed between the lengthening of the right and left ventricular MAPD times and the lengthening of the epicardial and endocardial MAPD times. This increase in dispersion of MAPD is thought to play an important role in the pathogenesis of ventricular arrhythmias, albeit that this is in the formation of re-entry circuits. Support for ET<sub>A</sub> receptors mediating these proposed electrophysiological effects of ET-1 comes from studies with the ET<sub>A</sub> antagonist LU 135.252. In these experiments, LU 135.252, was found to be anti-arrhythmic and was able to inhibit the main electrophysiological actions of ET-1 in the anaesthetised dog, namely MAPD prolongation and early after depolarisation (EAD) formation, while being unable to prevent the vasoconstriction caused by ET-1 suggesting that these direct electrophysiological effects of ET-1 are mediated through the ET<sub>A</sub> receptor [82].

In contrast to these *in vivo* studies, our group have examined the electrophysiological effects of ET-1 and S6c, the ET<sub>B</sub> receptor specific agonist, in an isolated paced working rabbit heart model which is devoid of neural and humoral influences. Over a wide concentration range, ET-1 reduced MAPD<sub>90</sub> but did not increase endocardial or epicardial MAPD<sub>90</sub> or refractoriness as has been reported in vivo. Furthermore, the abbreviation of MAPD<sub>90</sub> was, at all concentrations, associated with a concomitant reduction in coronary flow, providing no evidence of any direct electrophysiological effect of ET-1. S6c lacked any marked electrophysiological effect in either the endoor the epicardium suggesting that the ET<sub>B</sub> receptor does not play a major role in any observed electrophysiological action in the ventricles [83; In Press]. further investigated whether ET-1 or S6c could modify the electrophysiological changes induced during myocardial ischeamia/reperfusion. As shown in Fig 2, ET-1 but not S6c, shortened MAPD<sub>90</sub> before coronary artery occlusion but did not cause further shortening than that induced during myocardial ischaemia. This data is not, therefore, in agreement with the *in vivo* studies in the dog and may suggest that an intact nervous system or the presence of blood is necessary to observe electrophysiological effects that are independent of myocardial ischaemia.

There are, however, also *in vitro* studies in isolated cardiac tissue or cells which support the view that ET-1 has direct electrophysiological effects on the ventricles. In isolated canine cardiac tissue ET-1 causes a prolongation of the action potential duration, measured at the plateau phase, i.e. APD<sub>50</sub> in the right bundle branch, Purkinje cells and ventricle with the prolongation being most marked in the right bundle branch [70]. This study also demonstrated the development of EADs during

the prolonged plateau phase after ET-1 administration. Nicardipine was shown to abolish these EADs, suggesting that the voltage sensitive calcium current is involved in their genesis [70]. It should be noted, however, that the concentrations of ET-1 used in this study were more than 10 fold higher than the concentration of ET-1 (10<sup>-10</sup>M) required to cause a significant decrease in coronary flow in our study in the isolated rabbit heart. Thus, in the intact heart it may not have been possible to observe this ET-1 induced prolongation in action potential because of the counteracting effect of myocardial ischaemia. In neonatal rat ventricular myocytes, ET-1 was found to increase Ca<sup>2+</sup> entry through the sarcolemmal T-type Ca<sup>2+</sup> channel, possibly through a pathway involving PKC [84]. An increase in L-type Ca<sup>2+</sup> current (I<sub>CAL</sub>), which would be expected to prolong the plateau phase of the action potential and its duration has also been reported in adult rat ventricular myocytes. This effect was found to be ET receptor mediated and involved a PKC mediated pathway [85].

In contrast to the studies that have shown that ET-1 can stimulate  $Ca^{2+}$  current there have been several studies reporting an inhibition of  $Ca^{2+}$  current or lack of effect on basal  $Ca^{2+}$  current. Thus, ET-1 administration decreased the L-type  $Ca^{2+}$  current in isolated canine ventricular myocytes [86,87], in guinea pig [88], rabbit [89] and human myocytes [90]. In guinea pig ventricular myocytes, ET-1 had little effect on basal L-type  $Ca^{2+}$  current but it did reduce this current enhanced by  $\beta_1$  adrenoceptor stimulation [91]. This anti-adrenergic effect of ET-1 on  $I_{CAL}$  has been reported also in canine ventricular myocytes [86]. Thus, there is controversy in the literature about the role of  $I_{CAL}$  in the observed prolongation of action potential duration by ET-1 with the majority of studies supporting the view that an increase in this current is unlikely to be the main underlying mechanism.

The delayed rectifier K<sup>+</sup> current (I<sub>K</sub>), is an important determinant of action potential duration, which has been demonstrated to be regulated by ET-1. ET-1 has been shown either to enhance or decrease I<sub>K</sub> in guinea pig ventricular myocytes [92,93], to decrease it in human myocytes [90] and to have no effect on this current in canine myocytes [86]. ET-1 has a more clear-out effect to inhibit an enhanced I<sub>K</sub>, following β<sub>1</sub>-adrenoceptor stimulation in a range of species [86,93]. This effect was mediated by the ET<sub>A</sub> receptor and through a PTX sensitive G-protein/Protein Kinase A (PKA) pathway [93]. ET-1 has also been shown to modulate the ATP sensitive K<sup>+</sup> channel (K<sub>ATP</sub>). In guinea pig ventricular cells ET-1 caused a partial inhibition of the K<sub>ATP</sub> current, which was abolished by BQ-485, an ET<sub>A</sub> receptor antagonist [94], suggesting involvement of the ET<sub>A</sub> receptor. In the same study, ET-1 had no effect on normal action potential duration (measured in papillary muscle), but, it partially reversed the cromakalim, a K<sub>ATP</sub> channel opener, induced shortening of duration. Overall, ET-1 effects on potassium currents are also controversial with no clear explanation of an ionic basis for a prolongation of normal action potential duration. pronounced anti-adrenergic effect of ET-1 on both calcium and potassium current augmented by  $\beta_1$ -adrenoceptor stimulation it is possible that ET-1 will exhibit electrophysiological effects that are seen in vivo but not in vitro. That these antiadrenergic effects would, in turn, be pro-arrhythmic is hard to reconcile with the known pro-arrhythmic actions of sympathetic stimulation.

There are other possible mechanisms by which ET-1 might act to alter cardiac electrophysiology and arrhythmogenicity. In both rat and cat atrial myocytes ET-1 was found to exhibit an arrhythmogenic effect, which was suppressed by the inositol

1,4,5 trisphosphate (IP3) receptor antagonist, aminoethoxydiphenyl borate (2-APB), suggesting that activation of IP3 may be involved through an increase in Ca<sup>2+</sup> release from the sarcoplasmic reticulum [95,96]. Activation of the cardiac Na<sup>+</sup>-Ca<sup>2+</sup> exchanger (reverse mode activation) has been shown in guinea pig ventricular myocytes following ET-1 administration, which was found to be through a PKC dependent mechanism [97]. Reverse mode activation of the Na<sup>+</sup>-Ca<sup>2+</sup> exchanger results in Ca<sup>2+</sup> influx and may underlie arrhythmias involving after-depolarisations. The arrhythmogenic effects of ET-1 may also involve activation of the Na+-H+ exchanger leading to acidosis since activation of the Na<sup>+</sup>-H<sup>+</sup> exchanger has been observed in rabbit ventricular myocytes following ET-1 administration [98]. Acidosis, in turn, results in Ca<sup>2+</sup> overload through the activation of the Na<sup>+</sup>-Ca<sup>2+</sup> exchanger. Moreover, Na<sup>+</sup>-H<sup>+</sup> exchanger activation has been linked with a shortening of the monophasic action potential duration in pigs following coronary artery occlusion [99]. It is noteworthy, however, that these direct electrophysiological effects of ET-1, with the exception of the studies on Na<sup>+</sup>-H<sup>+</sup> exchanger, have been observed with concentrations that are greater than those which cause coronary vasoconstriction raising the question of their relevance to ET's pro-arrhythmic action in vivo.

#### Evidence for an antiarrhythmic effect of endothelin

Since ET-1 can mimic ischaemic preconditioning against myocardial injury (as described above), and given that ischaemic preconditioning can also protect against ventricular arrhythmias [100], it would seem feasible that ET-1 may also induce an anti-arrhythmic effect when given exogenously. In the aforementioned studies demonstrating an infarct-reducing effect of ET-1 when given prior to the onset of a

period of ischaemia the effect of this exogenous ET-1 administration on consequent arrhythmias was not reported. Our group undertook a study to specifically address this question and observed that ET-1 can indeed reduce ischaemic arrhythmias in an in vivo rat model of ischaemia and reperfusion [76]. In those studies we were unable to elucidate the receptor responsible for mediating the antiarrhythmic effect of ET-1, due to inherent antiarrhythmic effects of antagonists masking any blockade of the antiarrhythmic action of ET-1. However, in a subsequent study we demonstrated that the ET<sub>B</sub> agonist S6c was also able to reduce the incidence of ischaemic arrhythmias [101], implying that the antiarrhythmic action of ET-1 is mediated through the ET<sub>B</sub> receptor, rather than through the ET<sub>A</sub> receptor. Although the mechanisms underlying the antiarrhythmic effects of ET-1 remain undetermined, we have observed that its effects are much less evident in the isolated heart compared to the in vivo heart, signifying that the mechanisms are likely to be more complex than a direct effect on the myocardium, the coronary blood vessels, or both [102]. In contrast, in the studies with S6c we found that this was able to protect equally well in both the *in vivo* and *in* vitro heart. Taken together this suggests that the antiarrhythmic effect of ET-1 is no less complex than the cytoprotective effects, probably involving a number of mechanisms. However, more studies are required to fully elucidate this protective effect of endothelin.

#### **Concluding Comments**

As implied in the introduction to this review, the role of ET-1 in the setting of the ischaemic heart is much more complex than originally perceived. Clearly there are benefits of blocking the actions of ET-1, yet the strong evidence now pointing to protective effects of this peptide tells us that we need to resolve a number of issues

before the most appropriate form of anti-endothelin therapy can be developed. For example, can we block the injurious facet of ET-1's action on cardiomyocyte integrity, while at the same time harness its ability to contribute to tissue repair or change the homeostatic balance of the levels of the peptide in favour of tissue survival? Furthermore, with respect to the pro-arrhythmic effects of endothelin, will we ever be able to resolve the argument surrounding the direct versus indirect action of ET-1, or is the combined physiology of the ischaemic heart and of ET-1 too intricate to dissect out completely? If we can answer these questions then we should have a very powerful tool available to us to modulate ET-1 in the ischaemic heart for the best possible outcome.

# Acknowledgement

The authors are grateful to the British Heart Foundation for their support of studies into the effects of endothelin in the ischaemic heart.

#### References

- 1. Miyauchi T, Yanagisawa M, Tomizawa T et al. Increased plasma concentrations of endothelin-1 and big endothelin-1 in acute myocardial infarction. Lancet 1989; 2: 53-54.
- 2. Yasuda M, Kohno M, Tahara A et al. Circulating immunoreactive endothelin in ischaemic heart disease. Am Heart J 1990; 119: 801-806.
- 3. Naruse M, Naruse K, Kurimoto F et al. Radioimmunoassay for endothelin and immunoreactive endothelin in culture medium of bovine endothelial cells. Biochem Biophys Res Comm 1989; 160: 662-668.
- 4. Omland T, Lie RT, Aakvaag A, Aarsland T, Dickstein K. Plasma endothelin determination as a prognostic indicator of 1-year mortality after acute myocardial infarction. Circulation 1994; 89: 1573-1579.
- 5. Watanabe T, Suzuki N, Shimamoto N, Fujino M & Imada A. Contribution of endogenous endothelin to the extension of myocardial infarct size in rats. Circ Res 1991; 69: 370-371.
- 6. Tsuji S, Sawamura A, Watanabe H, Takihara K, Park SE, Azuma J. Plasma endothelin levels during myocardial ischaemia and reperfusion. Life Sci 1991; 48: 1745-1749.
- 7. Wang Q-D, Hemsen A, Li XS, Lundberg JM, Uriuda Y, Pernow J. Local overflow and enhanced tissue content of endothelin following myocardial ischaemia and reperfusion in the pig: modulation by L-arginine. Cardiovasc Res 1995; 29: 44-49.
- 8. Tonnessen T, Giaig A, Saleh D, Naess PA, Yanagisawa M, Christensen G. Increased in vivo expression and production of endothelin-1 by porcine cardiomyocytes subjected to ischaemia. Circ Res 1995; 76: 767-772.
- 9. Liu Y, Chen R, Casley DJ, Nayler WG. Ischemia and reperfusion increase 125I-labeled endothelin-1 binding in rat cardiac membranes. Am J Physiol 1990; 258 (3 Part 2): H829-H835.

- 10. Kurihara H, Yamaoki K, Nagai R, et al. Endothelin: a potent vasoconstrictor associated with vasospasm. Life Sci 1989; 44: 1937-1943.
- 11. Kurihara H, Yoshizumi M, Sugiyama T et al. The possible role of endothelin-1 in the pathogenesis of coronary vasospasm. J Cardiovasc Pharmacol 1989; 13 (Suppl 5): S132-S137.
- 12. Wang Q-D, Li X-S, Lundberg JM, Pernow J. Protective effects of non-peptide endothelin receptor antagonist bosentan on myocardial ischaemic amnd reperfusion injury in the pig. Cardiovasc Res 1995; 29: 805-812.
- 13. Grover GJ, Dzwonczynk S, Parham CS. The endothelin-1 receptor antagonist BQ123 reduces infarct size in a canine model of coronary occlusion and reperfusion. Cardiovasc Res 1993; 27: 1613-1618.
- 14. Gonon AT, Wang QD, Shimizu M, Pernow J. The novel non-peptide selective endothelin A receptor antagonist LU 135,252 protects against myocardial ischaemic ajnd reperfusion injury in the pig. Acta Physiol Scand 1998; 163: 131-137.
- 15. Ishida K, Takeshige K, Minakami S. Endothelin-1 enhances superoxide generation of human neutrophils stimulated by the chemotactic peptide N-formyl-mthionyl-leucyl-phenylalanine. Biochem Biophys Res Comm 1990; 173: 496-500.
- Lopez Farre A, Riesco A, Espinosa G et al. Effect of endothelin-1 on neutrophil adhesion to endothelial cells and perfused heart. Circulation 1993; 88: 1166-1171.
- 17. Gonon AT, Wang QD, Pernow J. The endothelin A receptor antagonist LU 135252 protects the myocardium from neutrophil injury during ischaemia/reperfusion. Cardiovasc Res 1998; 39: 674-682.
- 18. Pernow J, Wang Q-D. Endothelin in myocardial ischaemia and reperfusion. Cardiovasc Res 1997; 33: 518-526.

- 19. Maczewski M, Beresewicz A. The role of endothelin, protein kinase C and free radicals in the mechanism of post-ischaemic endothelial dysfunction in guinea pig hearts. J Mol Cell Cardiol 2000; 32: 297-310.
- Gonon AT, Gourine AV, Pernow J. Cardioprotection from ischaemia and reperfusion injury by an endothelin A-receptor antagonist in relation to nitric oxide production. J Cardiovasc Pharmacol 2000; 36: 405-412.
- 21. Gourine A, Gonon A, Pernow J. Involvement of nitric oxide in cardioprotective effect of endothelin receptor antagonist during ischaemia-reperfusion. Am J Physiol 2001; 280: H1105-H1112.
- 22. Han H, Neubauer S, Braeker B, Ertl G. ET1 contributes to ischaemia-reperfusion injury in isolated rat heart: attenuation of ischaemic-injury by the endothelin-1 antagonists BQ123 and BQ-610. J Mol Cell Cardiol 1995; 27: 761-766.
- 23. Illing B, Horn M, Han H et al. Protective effect of the specific endothelin-1 antagonist BQ-610 on mechanical function and energy metabolism during ischaemia reperfusion injury in isolated perfused rat hearts. J Cardiovasc Pharmacol 1996; 27: 487-494.
- 24. Richard V, Kaeffer N, Hogie M, Tron C, Blanc T, Thuillez C. Role of endogenous endothelin in myocardial and coronary endothelial injury after ischaemia and reperfusion in rats: studies with bosentan, a mixed ET<sub>A</sub>-ET<sub>B</sub> antagonist. Br J Pharmacol 1994; 113: 869-876.
- 25. Krause SM, Lynch JJ, Stabilito II, Woltmann RF. Intravenous administration of the endothelin-1 antagonist BQ-123 does not ameliorate myocardial ischaemic injury following acute coronary artery occlusion in the dog. Cardiovasc Res 1994; 28: 1672-1678.
- 26. Mertz TE, McClanahan TB, Flynn MA et al. Endothelin(A) receptor antagonism by PD 156707 does not reduce infarct size after coronary artery occlusion/reperfusion in pigs. J Pharmacol Exp ther 1996; 278: 42-49.

- 27. Burke SE, Nelson RA. Endothelin-receptor antagonist FR 139317 reduces infarct size when given before, but not after, coronary artery occlusion. J Cardiovasc Pharmacol 1997; 29: 87-92.
- Vitola JV, Forman MB, Holsinger JP et al. Role of endothelin in a rabbit model of acute myocardial infarction: Effects of receptor antagonists. J Cardiovasc Pharmacol 1996; 28: 774-783.
- 29. Li XS, Wang QD, Pernow J. Beneficial effects of the endothelin receptor antagonist bosentan on myocardial and endothelial injury following ischaemia/reperfusion in the rat. Eur J Pharmacol 1995; 283: 161-168.
- 30. Dagassan PH, Breu V, Clozel M, Clozel JP. Role of endothelin during reperfusion after ischaemia in isolated perfused rat heart. J Cardiovasc Pharmacol 1994; 24: 867-874.
- 31. Teerlink JR, Breu V, Sprecher U, Clozel M, Clozel J-P. Potent vasoconstriction mediated by endothelin ET<sub>B</sub> receptors in canine coronary arteries. Circ Res 1994; 74: 105-114.
- 32. Warner TD, Allcock GH, Corder R, Vane JR. Use of the endothelin antagonists BQ123 and PD142893 to reveal three endothelin receptors mediating smooth muscle contraction and the release of EDRF. Br J Pharmacol 1993; 110: 777-782.
- 33. Hide EJ, Piper J, Thiemermann C. Endothelin-1-induced reduction of myocardial infarct size by activation of ATP-sensitive potassium channels in a rabbit model of myocardial ischaemia and reperfusion. Br J Pharmacol 1995; 116: 2597-2602.
- 34. Wang, P, Gallagher KP, Downey JM et al. Pretreatment with endothelin-1 mimics ischaemic preconditioning against infarction in isolated rabbit heart. J Mol Cell Cardiol 1996; 28: 579-588.
- 35. Bugge E, Ytrehus K. Endothelin-1 can reduce infarct size through protein kinase C and  $K_{ATP}$  channels in the isolated rat heart. Cardiovasc Res 1996; 32: 920-929.

- 36. Fryer RM, Eels JT, Hsu AK, Henry MM, Gross GJ. Ischaemic preconditioning in rats: role of mitochondrial K(ATP) channel in preservation of mitochondrial function. Am J Physiol Heart Circ Physiol 2000; 278: H305-H312.
- 37. Gourine AV, Molosh AI, Poputnikov D, Bulhak A, Sjöquist P-O, Pernow J. Endothelin-1 exerts a preconditioning-like cardioprotective effect against ischaemia/reperfusion injury via the ET<sub>A</sub> receptor and the mitochondrial K<sub>ATP</sub> channel in the rat in vivo. Br J Pharmacol 2005; 144: 331-337.
- 38. Watanauki M, Hori M, Tsuchiya K et al. Endothelin-1 inhibition of cardiac ATP-sensitive K<sup>+</sup> channels via pertussis-toxin-sensitive G-proteins. Cardiovasc Res 1997; 33: 123-130.
- 39. Suzuki YJ, Evans T. Regulation of cardiac myocyte apoptosis by the GATA-4 transcription factor. Life Sci 2004, 74: 1829-1838.
- 40. Long X, Boluyt MO, Hipolito ML et al. p53 and the hypoxia-induced apoptosis of cultured neonatal rat cardiac myocytes. J Clin Invest 1997; 99: 2635-2643.
- 41. Fliss H, Gattinger D. Apoptosis in ischaemic and reperfused rat myocardium. Circ Res 1996; 79: 949-956.
- 42. Yaoita H, Ogawa K, Maehara K, Maruyama Y. Attenuation of ischaemia/reperfusion injury in rats by a caspase inhibitor. Circulation 1998; 97: 276-281.
- 43. Araki M, Hasegawa K, Iwai-Kanai E et al. Endothelin-1 as a protective factor against beta-adrenergic agonist-induced apoptosis in cardiac myocytes. J Amer Coll Cardiol 2000; 36: 1411-1418.
- 44. Kitta K, Clément SA, Remeika J, Blumberg JB, Suzuki YJ. Endothelin-1 induces phosphorylation of GATA-4 transcription factors in HL-1 atrial muscle cell line. Biochem J 2001; 278: 4705-4712.

- 45. Kakita T, Hasegawa K, Iwai-Kanai E et al. Calcineurin pathway is required for endothelin-1-mediated protection against oxidant stress-induced apoptosis in cardiac myocytes. Circ Res 2001; 88: 1239-1246.
- 46. Ehara N, Hasegawa K, Ono K et al. Activators of PPARgamma antagonize protection of cardiac myocytes by endothelin-1. Biochem Biophys Res Commun 2004; 321: 345-349.
- 47. Iwai-Kanai, Hasegawa K. Intracellular signalling pathways for norepinephrine- and endothelin-1-mediated regulation of myocardial cell apoptosis. Mol Cell Biochem 2004; 259: 163-168.
- 48. Iwai-Kanai E, Hasegawa K, Adachi S et al. Effects of endothelin-1 on mitochondrial function during the protection against myocardial cell apoptosis. Biochem Biophys Res Commun 2003; 305: 898-903.
- 49. Oie E, Vinge LR, Tonnessen T et al., Transient isopeptide-specific induction of myocardial endothelin-1 mRNA in congestive heart failure in rats. Basic Res Cardiol 2002; 97: 239-247.
- 50. Katwa LC. Cardiac myofibroblasts isolated from the site of myocardial infarction express endothelin de novo. Am J Physiol Heart Circ Physiol 2003; 285: H1132-H1139.
- 51. Ehrenreich H, Burd PR, Rottem M. Endothelins belong to the assortment of mast cell-derived and mast cell-bound cytokines. New Biol 1992; 4: 147-156.
- 52. Szalay L, Kaszaki J, Nagy S, Boros M. Endothelin-1 induces mucosal mast cell degranulation in then rat small intestine. Life Sci 2000; 67: 1947-1958.
- 53. Murray DB, Gardner JD, Brower GL, Janicki JS. Endothelin-1 mediates cardiac mast cell degranulation, matrix metalloproteinase activation and myocardial remodeling in rats. Am J Physiol Heart Circ Physiol 2004; 287: H2295-H2299.

- 54. Walsh S, Kane KA, McCaig, Wainwright CL. Endothelin-1 induces mast cell degranulation and increases tryptase level in the murine myocardium in vitro. J Mol Cell Cardio 2005; Abstract In Press.
- 55. Galli SJ, Biology of disease: new insights into "The riddle of the mast cells": microenvironmental regulation of mast cell development and phenotypic heterogeneity. Lab Invest 1990; 62: 5-33.
- 56. Matsushima H, Yamada N, Matsue H, Shimada S. The effects of endothelin-1 on degranulation, cytokine and growth factor production by skin-derived mast cells. Eur J Immunol 2004; 34: 1910-1919.
- 57. Hultner L, Ehrenrich H. Mast cells and endothelin-1: a life saving biological liaison? Trends Immunol 2005; 26: 235-238.
- 58. Maurer M, Wedemeyer J, Metz M et al. Mast cells promote homeostasis by olimiting endothelin-1-induced toxicity. Nature 2004; 432: 512-516.
- 59. Sargent CA, Liu EC, Chao CC et al. Role of endothelin receptor subtype B (ET-B) in myocardial ischaemia. Life Sci 1994; 55: 1833-1844.
- 60. Gan XT, Chakrabarti S, Karmazyn M. Increased endothelin-1 and endothelin receptor expression in myocytes of ischaemic and reperfused rat hearts and ventricular myocytes exposed to ischaemic conditions and its inhibition by nitric oxide generation. Can J Physiol Pharmacol 2003; 82: 105-113.
- 61. Crockett TR, Gray GA, Kane KA, Wainwright CL. Sarafotoxin 6c (S6c) reduces infarct size and preserves mRNA for the ET<sub>B</sub> receptor in the ischaemic/reperfused myocardium of anaesthetised rats. J Cardiovasc Pharmacol 2004; 44: 148-154.
- 62. D'Orleans-Juste PD, Labonté J, Bkailey G et al. Function of the endothelin B receptor in cardiovascular physiology and pathophysiology. Pharmacol Ther 2002; 95: 221-238.

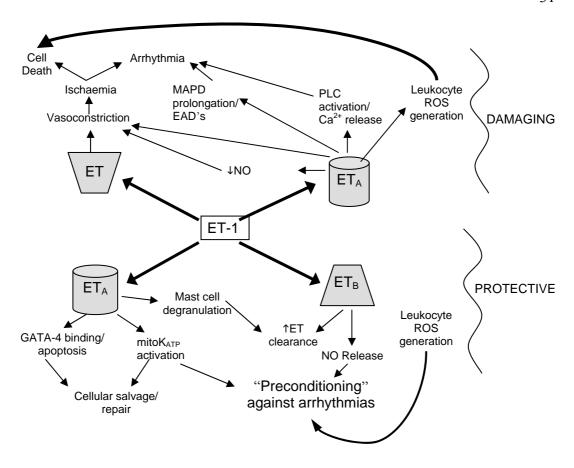
- 63. Chiariello M, Golino P, Cappelli-Bigazzi M et al. Reduction in infarct size by the prostacyclin analogue iloprost (ZK 36374) after experimental coronary artery occlusion-reperfusion. Am Heart J 1988; 115: 499-504.
- 64. Lefer AM, Siegfried MR, Ma X-L. Protection of ischemia-reperfusion injury by sydnonimine donors via inhibition of neutrophil-endothelium interaction. J Cardiovasc Pharmacol 1993; 22 (Suppl 7): S27-S33.
- 65. Lefer DJ, Scalia R, Campbell B et al. Peroxynitrite inhibits leukocyteendothelial cell interactions and protects against ischaemia-reperfusion injury in rats. J Clin Invest 1997; 99: 684-691.
- 66. Crockett TR, McBride M, Ness KF et al. The effect of S6c on platelet aggregation and leukocyte free radical generation in rat blood in vitro and ex vivo. Br J Pharmacol 2000; 131: 162P.
- 67. Ezra D, Goldstein RE, Czala JF, Feuerstein GZ. Lethal ischaemia due to intracoronary endothelin in pigs. Am J Physiol 1989; 257: H339-H343.
- 68. Yorikane R, Koike H. The arrhythmogernic action of endothelin in rats. Jpn J Pharmacol 1990; 53: 259-263.
- 69. Nichols AJ, Koster PF, Ohlstein EH. The effects of diltiazem on the coronary heamodynamics and cardiac functional effects produced by intracoronary administration of endothelin-1 in the anaesthetized dog. Br J Pharmacol 1990; 99: 597-601.
- 70. Yorikane R, Koike H, Miyake S. Electrophysiological effects of endothelin-1 on canine myocardial cells. J Cardiovasc Pharmacol 1991; 17 (suppl 7): S159-S162.
- 71. Garjani A, Wainwright CL, Zeitlin IJ, Wilson C, Slee SJ. Effects of endothelin-1 and the ETA-receptor antagonist BQ123, on ischaemic arrhythmias in anaesthetised rats. J Cardiovasc Pharmacol 1995; 25: 634-642.

- 72. Ercan ZS, Ilhan M, Kilinc M et al. Arrhythmogenic action of endothelin peptides in isolated perfused whole hearts from guinea pig and rats. Pharmacology 1996; 53: 234-240.
- 73. Duru F, Barton M, Lüscher TF, Candinas R. Endothelin and cardiac arrhythmias: do endothelin antagonists have a therapeutic potential as antiarrhythmic drugs? Cardiovasc Res 2001; 49: 272-280.
- 74. Vago H, Soos P, Zima E et al. The ET(A) receptor antagonist LU 135252 has no electrophysiological or anti arrhythmic effects during myocardial ischaemia/reperfusion in dogs. Clin Sci (Lond) 2002; 103 (Suppl 48): 223S-227S.
- 75. Alexiou K, Dschietzig T, Simsch O et al. Arrhythmogenic effects induced by coronary conversion of pulmonary big endothelin to endothelin: aggravation of this phenomenon in heritable hyperlipidaemia. J Am Coll Cardiol 1998; 32: 1773-1778.
- 76. Sharif I, Kane KA, Wainwright CL. Endothelin and ischaemic arrhythmias antiarrhythmic or arrhythmogenic? Cardiovasc Res 1998; 39: 625-632.
- 77. Di Filippo C, D'Amico M, Marfella R, Berrino L, Giugliano D, Rossi F. Endothelin-1 receptor antagonists reduce caqrdiac electrical instability induced by high glucose in rats. Naunyn Schmeidbergs Arch Pharmacol 2002; 366: 193-197.
- 78. Crockett TR, Scott GA, McGowan NW, Wainwright CL, Kane KA. Antiarrhythmic and electrophysiological effects of the endotheline receptor antagonists, BQ123 and PD161721. Eur J Pharmacol 2001; 432: 71-77.
- 79. Szabo T, Geller L, Merkely B et al. Investigating the dual nature of endothelin-1: Ischaemia or direct arrhythmogenic effect? Life Sciences 2000; 66(26): 2527-2541.
- 80. Becker R, Merkely B, Bauer A et al. Ventricular arrhythmias induced by endothelin-1 or by acute ischaemia: a comparative analysis using three-dimensional mapping. Cardiovasc Res 2001; 45: 310-320.

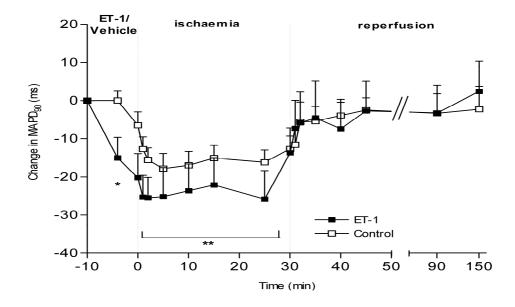
- 81. Geller L, Merkely B, Lang V et al. Increased monophasic action potential dispersion in Endothelin-1 induced ventricular arrhythmias. J Cardiovasc Pharmacol 1998; 31(Suppl. 1): S434-S436.
- 82. Kiss O, Geller L, Merkely B et al. (2000). Endothelin-A receptor antagonist LU 135.252 inhibits the formation of ventricular arrhythmias caused by intrapericardial infusion of endothelin-1. J Cardiovasc Pharmacol 2000; 36(Suppl 1): S317-S319.
- 83. McCabe C, Hicks MN, Kane KA et al. (2005). Electrophysiological and haemodynamic effects of endothelin ET<sub>A</sub> and ET<sub>B</sub> receptors in normal and ischaemia working rabbit hearts. Br J Pharmac. In Press.
- 84. Furukawa T, Ito H, Nitta J et al. Endothelin-1 enhances calcium entry through T-type calcium channels in cultured neonatal rat ventricular myocytes. Circ Res 1992; 71: 1242-1253.
- 85. He JQ, Pi YQ, Walker JW et al. Endothelin-1 and photoreleased diacylglycerol increase L-type Ca<sup>2+</sup> current by activation of protein kinase C in rat ventricular myocytes. J Physiol 2000; 524: 807-820.
- 86. Banyasz T, Magyar J, Kortvely A et al. Different effects of endothelin-1 on calcium and potassium currents in canine ventricular cells. Naunyn. Schmied. Arch. Pharmacol 2001; 363: 383-390.
- 87. Watanabe T & Endoh M. Antiadrenergic Effects of Endothelin-1 on the L-Type Ca<sup>2+</sup> Current in Dog Ventricular Myocytes. J Cardiovasc Pharmacol 2000; 36: 344-350.
- 88. Ono K., Eto K, Sakamoto A et al. Negative chronotropic effect of endothelin-1 mediated through ET<sub>A</sub> receptors in guinea pig atria. Circ Res 1995; 76: 284-292.
- 89. Kelso EJ, Spiers JP., McDermott BJ et al. Receptor-Mediated Effects of Endothelin on the L-Type Ca<sup>2+</sup> Current in Ventricular Cardiomyocytes. J Pharmacol Ex. Ther 1998; 286: 662-669.

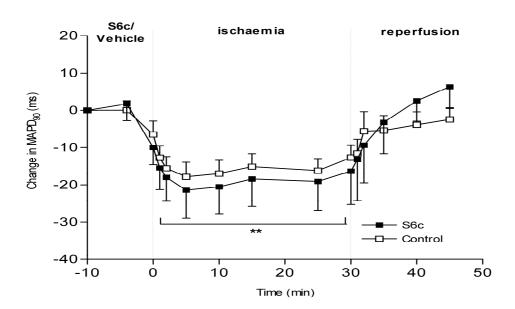
- 90. Magyar J, Iost N, Kortvely A et al. Effects of endothelin-1 on calcium and potassium currents in undiseased human ventricular myocytes. Plugers Arch 2000; 441(1): 144-149.
- 91. Thomas GP, Sims SM, Karmazyna M. Differential effects of endothelin-1 on basal and isoprenaline-enhanced Ca2+ current in guinea-pig ventricular myocytes. J Physiol 1997; 503 (Pt 1): 55-65.
- 92. Habuchi Y, Tanaka H, Furukawa T et al. Endothelin enhances delayed potassium current via PLC in guinea pig ventricular myocytes. Am J Physiol 1992; 262: H345-54.
- 93. Washizuka T, Horie M, Watanuki M & Sasayama S. Endothelin-1 inhibits the slow component of cardiac delayed rectifier K<sup>+</sup> currents via a pertussis toxinsensitive mechanism. Circ Res 1997; 81: 211-218.
- 94. Kobayashi S, Nakaya H, Takizawa T et al. (1996). Endothelin-1 partially inhibits ATP-sensitive K<sup>+</sup> current in guinea pig ventricular cells. J Cardiovasc Pharmacol 1996; 27: 12-19.
- 95. Mackenzie L, Bootman MD, Laine M et al. (2002). The role of inositol 1,4,5-trisphosphate receptors in Ca(2+) signalling and the generation of arrhythmias in rat atrial myocytes. J Physiol 2002; 541: 395-409.
- 96. Zima AV & Blatter LA. Inositol-1,4,5-trisphosphate-dependent Ca(2+) signalling in cat atrial excitation-contraction coupling and arrhythmias. J Physiol 2004; 555: 607-615.
- 97. Zhang YH, James AF, Hancox JC. Regulation by endothelin-1 of Na+-Ca2+ exchange current (I(NaCa)) from guinea-pig isolated ventricular myocytes. Cell Calcium 2001; 30: 351-360.
- 98. Wang H, Sakurai K, Endoh M. Pharmacological analysis by HOE642 and KB-R9032 of the role of Na(+)/H(+) exchange in the endothelin-1-induced Ca(2+) signalling in rabbit ventricular myocytes. Br J Pharmacol 2000; 131: 638-644.

- 99. Wirth KJ, Maier T, Busch AE. NHE1-inhibitor cariporide prevents the transient reperfusion-induced shortening of the monophasic action potential after coronary ischemia in pigs. Basic Res Cardiol 2001; 96: 192-197.
- 100. Parrat JR, Vegh A. Coronary vascular endothelium-myocyte interactions in protection of the heart by ischaemic preconditioning. J Physiol Pharmacol 1999; 50: 509-524.
- 101. Crockett TR, Sharif I, Kane KA, Wainwright CL. Sarafotoxin 6c protects against ischaemia induced cardiac arrhythmias in vivo and in vitro in the rat. J Cardiovasc Pharmacol 2000; 36 (Suppl 1): S2979-S299.
- 102. Sharif I, Crockett TR, Kane KA, Wainwright CL. The effects of endothelin-1 on ischaemic-induced ventricular arrhythmias in rat isolated hearts. Eur J Pharmacol 2001; 427: 235-242.



**Figure 1:** Paradigm for the complex actions of endothelin in the ischaemic heart that contribute to both cellular damage and to cardioprotection (see text for full details). MAPD – monophasic action potential duration; EAD's – early after depolarisations; mitoKATP – mitochondrial ATP-dependent potassium channel; ROS – reactive oxygen species.





**Figure 2.** Change in MAPD<sub>90</sub> in the endocardium of working rabbit hearts treated with vehicle, 10<sup>-10</sup>M ET-1 (top panel) or 10<sup>-8</sup>M S6c (bottom panel) before and during acute regional ischaemia and reperfusion. '\*' indicates significantly different from pre-drug value. '\*\*' indicates statistically significant differences from respective pre-ischaemic values, (P<0.05 in each case). (n=6-12).