



Queensland University of Technology
Brisbane Australia

This is the author's version of a work that was submitted/accepted for publication in the following source:

Frost, Ray L., Park, Yuri, & Ayoko, Godwin A. (2011) Characterisation of organoclays and adsorption of p-nitrophenol : environmental application. *Journal of Colloid and Interface Science*, 360(2), pp. 440-456.

This file was downloaded from: <http://eprints.qut.edu.au/42089/>

© Copyright 2011 Elsevier

Notice: *Changes introduced as a result of publishing processes such as copy-editing and formatting may not be reflected in this document. For a definitive version of this work, please refer to the published source:*

<http://dx.doi.org/10.1016/j.jcis.2011.04.085>

1 **Characterisation of organoclays and adsorption of p-nitrophenol: Environmental**
2 **application**

3

4 **Yuri Park, Godwin A. Ayoko, and Ray L. Frost***

5

6 Chemistry Discipline, Faculty of Science and Technology, Queensland University of Technology,
7 GPO 2434, Brisbane Queensland 4001, Australia

8

9 **Abstract**

10 Organoclays were synthesised through ion exchange of a single surfactant for sodium ions,
11 and characterised by a range of method including X-ray diffraction (XRD), BET, X-ray
12 photoelectron spectroscopy (XPS), thermogravimetric analysis (TGA), Fourier transform
13 infrared spectroscopy (FT-IR), and transmission electron microscopy (TEM). The change in
14 surface properties of montmorillonite and organoclays intercalated with the surfactant,
15 tetradecyltrimethylammonium bromide (TDTMA) were determined using XRD through the
16 change in basal spacing and the expansion occurred by the adsorbed p-nitrophenol. The
17 changes of interlayer spacing were observed in TEM. In addition, the surface measurement
18 such as specific surface area and pore volume was measured and calculated using BET
19 method, this suggested the loaded surfactant is highly important to determine the sorption
20 mechanism onto organoclays. The collected results of XPS provided the chemical
21 composition of montmorillonite and organoclays, and the high-resolution XPS spectra
22 offered the chemical states of prepared organoclays with binding energy. Using TGA and FT-
23 IR, the confirmation of intercalated surfactant was investigated. The collected data from
24 various techniques enable an understanding of the changes in structure and surface properties.
25 This study is of importance to provide mechanisms for the adsorption of organic molecules,
26 especially in contaminated environmental sites and polluted waters.

27 **Keywords** – Organoclay, X-ray photoelectron spectroscopy, montmorillonite, surfactant, BET

28 * Corresponding author: Ray L. Frost

29 Email address: g_r.frost@qut.edu.au

30 Address: 2 George Street, Brisbane Q 4001, Australia

31 **Introduction**

32 Clay minerals such as montmorillonite and smectite are abundant in nature. Clay minerals
33 are known as the swelling clays due to their tendency to swell and hydrate upon exposure to
34 water [1]. They are widely used in a variety of industries such as nano composites, catalysts,
35 photochemical reaction reagents, and adsorbents [2]. Among other applications, clays have
36 extensively been used as adsorbents in environmental systems, and the most commonly used
37 clay is montmorillonite because of its high cation exchange (CEC), swelling properties, high
38 surface areas, and consequential strong adsorption/absorption [3, 4]. Montmorillonite is a
39 typical 2:1 layered clay mineral that consists of two inward pointing tetrahedral sheets with a
40 central alumina octahedral sheet. The negatively charged clay layer surface is
41 counterbalanced by exchangeable cations such as Na^+ or Ca^{2+} in the interlayer space due to
42 the isomorphous substitution within the layers (e.g. the replacement of Mg^{2+} or Zn^{2+} for Al^{3+}
43 in the octahedral layer, and Al^{3+} for Si^{4+} in the tetrahedral sheets) [4, 5]. Because of the
44 hydration of inorganic cations on the exchange sites, the clay mineral surface is hydrophilic
45 in nature and this makes the natural clays ineffective sorbents for the removal of organic
46 compounds. Such a difficulty has been overcome through ion exchange of inorganic cations
47 with organic cations such as quaternary ammonium cations (QACs), represented as
48 $[(\text{CH}_3)_3\text{NR}]^+$, or $[(\text{CH}_3)_2\text{NR}_2]^+$, where R is a relatively short hydrocarbon substitution group.

49 By introducing cationic surfactant molecules into the interlamellar space through ion
50 exchange, the properties of clay minerals are changed from montmorillonite to organoclays.
51 The intercalation of cationic surfactant between clay layers exchanges the surface properties
52 from highly hydrophilic/lyophobic to increasingly hydrophobic/lipophilic [6]. As a result of
53 the modification of the properties of the swelling clay, the basal spacing in the layer increases
54 and new sorption sites of clays are exposed. Such changes in the properties of the surfactant
55 are important for the application of organoclays, and their significance in hydrophobic
56 organoclay is highlighted by their use for water purification. To date, the application of
57 organoclays to remove various organic compounds has been reported, and relative to
58 untreated clays, the adsorption capacity for the removal organic contaminants has been
59 improved by using organoclays [7-9].

60 In this study, tetradecyltrimethylammonium bromide (TDTMA) surfactant is used as an
61 example of a long alkyl cationic surfactant for the preparation of organoclays, since the effect
62 of the long alkyl surfactant with different loadings may increase the utilisation utility of the

63 modified clays. The present work was undertaken to investigate the changes of structure and
64 physical properties in the modified clays, and the interlayer configuration of intercalated
65 surfactant in montmorillonite was elucidated by using X-ray diffraction (XRD), transmission
66 electron microscopy (TEM), Brunauer, Emmett, and Teller (BET), thermogravimetric
67 analysis (TGA) and Fourier transform infrared spectroscopy (FT-IR). In addition, using X-
68 ray photoelectron spectroscopy (XPS), the surface characteristics of montmorillonite and
69 organoclays were investigated. As many literature articles have reported the effectiveness of
70 organoclays as adsorbents, this study will further investigate any structural or physical
71 changes, or adsorption properties of the resultant organoclays when intercalated with organic
72 compounds. P-nitrophenol was chosen as a test molecule in this research. The obtained
73 results will offer new insights into the structure and adsorption properties of organoclays, and
74 will enhance the potential industrial applications of organoclays as a filter material in water
75 purification.

76

77 **Experimental Methods**

78 **Materials**

79 The pure montmorillonite (MMT) was purchased from Sigma-Aldrich, and was used
80 without further purification. The cation exchange capacity (CEC) of this pure
81 montmorillonite is 76.4 meq/100g (according to the specification of its producer). The
82 surfactant selected for the organoclay complexes in this study is
83 tetradecyltrimethylammonium bromide (denoted as TDTMA, $C_{17}H_{38}NBr$, FW: 336.41) from
84 Sigma-Aldrich, and used without any further purification.

85

86 **Synthesis of Organoclays**

87 The synthesis of organoclay complexes was undertaken by the following procedure: 4 g of
88 montmorillonite was initially dispersed in 400 mL of deionised water with a Heidolph
89 magnetic stirrer for about 30 minutes. A predissolved stoichiometric amount of surfactant,
90 which was dissolved in 100 mL of deionised water was stirred for a further 30 minutes. The
91 dissolved surfactant was slowly added to the clay suspension at room temperature (about 28 ~
92 30 °C). The CEC of the montmorillonite is 76.4 meq/100g, which represents a measure of the

93 loading of the clay with the cationic surfactant. For instance, 1.0 CEC is applied when 76.4
94 meq/100g is intercalated into the montmorillonite. During the synthesis, a range of surfactant
95 concentration in terms of the CEC value from 0.25 CEC through 2.0 CEC was prepared and
96 labelled as 0.25 CEC-TDTMA, 0.5 CEC-TDTMA, 1.0 CEC-TDTMA, 1.5 CEC-TDTMA,
97 and 2.0 CEC-TDTMA. The mixtures were stirred for 3 hours at room temperature using a
98 Branson Ultrasonic model 250 sonifier with an output of 40 mW. All organoclay products
99 were washed free of bromide anions as determined by the use of the AgNO_3 , dried at room
100 temperature, and dried further in an oven (at 65 °C) for 12 hours. The dried organoclays were
101 ground in an agate mortar, and stored in a vacuum desiccators for a week.

102

103 **Adsorption of p-nitrophenol on the organoclays**

104 0.2 g of the prepared organoclay products were combined with 30 mL of p-nitrophenol
105 solution (4000 mg/L for the adsorption) in 50 mL centrifuged tube with plastic caps. The
106 tubes were shaken for 6 hours at 190 rpm on a shaker at the room temperature. The initial pH
107 of p-nitrophenol is 6.0. After shaking, the mixture was centrifuged at 3500 rpm for 20
108 minutes. After the centrifugation, the p-nitrophenol concentration in the aqueous phase was
109 determined by a UV-Vis spectrophotometer at 317 nm with a detection limit of 0.05 mg/L.

110 The p-nitrophenol uptake on montmorillonite and/or organoclays was calculated by the
111 following equation: $Q = (C_o - C_e) \cdot V / m$, Q is the uptake of p-nitrophenol, C_o is the initial
112 concentration, C_e is the equilibrium concentration, V is the volume of p-nitrophenol solution
113 and m is the mass of the adsorbent. The losses of the p-nitrophenol by both photochemical
114 decomposition and volatilisation were found to be negligible during adsorption [9]. The
115 obtained montmorillonite and organoclays adsorbed with p-nitrophenol were labelled as
116 montmorillonite-4000, 0.25 CEC-4000, 0.5 CEC-4000, 1.0 CEC-4000, 1.5 CEC-4000 and
117 2.0 CEC-4000.

118

119 **Characterisation methods**

120 **X-ray diffraction (XRD)**

121 The pure montmorillonite and synthesised organoclays were pressed in stainless and
122 samples holders. Powdered X-ray diffraction (XRD) patterns were recorded using $\text{CuK}\alpha$

123 radiation ($\lambda = 1.54\text{\AA}$) on a Philips PAN analytical X'pert PRO diffractometer operating at 40
124 kV and 40 mA with 0.25° divergence slit, 0.25° anti-scatter between 5 and 15° (2θ) at a step
125 size of 0.0167° . For XRD at low angle section, it was between 1.5 and 8° (2θ) at a step size of
126 0.0167° with variable divergence slit and 0.125° anti-scatter. In addition, the organoclays
127 adsorbed p-nitrophenol was applied as the same method prior to powder XRD analysis.

128

129 **Transmission electron microscopy (TEM)**

130 A JEOL 1010/2100 transmission electron microscopy was used to investigate
131 microstructure of organoclays. All samples were ultrasonically dispersed in absolute ethanol
132 solution and a small drops of suspension dissolved in ethanol solution was prepared on
133 carbon-coated films and dried in an oven at 60°C for 20 mins for TEM studies.

134

135 **Brunauer, Emmett, and Teller (BET)**

136 With a Micromeritics Tristar 3000 automated gas adsorption surface area analyses based
137 upon N_2 adsorption/desorption were carried out. From the measurement, pore structure
138 parameters were characterised at liquid nitrogen temperature. Before the measurement, the
139 samples were pre-heated at 90°C under the flow of N_2 on a Micromeritics Flowprep 060
140 degasser.

141

142 **X-ray photoelectron spectroscopy (XPS)**

143 The X-ray photoelectron spectroscopy (XPS) was applied using a Kratos AXIS Ultra with
144 a monochromatic Al X-ray source at 225W. A small amount of each fine powdered sample
145 was applied to the double sided adhesive tape on a standard Kratos Axis sample bar. This was
146 attached to the sample rod of the Load Lock system for initial evacuation to 10^{-6} Torr. The
147 sample bar was then transferred to the UHV sample analysis chamber (SAC) for collection of
148 X-ray photoemission spectra. A survey scan was run for each analysis from 0 to 1200eV with
149 a dwell time of 100 ms, pass energy of 160 eV at step of 1 eV with one sweep. In addition, a
150 high resolution analysis was undertaken at 40 eV at steps of 50 meV. The obtained spectra

151 were analysed using Casa XPS version 2.3.14 software, which enabled the baseline (Shirley
152 baseline applied) and various data handling procedures.

153

154 **Thermogravimetric analysis (TGA)**

155 Thermogravimetric analysis of the pure montmorillonite and synthesised organoclays were
156 obtained by using TA instruments Inc. Q500 high-resolution TGA operating at ramp
157 5 °C/min with resolution 6.0 °C from room temperature to 1000 °C in a high purity flowing
158 nitrogen atmosphere (40 cm³/min). Approximately 40 ~ 60 mg of finely dried ground sample
159 were heated in an open platinum crucible.

160

161 **Fourier transform infrared spectroscopy (FT-IR)**

162 Fourier transform infrared spectroscopy (FT-IR) was applied in this work, and the spectra
163 were obtained using a Nicolet Nexus 870 FT-IR spectrometer with a smart endurance single
164 bounce diamond ATR cell. Sixty four scans were collected for each measurement over the
165 spectral range of 550 ~ 4000 cm⁻¹. Spectral manipulation such as baseline adjustment,
166 smoothing, and normalisation were performed by using the GRAMS software package
167 (Galactic Industries Corporation, Salem, NH, USA). Band component analysis was
168 undertaken using the Jandel ‘Peakfit’ software package, which enables the type of fitting
169 function to be selected and allows specific parameters to be fixed or varied accordingly. Band
170 fitting was performed using a Lorentzian-Gaussian cross-product function with the minimum
171 number of component bands used for the fitting process. The peak fitting was produced
172 squared correlation (R^2) greater than 0.991.

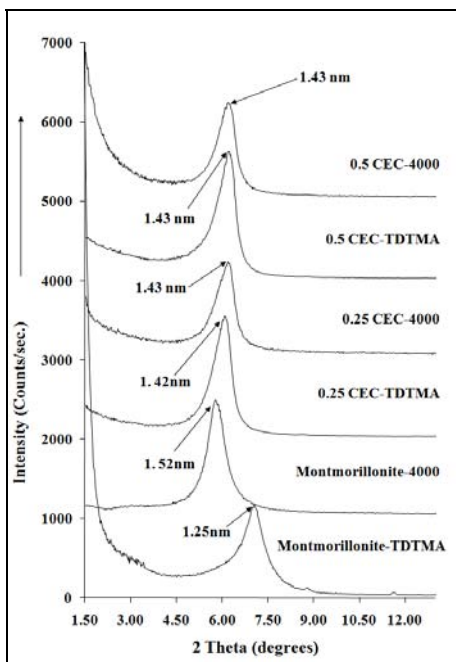
173

174 **Results and Discussion**

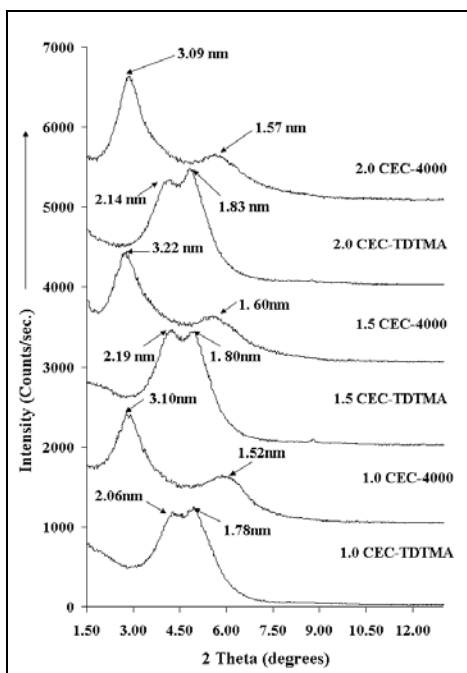
175 **X-ray diffraction (XRD)**

176 X-ray diffraction (XRD) is one of the most useful techniques to probe the structural
177 geometry and texture of organoclays. The basal spacing information of organoclays obtained
178 from XRD provides the intercalation and molecular structure configuration of the organic
179 surfactant into the clay layers. The XRD patterns of montmorillonite, and organoclays

180 prepared at different surfactant loadings with and without adsorbed p-nitrophenol are
181 presented in Figures 1 and 2. The increasing basal spacing of untreated montmorillonite and
182 organoclays is clearly seen Figures 3 and 4.



183
184 **Figure 1.** XRD patterns of montmorillonite, 0.25 CEC, and 0.5 CEC with and without
185 adsorbed p-nitrophenol

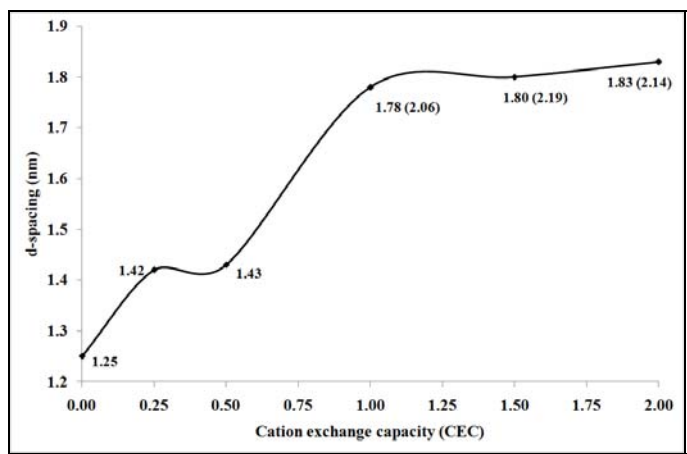


186
187 **Figure 2.** XRD patterns of 1.0 CEC, 1.5 CEC, and 2.0 CEC with and without adsorbed p-
188 nitrophenol

189 The sodium exchanged montmorillonite has a d-spacing of 1.25 nm, which was expanded to
190 1.52 nm upon adsorption of p-nitrophenol. This expansion indicated that p-nitrophenol is
191 adsorbed between the clay layers. For an ion exchanged of 0.25 CEC surfactant concentration
192 level, a basal spacing of 1.42 nm is observed, and the value of 1.43 nm is observed upon the
193 adsorption of p-nitrophenol. The basal spacing of 1.43 nm is observed at the 0.5 CEC-
194 TDTMA and the expansion remains at 1.43 nm after adsorption of p-nitrophenol on the 0.5
195 CEC organoclay. Upon increasing the surfactant loading to 1.0 CEC surfactant concentration,
196 two d(001) spacings (1.78 nm and 2.06 nm) are observed, and there are two spacings at 1.52
197 and 3.10 nm observed upon adsorption of p-nitrophenol. At 1.5 CEC surfactant concentration
198 level, the organoclays also have the double overlapping expansion peaks at 1.80 and 2.19 nm,
199 which have a slightly more intense peak than that of organoclays at 1.0 CEC surfactant
200 concentration level. Upon adsorption of p-nitrophenol, the d spacing changed to 1.60 and
201 3.22 nm, respectively. The basal spacings at 2.0 CEC resulted in basal spacings of 1.83 and
202 2.14 nm. Where upon adsorption of p-nitrophenol, spacings of 1.57 and 3.09 nm are observed.
203 Depending on the concentration of surfactant (TDTMA), the expansion of the layers in each
204 step occurred. Based on the result, the configuration of the structure of the molecules between
205 clay unit layers is suggested.

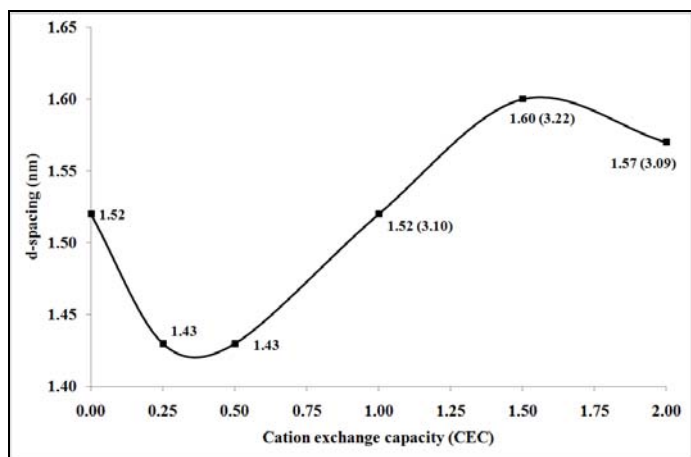
206 In 0.25 CEC-TDTMA and 0.5 CEC-TDTMA, the d values are 1.42 and 1.43 nm,
207 respectively, and this implies monolayer arrangement in the interlayer space of
208 montmorillonite. From 1.0 CEC-TDTMA to 2.0 CEC-TDTMA, the two basal spacings at
209 1.78 nm (with a shoulder of 2.06 nm), 1.80 nm (with a shoulder of 2.19 nm) and 1.83 nm
210 (with a shoulder of 2.14 nm) indicate the two arrangement of the TDTMA surfactant
211 molecule within the clay mineral layers; the first basal spacing of 1.78 nm, 1.80 nm, and 1.83
212 nm is attributed to the arrangement of the surfactant molecule within the clay layers either
213 flat or perpendicular to the clay siloxane surface, while 2.06 nm, 2.19 nm, and 2.14 nm basal
214 spacings are ascribed to the surfactant molecules at right angles to the clay mineral surface.
215 Hence, at the higher CEC values from 1.0 to 2.0 CEC, both lateral bilayer and pseudo-
216 trimolecular layer structures of surfactant are observed. Upon adsorption of p-nitrophenol on
217 the organoclay the basal spacing for untreated montmorillonite was a slight expansion of the
218 clay layers from 1.25 nm to 1.52 nm, while 0.25 CEC-4000 and 0.5 CEC-4000 organoclays
219 both have spacing of 1.43 nm. This unchanged value may suggest that p-nitrophenol has
220 penetrated and bonded to the surfactant molecules in the clay layers. However, for the 1.0
221 CEC-4000 organoclays, there are two distinct basal spacings at 1.52 and 3.10 nm. The first

222 spacing of 1.52 nm is assigned to the organoclay with minimal p-nitrophenol adsorption,
 223 while the expansion of 3.10 nm is assigned to surfactant molecules with adsorbed p-
 224 nitrophenol between clay layers. Similar values of basal spacings are also observed upon
 225 reaching the 1.5 CEC-4000. By comparison with the previous studies by Xi et al.,[10] and
 226 Liu et al.,[11] similar configuration changes were obtained. As the surfactant concentration
 227 increased, the configuration structure changes from a lateral monolayer to lateral
 228 bilayer/pseudo-trimolecular layer structure were formed. However, the paraffin layer
 229 arrangement was not observed at the higher surfactant concentration in this study, and this
 230 may be due to the different molecular sizes of the surfactants. In addition, this result may be
 231 affected by the differences in the measurements and in the control humidity.



232

233 **Figure 3.** d(001) basal spacing of montmorillonite and organoclays



234

235 **Figure 4.** d(001) basal spacing of montmorillonite and organoclays with adsorbed p-
 236 nitrophenol

237

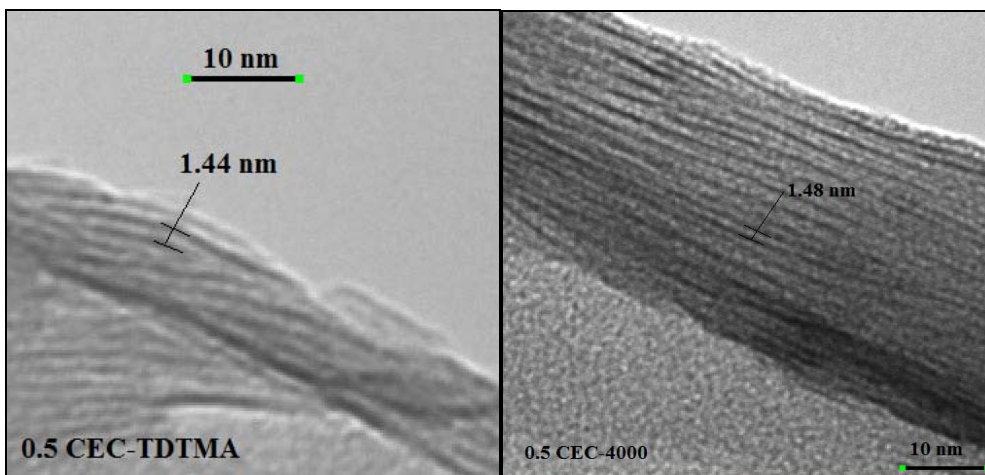
238 **Transmission electron microscopy (TEM)**

239 A selection of TEM images of 0.5 CEC-TDTMA, 2.0 CEC-TDTMA, 0.5 CEC-4000, and
240 2.0 CEC-4000 organoclays are present in Figure 5. The images of 0.5 CEC-TDTMA shows
241 that the spacing of 1.44 nm, which is in good agreement with the value of 1.43 nm obtained
242 from XPD data. The 0.5 CEC-4000 shows that the spacing of 1.48 nm, which is slightly
243 bigger than the spacing of 1.44 nm measured by XRD. There are multiple interlayer spacings
244 are found at 2.20, 2.15, 1.85, 1.80 nm for 2.0 CEC-TDTMA. In comparison with the value of
245 2.14 and 1.83nm obtained from XRD, there are similar spacings obtained. In addition, upon
246 the adsorption of p-nitrophenol, 2.0 CEC-4000 has also several spacings at 1.32, 1.50, 1.52,
247 1.78, and 3.02 nm. The observed 3.02 and 1.52 nm are well matched with that of XRD values.
248 Compared to TEM, where various basal spacings were observed, only average were shown
249 by XRD. From the results of the TEM technique, it is proposed that layers are not only linear
250 but also curved or bent. Due to the ununiformed structure, several different interlayer
251 spacings within organoclays are observed.

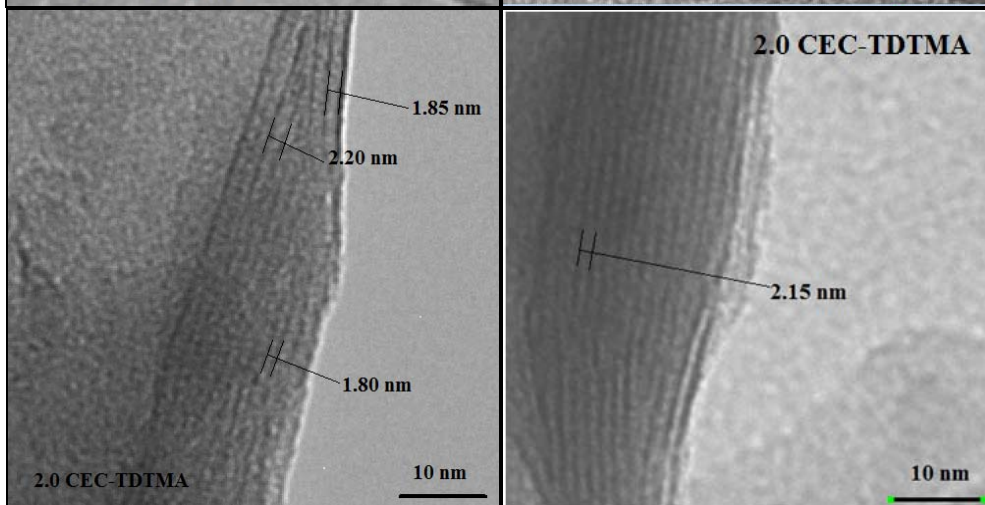
252

253

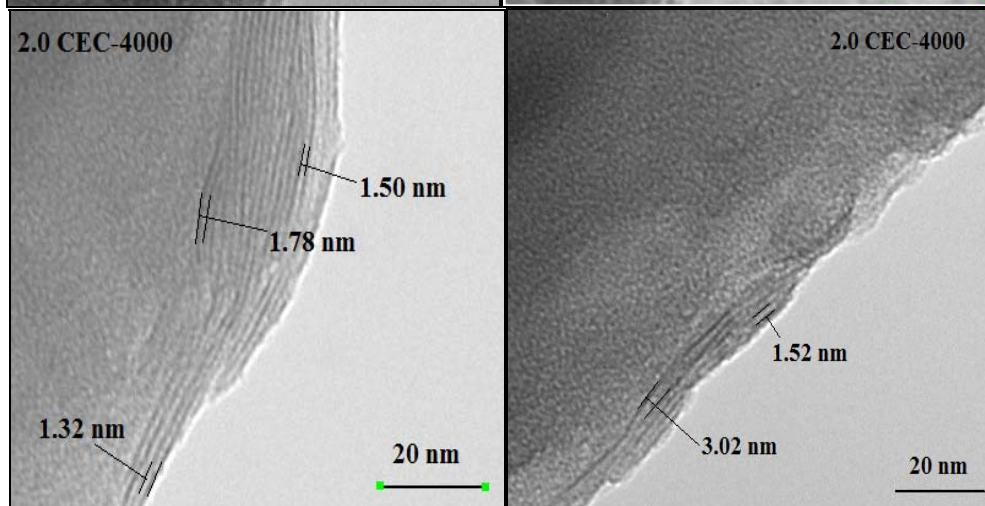
254



255



256



257

Figure 5. TEM images of 0.5 CEC-TDTMA, 2.0 CEC-TDTMA, 0.5 CEC-4000, and 2.0

258

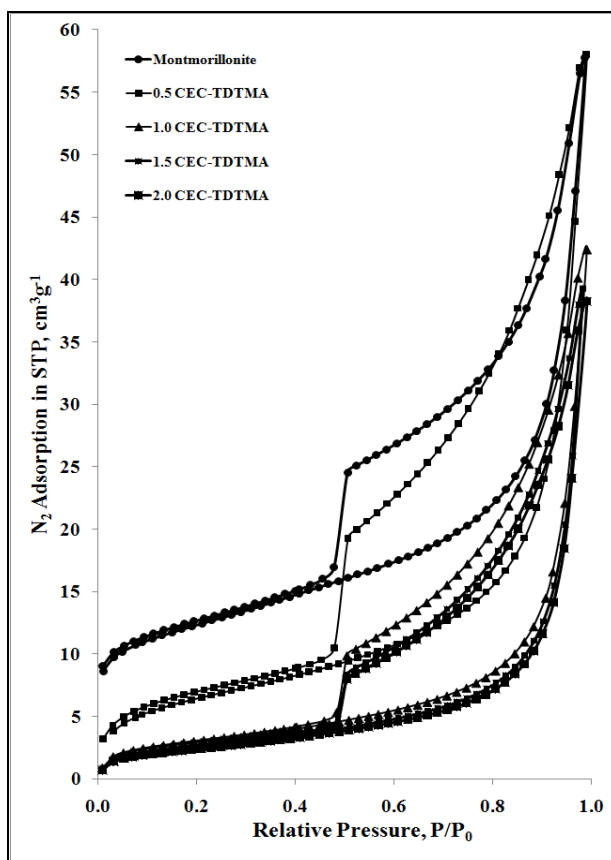
CEC-4000

259

260

261 **Surface area measurement**

262 The BET method, the N₂ adsorption-desorption measurements of the clay and organoclays,
263 was used to study the porosity and textual properties at liquid N₂ temperature. The results are
264 shown in Figure 6. The Figure indicates that N₂ adsorption isotherms of clay and organoclays
265 exhibit a Type II sorption behaviour in the classification of Brunauer, Deming, Deming and
266 Teller (BDDT) [12]. It is clearly seen that the same form of isotherms and hysteresis loop are
267 seen in both montmorillonite and organoclays. The large uptake of nitrogen is observed close
268 to the saturation pressure, and this apparent step in adsorption branch with a sharp decline in
269 the desorption branch implies the presence of mesoporosity [13-15]. This is in agreement
270 with the pore diameter calculated from the Barrett-Joyner-Halenda (BJH) desorption isotherm
271 as shown in Table 1. As shown in Figure 6, the adsorption of N₂ of montmorillonite is much
272 higher than that of the other organoclays. This indicates that montmorillonite possesses a
273 higher specific surface area. It is known that the specific surface area (S_{BET}), pore volume
274 (V_p), and pore size distribution can be calculated and the calculated parameters are
275 summarised in Table 1.



276

277 **Figure 6.** Nitrogen adsorption-desorption isotherms of montmorillonite and organoclays

278
279

Table 1 BET specific surface area (S_{BET}), pore volume (V_p) and pore diameter for montmorillonite and organoclays

Sample ID	$S_{\text{BET}}, \text{m}^2 \cdot \text{g}^{-1}$	$V_p, \text{cm}^3 \cdot \text{g}^{-1}$	Mean D, nm	
			BET ^b , nm	BJH ^c , nm
Montmorillonite	42.7074	0.089560	8.3644	7.6372
0.5 CEC-TDTMA	23.7631	0.086054	14.5296	7.2432
1.0 CEC-TDTMA	10.8419	0.065352	24.2003	8.2589
1.5 CEC-TDTMA	9.4323	0.060594	25.7665	8.6636
2.0 CEC-TDTMA	8.8507	0.059009	26.7219	8.7463

280 ^a BJH desorption cumulative pore volume of pores between 1.7 and 300 nm in diameter.

281 ^b Adsorption average pore diameter ($4V/A$ by BET).

282 ^c Barrett-Joyner-Halenda (BJH) desorption average pore diameter ($4V/A$).

283

284 As shown in Table 1, the BET surface area of montmorillonite and organoclays decreases
285 in order: Montmorillonite >> 0.5 CEC-TDTMA > 1.0 CEC-TDTMA > 1.5 CEC-TDTMA >
286 2.0 CEC-TDTMA. Especially, the surface area drops dramatically from montmorillonite to
287 0.5 CEC-TDTMA. It is also found that the pore volume for organoclays decreases with an
288 increase of loaded surfactants and this will be further discussed using TGA results. The pore
289 size is also related to the loading of surfactant, and organoclays with high surfactant loadings
290 have an enlarge pore size than that with low surfactant loadings. As shown in Table 1, using
291 specific surface area and pore volume, there are two different groups made. The surfactant
292 loading concentration up to 0.5 CEC-TDTMA with 23.7631 m^2/g and pore volume of
293 0.086054 cm^3/g . The other group includes 1.0 CEC-TDTMA to 2.0 CEC-TDTMA with
294 similar surface area of 8.8507 ~ 10.8419 m^2/g and the pore volume of 0.059 ~ 0.065 cm^3/g .
295 The organoclays at higher concentrations (1.0 ~ 2.0 CEC TDTMA) have the lower BET
296 surface area and pore volume, whereas montmorillonite has a largest BET surface area and
297 pore volume. From the result, it is assumed that the pore volume and surface area are not a
298 key factor in terms of control the affinity between organoclays and organic pollutants[16].
299 Hence, the loaded surfactant is highly important to determine the sorption mechanism onto
300 organoclays.

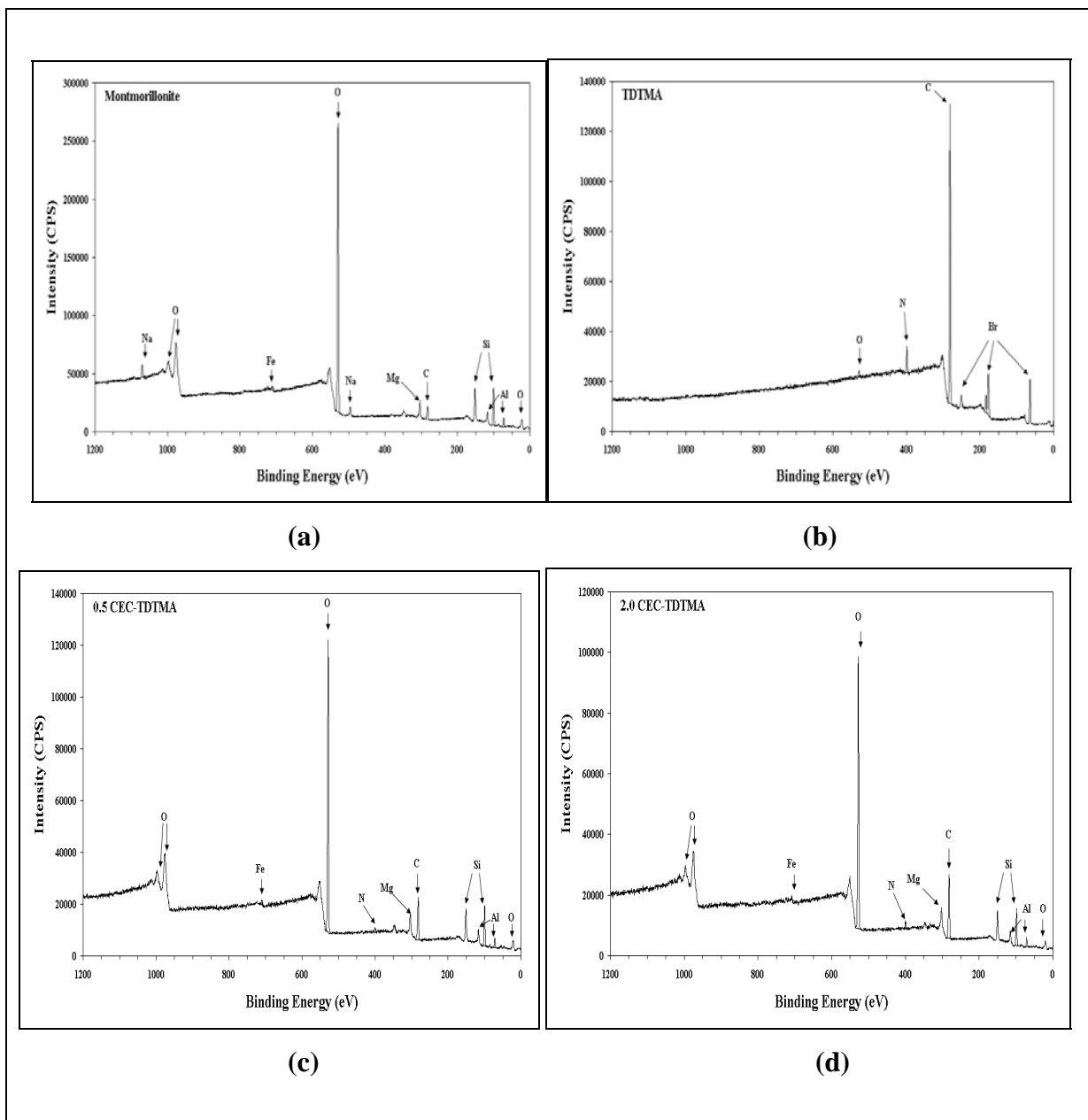
301

302

303 **X-ray photoelectron spectroscopy (XPS)**

304 X-ray photoelectron spectroscopy (XPS) is used to further determine the surface chemical
305 composition and chemical states of prepared materials. This technique has been widely
306 applied for the investigation of top few layers of material surface with partially filled valance
307 band. Figure 7 shows the XPS survey scans of montmorillonite, surfactant (TDTMA), and
308 representative organoclays (0.5 CEC-TDTMA and 2.0 CEC-TDTMA). The survey scan has
309 an obvious verification in the presence of sodium, silicon, aluminium, iron, and magnesium
310 in the montmorillonite. It has also indentified the presence of carbon, nitrogen, and bromine
311 in TDTMA. A minor amount of carbon in montmorillonite and oxygen in TDTMA is
312 observed as a result of adsorbed CO₂ [17]. The peaks of magnesium and iron (trace amount)
313 are seen in the scans of organoclays, whereas the sodium peak is disappeared. Both
314 magnesium and iron are located in the montmorillonite structure rather than in the interlayer.
315 The absence of sodium ions in the organoclays confirms the sodium ions are exchanged when
316 cationic surfactants are introduced in the interlayer. The prominent peaks of carbon and
317 silicon are recorded in the montmorillonite and organoclays, the ratio of C/Si has been found
318 and the ratio increased in the following order: 0.34 (montmorillonite) < 1.58 (0.5 CEC-
319 TDTMA) < 1.70 (1.0 CEC-TDTMA) < 1.82 (1.5 CEC-TDTMA) < 2.55 (2.0 CEC-TDTMA).
320 The intercalation of surfactant increases with an increased loading. Meanwhile, the Al/Si
321 ratio dropped from 0.42 (montmorillonite) to 0.35 (1.0 CEC-TDTMA) and further 0.34 (2.0
322 CEC-TDTMA). As the intercalation of surfactants increases in the interlayer, the distance
323 between Al-O(OH) octahedral sheets and two Si-O tetrahedral sheets in the structure of
324 montmorillonite is expanded, and hence, the detecting ratio of Al and Si is lower. This is
325 further investigated by the high-resolution XPS scans.

326



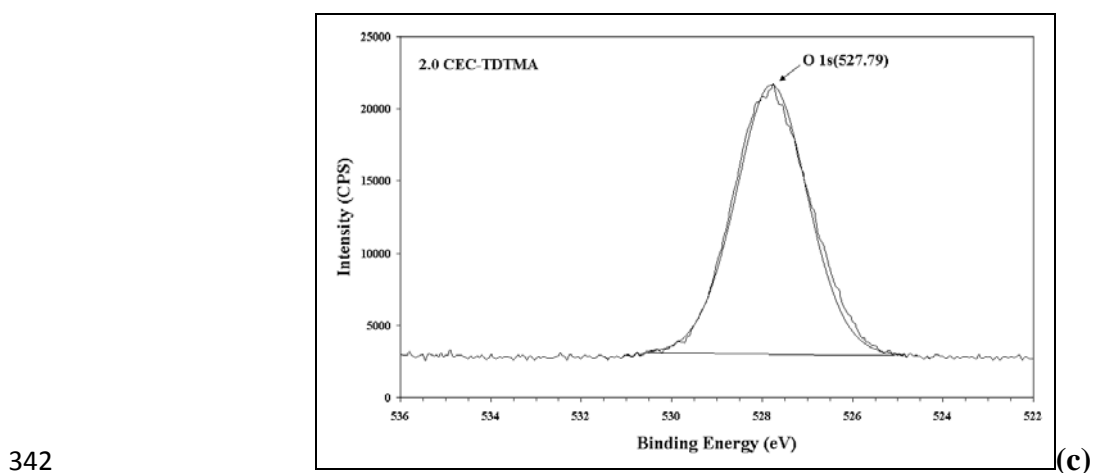
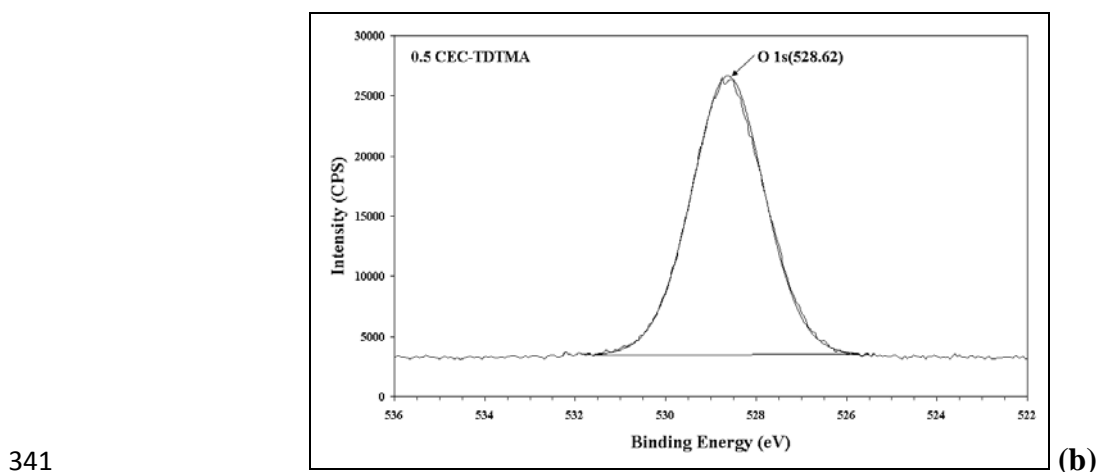
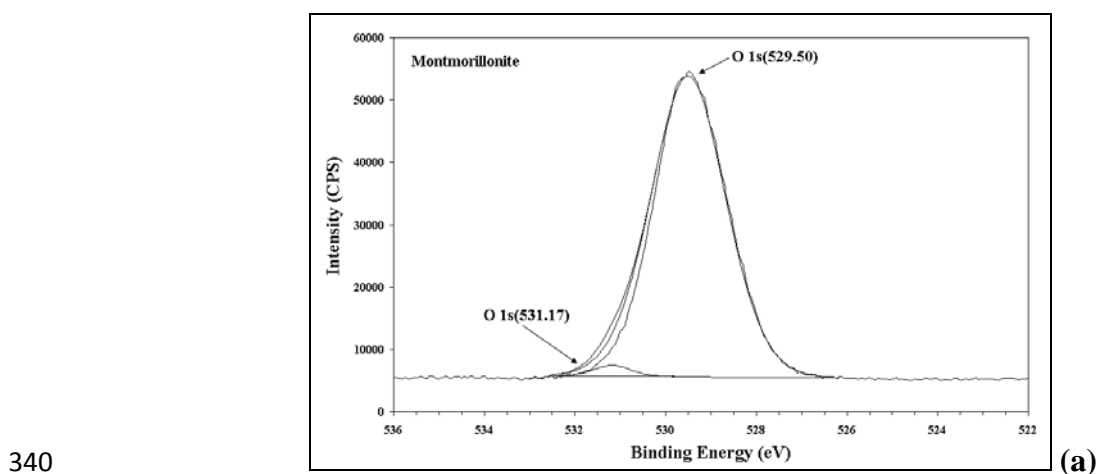
327

328 **Figure 7.** XPS survey scan spectra of montmorillonite (a), surfactant (b), and organoclays (c
 329 and d)

330

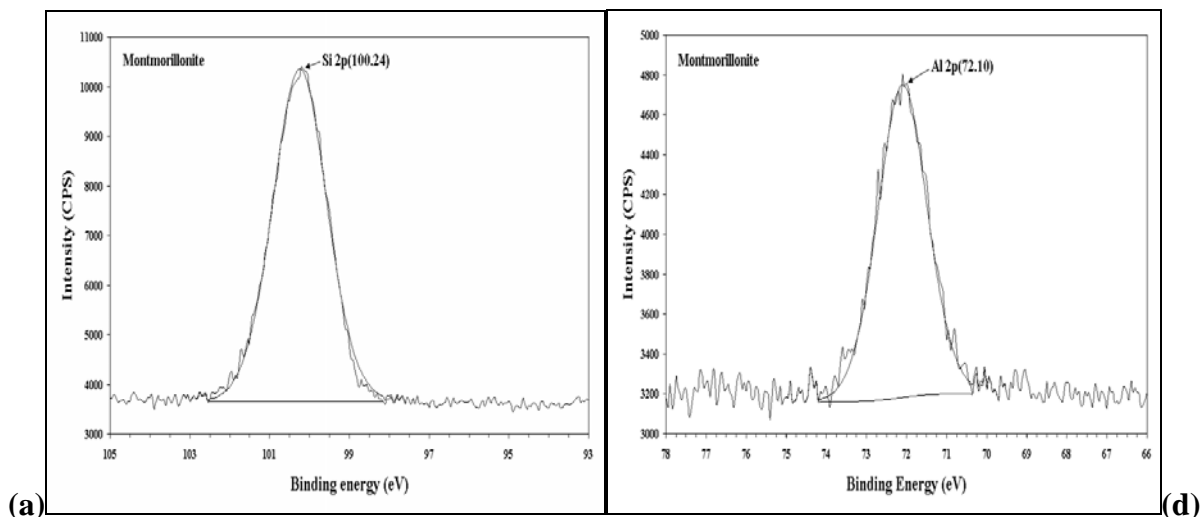
331 The high resolution XPS spectra of O 1s of montmorillonite and organoclays are compared
 332 in Figure 8. It hardly distinguishes between O and OH in the montmorillonite. The intense
 333 main peak with the binding energy of 529.42 eV corresponds to O(OH) in the
 334 montmorillonite structure, while the other small peak at 531.17 eV is considered as oxygen in
 335 H₂O. The small trace of an oxygen peak is not observed in organoclays due to their
 336 hydrophobic characteristics and the binding energy has decreased from 528.64 to 527.59 eV

337 when the surfactant loading increased from 0.5 to 2.0 CEC. This reduced binding energy is
338 also observed in the high resolution Si 2p and Al 2p spectra (Figure 9). The decreased
339 binding energy of both O 1s and Si 2p shows the change of interlayer structure in organoclays.

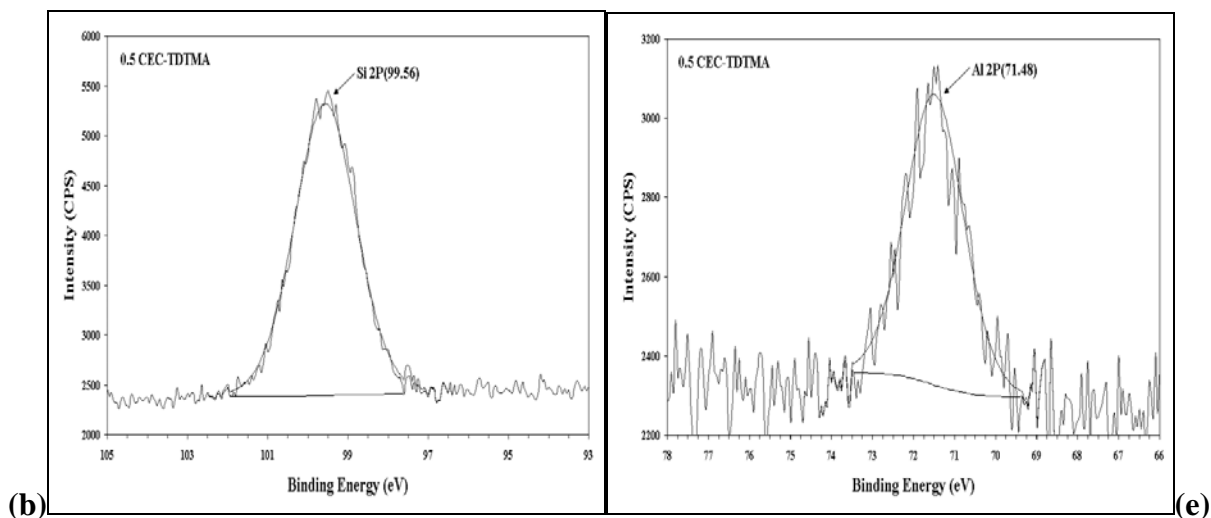


343 **Figure 8.** XPS high resolution spectra of O 1s for montmorillonite (a), organoclays (b, c)

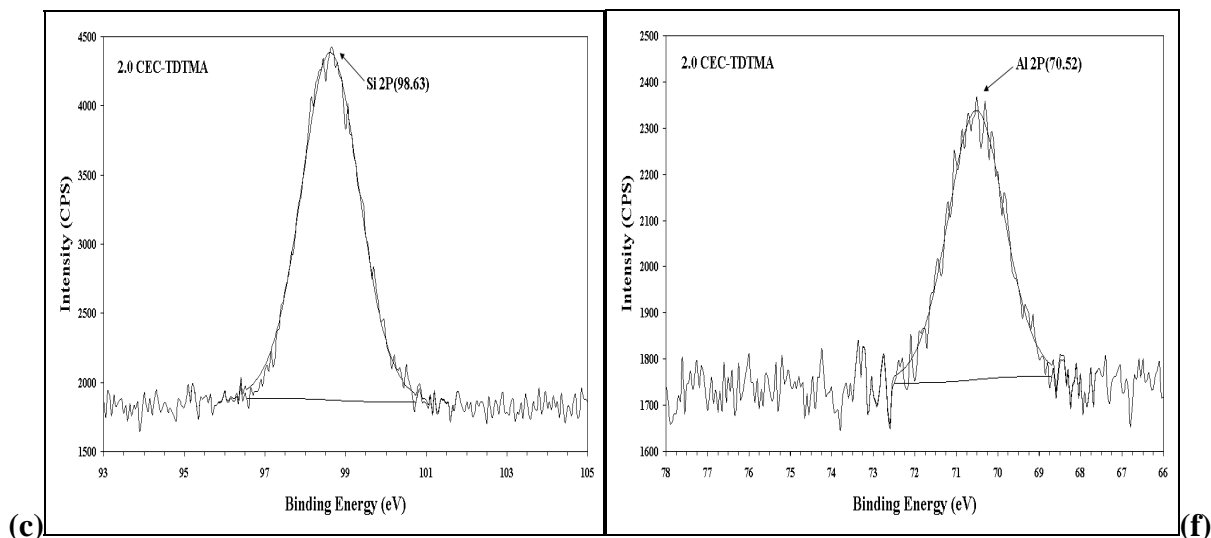
344



345



346



347

Figure 9. XPS high resolution spectra of Si 2p and Al 2p for montmorillonite (a and d),
348 organoclays (b, e and c, f)

349

350 In the high-resolution XPS spectra, C 1s of organoclays is characterised by two transitions
 351 at 281 and 283 eV (Figure 10). The transition peak at about 281 eV is ascribed to the C-C
 352 bond in the long chain of the surfactant, and the C-N bond has a binding energy of about 283
 353 eV. Table 2 shows the results for curve fitted binding energy and their atomic contents (at. %) of
 354 the highly resolved C 1s XPS spectra present in Figure 10. The binding energy for C-C
 355 bond increases gradually as the surfactant loading increases from 0.25 to 2.0 CEC in
 356 organoclays, while C 1s binding energy of C-N bonds remain similar. The result of C 1s
 357 binding energy for C-C bond shows that the loading surfactant affects the C 1s binding
 358 energy in organoclays. When the cationic surfactant is intercalated in the interlayer structure,
 359 the nitrogen as a head group in the alkyl surfactant interacts with the negative clay due to the
 360 electrostatic interaction. From the interaction between nitrogen head group and negative clay
 361 surface, the binding energy for C-N remains similar and the binding energy is not changed
 362 with an increase of the loading surfactant. However, the increasing binding energy for C-C
 363 bond may be explained by the arrangement of surfactant in the interlayer. At the lower
 364 concentration of the surfactant, the alkyl chains are parallel to the silica clay surface within
 365 the interlayer (e.g. monolayer or bilayer). As a result of an increase amount of loading
 366 surfactant, the arrangement of surfactant is at right angle to the clay mineral surface such as
 367 pseudo-trilayer or paraffin structures, which can be described using XRD. The increased
 368 packing density within the interlayer causes the increase of C-C binding energy in the alkyl
 369 chain.

370 **Table 2 Binding energy and their atomic contents of C 1s for organoclays**

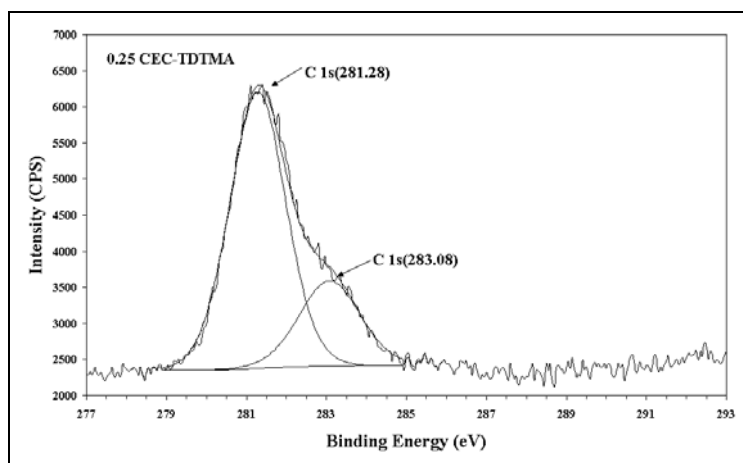
C 1s (eV)	0.25 CEC	0.5 CEC	1.0 CEC	1.5 CEC	2.0 CEC
C-C bond	281.28 (74.91)	281.34 (72.83)	281.33 (76.09)	281.34 (77.55)	281.38(84.31)
C-N bond	283.08 (25.09)	283.09 (27.17)	283.04 (23.91)	283.08 (22.45)	283.09 (15.69)

371

372

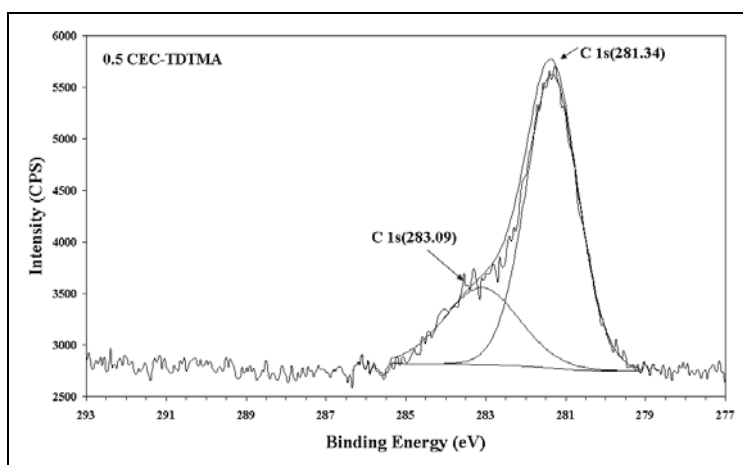
373

374



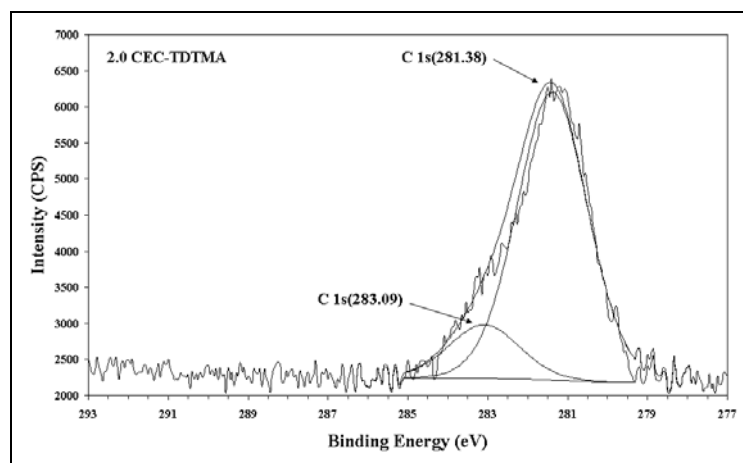
375

(a)



376

(b)



377

(c)

Figure 10. XPS high resolution spectra of C 1s for organoclays (a, b, c)

378

379

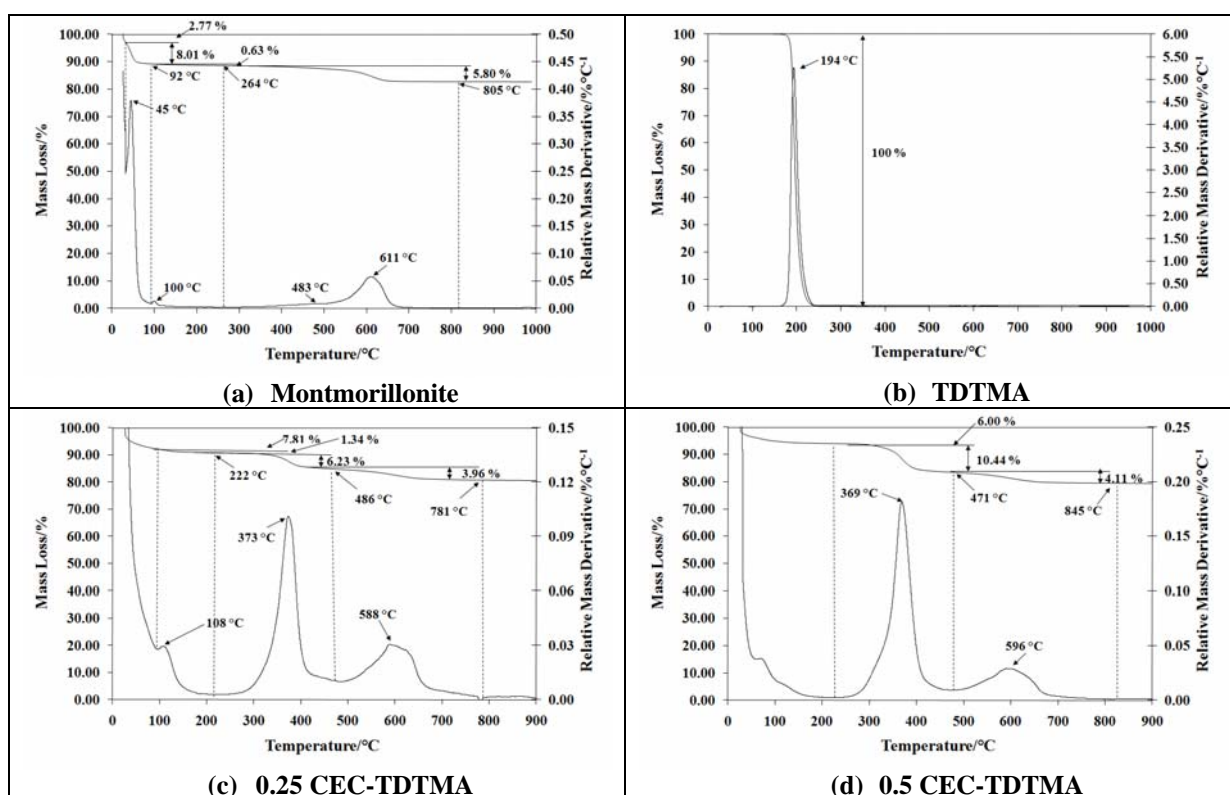
380 Bromine peaks were observed from the scan of the organoclays. However, the highly well
 381 resolved Br peaks are poorly obtained (not shown) and the content of bromine in organoclays
 382 is limited. The result indicates that the preparation of organoclays, which are free of Br ions

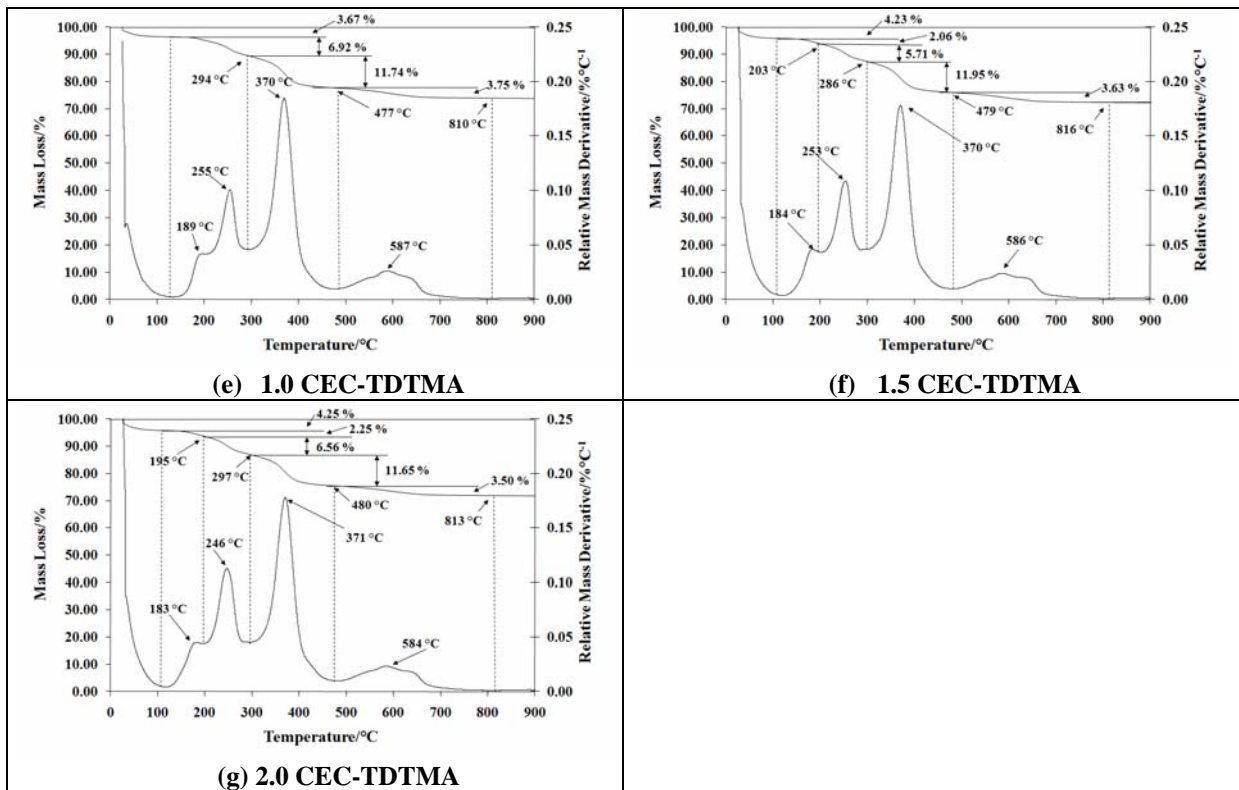
383 was successful and that the surfactant TDTMA⁺ was highly exchanged with Na⁺ within the
384 interlayer [18, 19].

385

386 Thermogravimetric analysis (TGA)

387 Thermal stability of organoclays and packing arrangement of organic cationic surfactant
388 within the organoclays at an elevated temperature were determined by TGA [4, 20, 21]. The
389 TG and DTG curves for montmorillonite and the organoclays with and without p-nitrophenol
390 are shown in Figures 11 and 12.





391

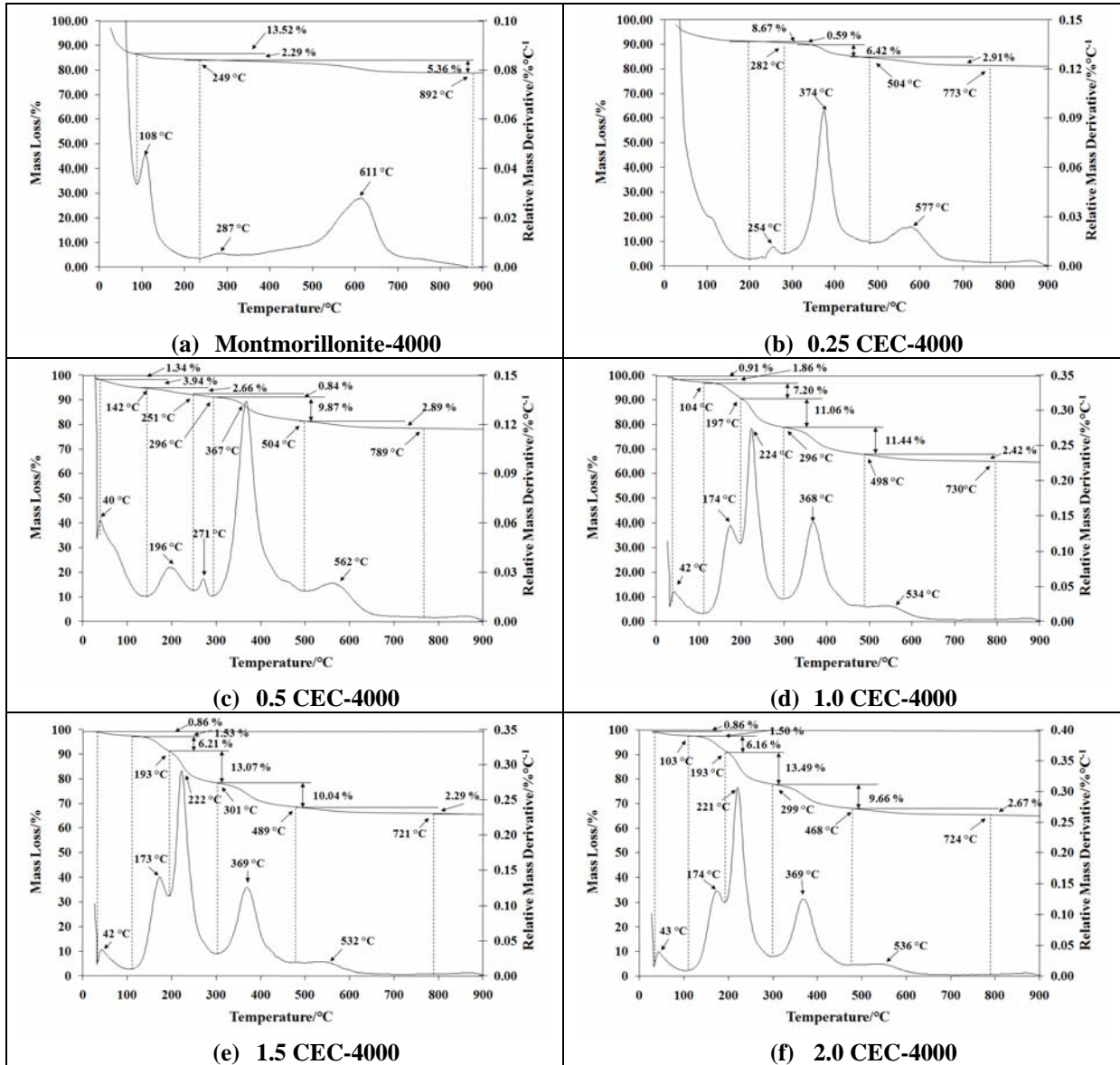
392 **Figure 11.** Thermogravimetric analyses of montmorillonite, Surfactant (TDTMA) and
 393 organoclays

394

395 In Figure 11, there are several mass loss steps observed. The first two mass loss steps of
 396 montmorillonite involve dehydration and/or dehydroxylation from room temperature and
 397 before 110 °C. The two steps are attributed to the dehydration of adsorbed water, and water
 398 molecules around metal cations such as Na^+ or Ca^{2+} in the exchangeable sites of
 399 montmorillonite. The experimental mass loss peak during the second hydration and
 400 hydroxylation of adsorption water appears only for untreated montmorillonite and 0.25 CEC-
 401 TDTMA. The montmorillonite (Figure 11(a)) shows two thermal decomposition steps at both
 402 483 and 611 °C, which were caused by the dehydroxylation of the montmorillonite. The
 403 decomposition at a lower temperature 483 °C is assigned to the loss of OH units at the end of
 404 clay layers, while the temperature at 611 °C is contributed by dehydroxylation of clay.
 405 However, the thermal decomposition of montmorillonite with adsorbed p-nitrophenol is
 406 differently shown. Figure 12(a) shows that the decomposition at 287 °C is occurred and this
 407 may be assigned to the adsorbed p-nitrophenol on the montmorillonite. It is reported by Zhou
 408 and colleagues that p-nitrophenol sublimates at 131 °C [22]. The reason for obtaining higher

409 temperature than 131 °C may be chemical reactions between the adsorbed p-nitrophenol and
 410 clay surface of montmorillonite.

411



412

413 **Figure 12.** Thermogravimetric analyses of montmorillonite and organoclays adsorbed p-
 414 nitrophenol

415

416 One small dehydration peak for 0.25 CEC-TDTMA and 0.5 CEC –TDTMA is observed at
 417 373 and 369 °C, respectively. Meanwhile, there are three peaks centred at 189/255/370 °C for
 418 1.0 CEC-TDTMA, 184/253/370 °C for 1.5 CEC-TDTMA, and 183/246/371 °C for 2.0 CEC-

419 TDTMA. The third step is considered as the loss of surfactant by comparing the peak of pure
420 surfactant, which decomposes at 194 °C, with that of untreated montmorillonite. From the
421 results, it is proposed that the number of peaks in the third step is affected by the loading of
422 the surfactant. When the concentration of the organoclay is relatively low (e.g. 0.25 CEC and
423 0.5 CEC), there is a single peak observed (the peak position around 370 °C). The reason for
424 the single peak is that the organic cations exchange with Na⁺ ions, which is mainly adhered to
425 the surface sites via electrostatic interactions [21]. With the increase of surfactant
426 concentration loading, some surfactant molecules tend to attach to the clay surface, which
427 results in the appearance of the second peak (the peak position around 250 °C). When the
428 surfactant loading has exceeded the CEC of the clay (1.0, 1.5, and 2.0 CEC), the surfactant
429 molecules strongly adhere to the clay surface by van der Waals forces, and this causes a new
430 peak to appear (at 183 ~ 185 °C). In addition, the intensity of the peaks increase with
431 increased surfactant loading. The effect causes the temperature of the decomposition
432 surfactant in the third step to increase. The second and third peaks for 1.0 CEC-TDTMA, 1.5
433 CEC-TDTMA, and 2.0 CEC-TDTMA compare closely to the pure surfactant peak at 194 °C,
434 and the temperature of the two peaks is gradually decreased.

435 The thermal decomposition peaks for organoclays with adsorbed p-nitrophenol are
436 observed in Figure 12. The two decomposition peaks of 0.25 CEC-4000 are observed at 254
437 and 374 °C. The former is attributed to the adsorbed p-nitrophenol on organoclay, while the
438 later decomposition temperature is ascribed to the combustion of the surfactant. The thermal
439 decomposition of 0.5 CEC-4000 is very different from that of the organoclays. The main
440 difference is the presence of two peaks for the loss of adsorbed p-nitrophenol. Two thermal
441 decomposition steps at 196 and 271 °C are observed with mass losses of 2.66 and 0.84 % and
442 these may be formed from the result of the removal and desorption of p-nitrophenol in the
443 clay layers[22]. The mass loss at 367 °C of 9.87 % is the loss of the surfactant. When the
444 surfactant loading is increased above 0.5 CEC, three decomposition steps are observed for 1.0,
445 1.5, and 2.0 CEC adsorbed p-nitrophenol. The three decomposition temperatures are 174, 224,
446 and 368 °C for 1.0 CEC-4000, 173, 222, and 369 °C for 1.5 CEC-4000, and 174, 221, and
447 369 °C for 2.0 CEC-4000. The first two decomposition steps are assigned to the loss of
448 surfactant, and the later decomposition temperature is attributed to the loss of intercalated
449 surfactant molecules. The adsorbed p-nitrophenol peaks, which were found at 0.25 and 0.5
450 CEC, are not observed and this is assumed that the adsorbed p-nitrophenol is decomposed
451 simultaneously with the surfactant. During the desurfactant procedure, the experimental

452 mass loss increases from 6.23 % (0.25 CEC-TDTMA) to 20.46 % (2.0 CEC-TDTMA), and
 453 100 % for the pure surfactant. The last mass loss over the temperature from 611 to 584 °C is
 454 ascribed to the loss of hydroxylation from the structural inner OH units of the
 455 montmorillonite. The dehydroxylation temperature in the organoclays upon the adsorption of
 456 p-nitrophenol is lower because of the chemical reaction between adsorbed p-nitrophenol and
 457 siloxane clay surfaces. The chemical binding of the p-nitrophenol to the inner OH units
 458 reduces the dehydroxylation temperature. The intensity of the peak decreases with increase in
 459 the surfactant concentrations.

460 **Table 3** TGA results of montmorillonite (MMT), surfactant (TTMAB), and organoclays with
 461 and without adsorbed p-nitrophenol

Sample	Dehydration/ hydroxylation		Dehydration/ dehydroxylation		Desurfactant		Dehydroxylation (OH unit)	
	% mass loss (step 1)	Temp. (°C)	% mass loss (step 2)	Temp. (°C)	% mass loss (step 3)	Temp. (°C)	% mass loss (step 4)	Temp. (°C)
Montmorillonite	10.78	-	0.63	100	-	-	5.80	483,611
0.25 CEC-TDTMA	7.81	-	1.34	108	6.23	373	3.96	588
0.5 CEC-TDTMA	6.00	-	-	-	10.44	369	4.11	596
1.0 CEC-TDTMA	3.67	-	-	-	18.66	189, 255, 370	3.75	587
1.5 CEC-TDTMA	4.23	-	-	-	19.72	184, 253, 370	3.63	586
2.0 CEC-TDTMA	4.25	-	-	-	20.46	183, 246, 371	3.50	584
TTMAB	-	-	-	-	100	194	-	-
MMT-4000	13.52	-	2.29	108			5.36	611
0.25 CEC-4000	8.67	-	-	-	0.59 ^a ,6.42	254 ^b ,374	2.91	577
0.5 CEC-4000	5.28	-	-	-	13.37	196 ^b , 271 ^b , 367	2.89	562
1.0 CEC-4000	2.77	-	-	-	29.70 ^c	(174, 224, 368) ^d	2.42	534
1.5 CEC-4000	2.39	-	-	-	29.32 ^c	(173, 222, 369) ^d	2.29	532
2.0 CEC-4000	2.36	-	-	-	29.31 ^c	(174, 221, 369) ^d	2.67	536

- 462 ^a Mass loss of adsorbed p-nitrophenol
463 ^b Thermal decomposition of adsorbed p-nitrophenol
464 ^c Mass loss of adsorbed p-nitrophenol and surfactant
465 ^d Thermal decomposition of adsorbed p-nitrophenol and surfactant

466

467 It is summarised that four steps are discernible from DTG figure (Figures 11 and 12, and
468 Table 3). The first two steps involve the dehydration and/or dehydroxylation of adsorbed
469 water and water adsorbed by the metal cations. The decomposition of the surfactant at the
470 temperature between 184 and 373 °C is observed in the third step. Upon the adsorption of p-
471 nitrophenol, the decomposition of adsorbed p-nitrophenol is determined and the organoclays
472 above 0.5 CEC, p-nitrophenol is decomposed simultaneously with the surfactant. In the last
473 step, the dehydroxylation of OH structural units in the clay is observed at the temperature
474 from 611 to 584 °C. It is noted that the temperatures for the loss of p-nitrophenol are
475 relatively greater than that for the pure p-nitrophenol, and this indicates the p-nitrophenol
476 molecules are strongly bonded to the organoclay, which leads to lower dehydroxylation
477 temperature.

478

479 **Fourier transform Infrared spectroscopy (FT-IR)**

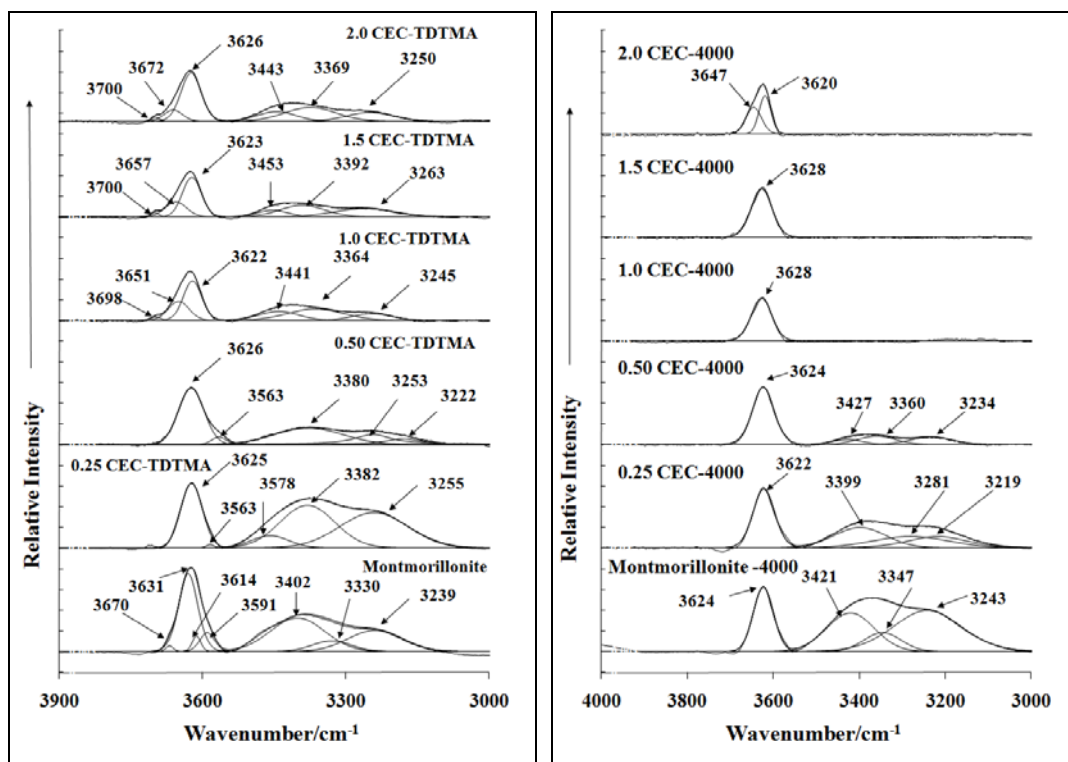
480 Infrared spectroscopy is an essential method to probe the molecular environment of the
481 intercalated surfactant for the organoclays [23]. From the observed infrared spectra, there are
482 several distinct regions: OH stretching region (3700 ~ 3000 cm⁻¹), CH stretching region
483 (2900 ~ 2800 cm⁻¹), HOH bending vibration region (1700 ~ 1600 cm⁻¹) and HCH bending
484 vibration region (1520 ~ 1400 cm⁻¹).

485 (i) OH stretching region

486

487 As shown in Figure 13, the infrared spectrum of montmorillonite and organoclays
488 (a) and with adsorbed p-nitrophenol (b) in the OH stretching region is characterised
489 by two bands at 3700 ~ 3000 cm⁻¹.

490



(a)

(b)

Figure 13. Infrared spectroscopy of montmorillonite and organoclays with and without p-nitrophenol in 3900 ~ 3000 cm⁻¹ spectral range

A sharp and intense peak at 3631cm⁻¹ for montmorillonite is assigned to OH stretching vibrations of the structural hydroxyl group, while the broad bands at 3402, 3330, and 3230 cm⁻¹ are ascribed to other water hydrogen bonded to water molecules adsorbed within the interlayer of the clay (see Figure 13a). The infrared spectrum of montmorillonite with adsorbed p-nitrophenol shows peaks in similar position to that of non-reacted montmorillonite (see Figure 13b). The bands at 3421, 3347, and 3243cm⁻¹ for montmorillonite-4000 shows an increased intensity than that of montmorillonite. However, the bands for water hydrogen bonded to other water molecules no longer existed when the surfactant loading has exceeded the CEC of the clay. This is in great agreement with the study by Zhou et. al. There is a hydrogen bond between p-nitrophenol with water for untreated montmorillonite and organoclays at low CEC concentrations [6].

509 The position of the sharp band corresponding to the OH vibrations of the structural
510 hydroxyl groups is relatively independent of the surfactant concentration. However,
511 the broad bands between $3500 \sim 3200 \text{ cm}^{-1}$ are strongly dependent on the surfactant
512 concentration loading. At low concentrations of surfactant, the infrared spectrum of
513 0.25 CEC-TDTMA showed the broad adsorption band near 3382 cm^{-1} with
514 shoulders at 3578 and 3255 cm^{-1} . The band shifted to lower wavenumber from 3382
515 to 3364 cm^{-1} with an increase in the surfactant loading up to 1.0 CEC-S. The change
516 in wavenumber is related to the environment of water. In addition, additional bands
517 at 3672 and 3700 cm^{-1} are observed above 1.0 CEC-TDTMA and this is suggested
518 that the water in the montmorillonite is gradually displaced with the surfactant
519 (TDTMA), and that the surfactant is adsorbed on other surfactant molecules, which
520 have already been adsorbed in the clay interlayer.

521

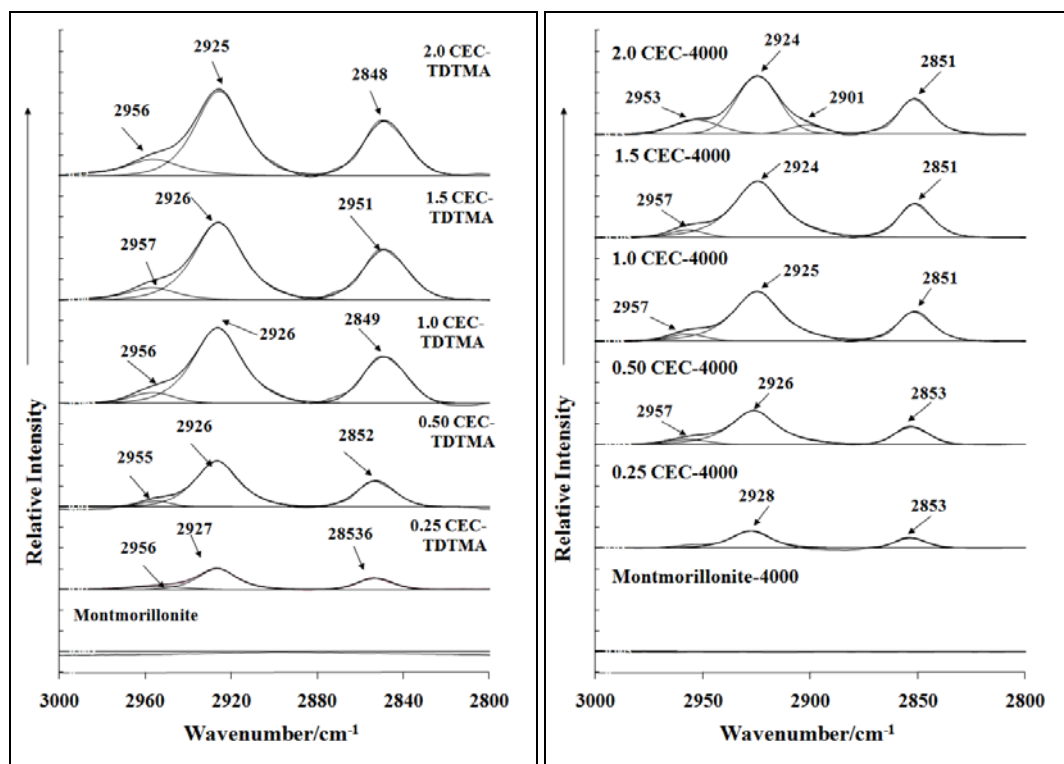
522 (ii) CH stretching region

523

524 The spectra of the CH stretching region for montmorillonite and organoclays at
525 different surfactant loadings and with p-nitrophenol are shown in Figure 14. No
526 bands attributable to the CH stretching vibrations were observed for montmorillonite
527 and montmorillonite-4000. However, infrared bands in the region between 2700 and
528 2900 cm^{-1} are observed for organoclays modified with TDTMA, and these are
529 ascribed to the asymmetric $\nu_{\text{as}}(\text{CH}_2)$ and symmetric $\nu_{\text{s}}(\text{CH}_2)$ stretching modes of the
530 surfactant. It is observed that the CH_2 asymmetric stretching modes slightly shifted
531 to lower wavenumbers upon intercalation of the surfactant molecules (from 2927 to
532 2925 cm^{-1}). The wavenumber of the symmetric CH_2 stretching modes is shifted to
533 2848 cm^{-1} from 2853 cm^{-1} . From the spectra, the wavenumbers of the asymmetric
534 and symmetric CH stretching modes of amine chains in the region indicated the
535 conformational difference between organoclays intercalated with TDTMA at the
536 various CEC concentrations. Li and Ishida [24] reported that the chains are highly
537 ordered (all-trans conformation), the wavenumbers are decreased with in accordance
538 to the increase of ordered conformers within the clay interlayers. Similarly, the
539 asymmetric and symmetric CH stretching modes shifted to low wavenumbers for
540 organoclays adsorbed with p-nitrophenol. It is suggested that these two CH
541 stretching peaks are extremely sensitive to the conformational changes of the chain
542 within the interlayer, where the wavenumber decreases as the loading of the

543 surfactant increase from 0.25 to 2.0 CEC. Similar results have been shown in some
 544 previous studies [25, 26]. Thus the wavenumbers of the CH stretching bands are
 545 highly sensitive to the conformational changes of the chain, which can be
 546 qualitatively monitored by the wavenumber shifts. In addition, a significant
 547 wavenumber shift indicates more gauche conformational molecules introduced into
 548 alkyl chain with the decrease of alkyl chain length. The conformation of adsorbed
 549 long alkyl chain surfactants progressively forms a solid like molecular environment
 550 (with high packing density and ordering), and this increase in surfactant packing
 551 density can lead to the expansion of clay mineral interlayer spaces, which can be
 552 characterised by XRD.

553



554

555

(a)

(b)

556 **Figure 14.** Infrared spectroscopy of montmorillonite and organoclays with and without p-
 557 nitrophenol in the 3000 ~ 2800 cm⁻¹ spectral range

558

559

560

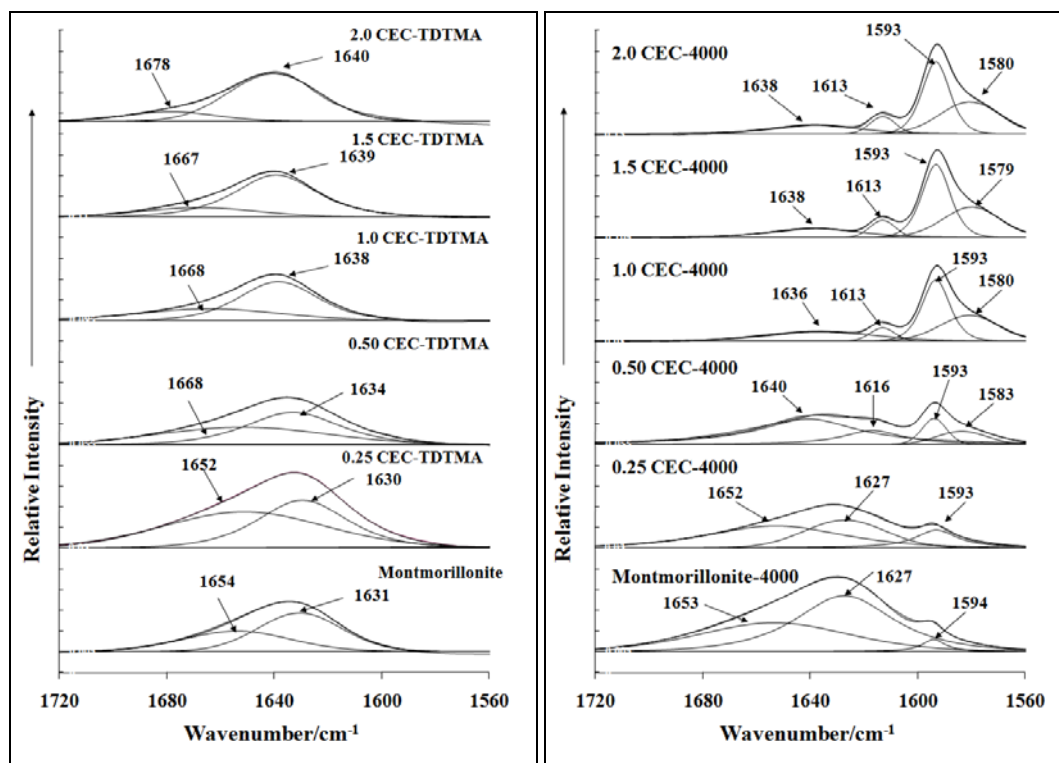
561

562 (iii) HOH and HCH bending vibrations

563

564 Figure 15a presents the changes in the adsorption bands related to the HOH bending
565 vibrations of the water molecules adsorbed on montmorillonite and organoclays.

566



567

568

(a)

(b)

569 **Figure 15.** Infrared spectroscopy of montmorillonite and organoclays with and without p-
570 nitrophenol in the 1720 ~ 1560 cm⁻¹ spectral range

571

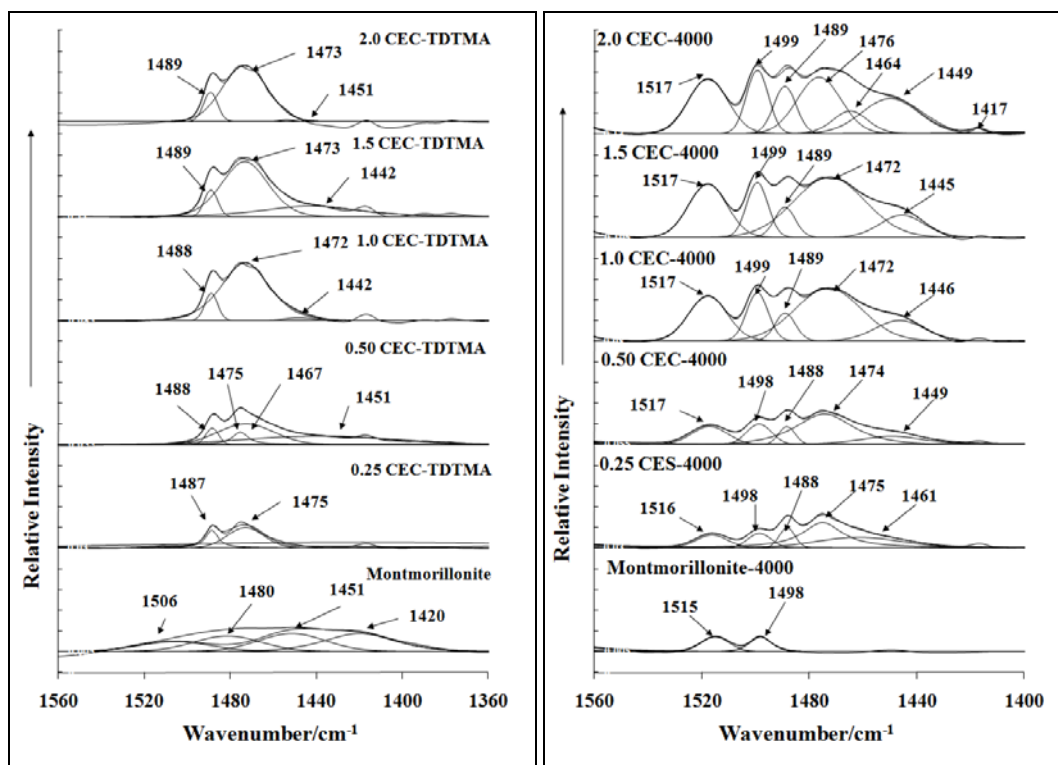
572 The positions at these bands for montmorillonite are at 1631 cm⁻¹ with a shoulder at
573 1654 cm⁻¹. It is apparent from Figure that the wavenumber of the water bending
574 peaks shifted significantly to higher wavenumbers from 1630 cm⁻¹ with the increase
575 in surfactant concentration. However, the intensity of this adsorption band decreased,
576 and this can be attributed to the amount of hydrogen bonded water molecules at the
577 high surfactant concentration level (for example, 2.0 CEC-TDTMA) is less than that
578 at lower surfactant concentrations (for example, 0.25 CEC-TDTMA).

579

580 The IR spectra of montmorillonite and organoclays with adsorbed p-nitrophenol are
581 shown in Figure 15b. The HOH bending modes for montmorillonite-4000 are
582 observed at 1627 cm^{-1} with a shoulder of 1653 cm^{-1} . An additional small peak at
583 1594 cm^{-1} appears, and this is assigned to adsorbed p-nitrophenol compared to the
584 spectra of non-reacted montmorillonite. From the literature[6], the peak at 1599 cm^{-1}
585 is ascribed to C=C aromatic stretching vibration, and the OH deformation modes of p-
586 nitrophenol is present at 1623 cm^{-1} . The wavenumber of OH bending peaks of
587 organoclays with adsorbed p-nitrophenol are gradually shifted from 1627 cm^{-1} to
588 1613 cm^{-1} with an increase of loading surfactant. The intensity of the water bending
589 modes is gradually decreased with the increase of surfactant loading, whereas the
590 intensity of p-nitrophenol at 1594 cm^{-1} is significantly increased. It is proposed that
591 p-nitrophenol is highly reacted with clay surface in the organoclays, and that as a
592 result of the intercalation of the surfactant, the surface property of montmorillonite is
593 modified from hydrophilic to hydrophobic.

594
595 Major peaks in the region between 1360 and 1560 cm^{-1} appeared to be due to the
596 methylene scissoring modes and the HCH deformation region of the TDTMA
597 intercalated montmorillonite as shown in Figure 16a.

598
599
600
601
602
603
604
605
606
607
608
609
610
611



612

613

(a)

(b)

614 **Figure 16.** Infrared spectroscopy of montmorillonite and organoclays without p-nitrophenol
 615 in the 1560 ~ 1360 cm⁻¹ spectral range

616

617 The bands in this region for montmorillonite are found at 1506, 1480, 1451, and
 618 1420 cm⁻¹. However, the band at 1506 and 1420 cm⁻¹ are not observed in the
 619 surfactant intercalated with montmorillonite. At low CEC surfactant concentration
 620 (0.25 CEC-TDTMA), two small peaks at 1487 and 1475 cm⁻¹ appeared and the
 621 intensity of these peaks increases with increase in the surfactant loading. The IR
 622 spectra of montmorillonite and organoclays with adsorbed p-nitrophenol are shown
 623 in Figure 16b. The peak at 1515 cm⁻¹ in montmorillonite is drawn by the adsorbed p-
 624 nitrophenol as antisymmetric NO₂ stretching vibrations (at 1523 cm⁻¹). The
 625 wavenumber from 1523cm⁻¹ to 1515 cm⁻¹ provides again a strong evidence for the
 626 reaction between p-nitrophenol with the montmorillonite clay surfaces. The intensity
 627 of peaks at 1498, 1488, 1475, and 1461 cm⁻¹ increases with increase in the surfactant
 628 loading.

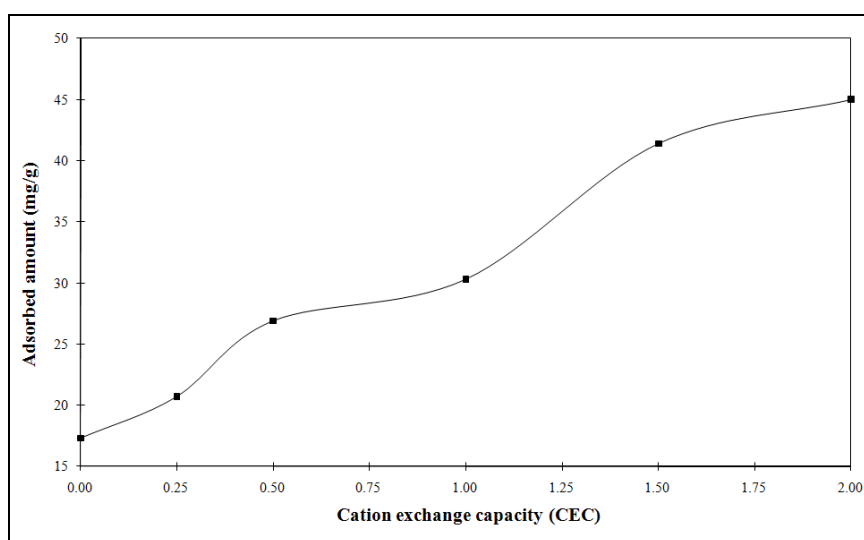
629

630

631 **Adsorption of p-nitrophenol on organoclays**

632 The prepared montmorillonite and organoclays are used for the adsorption study of p-
633 nitrophenol in water, and the results are given in Figure 17. The adsorbed amounts of p-
634 nitrophenol onto montmorillonite and organoclays increase in order: montmorillonite < 0.25
635 CEC-TDTMA < 0.5 CEC-TDTMA < 1.0 CEC-TDTMA < 1.5 CEC-TDTMA < 2.0 CEC-TDTMA.
636 The amount of adsorbed p-nitrophenol on organoclays is larger than that on montmorillonite.
637 It is also found that the addition of surfactants in organoclays would increase the amount of
638 adsorbed p-nitrophenol on organoclays. Therefore, the amount of loaded surfactant is
639 important factor to determine the adsorbed amount of p-nitrophenol onto organoclays.

640



641

642 **Figure 17.** Uptake of p-nitrophenol on montmorillonite and organoclays

643

644 From the characterisation studies of montmorillonite and organoclays, the mechanisms of
645 adsorbed p-nitrophenol are suggested. Montmorillonite has mainly electrostatic attraction
646 between p-nitrophenol and hydrated cations in interlayer space of the clay and therefore, the
647 larger d-spacing is observed in XRD pattern [27]. As the loaded surfactant molecules in the
648 interlayer space of clay, the surface properties for organoclays convert from hydrophilic to
649 hydrophobic and the intercalated surfactant molecules are also sorbed outside the clay layer.
650 From the study of BET, there are two groups of organoclays observed. For organoclays
651 prepared at relatively low surfactant loadings (0.25 and 0.5 CEC), surfactant molecules are
652 preferred to intercalated in the interlayer space of clays, hence, p-nitrophenol molecules have

653 adsorbed in the interlayer space of organoclays and the organoclays intercalated with
654 surfactant molecules are an excellent medium for partition of p-nitrophenol [28]. However,
655 organoclays prepared at surfactant loadings exceed 1.0 CEC, surfactant molecules are highly
656 intercalated in the clay interlayer space as well as interparticle pores, which lead to decreased
657 surface area and pore volume. Hence, the clay layers are almost completely enclosed, and the
658 pores are enlarged by the loaded surfactant molecules. The p-nitrophenol molecules can be
659 captured not only within the clay layer space but also in the enlarged pores. The organoclay
660 as a partition medium has strong affinity on p-nitrophenol molecules [29] and its efficiency
661 is shown in the adsorption curves in Figure 17.

662 Accordingly, the sorption efficiency of the corresponding organoclays is higher than
663 montmorillonite. Based on the distribution and arrangement of surfactant in organoclays, the
664 mechanism is suggested as: the sorption mechanism of organic molecules at the organoclays
665 (1.0 CEC ~ 2.0 CEC) is controlled by partition, whereas the organoclays prepared at lower
666 surfactant loadings are mainly adsorption [7, 30-32].

667

668 **Conclusions**

669 Organoclays were prepared using montmorillonite and tetradecyltrimethylammonium
670 bromide (TDTMA) as an example of a surfactant with a long alkyl chain. The prepared
671 organoclays were characterised by modern physical techniques. Based on the basal spacing
672 from the XRD pattern, information of the molecular arrangement within the clay interlayer
673 space has been obtained as a function of the surfactant concentration. The configurations of
674 the surfactant within the organoclays take a lateral monolayer arrangement at lower surfactant
675 concentration (0.25 and 0.5 CEC). At higher surfactant concentrations (1.0 ~ 2.0 CEC), both
676 configurations of the surfactant (a lateral bilayer and pseudotrimolecular layer structures)
677 were observed. Moreover, the XRD pattern of organoclays adsorbed p-nitrophenol indicated
678 the intercalation of p-nitrophenol within organoclays, and the expanded interlayer spacing of
679 organoclays were measured and identified using TEM. Using BET method, it is found that
680 the surface area is inversely related to the pore volume for organoclays. As the loading of the
681 surfactant increase in organoclays, the pore size becomes smaller with an increase of surface
682 area. Hence, it is concluded that the pore parameters are significantly related to the
683 distribution and arrangement of surfactant intercalated into the clay surface. The loaded
684 surfactant in the interlayer space were further analysed by TGA. Throughout the XPS study,

685 the chemical composition of montmorillonite and organoclays were analysed and the binding
686 energy of C-C is relatively sensitive to determine the structural changes in organoclays. In
687 infrared spectra, the CH stretching region was extremely sensitive to conformation changes
688 of the chain within the interlayer and a significant wavenumber shift indicates that more
689 gauche conformational molecules introduced into alkyl chain with the decrease of alkyl chain
690 length. From the results, the structural properties of organoclays intercalated with surfactant
691 were changed from hydrophilic to hydrophobic, and the adsorbed p-nitrophenol intercalates
692 the organoclay and displaces the surfactant molecules or rearranges the structure of surfactant
693 molecules within the organoclay interlayer. In addition, the different sorption mechanisms
694 involving adsorption and/or partition may affect the sorption of p-nitrophenol onto
695 organoclays. This type of organoclays will potentially be used as good adsorbents for
696 recalcitrant organic molecules in the industry and this organoclay will be applied to test in
697 adsorbing efficiency from p-nitrophenol and further removal of herbicides and pesticides
698 from contaminated samples.

699

700 **Acknowledgment**

701 Authors gratefully acknowledge and express gratitude to Dr. Barry Wood from the XPS
702 facilities at University of Queensland for his assistance and availability. We also would like
703 to thank Jin Cheng for his assistance with the TEM analysis.

704

705

706

707

708

709

710

711

712

713 **References**

- 714 [1] J.A. Smith, P.R. Jaffe, C.T. Chiou, *Environmental Science & Technology* 24 (1990)
715 1167.
- 716 [2] L.B. de Paiva, A.R. Morales, F.R. Valenzuela Díaz, *Applied Clay Science* 42 (2008) 8.
- 717 [3] M.O. Adebajo, R.L. Frost, J.T. Kloprogge, O. Carmody, S. Kokot, *Journal of Porous*
718 *Materials* 10 (2003) 159.
- 719 [4] Y. Xi, R.L. Frost, H. He, T. Kloprogge, T. Bostrom, *Langmuir* 21 (2005) 8675.
- 720 [5] H. He, J. Duchet, J. Galy, J.-F. Gérard, *Journal of Colloid and Interface Science* 295
721 (2006) 202.
- 722 [6] Q. Zhou, R.L. Frost, H. He, Y. Xi, M. Zbik, *Journal of Colloid and Interface Science*
723 311 (2007) 24.
- 724 [7] M. Cruz-Guzman, R. Celis, M.C. Hermosin, W.C. Koskinen, J. Cornejo, *Journal of*
725 *Agricultural and Food Chemistry* 53 (2005) 7502.
- 726 [8] G. Yuan, *Journal of Environmental Science and Health, Part A: Toxic/Hazardous*
727 *Substances and Environmental Engineering* 39 (2005) 2661
- 728 [9] L. Zhu, B. Chen, X. Shen, *Environmental Science & Technology* 34 (1999) 468.
- 729 [10] Y. Xi, M. Mallavarapu, R. Naidu, *Applied Clay Science* 48 (2010) 92.
- 730 [11] R. Liu, R.L. Frost, W.N. Martens, Y. Yuan, *Journal of Colloid and Interface Science*
731 327 (2008) 287.
- 732 [12] S. Brunauer, L.S. Deming, W.E. Deming, E. Teller, *Journal of the American*
733 *Chemical Society* 62 (1940) 1723.
- 734 [13] S.J. Gregg, K.S.W. Sing, *Adsorption, Surface Area and Porosity*. Academic Press,
735 London and New York, 1967.
- 736 [14] M.A. Vicente, M. Suárez, J.d.D. López-González, M.A. Bañares-Muñoz, *Langmuir*
737 12 (1996) 566.
- 738 [15] C.-C. Wang, L.-C. Juang, C.-K. Lee, T.-C. Hsu, J.-F. Lee, H.-P. Chao, *Journal of*
739 *Colloid and Interface Science* 280 (2004) 27.
- 740 [16] H. He, Q. Zhou, W.N. Martens, T.J. Kloprogge, P. Yuan, Y. Xi, J. Zhu, R.L. Frost,
741 *Clays and Clay Minerals* 54 (2006) 689.
- 742 [17] L. Bugyi, A. Oszkó, F. Solymosi, *Surface Science* 461 (2000) 177.
- 743 [18] H. He, Q. Zhou, R.L. Frost, B.J. Wood, L.V. Duong, J.T. Kloprogge, *Spectrochimica*
744 *Acta Part A: Molecular and Biomolecular Spectroscopy* 66 (2007) 1180.
- 745 [19] Z. Klapysa, T. Fujita, N. Iyi, *Applied Clay Science* 19 (2001) 5.
- 746 [20] H. He, Z. Ding, J. Zhu, P. Yuan, Y. Xi, D. Yang, R.L. Frost, *Clays and Clay Minerals*
747 53 (2005) 287.
- 748 [21] Y. Xi, Q. Zhou, R.L. Frost, H. He, *Journal of Colloid and Interface Science* 311 (2007)
749 347.
- 750 [22] Q. Zhou, H. He, R.L. Frost, Y. Xi, *The Journal of Physical Chemistry C* 111 (2007)
751 7487.
- 752 [23] J. Zhu, H. He, L. Zhu, X. Wen, F. Deng, *Journal of Colloid and Interface Science* 286
753 (2005) 239.
- 754 [24] Y. Li, H. Ishida, *Langmuir* 19 (2003) 2479.
- 755 [25] Y. Ma, J. Zhu, H. He, P. Yuan, W. Shen, D. Liu, *Spectrochimica Acta Part A:*
756 *Molecular and Biomolecular Spectroscopy* 76 (2010) 122.
- 757 [26] R. Zhu, L. Zhu, J. Zhu, L. Xu, *Applied Clay Science* 42 (2008) 224.
- 758 [27] Q. Zhou, H.P. He, J.X. Zhu, W. Shen, R.L. Frost, P. Yuan, *Journal of Hazardous*
759 *Materials* 154 (2008) 1025.
- 760 [28] H. He, Q. Zhou, W.N. Martens, T.J. Kloprogge, P. Yuan, Y. Xi, J. Zhu, R.L. Frost,
761 *Clays and Clay Minerals* 54 (2006) 689.

762 [29] S.A.M. Boyd, M.M.; Chiou, C.T., Soil Science Society of America journal 52 (1988)
763 652.
764 [30] M. Cruz-Guzmán, R. Celis, M.C. Hermosín, J. Cornejo, Environmental Science &
765 Technology 38 (2003) 180.
766 [31] J.-F. Lee, M.M. Mortland, C.T. Chiou, D.E. Kile, S.A. Boyd, Clays and Clay
767 Minerals 38 (1990) 113.
768 [32] W.F.M.S.U. Jaynes, East Lansing, MI); Boyd, S.A., Soil Science Society of America
769 journal 55 (1991) 43.

770
771

772

773

774

775

776

777

778

779

780

781

782

783

784

785

786

787

788

789

790 **List of Tables**

791 Table 1. BET specific surface area (S_{BET}), pore volume (V_p) and pore diameter for
792 montmorillonite and organoclays

793 Table 2. Binding energy and their atomic contents of C 1s for organoclays

794 Table 3. TGA results of Montmorillonite (MMT), surfactant (TTMAB), and organoclays with
795 and without adsorbed p-nitrophenol

796

797

798

799

800

801

802

803

804

805

806

807

808

809

810

811

812

813 **List of Figures**

814 Figure 1. XRD patterns of montmorillonite, 0.25 CEC, and 0.5 CEC with and without
815 adsorbed p-nitrophenol

816 Figure 2. XRD patterns of 1.0 CEC, 1.50 CEC, and 2.0 CEC with and without adsorbed p-
817 nitrophenol

818 Figure 3. d(001) basal spacing of montmorillonite and organoclays

819 Figure 4. d(001) basal spacing of montmorillonite and organoclays with adsorbed p-
820 nitrophenol

821 Figure 5. TEM images of 0.5 CEC-TDTMA, 2.0 CEC-TDTMA, 0.5 CEC-4000, and 2.0
822 CEC-4000

823 Figure 6. Nitrogen adsorption-desorption isotherms of montmorillonite and organoclays

824 Figure 7. XPS survey scan spectra of montmorillonite (a), surfactant (b), and organoclays (c
825 and d)

826 Figure 8. XPS high resolution spectra of O 1s for montmorillonite (a), organoclays (b, c)

827 Figure 9. XPS high resolution spectra of Si 2p and Al 2p for montmorillonite (a and d),
828 organoclays (b, c and e, f)

829 Figure 10. XPS high resolution spectra of C 1s for organoclays (a, b, c)

830 Figure 11. Thermogravimetric analyses of Montmorillonite and surfactant (TDTMA), and
831 organoclays

832 Figure 12. Thermogravimetric analyses of Montmorillonite and organoclays adsorbed p-
833 nitrophenol

834 Figure 13. Infrared spectroscopy of Montmorillonite and organoclays with and without p-
835 nitrophenol in 3900 ~ 3000 cm^{-1} spectral range

836 Figure 14. Infrared spectroscopy of Montmorillonite and organoclays with and without p-
837 nitrophenol in the 3000 ~ 2800 cm^{-1} spectral range

838 Figure 15. Infrared spectroscopy of Montmorillonite and organoclays with and without p-
839 nitrophenol in the 1720 ~ 1560 cm^{-1} spectral range

840 Figure 16. Infrared spectroscopy of Montmorillonite and organoclays with and without p-
841 nitrophenol in the 1560 ~ 1360 cm⁻¹ spectral range

842 Figure 17. Uptake of p-nitrophenol on montmorillonite and organoclays

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

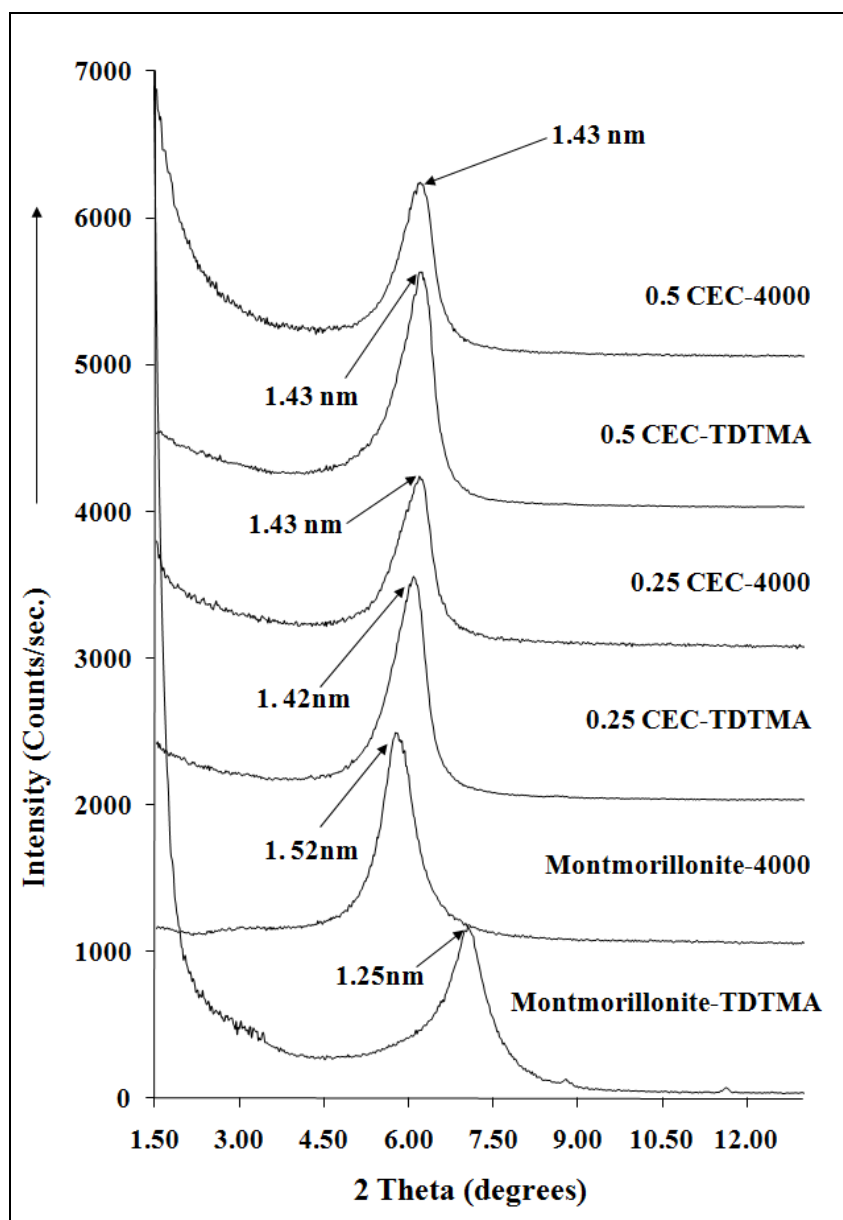
858

859

860

861

862



864

865 **Figure 1 XRD patterns of montmorillonite, 0.25 CEC, and 0.5 CEC with and without**
 866 **adsorbed p-nitrophenol**

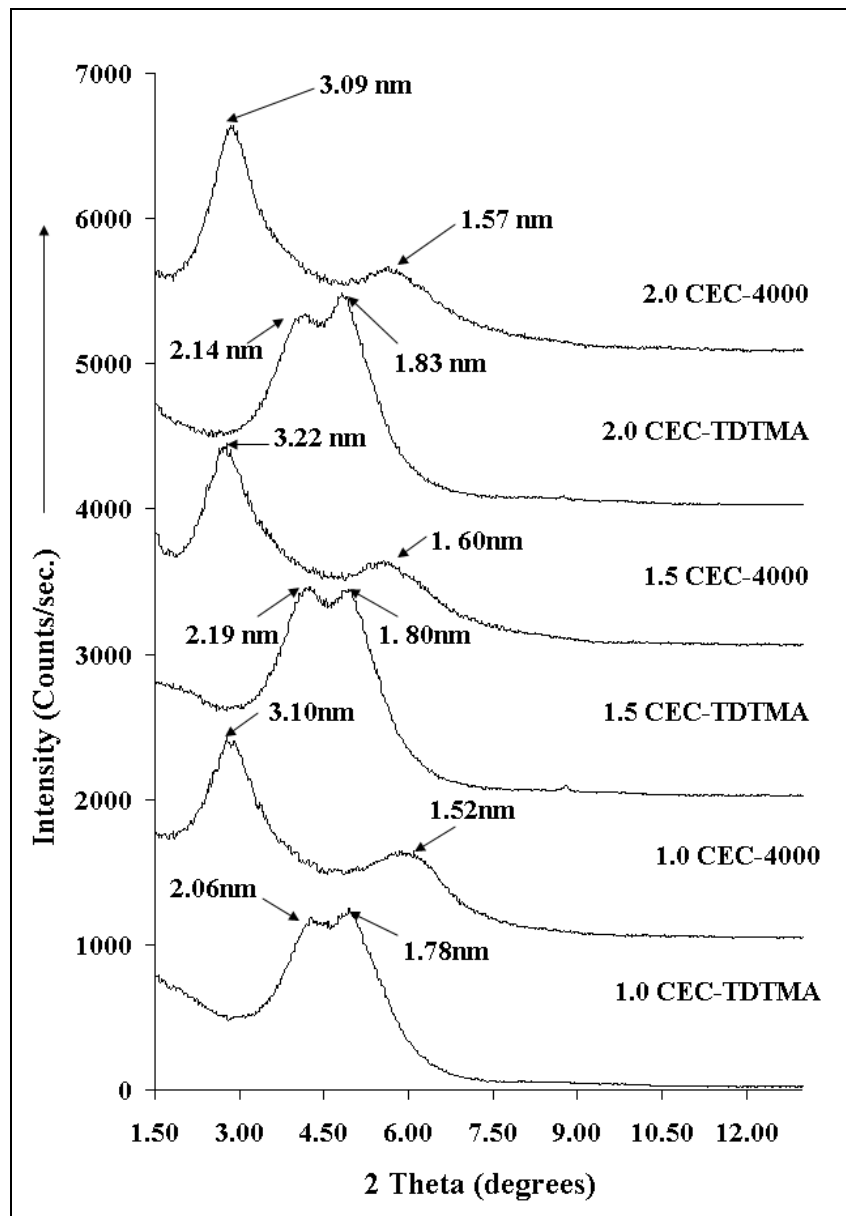
867

868

869

870

871



873

874 **Figure 2 XRD patterns of 1.0 CEC, 1.50 CEC, and 2.0 CEC with and without adsorbed**
 875 **p-nitrophenol**

876

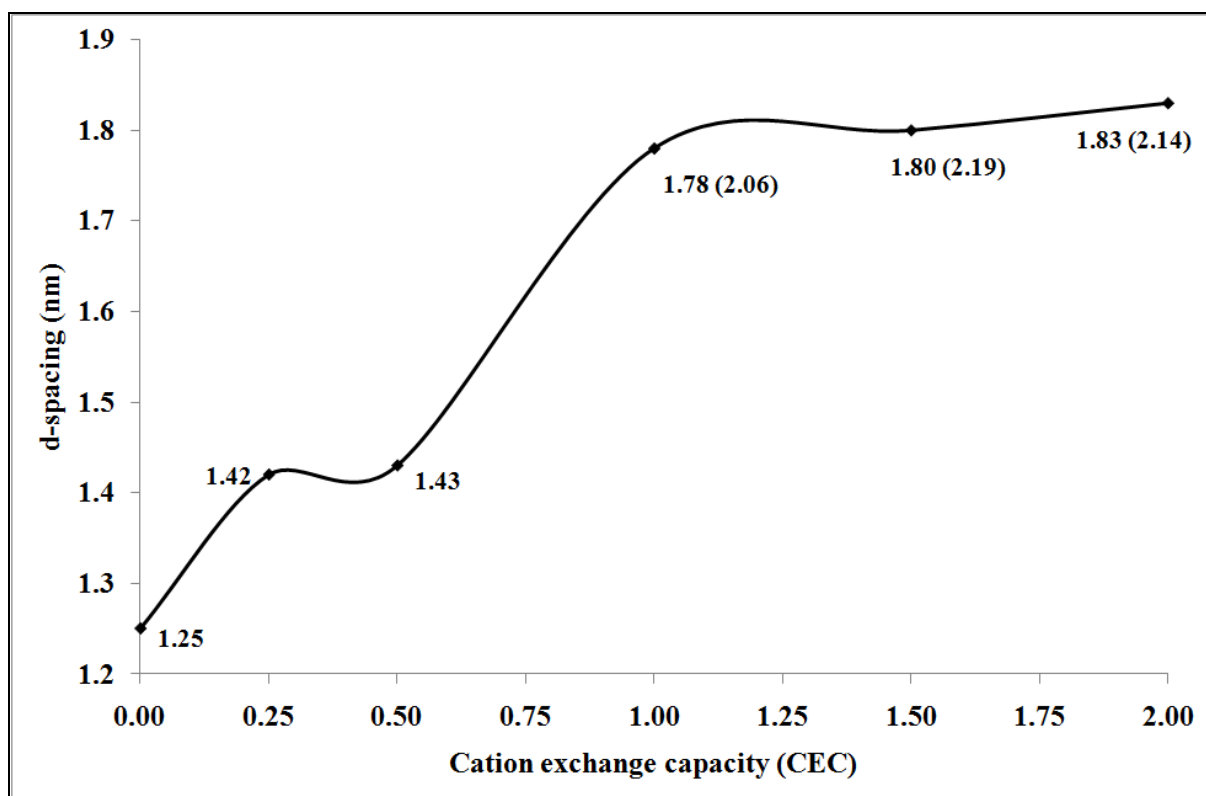
877

878

879

880

881



882

883 **Figure 3 d(001) basal spacing of montmorillonite and organoclays**

884

885

886

887

888

889

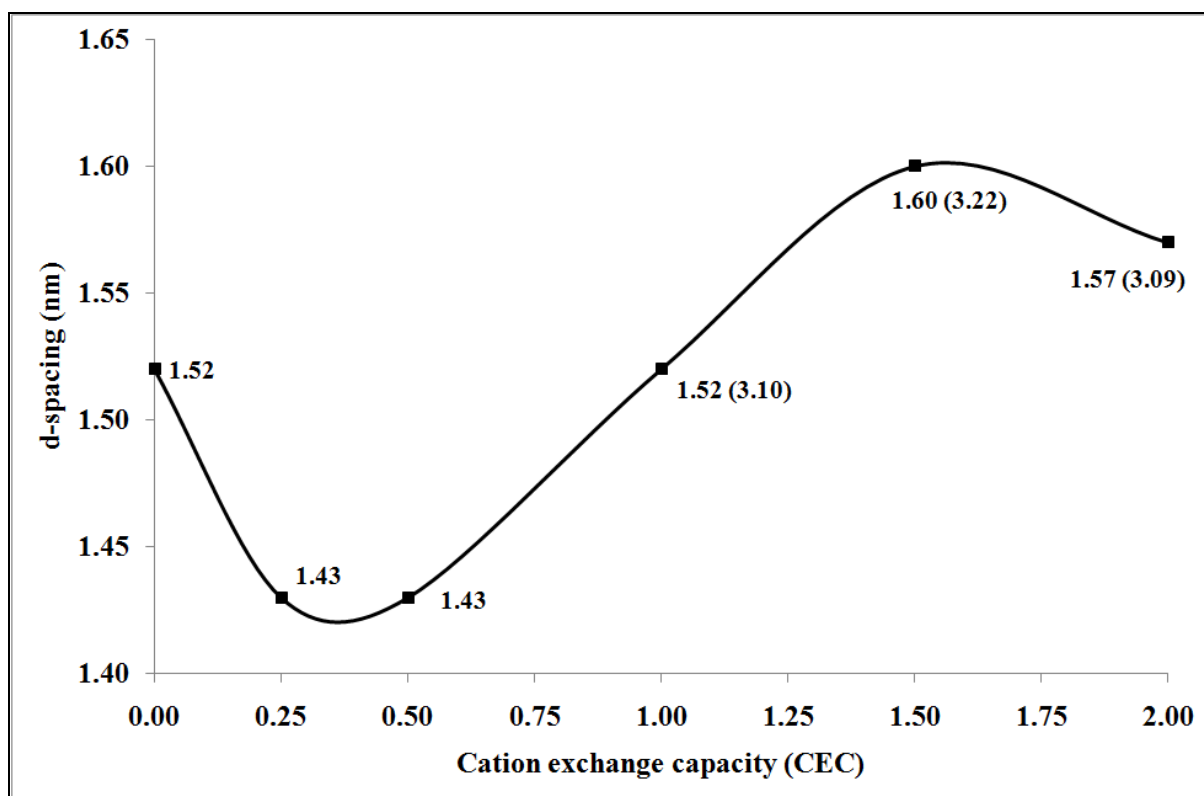
890

891

892

893

894



896

897 **Figure 4 d(001) basal spacing of montmorillonite and organoclays with adsorbed p-**
898 **nitrophenol**

899

900

901

902

903

904

905

906

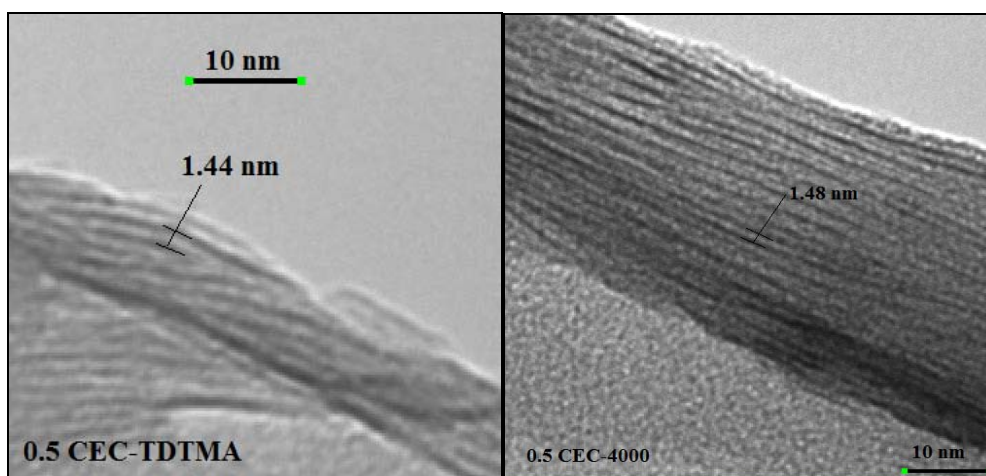
907

908

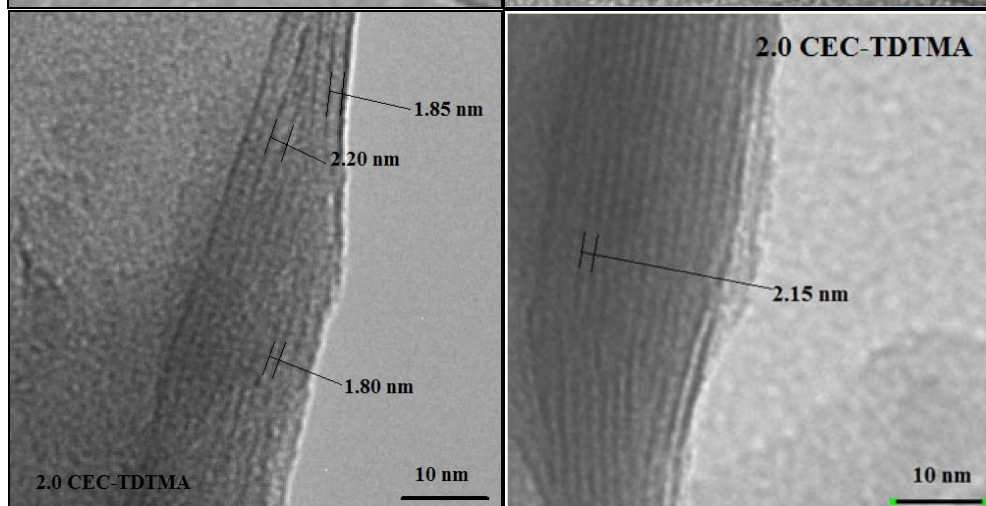
909

910

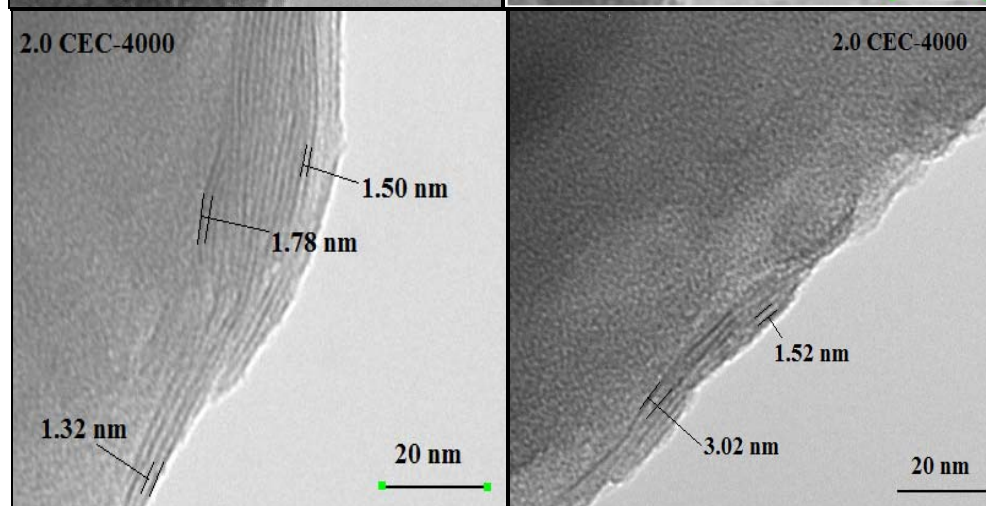
911



912



913



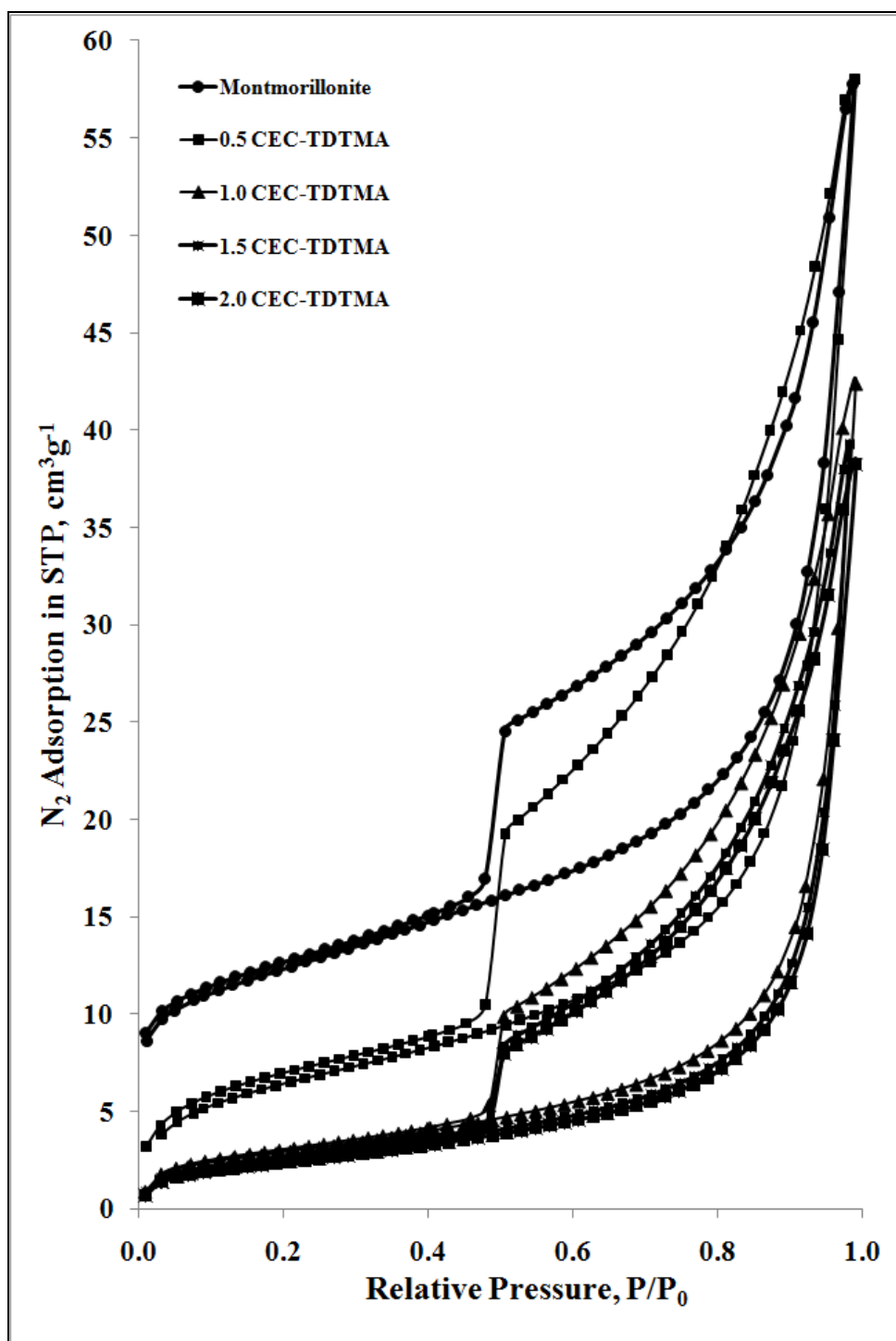
914

915

Figure 5 TEM images of 0.5 CEC-TDTMA, 2.0 CEC-TDTMA, 0.5 CEC-4000, and 2.0 CEC-4000

916

917



918

919 **Figure 6 Nitrogen adsorption-desorption isotherms of montmorillonite and organoclays**

920

921

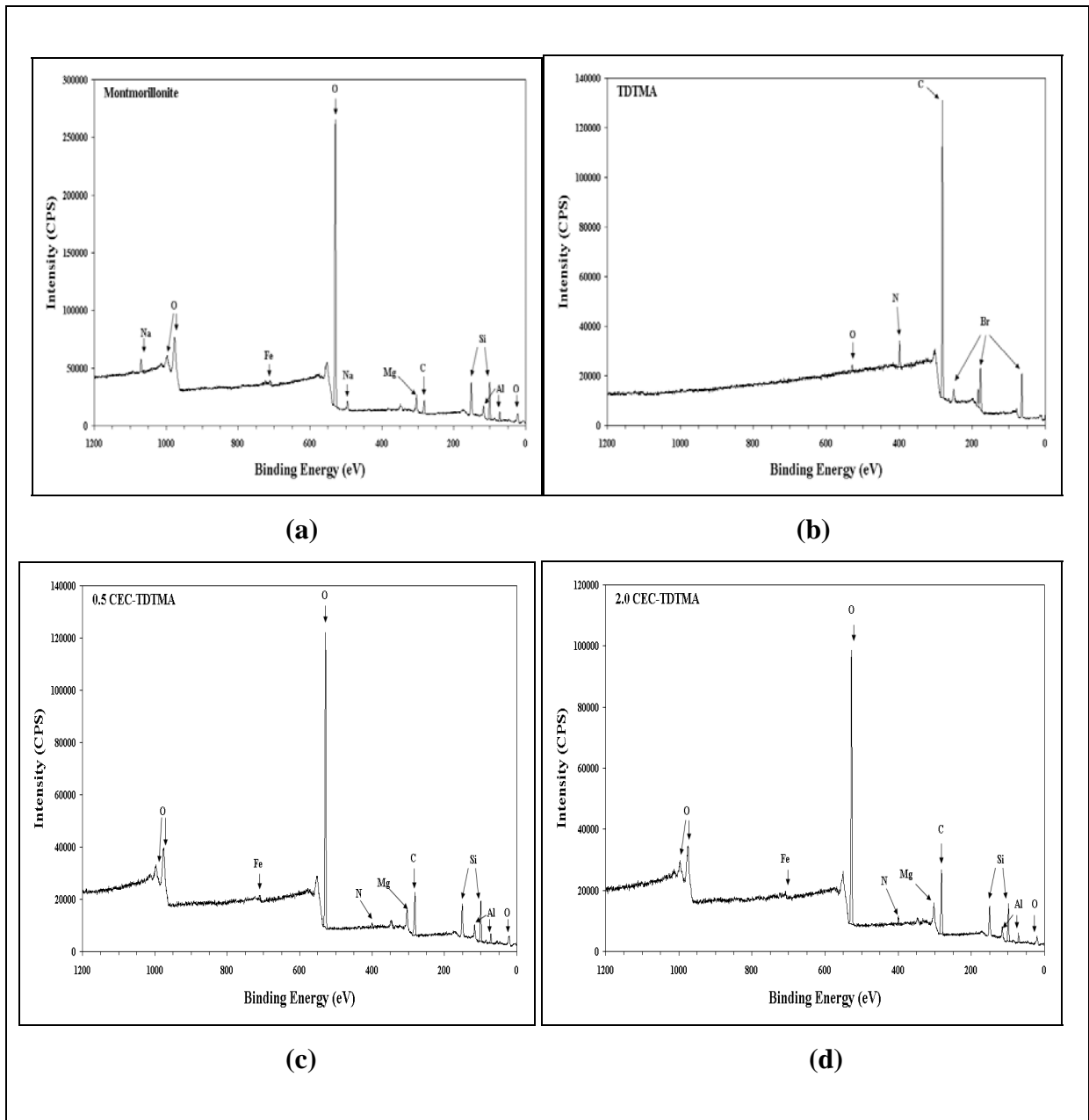


Figure 7 XPS survey scan spectra of montmorillonite (a), surfactant (b), and organoclays (c and d)

923

924

925

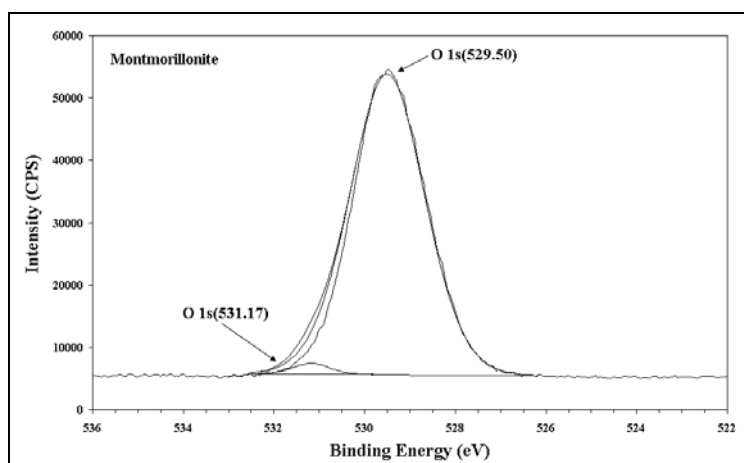
926

927

928

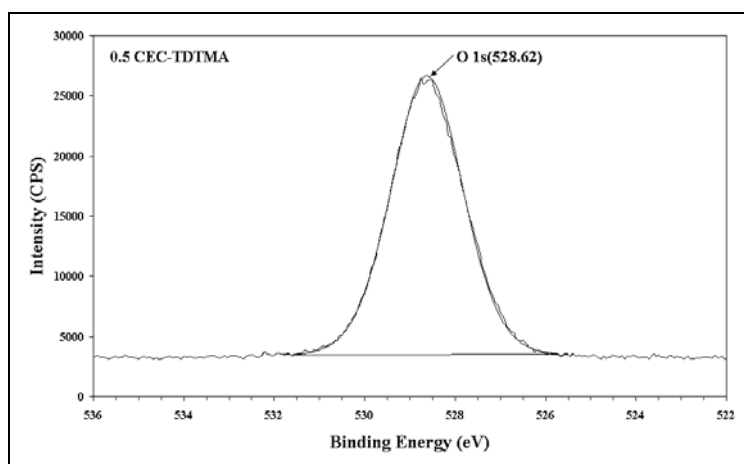
929

930



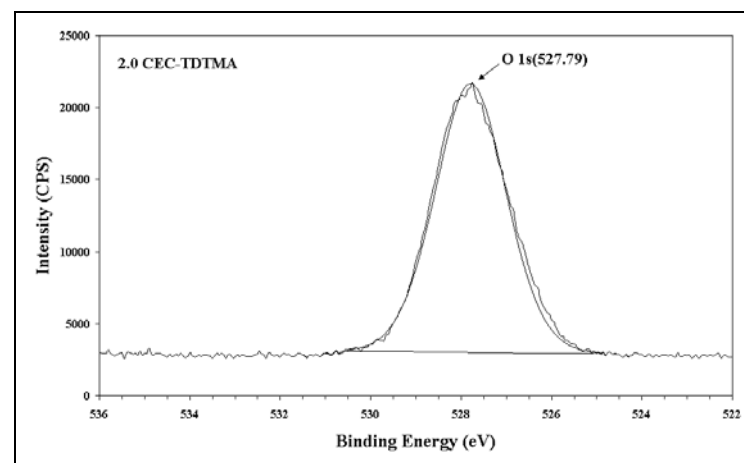
931

(a)



932

(b)



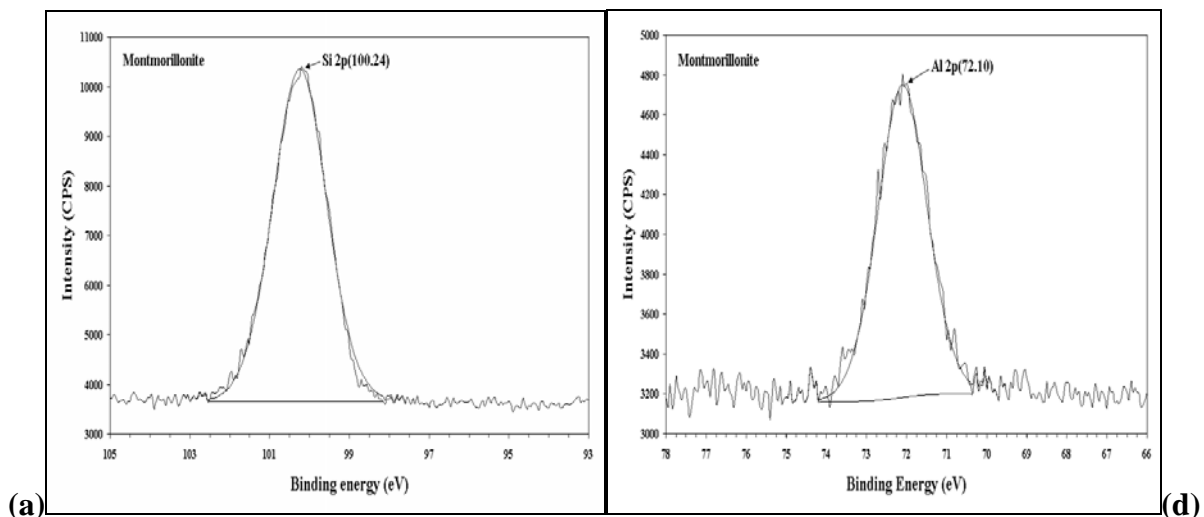
933

(c)

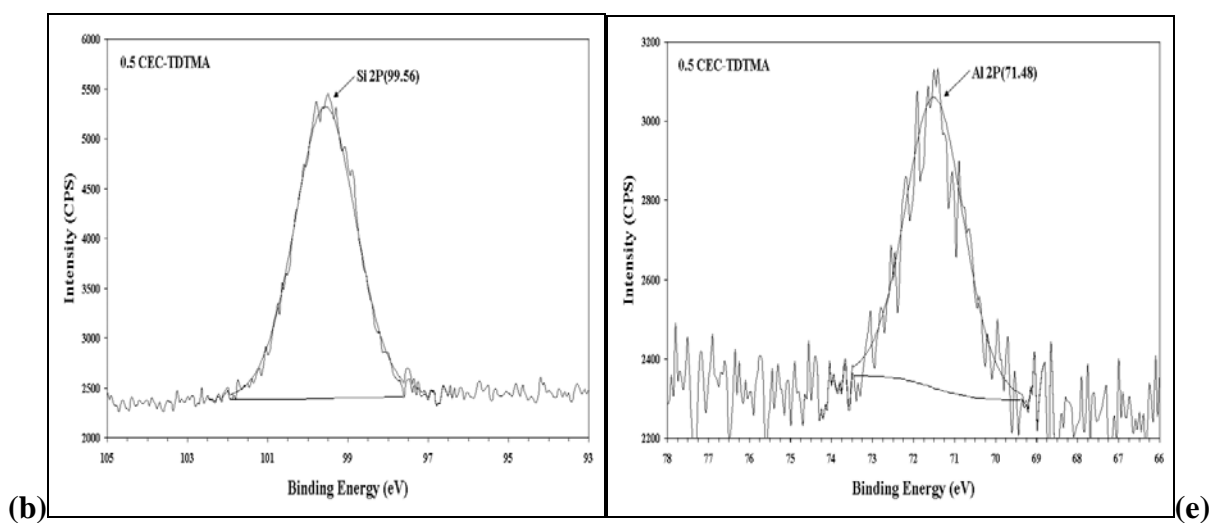
934 **Figure 8 XPS high resolution spectra of O 1s for montmorillonite (a), organoclays (b, c)**

935

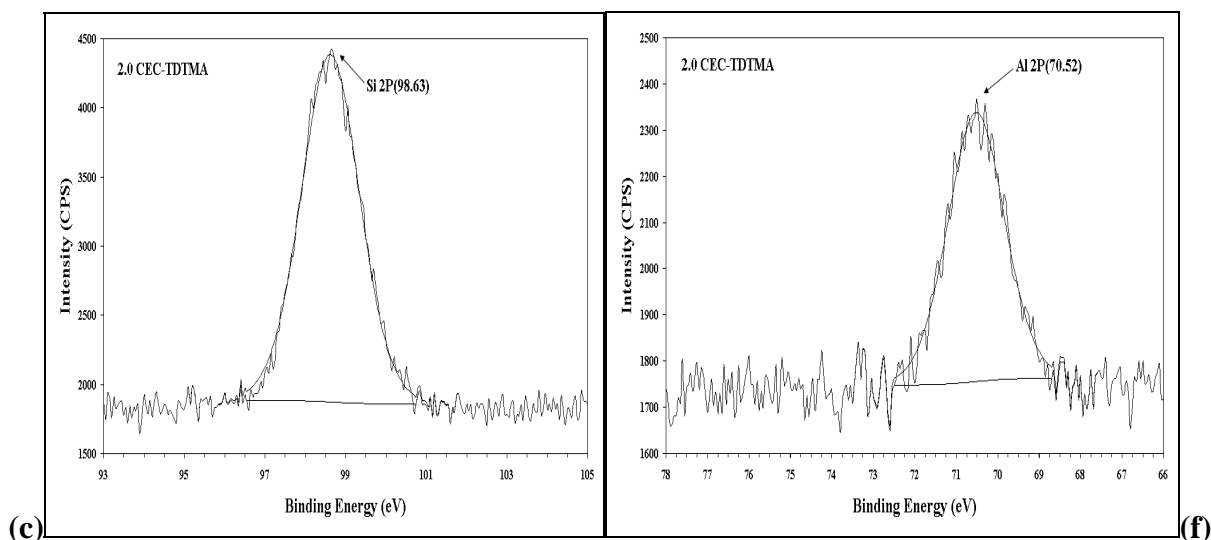
936



937



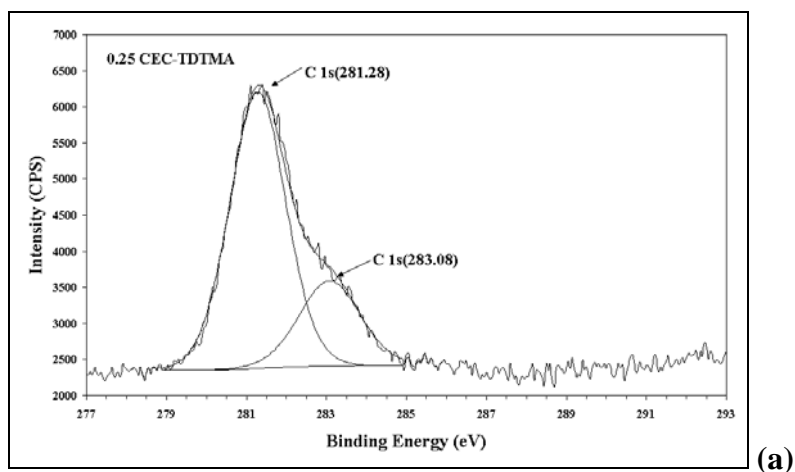
938



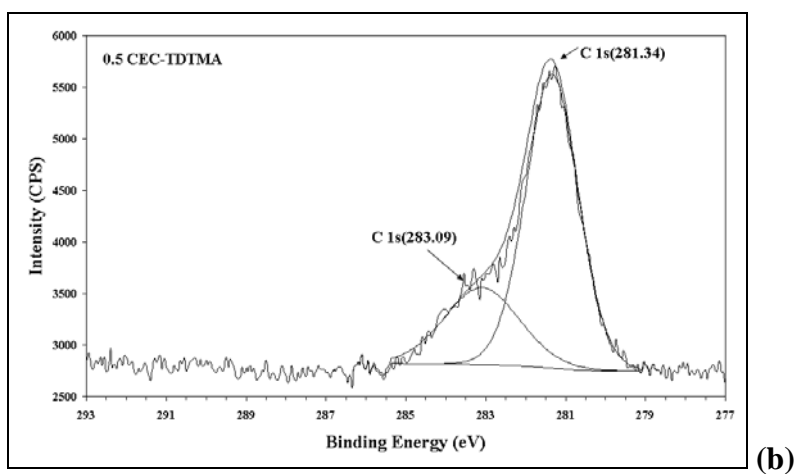
939 **Figure 9 XPS high resolution spectra of Si 2p and Al 2p for montmorillonite (a and d),**
940 **organoclays (b, e and c, f)**

941

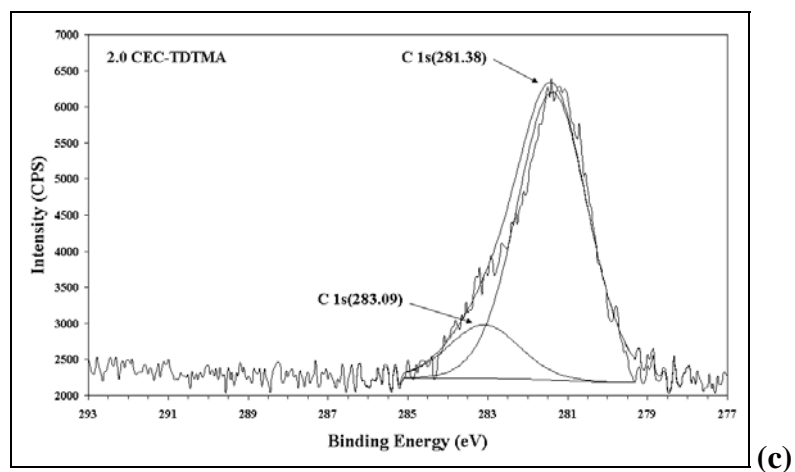
942



943



944



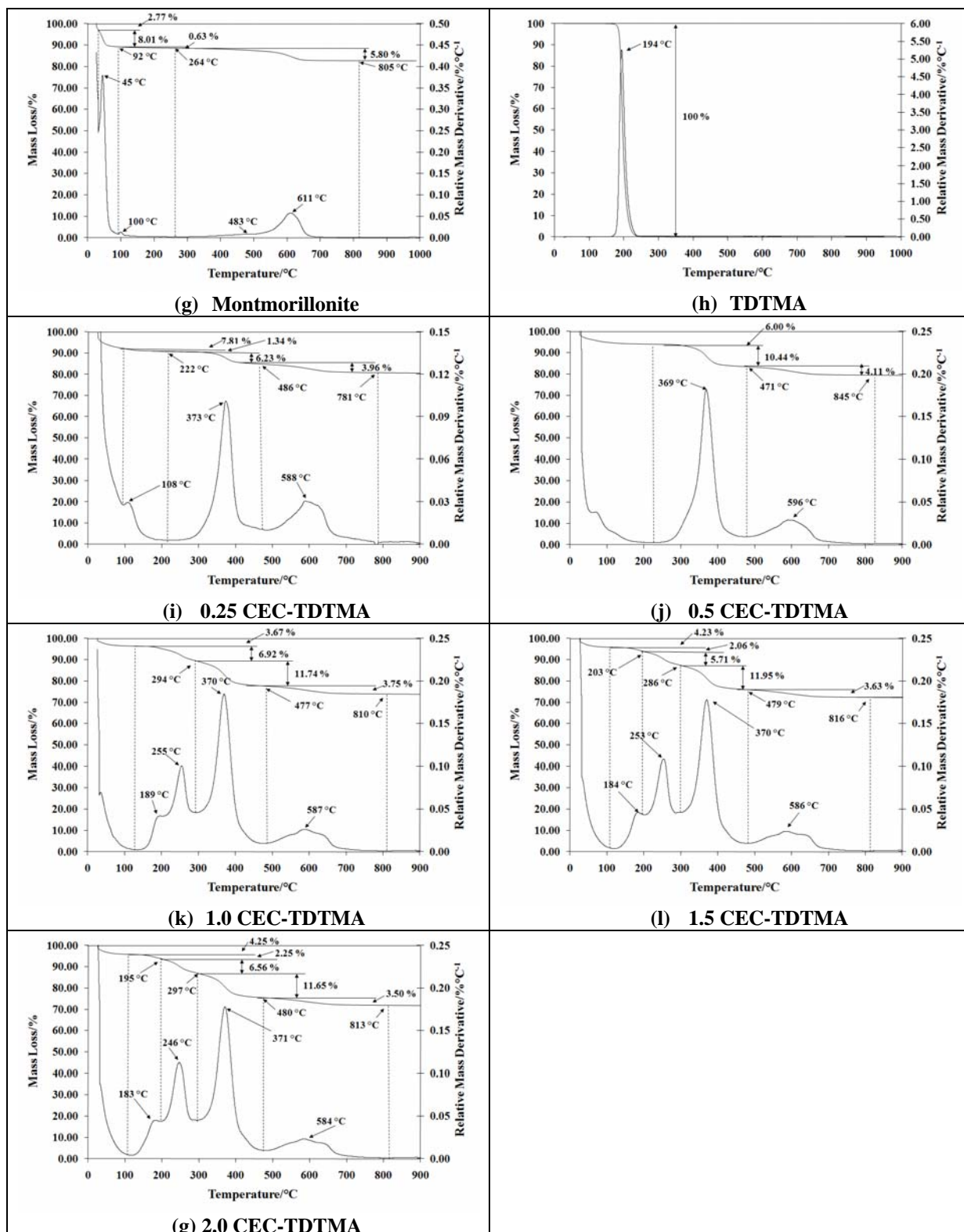
945

Figure 10 XPS high resolution spectra of C 1s for organoclays (a, b, c)

946

947

948



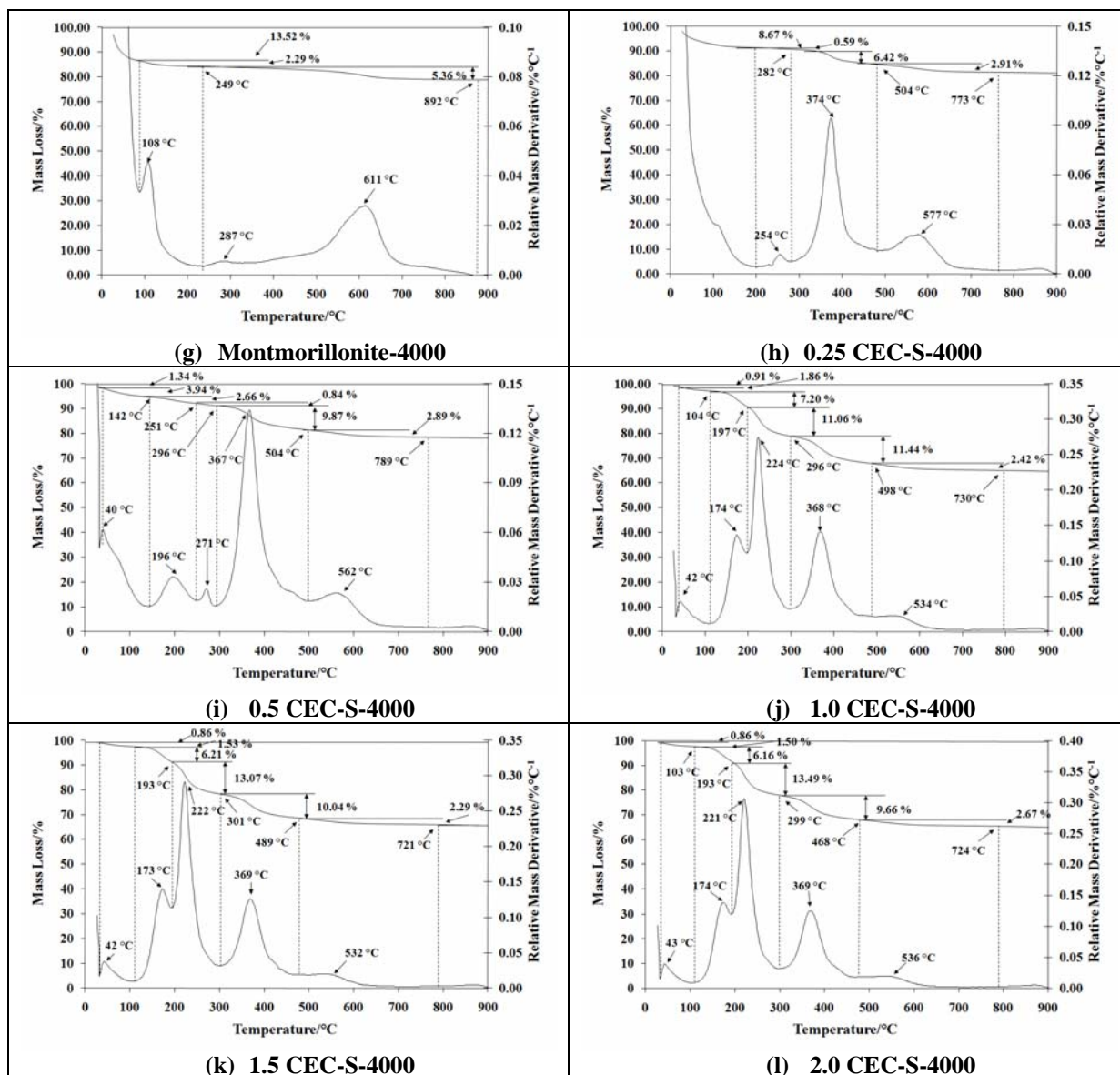
950

951

952

Figure 11 Thermogravimetric analyses of montmorillonite, Surfactant (TDTMA) and organoclays

953



955

956 **Figure 12 Thermogravimetric analyses of montmorillonite and organoclays adsorbed p-**
 957 **nitrophenol**

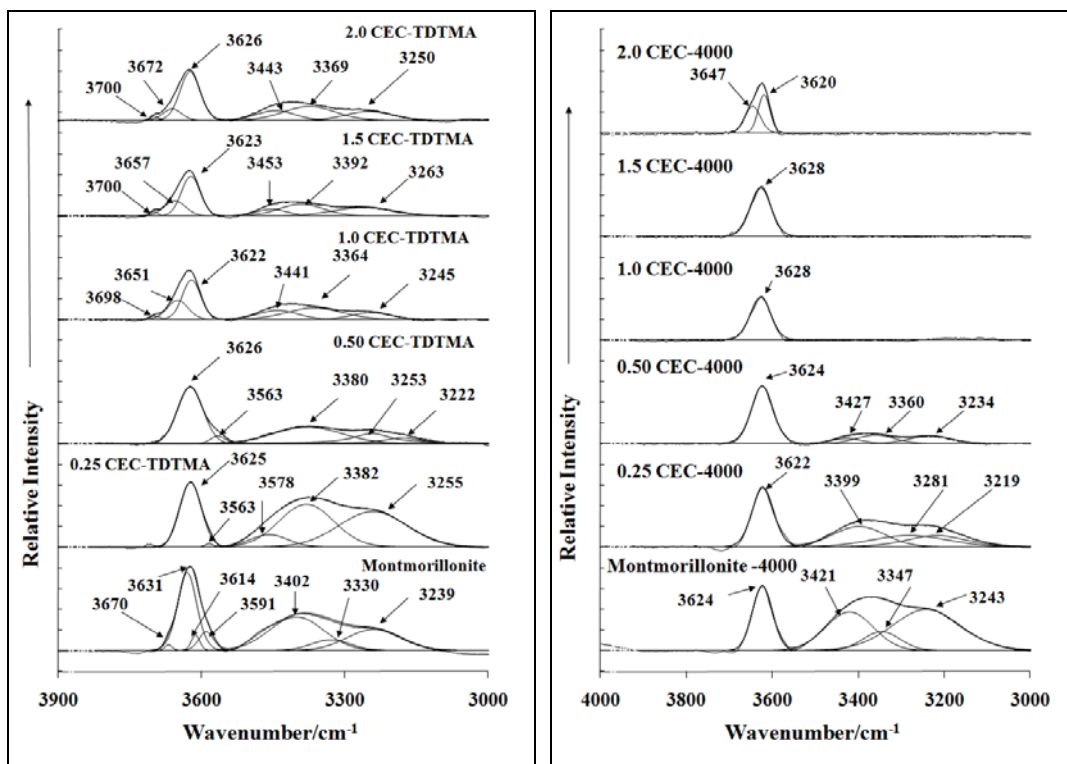
958

959

960

961

962



964

965

(a)

(b)

966 **Figure 13 Infrared spectroscopy of montmorillonite and organoclays with and without**
967 **p-nitrophenol in 3900 ~ 3000 cm⁻¹ spectral range**

968

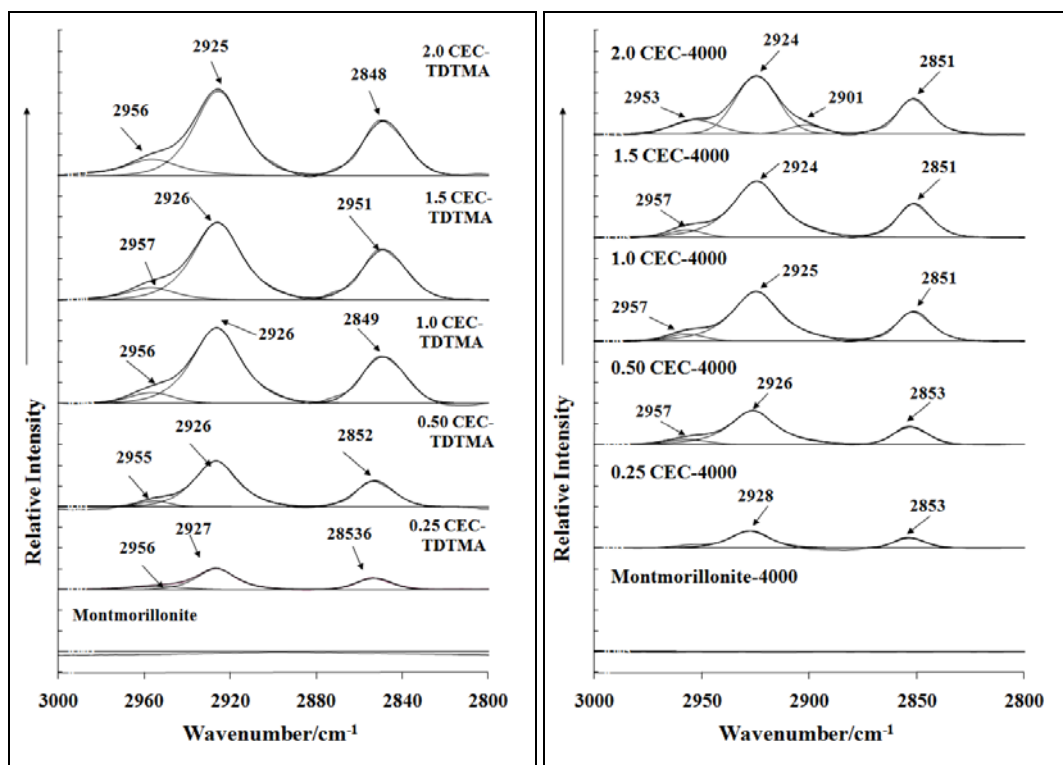
969

970

971

972

973



974

975

(a)

(b)

976 **Figure 14 Infrared spectroscopy of montmorillonite and organoclays with and without**
 977 **p-nitrophenol in the 3000 ~ 2800 cm⁻¹ spectral range**

978

979

980

981

982

983

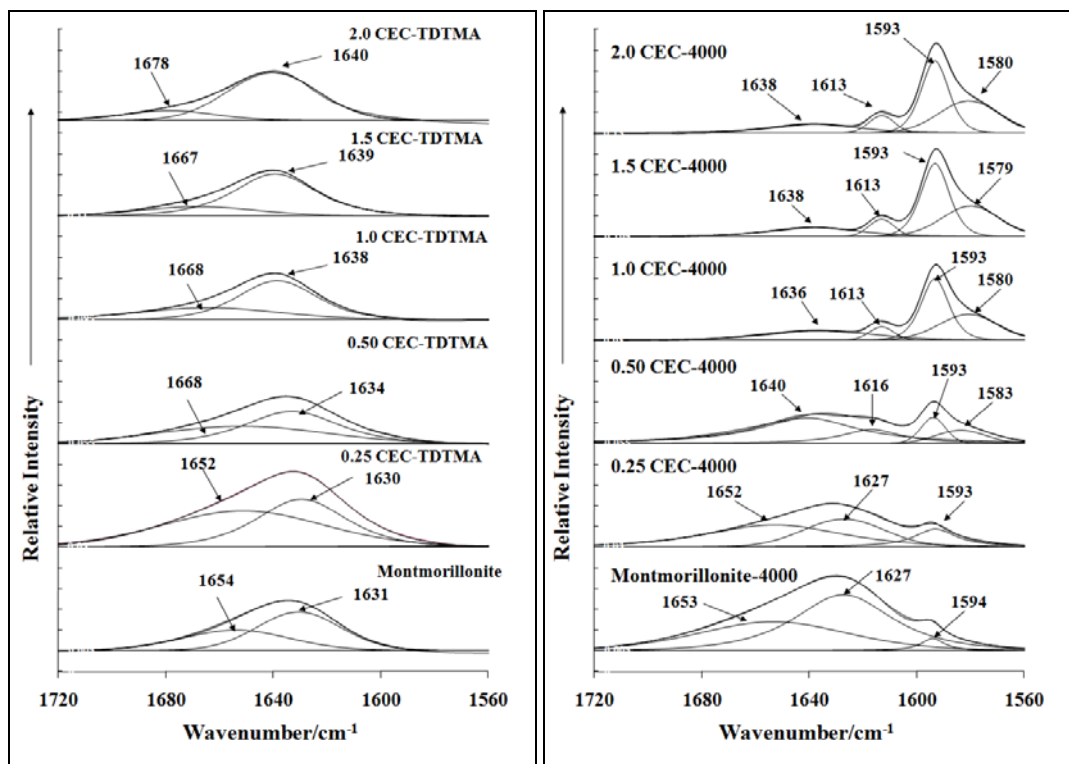
984

985

986

987

988



990

991

(a)

(b)

992 **Figure 15 Infrared spectroscopy of montmorillonite and organoclays with and without**
993 **p-nitrophenol in the 1720 ~ 1560 cm⁻¹ spectral range**

994

995

996

997

998

999

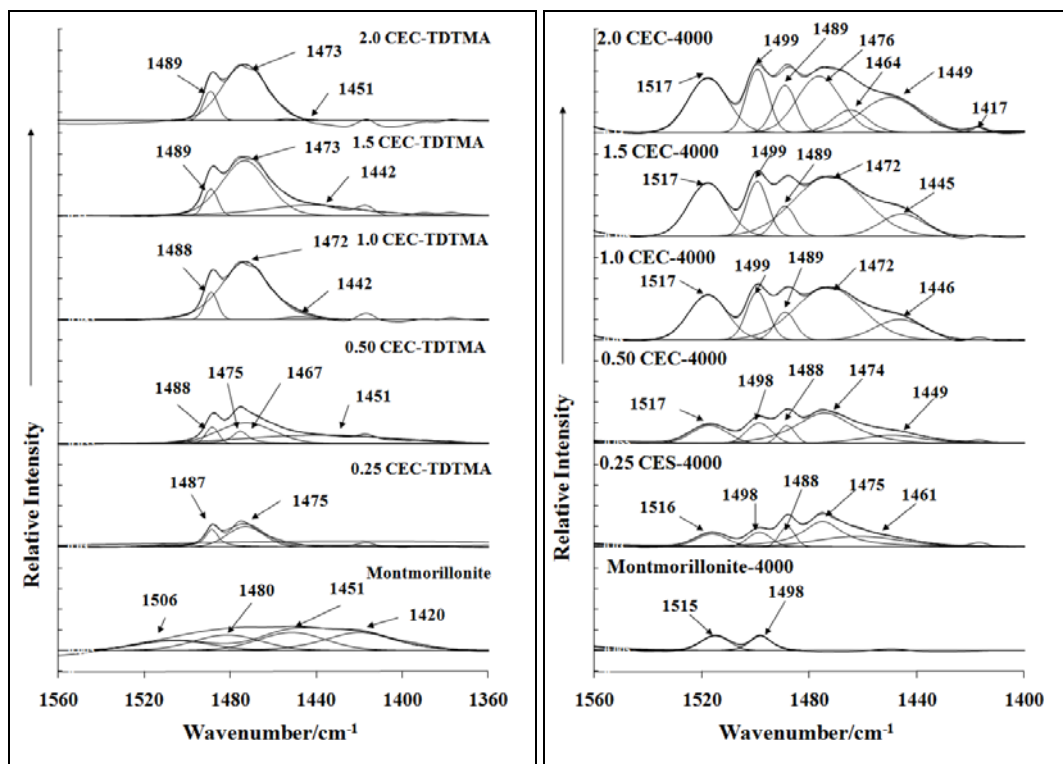
1000

1001

1002

1003

1004



1005

1006

(a)

(b)

1007 **Figure 16 Infrared spectroscopy of montmorillonite and organoclays without p-**
1008 **nitrophenol in the 1560 ~ 1360 cm⁻¹ spectral range**

1009

1010

1011

1012

1013

1014

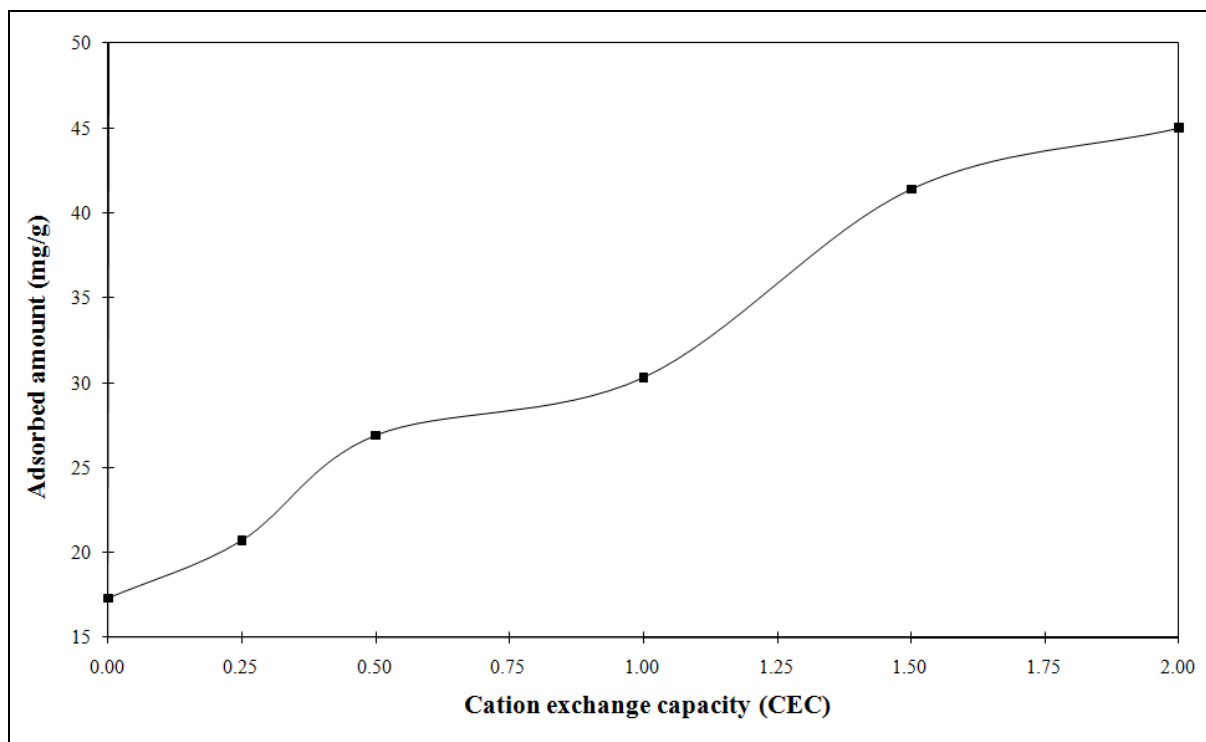
1015

1016

1017

1018

1019



1020

1021

Figure 17. Uptake of p-nitrophenol on montmorillonite and organoclays

1022

1023

1024

1025

1026

1027

1028

1029

1030

1031

1032