



Queensland University of Technology
Brisbane Australia

This is the author's version of a work that was submitted/accepted for publication in the following source:

Cowin, Allison J, Lei, Nazi, Linda, Franken, Ruzehaji, Nadira, Offenhäuser, Carolin, Kopecki, Zlatko, & Murray, Rachael (2012) Lysosomal secretion of Flightless I upon injury has the potential to alter inflammation. *Communicative & Integrative Biology*, 5(6), pp. 546-549.

This file was downloaded from: <http://eprints.qut.edu.au/56390/>

© Copyright 2012 Open access

- author can archive post-print (ie final draft post-refereeing) - subject to Restrictions below, author can archive publisher's version/PDF •Authors final version only •On Institutional Repository •Must link to publisher version •Published source must be acknowledged •Landes Bioscience will deposit in PubMed Central or Europe PMC within 6-12 months of publication, depending on funding agency policy •Embargoes on funding agency requirements, can be removed by payment of Open Access fee •Publisher's version/PDF may be used upon payment of Open Access fee

Notice: *Changes introduced as a result of publishing processes such as copy-editing and formatting may not be reflected in this document. For a definitive version of this work, please refer to the published source:*

<http://dx.doi.org/10.4161/cib.21928>

Lysosomal secretion of Flightless I upon injury has the potential to alter inflammation

Allison J. Cowin,^{1,2} Nazi Lei,³ Linda Franken,³ Nadira Ruzehaji,¹ Carolin Offenhäuser,³ Zlatko Kopecki¹ and Rachael Z. Murray^{3,4,*}

¹Women's and Children's Health Research Institute; North Adelaide; SA Australia; ²Discipline of Paediatrics; The University of Adelaide; Adelaide, SA Australia; ³CHBRI Wound Healing Laboratory; Kids Research Institute; Children's Hospital at Westmead; Sydney, NSW Australia; ⁴Tissue Repair and Regeneration Program; Institute of Health and Biomedical Innovation; Queensland University of Technology; Brisbane, QLD Australia

Keywords: Flightless I, late endosome, lysosome, secretion, Cathepsin D, Rab 7 and Stx11

Intracellular Flightless I (Flii), a gelsolin family member, has been found to have roles modulating actin regulation, transcriptional regulation and inflammation. In vivo Flii can regulate wound healing responses. We have recently shown that a pool of Flii is secreted by fibroblasts and macrophages, cells typically found in wounds, and its secretion can be upregulated upon wounding. We show that secreted Flii can bind to the bacterial cell wall component lipopolysaccharide and has the potential to regulate inflammation. We now show that secreted Flii is present in both acute and chronic wound fluid.

Introduction

Flightless I (Flii) is a multifunctional protein of the gelsolin family of actin-remodelling proteins. There are eight members of this family in mouse and humans, each containing one or two gelsolin domains consisting of three repeated gelsolin motifs of between 125 and 150 amino acids in length.¹ Flii contains 6 repeat gelsolin motifs and an additional 11 Leucine Rich Repeat (LRR) domains not present in other family members.^{2,3} It has been found to play roles in many processes including actin regulation, transcription and inflammation. Flii is distributed across many cellular compartments, as would be expected for such a multifunctional protein, including the nucleus, cytosol and lysosomes.³ In vivo, Flii negatively regulates excisional wound, blister wound and burn injury repair.^{4–6} Flii overexpressing mice (Flii^{Tg/Tg}) have impaired healing with larger, less contracted wounds, reduced cell proliferation and delayed epithelial migration. In contrast, mice with reduced levels of Flii (Flii^{-/-}) have improved wound healing with increased epithelial migration and enhanced wound contraction.⁵ Wounds in Flii overexpressing mice also show significantly elevated levels of collagen I and overexpression of collagen is a major contributing factor to hypertrophic, or excessive scar formation.⁵ Given Flii's known role in processes that are all involved in regulating tissue repair it is not surprising that it could alter the course of wound healing and scar formation.

Regulation of Actin and Transcription

Unlike many gelsolin family members, which enhance actin polymerisation and cap actin filaments, Flii binds actin filaments and actin monomers and inhibits actin polymerisation.^{7,8}

Flii caps but does not sever actin filaments and thereby retards actin filament turnover.^{7,8} Flii associates with focal adhesions, which are specialized structures that link the actin cytoskeleton to the surface integrin receptors and anchor cells to the extracellular matrix, and can alter their formation.^{7,9} Cells with reduced levels of gelsolin migrate slower, while a reduction in Flii levels leads to an increase in migration.⁵ Both fibroblasts and keratinocytes, cells typically found in wounds, that have less Flii migrate faster in vitro and in vivo and vice versa.^{7,10} The impaired wound healing seen in mice overexpressing Flii could in part be due to the ability of Flii to inhibit cell adhesion and migration. However, Flii has several other functions, which could also contribute to its negative role in tissue repair. Like gelsolin, Flii has the ability to regulate transcription.¹¹ Flii binds to hormone-activated nuclear receptors, including the estrogen and thyroid hormone receptor, as well as the coactivators GRIP1 and CARM1.¹² Flii positively regulates hormone-stimulated gene expression by the estrogen receptor through its gelsolin region and is involved in the recruitment of the chromatin remodelling complex SW1/SNF to estrogen-responsive promoters.^{12,13} Flii also inhibits β -catenin and LEF1/TCF-mediated transcription.¹⁴ Thus, Flii has the ability to alter transcription in part through its gelsolin domains. Our new data has shown that in fibroblasts Flii is found in the nucleus of some but not all cells (Figs. 1 and 2)³ and it is probably this pool of Flii which may well be responsible for regulating transcription in these cells.¹⁵ Our new data further suggests that Flii may have distinct roles in different cell types or under specific conditions. We show that unlike fibroblasts, macrophages have little to no Flii in the nucleus whether they were activated or not suggesting that its roles could differ depending on cell type or stimulus.³

*Correspondence to: Rachael Murray; Email: rachael.murray@qut.edu.au
Submitted: 07/31/12; Accepted: 08/22/12
<http://dx.doi.org/10.4161/cib.21928>

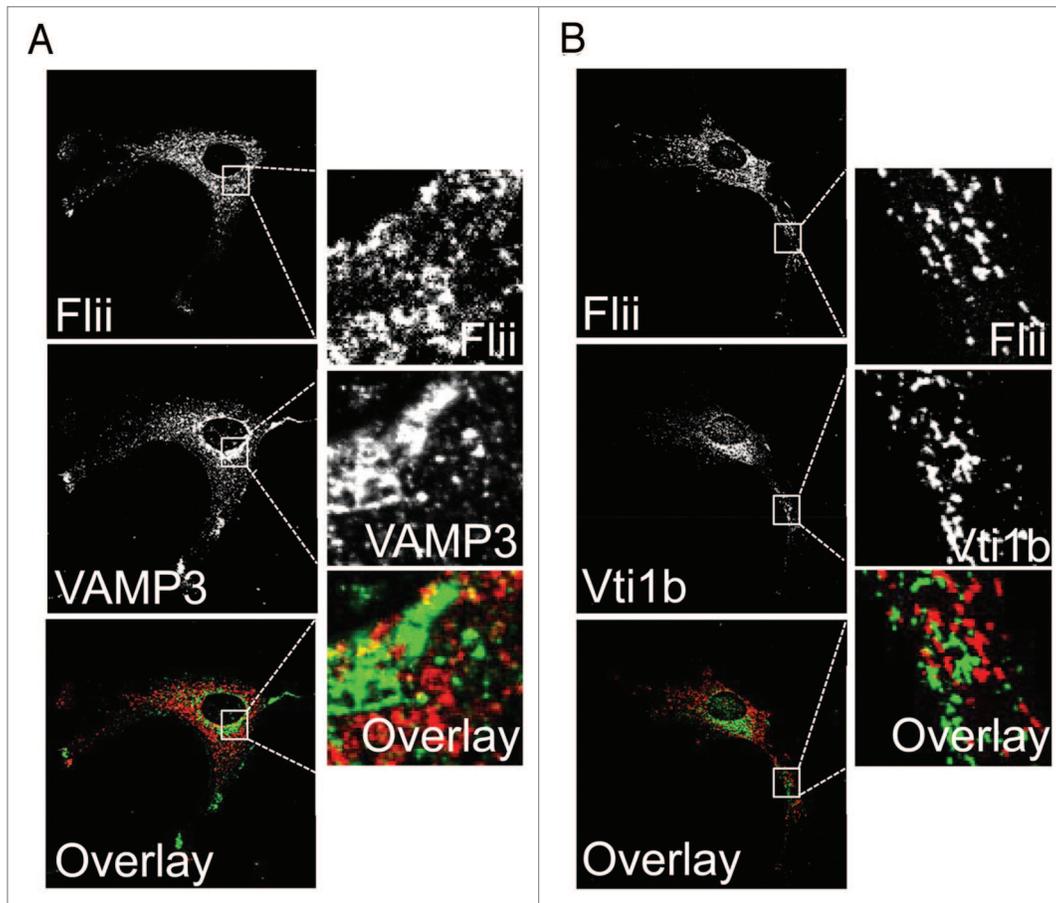


Figure 1. Flii is not found in the classical secretory pathway in fibroblasts. Primary fibroblasts were fixed with methanol, immunostained for Flii (mouse anti-Flii antibody) in combination with the recycling endosome SNARE protein VAMP3 (A) or the trans-Golgi complex and recycling endosome SNARE Vti1b (B). Flii is not located in compartments of the classical secretory pathway in fibroblasts.

Flii's Role in Regulating Inflammation

In recent years it has becoming increasingly obvious that Flii has an important role in dampening inflammation.^{3,16-19} Unlike other gelsolin family members Flii has 11 LRR domains in the N-terminus.³ These LRR domains share nearly 50% similarity to the LRR domains of the immune related receptor toll-like receptor (TLR) 4. The immune system detects infection or injury via the LRR domains of TLRs.²⁰ They can bind to pathogen-associated molecular pattern (PAMPs) molecules, such as the gram-negative bacteria cell wall component lipopolysaccharide or to damage-associated molecular pattern (DAMPs) molecules such as HMGB1 that are released from damaged and dying cells, as well as extracellular matrix cleavage products.²¹ Their binding in turn activates intracellular TLR signaling pathways that ultimately lead to the secretion of cytokines. Typically both DAMPs and PAMPs are present in wounds. The importance of these domains in Flii was first hinted at by mouse knockout studies of Flii and other gelsolin family members.¹⁵ Apart from Flii, mice lacking members of the gelsolin family are viable, but with actin defects.¹⁵ In contrast, homozygous disruption of the *Flii* gene in mice leads to very early failure of embryonic development with

impaired cellularization and gastrulation of the embryo indicating that Flii is essential developmental regulator and has additional important functions, which could be related to the role of the LRR domains.²²

We have recently shown a pool of Flii is located in the cytosol and this pool may in part be responsible for its role in dampening inflammation.^{3,16,19} Although inflammation appears to be a necessary part of the normal adult wound healing process, excessive activation of TLR receptors and the subsequent increased or prolonged inflammatory response can induce considerable tissue damage which can lead to impaired healing. Flii is upregulated in mouse wounds peaking around day 7 when the inflammatory stage of tissue repair is being switched off.⁵ Whether Flii is playing a role in dampening inflammation during tissue repair has yet to be tested in vivo. However in vitro Flii has been shown to dampen inflammation in multiple ways. In macrophages Flii is located in a complex with the TLR adaptor protein MyD88 through its interaction with nucleoredoxin.^{17,19} Binding of Flii to this complex inhibits MyD88 binding to TLR4, which results in the inhibition TLR signaling pathways and reduces cytokine secretion.^{17,19} We recently found a pool of Flii localized to late endosomes/lysosomes in fibroblasts and macrophages where it may also have a role in dampening inflammation.^{3,18} Flii binds

to caspase-1 and in doing so inhibits the maturation of the cytokine pro-interleukin-1 β to pro-interleukin-1 β in macrophages, thus reducing its secretion.¹⁸ Exactly where in the cell pro-interleukin-1 β is cleaved to form the mature secreted form is controversial; however, there is data to suggest it may take place in the lysosome and that the mature form can be secreted from this compartment.²³

Flii is Secreted and the Secreted Form also has the Potential to Alter Inflammation

Flii was thought to be solely an intracellular protein, however, our recent study has shown that *in vitro* Flii is constitutively secreted by at least two cell types typically found in wounds; macrophages and fibroblasts.³ This secretion from fibroblasts can be upregulated in response to wounding and by macrophages in response to LPS stimulation.³ The upregulation in secretion after wounding suggest this pool of Flii may play a role during the repair process. In macrophages Flii localizes to late endosomes/lysosomes but not to compartments typically associated with the classical secretory pathway, for example the Golgi complex in macrophages.³ Similar results are shown here for primary fibroblasts (Figs. 1 and 2). Flii does not colocalize with the trans-Golgi network and recycling endosome-associated SNARE protein Vti1b in primary fibroblasts (Fig. 1A) or with the recycling endosome associated SNARE protein VAMP3 (Fig. 1B). However, a pool of Flii colocalizes with the lysosomal enzyme Cathepsin D in these cells (Fig. 2A). Our recent data shows that Flii is secreted from macrophages via late endosomes/lysosomes in a manner regulated by Rab7 and Stx11.³ This data suggests that Flii may therefore not only affect cellular activities via its intracellular/nuclear functions but it may also have important extracellular activities.

In vivo, we have shown that Flii is also present in human plasma samples.³ Given Flii's important role in wound healing we have now looked to see whether Flii is secreted into wound fluid from a number of sources. We find that secreted Flii is also present in acute wound fluid from patients undergoing abdominoplasty and in blister fluid as well as in chronic wound fluid taken from patients with venous leg ulcers (Fig. 3A). Exactly what role Flii is playing in the wound fluid is currently unclear; however our results suggest that Flii can form a complex with the bacterial cell wall protein lipopolysaccharide (LPS).³ An antibody to the first LLR domain is able to inhibit formation of this complex suggesting that the LRR region might be involved in this process.³ Altering the level of secreted Flii in the media showed that Flii can negatively influence the LPS induced production and secretion of cytokine, such that cells stimulated with LPS in the presence of media with higher levels of Flii have reduced production and secretion of TNF.³ Thus, it would appear that secreted Flii has the potential to dampen inflammation.

Prolonged or augmented inflammation can induce significant tissue damage unfavorable to the repair process thus the body has developed mechanisms that finely tune and regulate TLR activation. For example, surface TLR expression is downregulated after activation and at the same time soluble TLRs are secreted that compete with their membrane-associated forms for binding

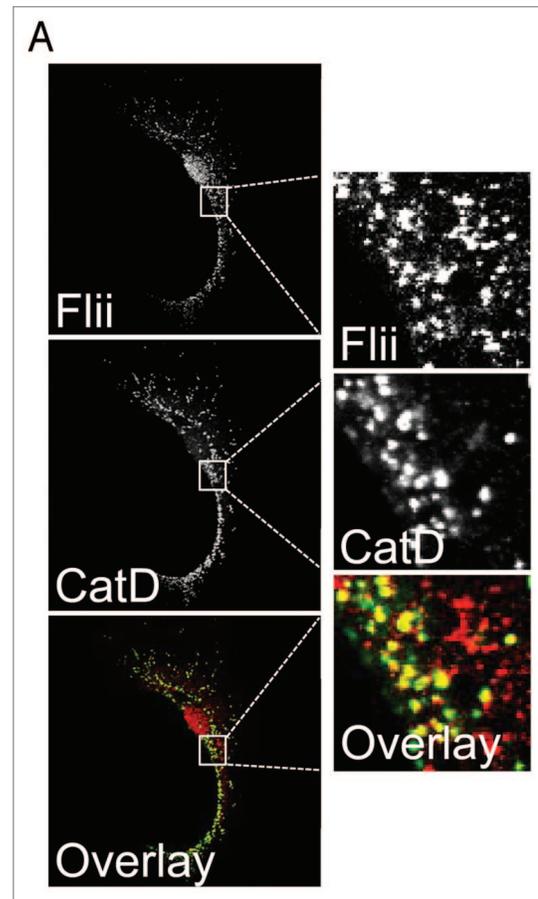


Figure 2. Flii is located in late endosomes/lysosomes in fibroblasts. (A) Primary fibroblasts were fixed with methanol, immunostained for Flii (mouse anti-Flii antibody) and the late endosomal/lysosomal enzyme cathepsin D (CatD). Flii co-localizes with cathepsin D in late endosomes/lysosomes in fibroblasts.

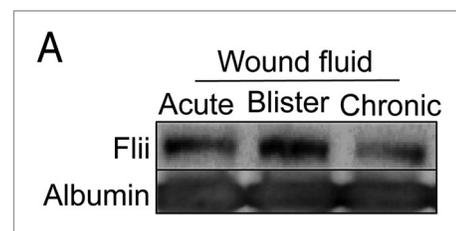


Figure 3. Flii is present in wound fluid from acute and chronic wounds. The clinical investigations were conducted under approval from the Women's and Children's Health Network Human Research Ethics Committee, Adelaide, Australia, in accordance to the Declaration of Helsinki principles and with written informed consent. (A) Wound fluid collected from patients undergoing abdominoplasty, from blister fluid and from patients with venous leg ulcers was immunoblotted for Flii and albumin. Flii is secreted into both acute and chronic wounds.

to ligands, thus limiting the stimulation of TLR signaling and inflammation. It is possible given the timing of Flii's upregulation after injury that Flii could be potentially playing a role in limiting the inflammatory response both from within the cells

and in the extracellular matrix. It will be interesting in the future to see whether this is the case.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

References

1. Kwiatkowski DJ, Janmey PA, Yin HL. Identification of critical functional and regulatory domains in gelsolin. *J Cell Biol* 1989; 108:1717-26; PMID:2541138; <http://dx.doi.org/10.1083/jcb.108.5.1717>.
2. Kopecki Z, Cowin AJ. Flightless I: an actin-remodelling protein and an important negative regulator of wound repair. *Int J Biochem Cell Biol* 2008; 40:1415-9; PMID:17526423; <http://dx.doi.org/10.1016/j.biocel.2007.04.011>.
3. Lei N, Franken L, Ruzehaji N, Offenhäuser C, Cowin AJ, Murray RZ. Flightless, secreted through a late endosome/lysosome pathway, binds LPS and dampens cytokine secretion. *J Cell Sci* 2012; In press; PMID:22718342; <http://dx.doi.org/10.1242/jcs.099507>.
4. Adams DH, Ruzehaji N, Strudwick XL, Greenwood JE, Campbell HD, Arkell R, et al. Attenuation of Flightless I, an actin-remodelling protein, improves burn injury repair via modulation of transforming growth factor (TGF)-beta1 and TGF-beta3. *Br J Dermatol* 2009; 161:326-36; PMID:19519830; <http://dx.doi.org/10.1111/j.1365-2133.2009.09296.x>.
5. Cowin AJ, Adams DH, Strudwick XL, Chan H, Hooper JA, Sander GR, et al. Flightless I deficiency enhances wound repair by increasing cell migration and proliferation. *J Pathol* 2007; 211:572-81; PMID:17326236; <http://dx.doi.org/10.1002/path.2143>.
6. Kopecki Z, Arkell RM, Strudwick XL, Hirose M, Ludwig RJ, Kern JS, et al. Overexpression of the Flii gene increases dermal-epidermal blistering in an autoimmune ColVII mouse model of epidermolysis bullosa acquisita. *J Pathol* 2011; 225:401-13; PMID:21984127; <http://dx.doi.org/10.1002/path.2973>.
7. Mohammad I, Arora PD, Naghibzadeh Y, Wang Y, Li J, Mascarenhas W, et al. Flightless I is a focal adhesion-associated actin-capping protein that regulates cell migration. *FASEB J* 2012; 26:3260-72; PMID:22581781; <http://dx.doi.org/10.1096/fj.11-202051>.
8. Silacci P, Mazzolai L, Gauci C, Stergiopoulos N, Yin HL, Hayoz D. Gelsolin superfamily proteins: key regulators of cellular functions. *Cell Mol Life Sci* 2004; 61:2614-23; PMID:15526166; <http://dx.doi.org/10.1007/s00018-004-4225-6>.
9. Kopecki Z, O'Neill GM, Arkell RM, Cowin AJ. Regulation of focal adhesions by flightless i involves inhibition of paxillin phosphorylation via a Rac1-dependent pathway. *J Invest Dermatol* 2011; 131:1450-9; PMID:21430700; <http://dx.doi.org/10.1038/jid.2011.69>.
10. Kopecki Z, Arkell R, Powell BC, Cowin AJ. Flightless I regulates hemidesmosome formation and integrin-mediated cellular adhesion and migration during wound repair. *J Invest Dermatol* 2009; 129:2031-45; PMID:19212345; <http://dx.doi.org/10.1038/jid.2008.461>.
11. Archer SK, Claudianos C, Campbell HD. Evolution of the gelsolin family of actin-binding proteins as novel transcriptional coactivators. *Bioessays* 2005; 27:388-96; PMID:15770676; <http://dx.doi.org/10.1002/bies.20200>.
12. Lee YH, Campbell HD, Stallcup MR. Developmentally essential protein flightless I is a nuclear receptor coactivator with actin binding activity. *Mol Cell Biol* 2004; 24:2103-17; PMID:14966289; <http://dx.doi.org/10.1128/MCB.24.5.2103-2117.2004>.
13. Jeong KW, Lee YH, Stallcup MR. Recruitment of the SWI/SNF chromatin remodeling complex to steroid hormone-regulated promoters by nuclear receptor coactivator flightless-I. *J Biol Chem* 2009; 284:29298-309; PMID:19720835; <http://dx.doi.org/10.1074/jbc.M109.037010>.
14. Lee YH, Stallcup MR. Interplay of Fli-I and FLAP1 for regulation of beta-catenin dependent transcription. *Nucleic Acids Res* 2006; 34:5052-9; PMID:16990252; <http://dx.doi.org/10.1093/nar/gkl652>.
15. Archer SK, Behm CA, Claudianos C, Campbell HD. The flightless I protein and the gelsolin family in nuclear hormone receptor-mediated signalling. *Biochem Soc Trans* 2004; 32:940-2; PMID:15506930; <http://dx.doi.org/10.1042/BST0320940>.
16. Dai P, Jeong SY, Yu Y, Leng T, Wu W, Xie L, et al. Modulation of TLR signaling by multiple MyD88-interacting partners including leucine-rich repeat Fli-I-interacting proteins. *J Immunol* 2009; 182:3450-60; PMID:19265123; <http://dx.doi.org/10.4049/jimmunol.0802260>.
17. Hayashi T, Funato Y, Terabayashi T, Morinaka A, Sakamoto R, Ichise H, et al. Nucleoredoxin negatively regulates Toll-like receptor 4 signaling via recruitment of flightless-I to myeloid differentiation primary response gene (88). *J Biol Chem* 2010; 285:18586-93; PMID:20400501; <http://dx.doi.org/10.1074/jbc.M110.106468>.
18. Li J, Yin HL, Yuan J. Flightless-I regulates proinflammatory caspases by selectively modulating intracellular localization and caspase activity. *J Cell Biol* 2008; 181:321-33; PMID:18411310; <http://dx.doi.org/10.1083/jcb.200711082>.
19. Wang T, Chuang TH, Ronni T, Gu S, Du YC, Cai H, et al. Flightless I homolog negatively modulates the TLR pathway. *J Immunol* 2006; 176:1355-62; PMID:16424162.
20. Bell JK, Mullen GE, Leifer CA, Mazzoni A, Davies DR, Segal DM. Leucine-rich repeats and pathogen recognition in Toll-like receptors. *Trends Immunol* 2003; 24:528-33; PMID:14552836; [http://dx.doi.org/10.1016/S1471-4906\(03\)00242-4](http://dx.doi.org/10.1016/S1471-4906(03)00242-4).
21. Bianchi ME. DAMPs, PAMPs and alarmins: all we need to know about danger. *J Leukoc Biol* 2007; 81:1-5; PMID:17032697; <http://dx.doi.org/10.1189/jlb.0306164>.
22. Campbell HD, Fountain S, McLennan IS, Berven LA, Crouch MF, Davy DA, et al. Fliih, a gelsolin-related cytoskeletal regulator essential for early mammalian embryonic development. *Mol Cell Biol* 2002; 22:3518-26; PMID:11971982; <http://dx.doi.org/10.1128/MCB.22.10.3518-3526.2002>.
23. Eder C. Mechanisms of interleukin-1beta release. *Immunobiology* 2009; 214:543-53; PMID:19250700; <http://dx.doi.org/10.1016/j.imbio.2008.11.007>.

Acknowledgments

This work was supported by fellowships to R.Z.M. (#457247) and A.J.C. (#1002009) from the National Health and Medical Research Council of Australia and a University of Queensland International Postgraduate Research Scholarship to C.O.