

Development of two-color fluorescent imager and integrated fluidic system for nanosatellite biology applications

Diana Wu *et al* NASA Ames Research Center ASGSR December 1, 2012



Technologies (MisST)



Fluidic

System

NanoSatellite Science Payload

22 cm

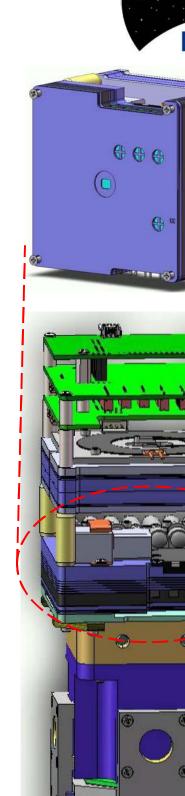






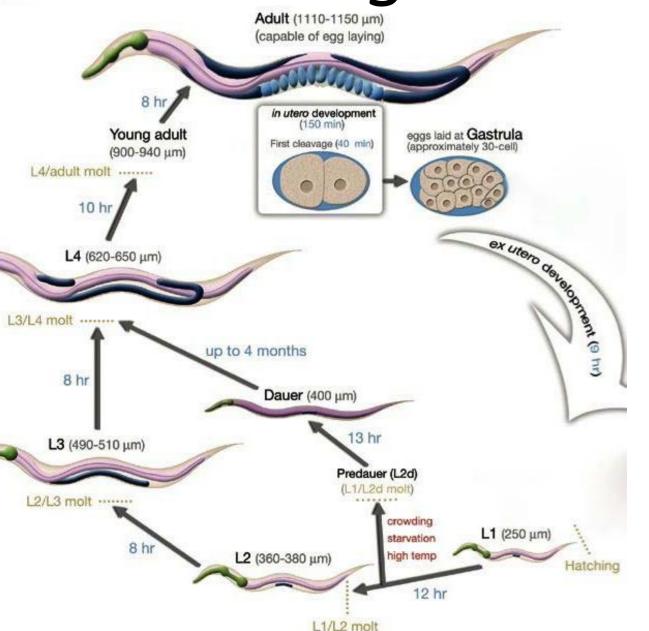
Overview

- ve: Technology demonstration of 2-color cent imager for space biology applications
- model organisms, resolve major features/organs, interest & control
- entation: 2-cube integrated culture/imaging ent
- . elegans model organism
- uidics: culture wells, imaging zone, reagent storage
- nager: fixed focus, 300 kpixel CMOS chip
- nage storage: 2 GB/6,000 images/8 bits per color elemetry of 1 - 20 images)
- lermetic containment vessel

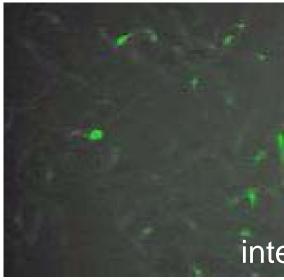


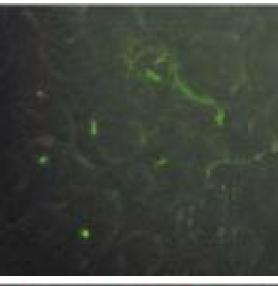
C. elegans Overview

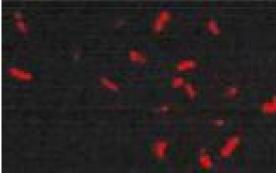




- ding and short life cycle make it inexpensive to and ideal for research
- study spaceflight effects and experiments using C.









C. elegans Imaging



White

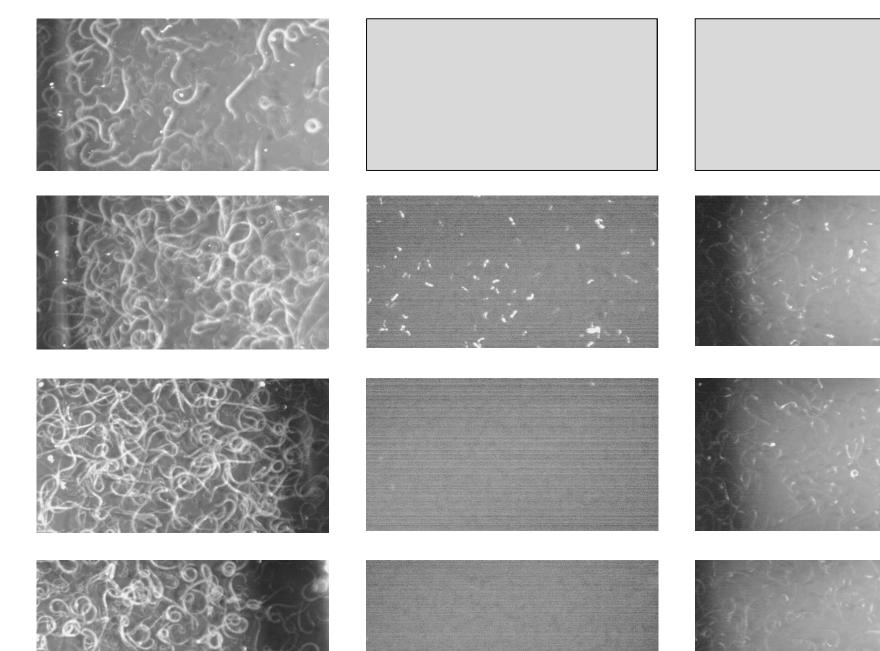
Amber

Blue

t) rescent

- nato uscle cells
- uscle cells
- **4** nerve cells

12 intestinal cells

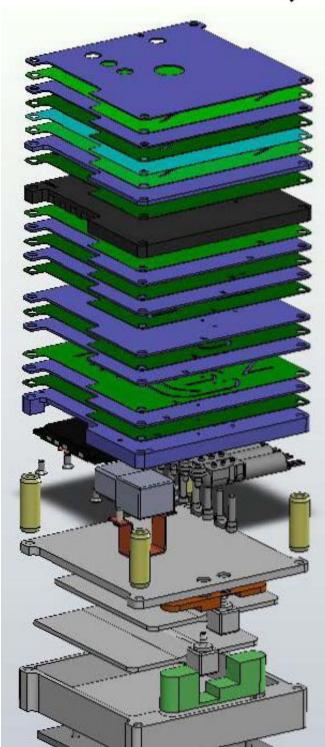




Fluidics Overview

ach: Multilayer polymer microfluidic card

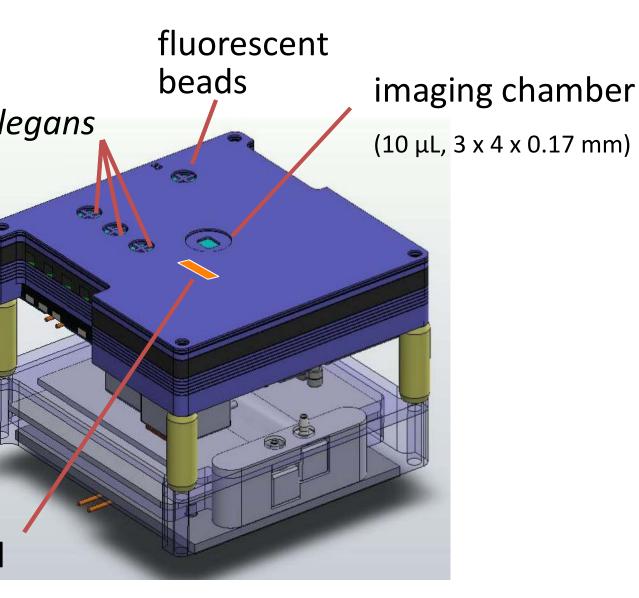
- Multi-layer lamination to form 3-D fluidic network
- Laser-cut fluidic channels in pressuresensitive adhesive (PSA)
- Lamination of machined polycarbonate sheets with via holes, filters, polystyrene using PSA
- ers, adhesives, sterilization, biocompatibility, all have GeneSat, PharmaSat, O/OREOS-
- EcAMSat heritage



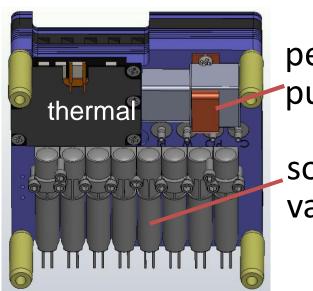


Fluidic Card and Reservoir



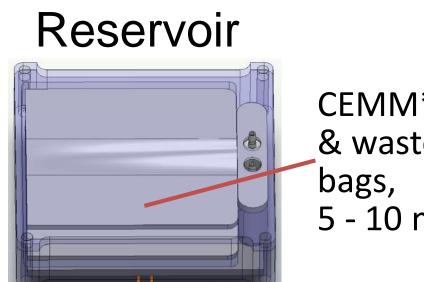


Fluidic Card



perista pump

soleno valves



Experimental Timeline

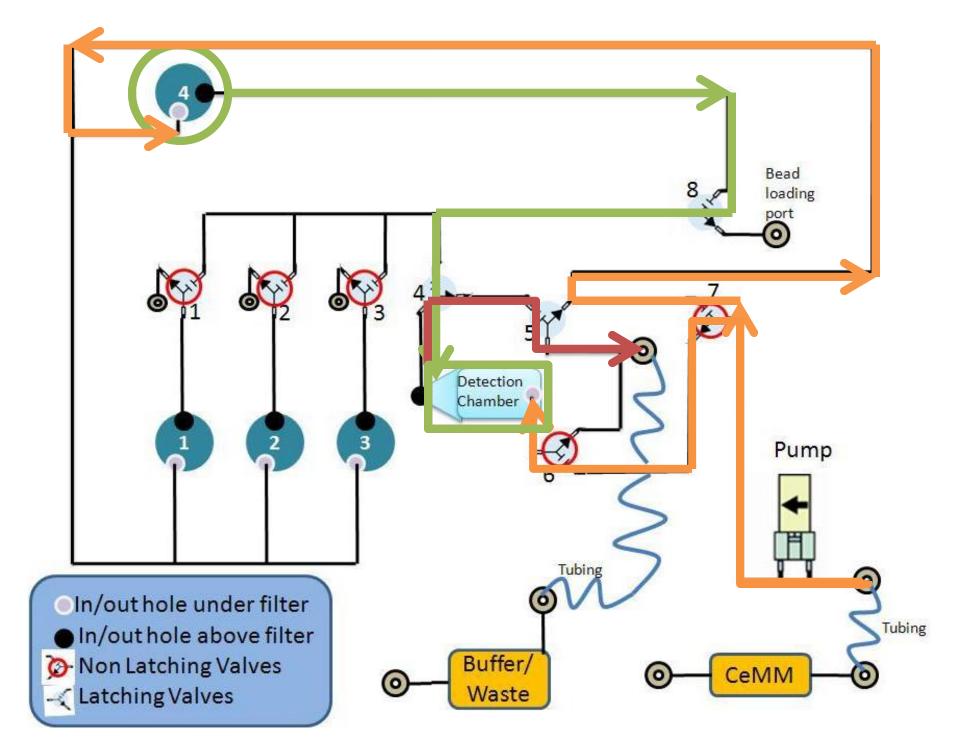






Fluidic Sequence - Beads

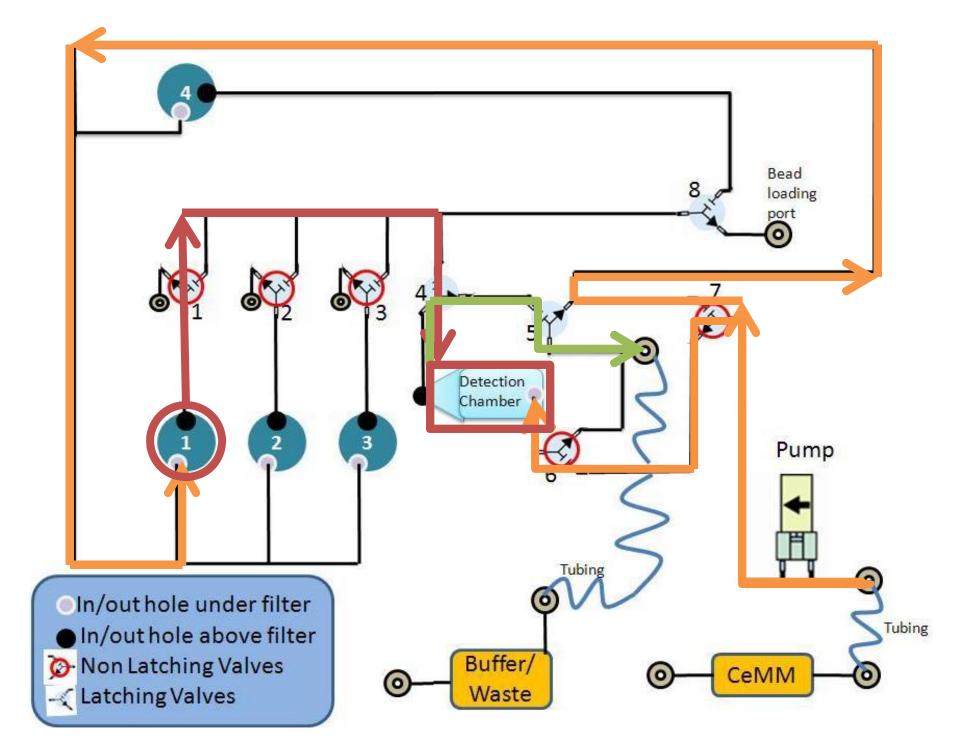






Fluidic Sequence- Worms







Fluorescence Imager



Imager Components:

Luxeon Rebel LEDs:

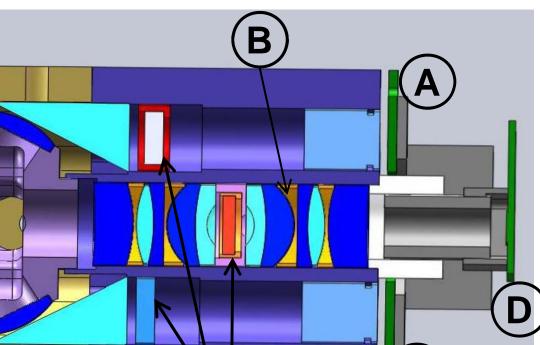
- A B C
- Edmunds relay lens
- Semrock filters: blue excitation, amber excitation,

2 amber, 1 blue, 1 white

dual band-pass emission

Camera:

COMedia C328-7221



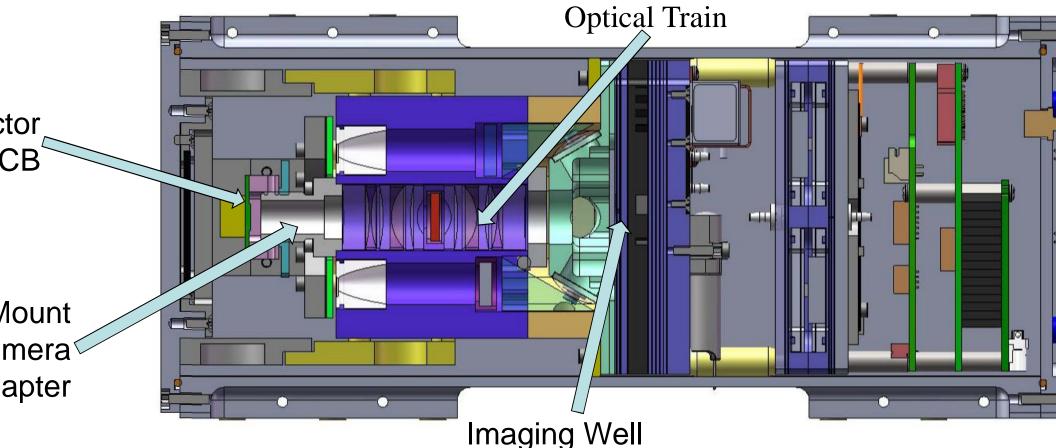
Imager Specifications:

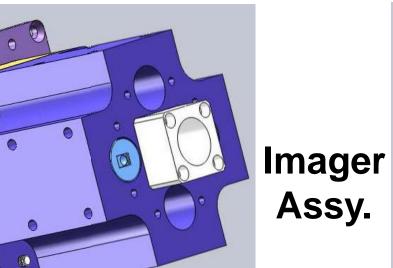
- 2-color fluorescence: Green excite/emit peaks at: 487/ Red excite/emit peaks at: 587/61
- Cool white for standard imaging
- Lateral resolution: ~ 8 μm
- Magnification: 1:1
- Field of View: ~3 x 4 mm
- Depth of Focus: ~150 μm

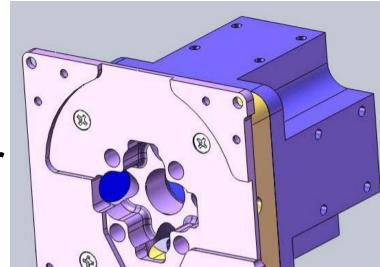


Fluorescence Imager









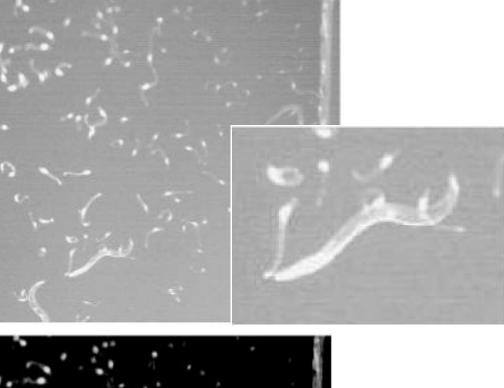
- Fluidic well depth:
 0.17 mm
- Fine pitch thread of S-mount (0.5mm/th
- Camera adjustmer
 ±5 mm
- Locknut/set screw

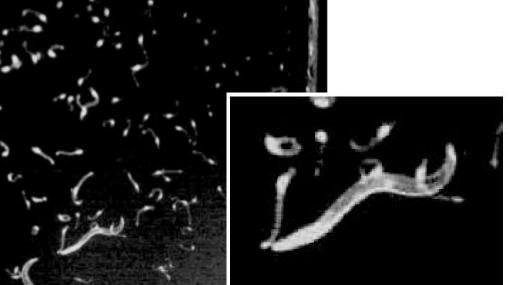


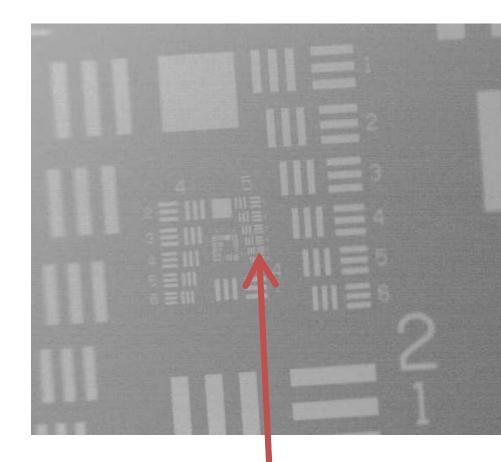
Camera chip



orescently labeled C. elegans





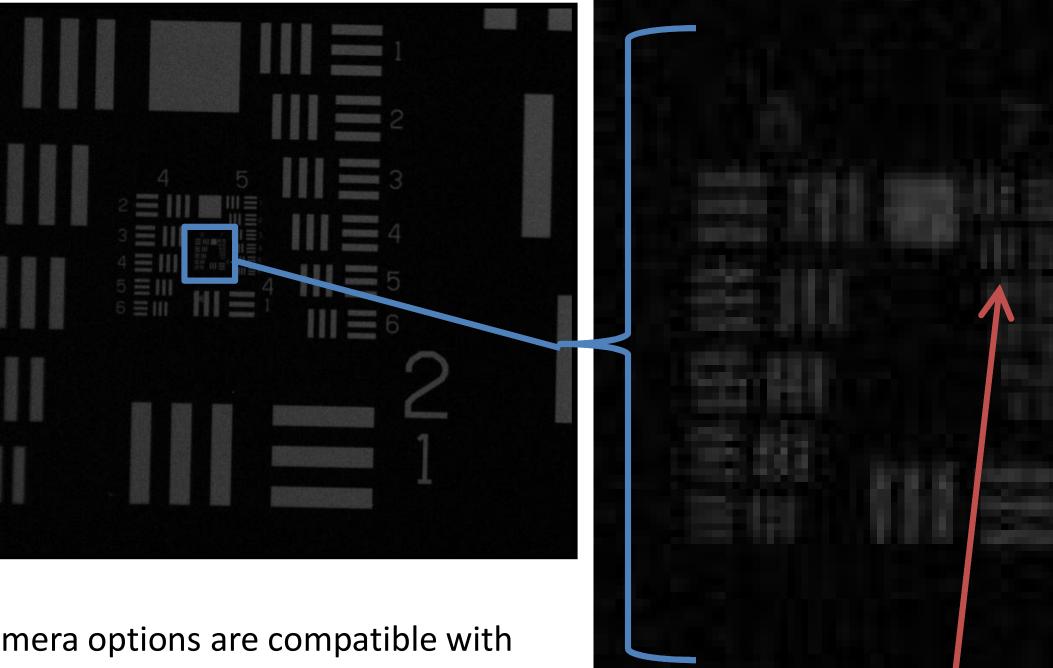


Resolves ~8 μ m feature size



CCD imaging chip





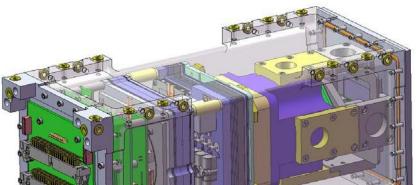
d lens system



Payload Functional Overview



S	~1.5 – 2kg
ors	Pressure : accurate to within 2% of 1 atm. Humidity: accurate to within 4% from true value after temp. correc Temperature accurate to within +/- 0.5ºC.
essor	PIC 32
perature rol	Kept at 4°C to 30°C during the experiment phase.
ronmental nber	Maintains an atmospheric pressure in the pressurized payload cham volume within 11.7 to 15.7 psia.
em lization	Material selections have heritage to past missions for ability to be sterilized and low outgassing.

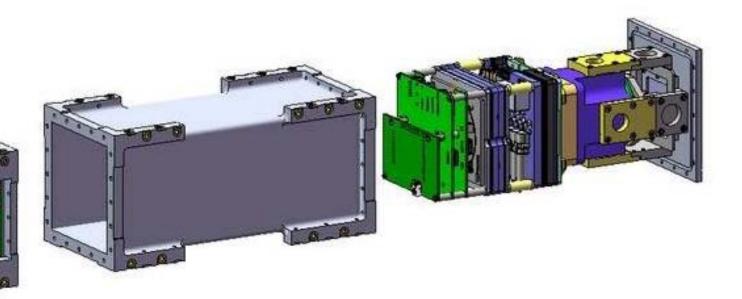




Payload Design Overview



Integrated Payload - Exploded View -









Led en

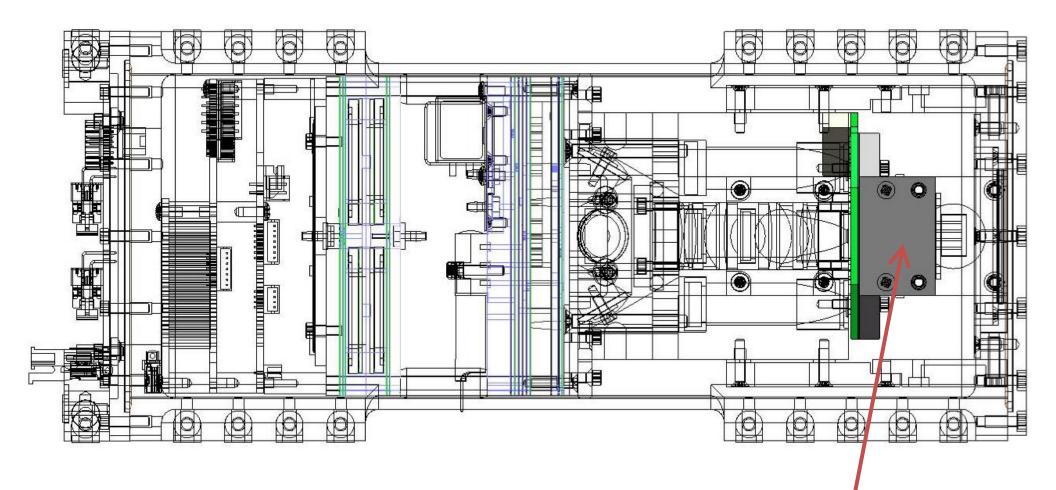
Integrated Payload - Sectional View -

Focal **Imaging field**





LED Assembly

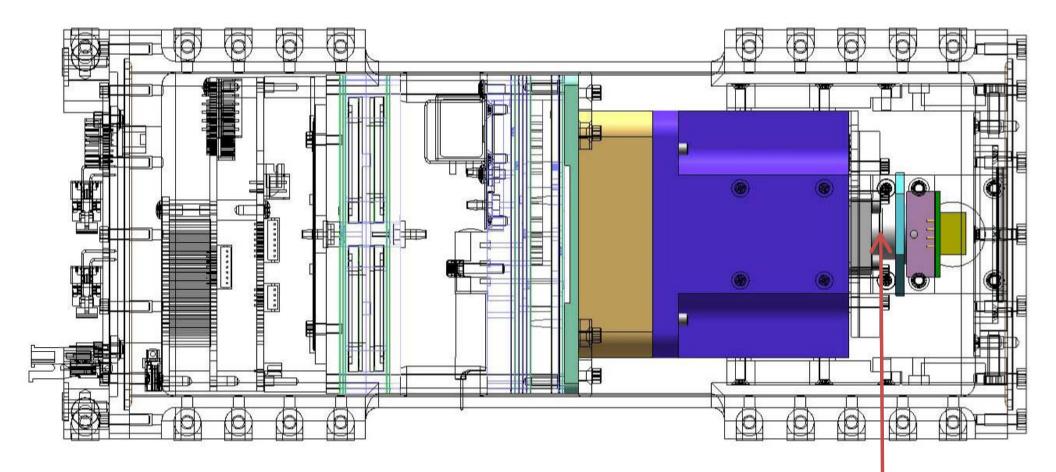


LED heat sink





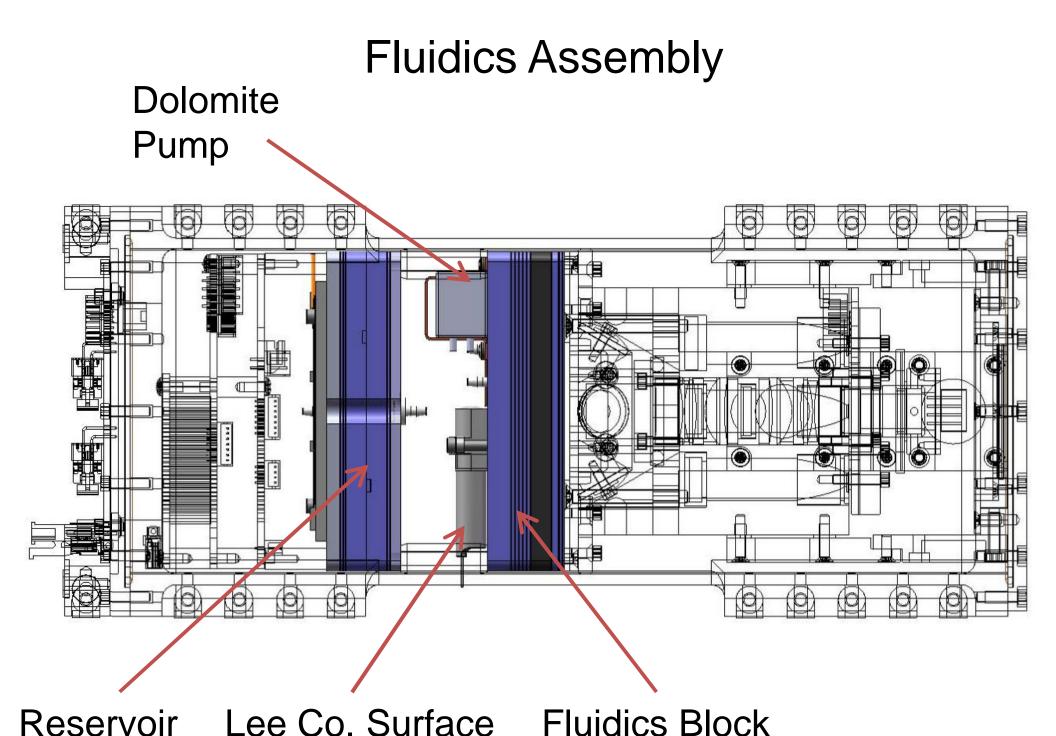
Camera Assembly



Camera Adapter/Focus Adju



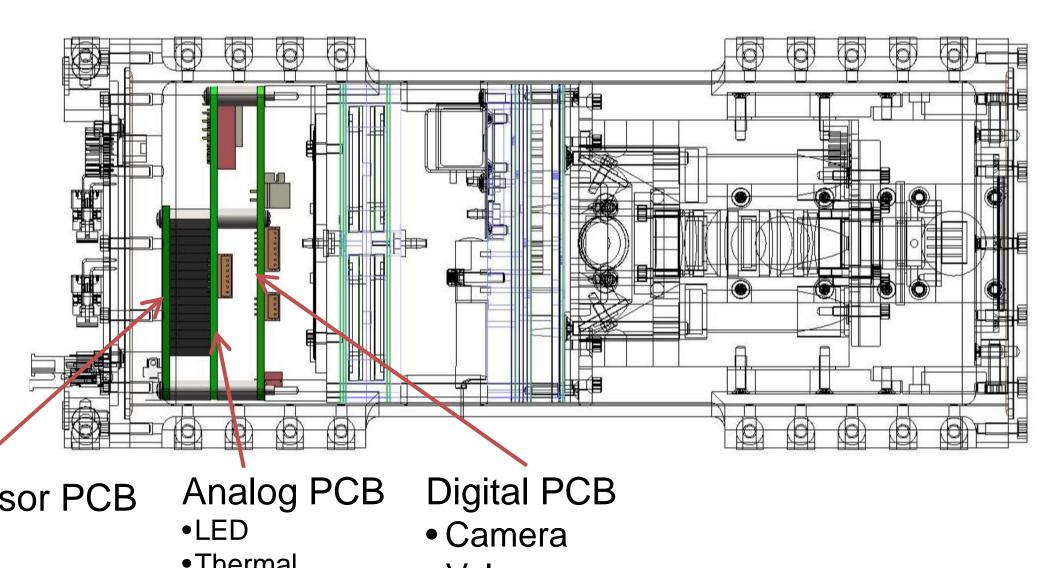








Electronics Assembly





Conclusion



- echnology built:
- integrated fluidics system that maintains
 C.elegans viability and supports growth
- fixed-focus imager with fluorescence and scattered-light imaging capabilities
- Biocompatibility testing complete

Vaiting to partner with a principle nvestigator and launch



Acknowledgements



- dics: Ming Tan, Matthew Piccini
- ogy: Matthew Lera, Macarena Parra
- ging: Linda Timucin
- hanical: Abraham Rademacher, Giovanni Minelli, Chris Beasle
- trical: Aaron Schooley
- agement: Andres Martinez
- nology: Antonio Ricco

ding: NASA/Exploration Systems Mission Directorate





Thank You

Questions?



ConOps



lmager Test A, B, W)	Heating	Image Red Beads	Move Red Beads Out	Image	Move Green Beads In	lmage Green Beads	Feed all worms	Move Green Beads Out	Image	Move Red Worms In	Red Worm Imaging	Move Red Worms Out	Image	Move Green Worms In	Green Worm Imaging	Move Green Worms Out	Imaç
O	ff		Move Red Out	Off	Move Green In	Off	Feed all worms	Move Green Out	Off	Move Worms In	Off	Move Worms Out	Off	Move Worms In	Off	Move Worms Out	Ofl

Validate Imager Cycle	No imaging	3 Cycles A, B, W	No imaging	1 Cycle A, B, W		4 Cycles A, B, W	No Imaging	1 Cycle A, B, W	No imaging	~ 6 Cycles & back to back with same color	No Imaging	1 Cycle A, B, W	No Imaging	~ 6 Cycles & back to back with same color	No Imaging	1 Сус А, В,
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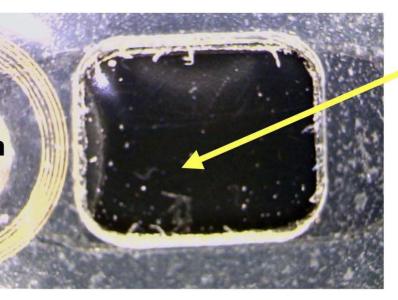
Heat to (22C)	Temp Maintain (22C)

Alive C)	Heat to (22C)	Temp Maintain (22C)
)	(22C)	

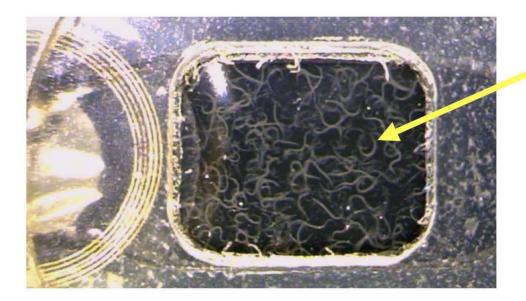
Alive	Heat				
	to				
;)	(17C)				

Temp Maintain (17C)





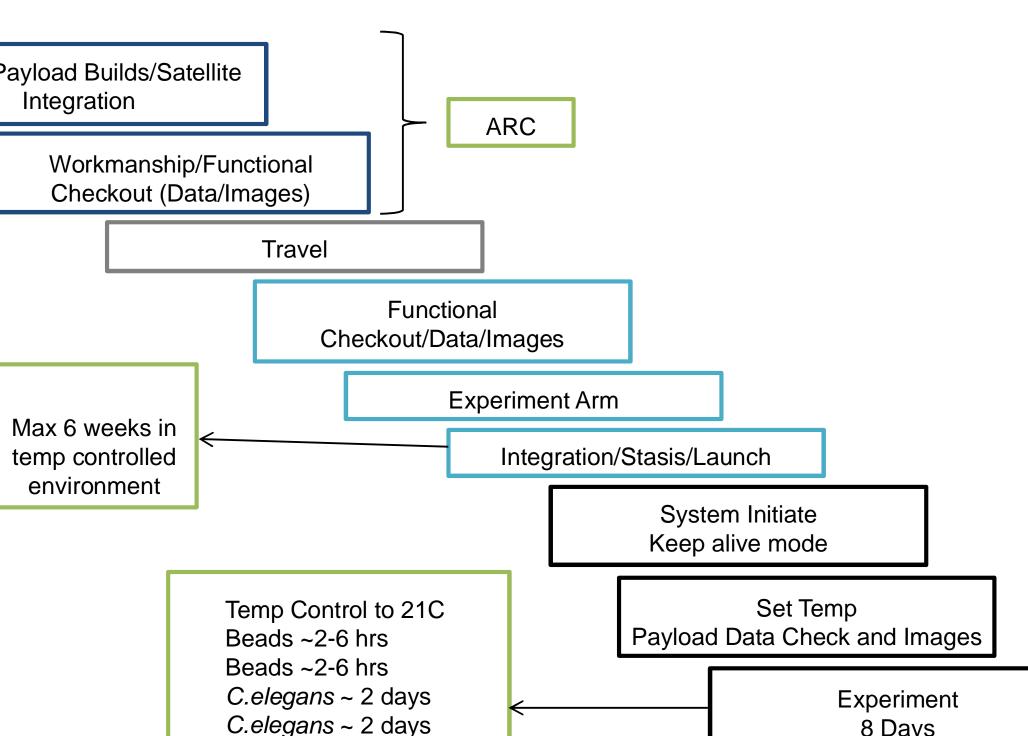
After pumping worms into imaging chamber





Payload ConOps







act with C. elegans

- b. (machined)
- b. porous membrane
- e-sensitive adhesive (PSA) (cut edges only)
- acrylic adhesive, polyester carrier
- laser-cut edges
- rene gas-permeable cover film

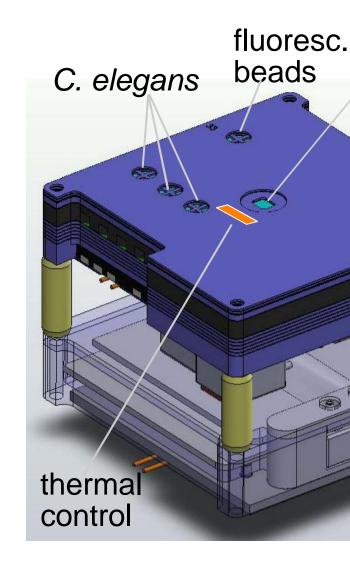
vith growth medium only (reservoir/tubing)

- nylene vinylacetate) [EVA] bag
- fone barbed ports
- thane tubing

ntact with C. elegans and growth medium

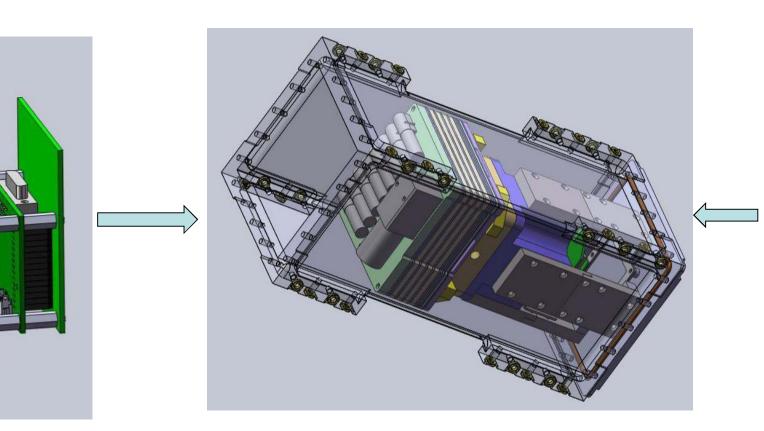
- wetted valve materials
- .PPS = Polyphenylene sulfide ("Fortron")
- .PBT = Polybutylene terephthalate ("Valox")
- .316 SS
- .FeCr Alloy
- Silicone Rubber

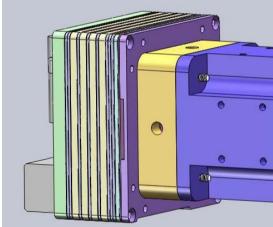


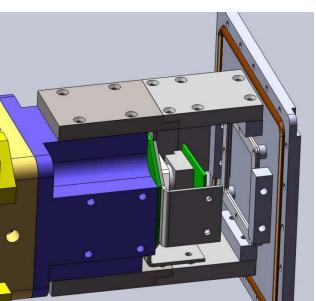


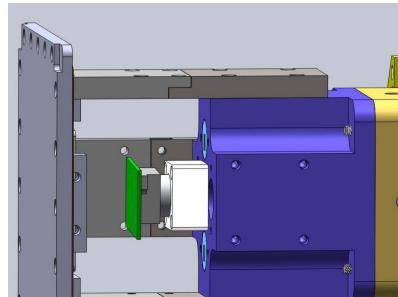








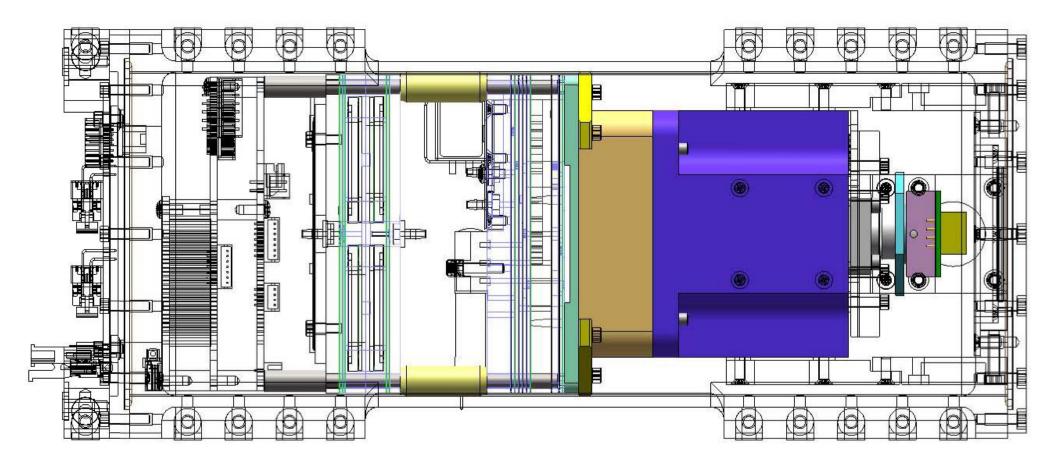






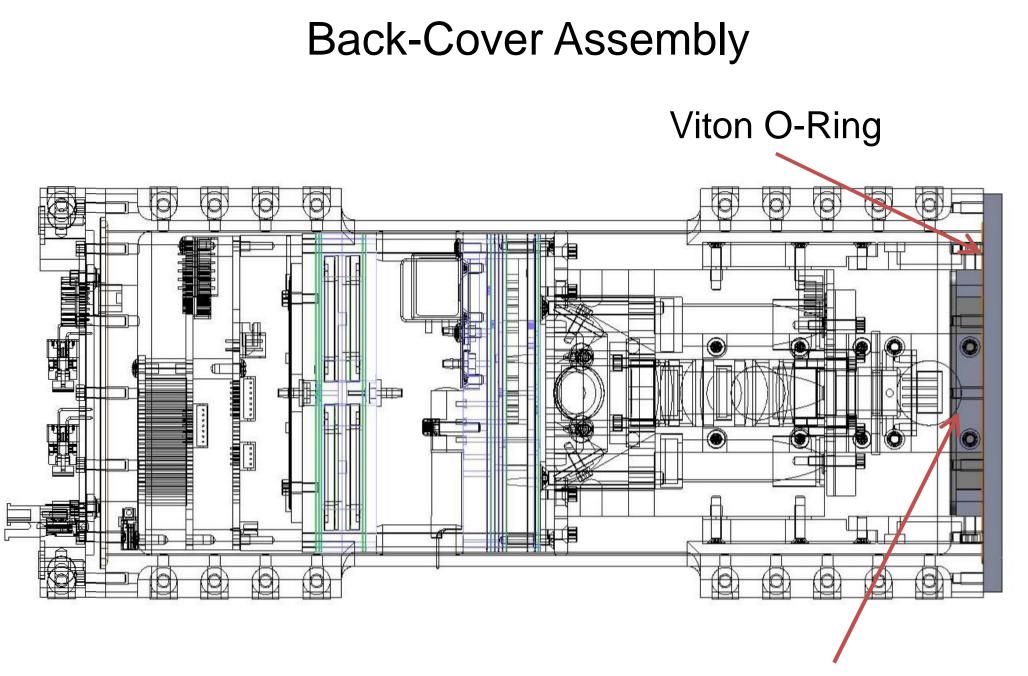


Fluidics Stack Pins - Detail -







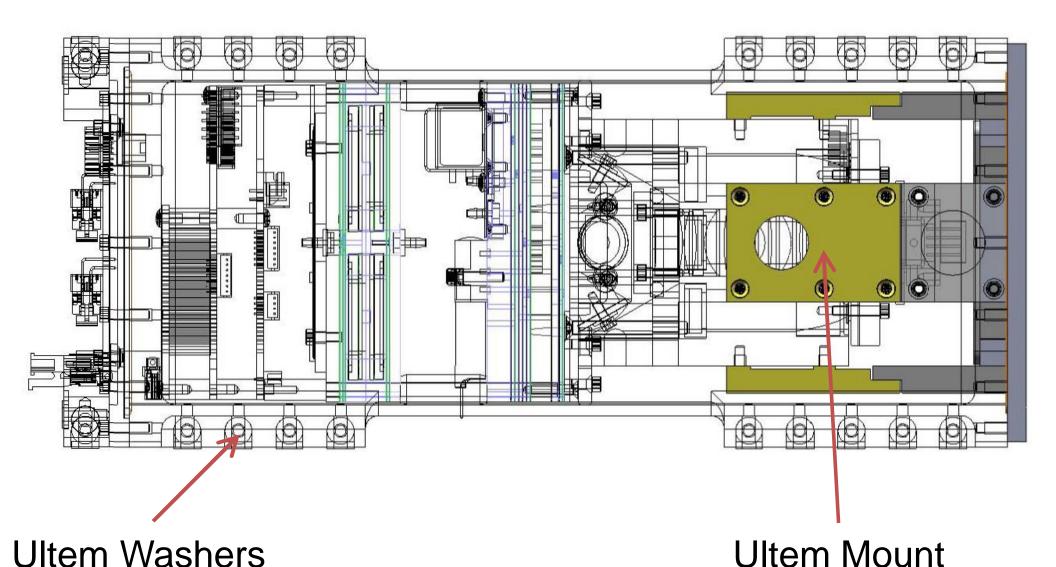


Activated Carbon





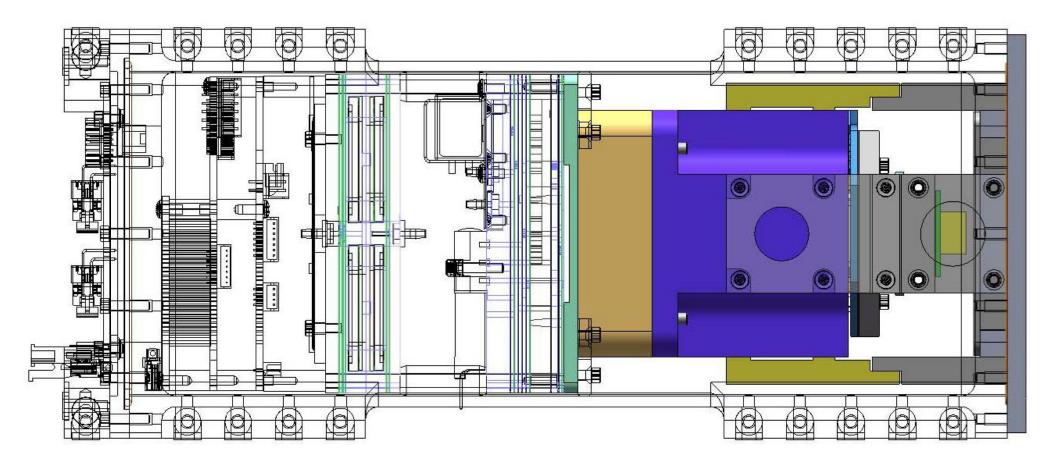
Camera Mount - Mated to Back Panel -





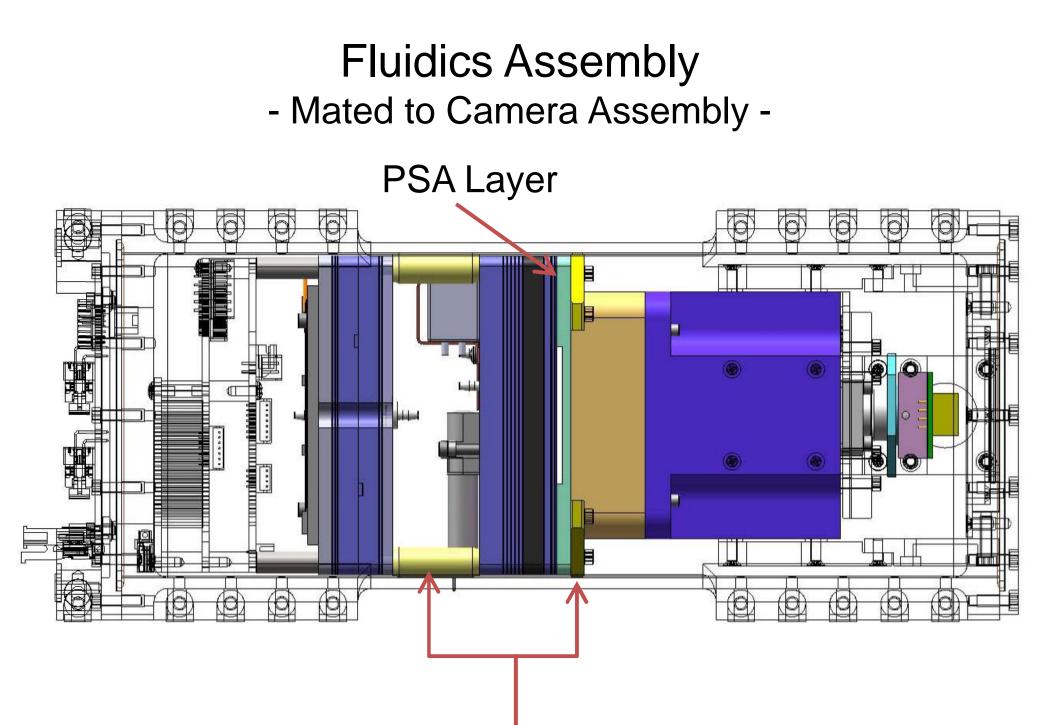


Camera Assembly - Mated to Camera Mount -









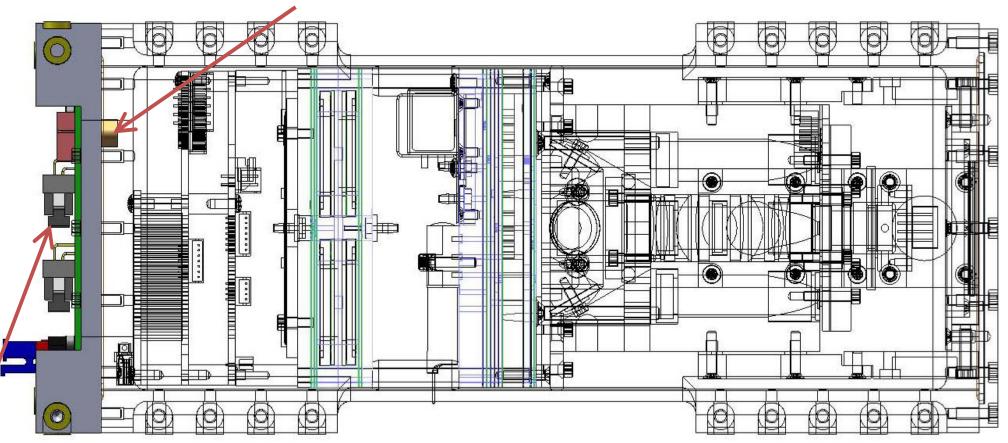




Front Panel Assembly

Hermetic Connector

•Laser Welded

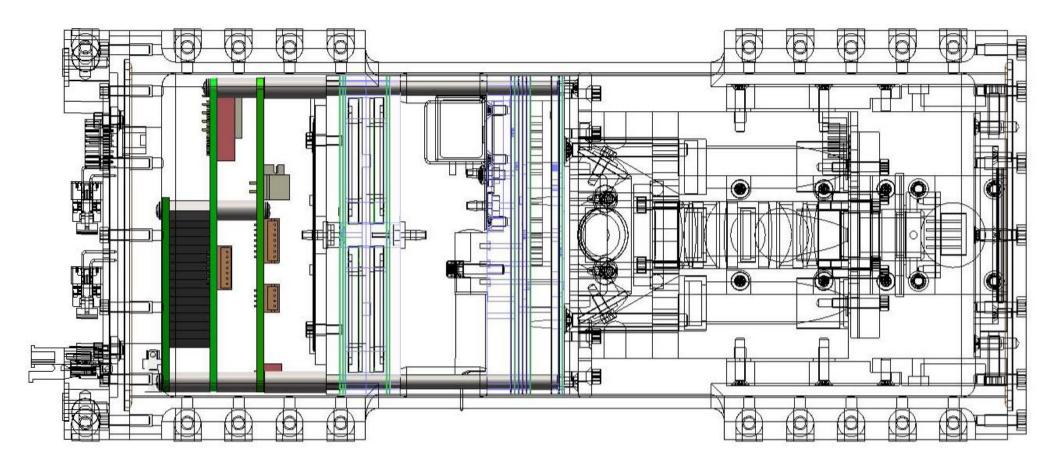


er Interface Board





Electronics Assembly - Mated to Fluidics Stack Pins -

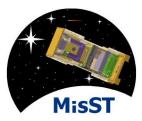






	Description
0 Imager Interface Board	Translates the bus 50pin cable to the hermetic connect the payload can. Also has connection for the remove k flight "kill switch"
0 Imager Payload Analog	Contains all analog payload circuits. Temperature ser circuits, LED current drivers, RH sensor, pressure ser circuitry, Heater circuitry
2 Imager Payload Digital	Contains all digital and inductive load circuits. Valve bridges, motor switches, memory chips, camera interf
0 Imager LED	Contains the 4 high power imager illumination led's <i>A</i> to a heat sink and devices are properly thermally sunfor continuous use.
0 Imager Processor	Contains the payload microprocessor and associated uC LDO, RTC, FRAM, ADC reference



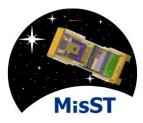


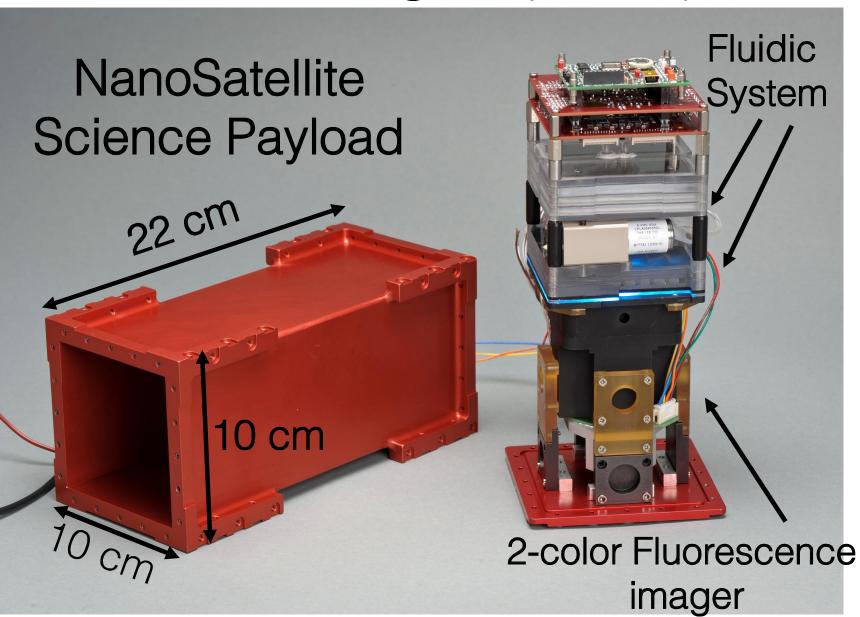
Development of two-color fluorescent imager and integrated fluidic system for nanosatellite biology applications

Diana Wu *et al* NASA Ames Research Center ASGSR December 1, 2012



Microsatellite *in-situ* Space Technologies (MisST)







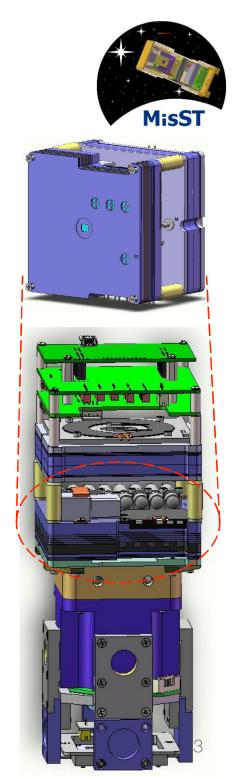
Payload Science & Technology Overview

Objective: Technology demonstration of 2-color fluorescent imager for space biology applications

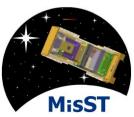
Target: model organisms, resolve major features/organs, gene of interest & control

Implementation: 2-cube integrated culture/imaging instrument

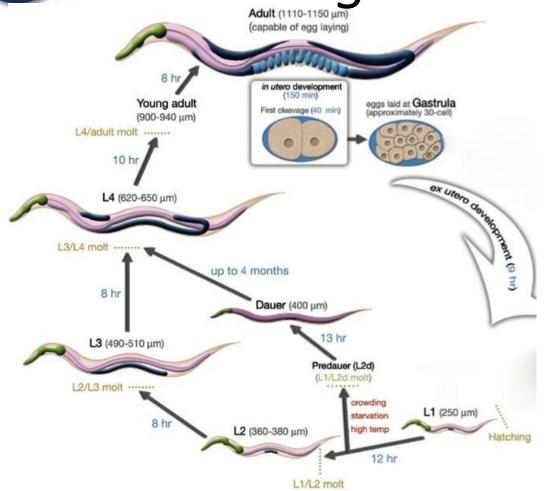
- C. elegans model organism
- •Fluidics: culture wells, imaging zone, reagent storage
- •Imager: fixed focus, 300 kpixel CMOS chip
- •Image storage: 2 GB/6,000 images/8 bits per color (telemetry of 1 20 images)
- Hermetic containment vessel





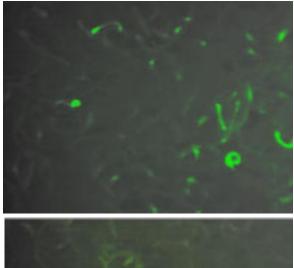


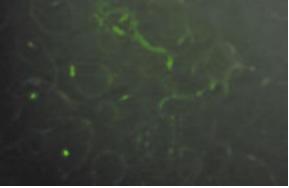
C. elegans Overview

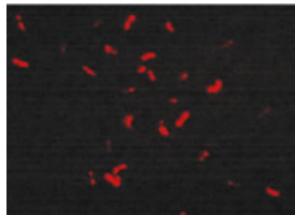


•Easy feeding and short life cycle make it inexpensive to maintain and ideal for research

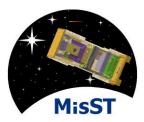
•Used to study spaceflight effects and experiments using *C. elegans* have flown numerous times including on STS-107.









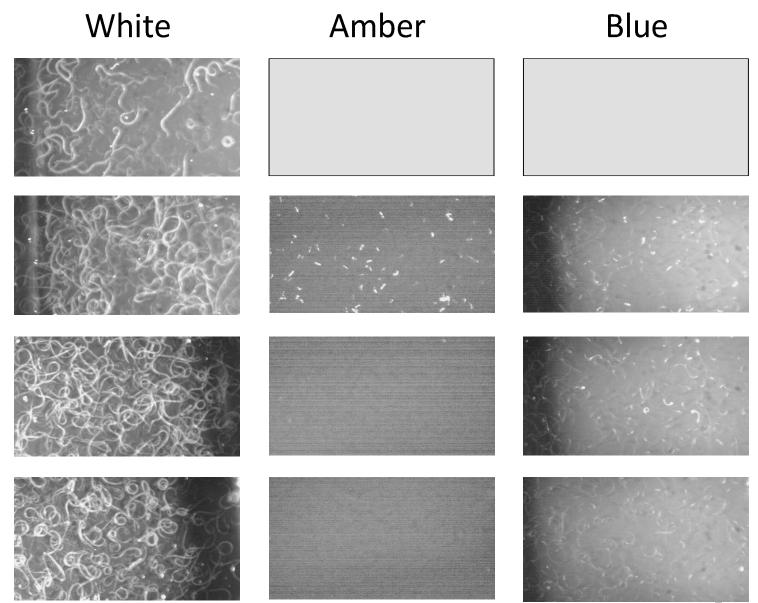


N2 (wt) not fluorescent

tdTomato red – muscle cells

IM324 green – nerve cells

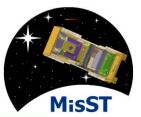
MR142 green – intestinal cells



C. elegans Imaging



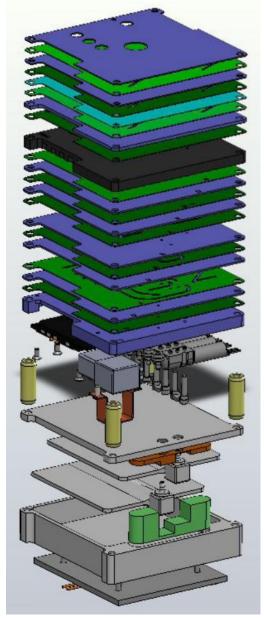
Fluidics Overview



Approach: Multilayer polymer microfluidic card

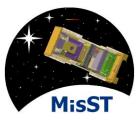
- Multi-layer lamination to form 3-D fluidic network
- Laser-cut fluidic channels in pressuresensitive adhesive (PSA)
- Lamination of machined polycarbonate sheets with via holes, filters, polystyrene using PSA

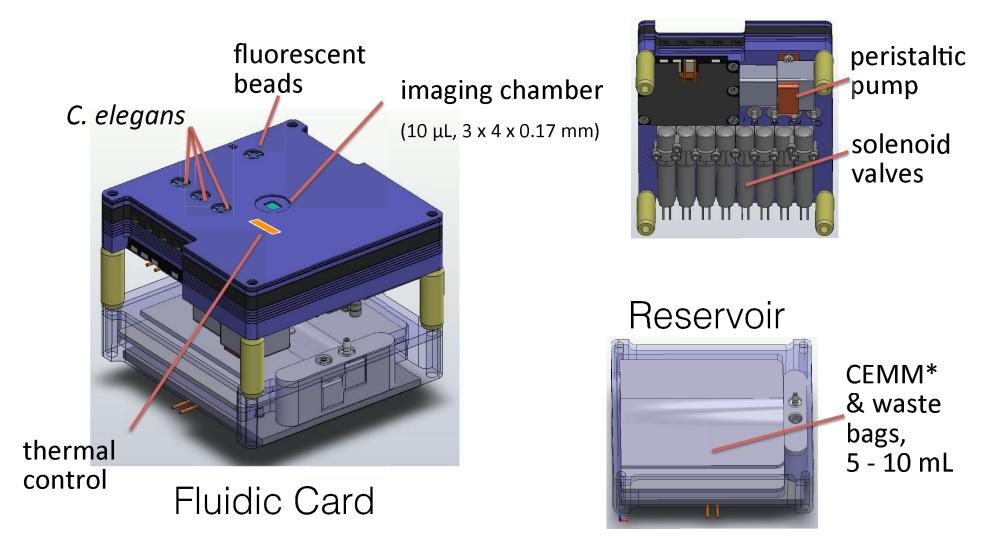
Polymers, adhesives, sterilization, biocompatibility, & fab. all have GeneSat, PharmaSat, O/OREOS-SESLO, EcAMSat heritage





Fluidic Card and Reservoir



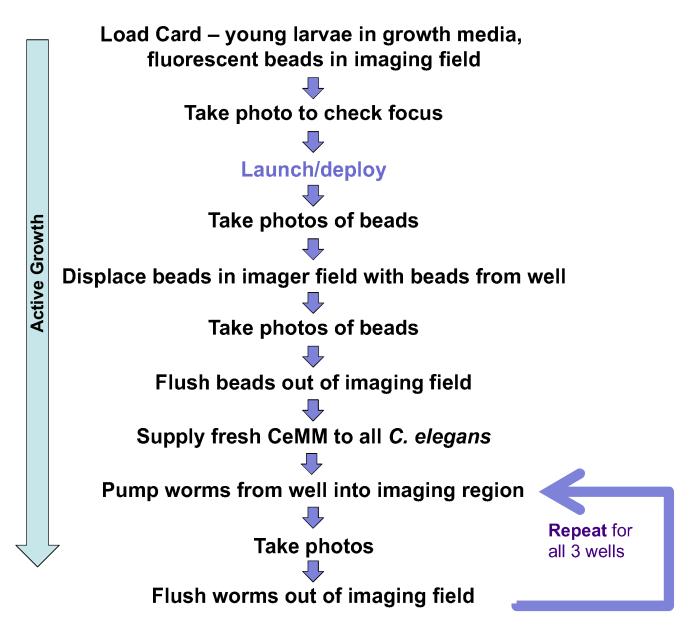


*C. elegans maintenance medium



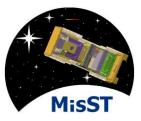


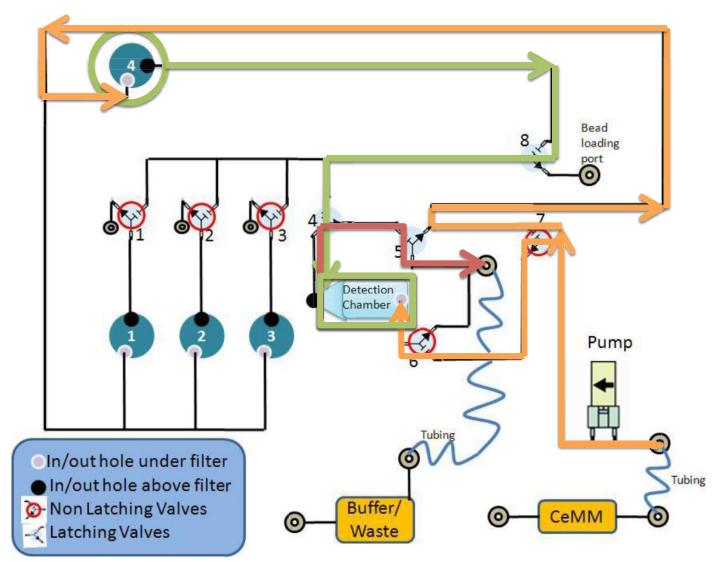
Experimental Timeline





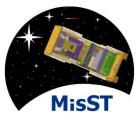
Fluidic Sequence - Beads

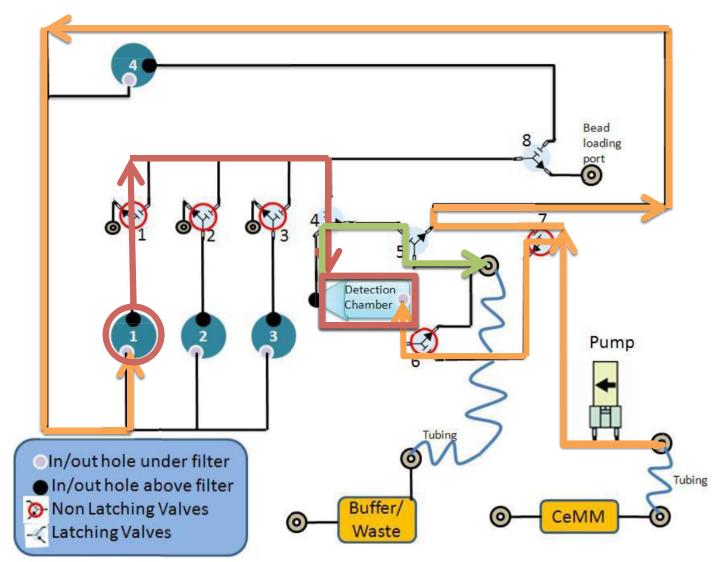






Fluidic Sequence- Worms



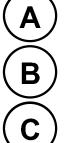




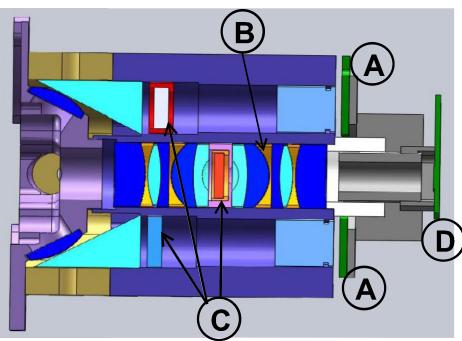
Fluorescence Imager



Imager Components:

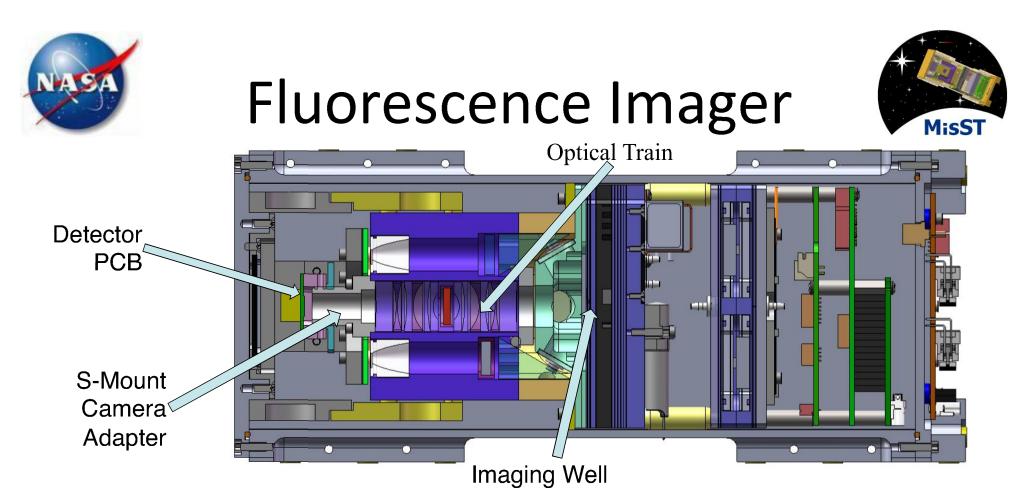


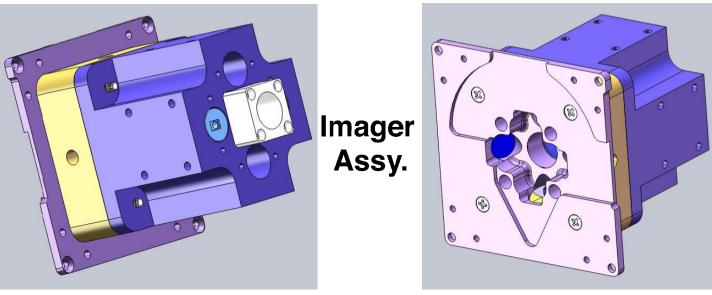
- Luxeon Rebel LEDs: 2 amber, 1 blue, 1 white
- Edmunds relay lens
- Semrock filters:
- blue excitation, amber excitation, dual band-pass emission
-) Camera:
 - COMedia C328-7221



Imager Specifications:

- 2-color fluorescence: Green excite/emit peaks at: 487/509nm Red excite/emit peaks at: 587/610nm
- Cool white for standard imaging
- Lateral resolution: ~ 8 μ m
- Magnification: 1:1
- Field of View: ~3 x 4 mm
- Depth of Focus: ~150 μm





- Fluidic well depth:
 0.17 mm
- Fine pitch thread on
 S-mount (0.5mm/thread)
- Camera adjustment:
 ±5 mm

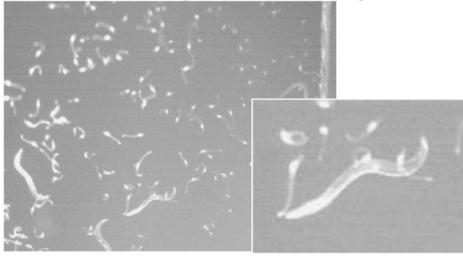
• Locknut/set screws hold detector in place

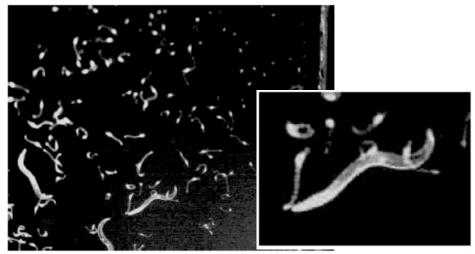


COMedia C328-7221 Camera chip

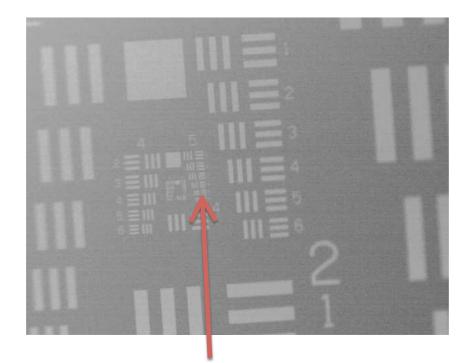


green fluorescently labeled C. elegans





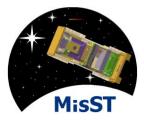
Post Processing [Contrast]

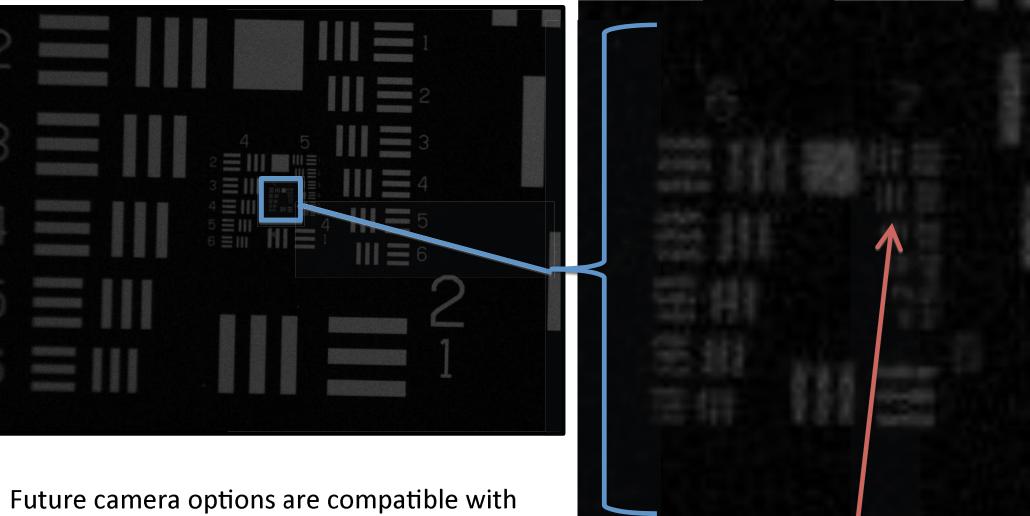


Resolves ~8 µm feature size



CCD imaging chip





Future camera options are compatible with developed lens system (e.g., Omnivision 7141 CMOS)

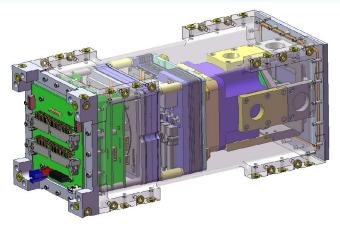
Resolves ~ 4 μm feature size







Mass	~1.5 – 2kg	
Sensors	Pressure : accurate to within 2% of 1 atm. Humidity: accurate to within 4% from true value after temp. correction Temperature accurate to within +/- 0.5°C.	
Processor	PIC 32	
Temperature Control	Kept at 4°C to 30°C during the experiment phase.	
Environmental Chamber	Maintains an atmospheric pressure in the pressurized payload chamber volume within 11.7 to 15.7 psia.	
System Sterilization	Material selections have heritage to past missions for ability to be sterilized and low outgassing.	

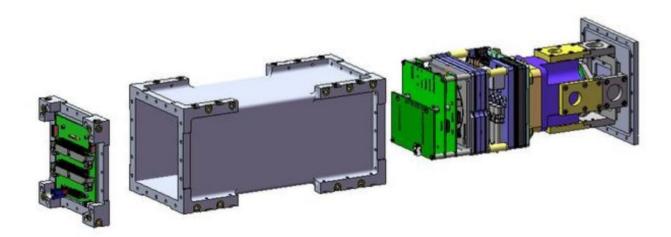




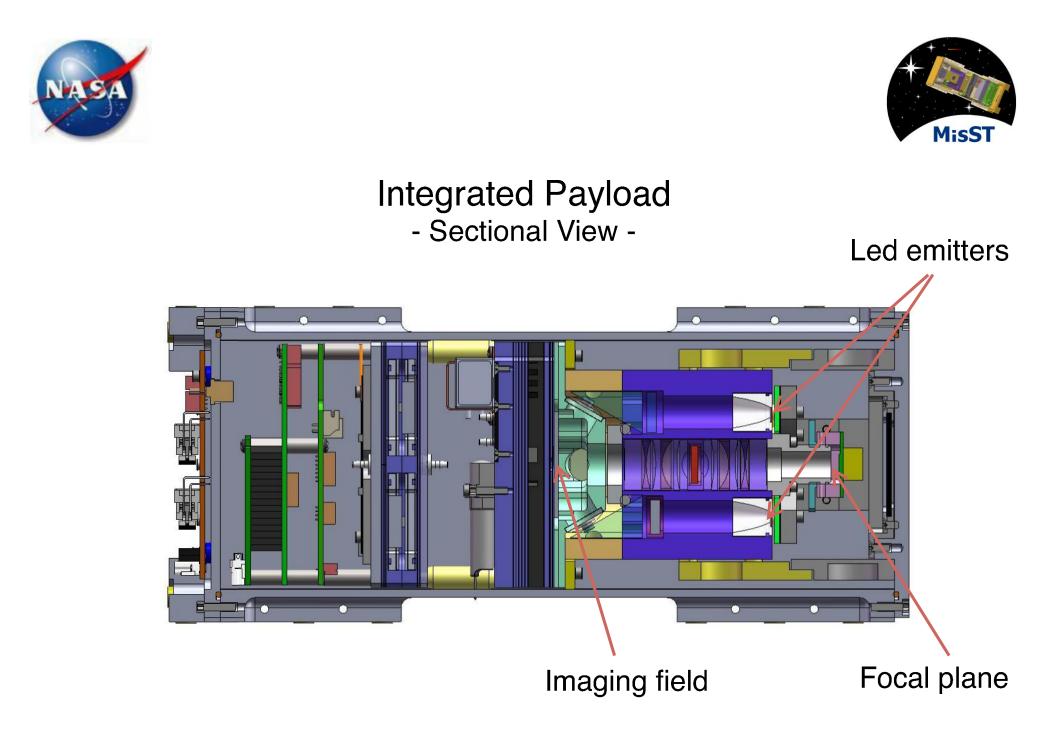


Payload Design Overview

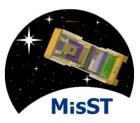
Integrated Payload - Exploded View -



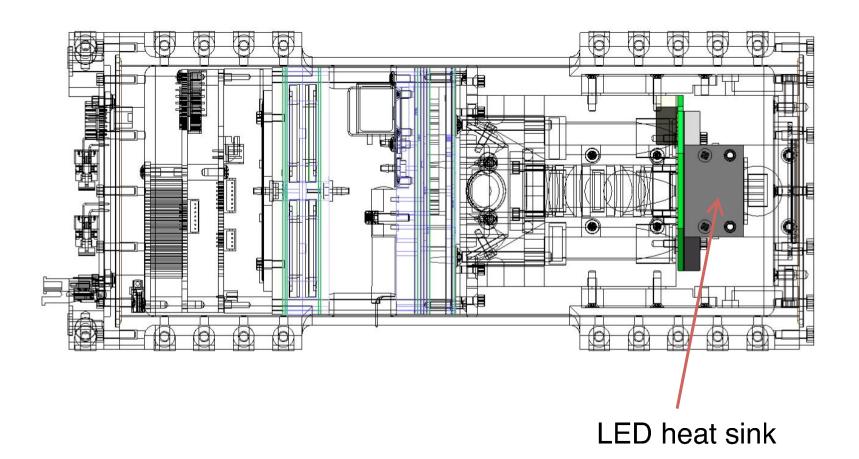








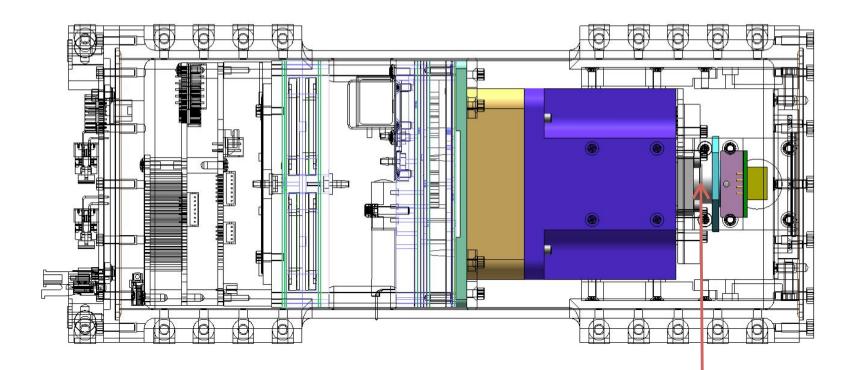
LED Assembly





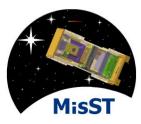


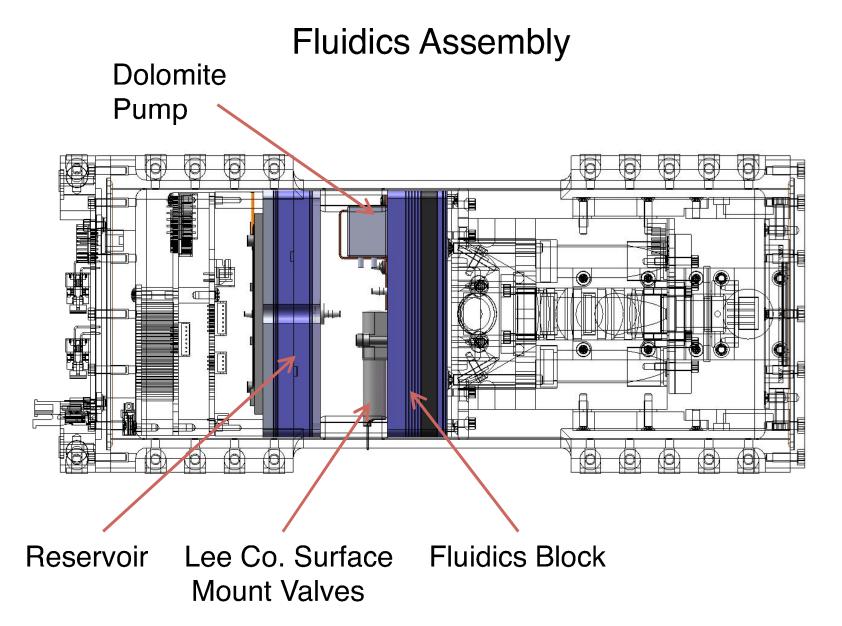
Camera Assembly

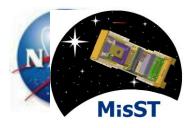


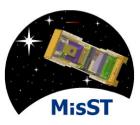
Camera Adapter/Focus Adjustment With Locknut



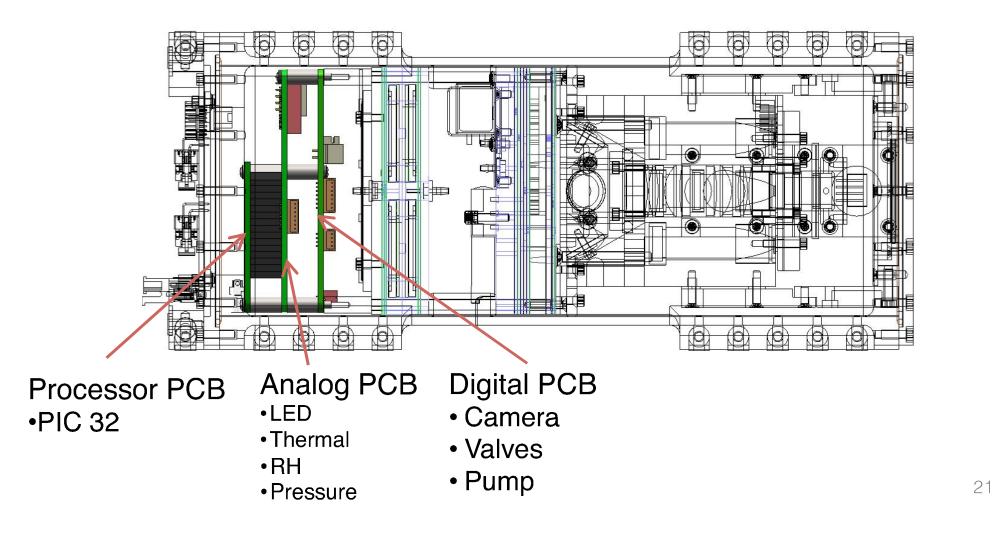






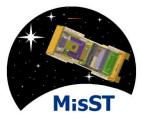


Electronics Assembly





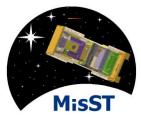
Conclusion



- Technology built:
 - integrated fluidics system that maintains
 C.elegans viability and supports growth
 - fixed-focus imager with fluorescence and scattered-light imaging capabilities
- Biocompatibility testing complete
- Waiting to partner with a principle investigator and launch



Acknowledgements



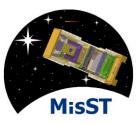
Fluidics: Ming Tan, Matthew Piccini

Biology: Matthew Lera, Macarena Parra

- Imaging: Linda Timucin
- Mechanical: Abraham Rademacher, Giovanni Minelli, Chris Beasley
- Electrical: Aaron Schooley
- Management: Andres Martinez
- Technology: Antonio Ricco

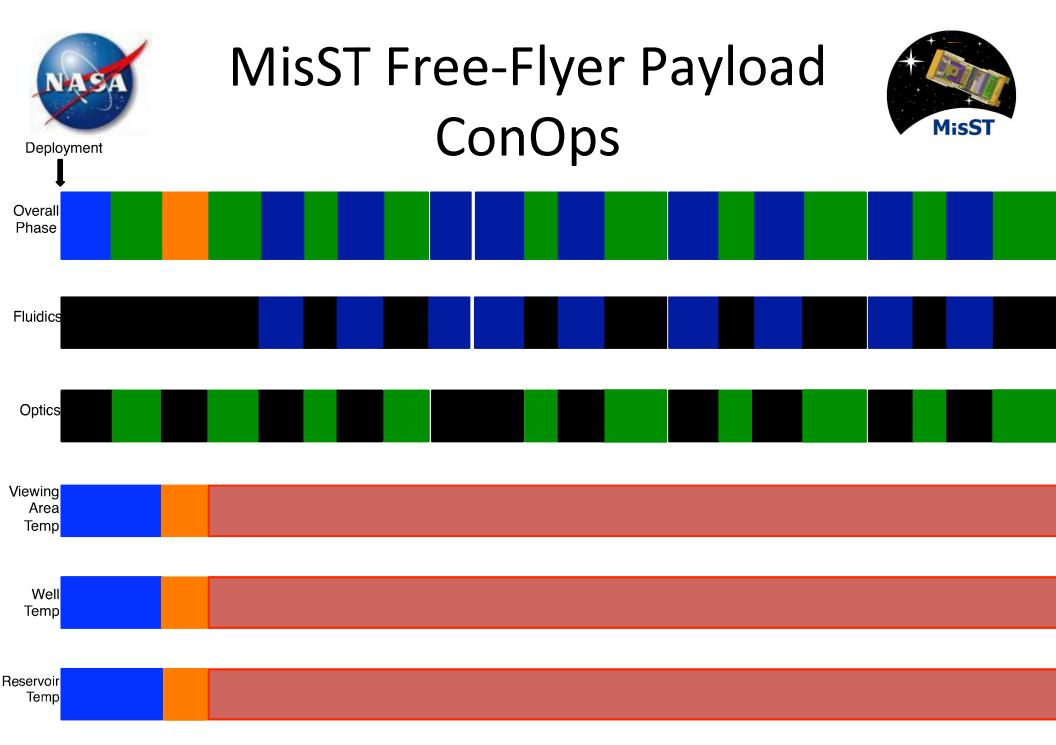
Funding: NASA/Exploration Systems Mission Directorate





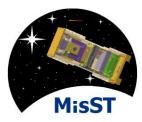
Thank You

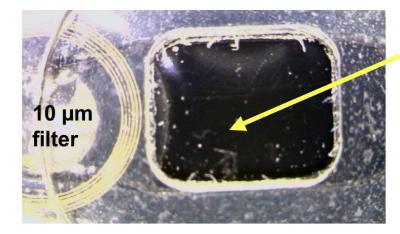
Questions?



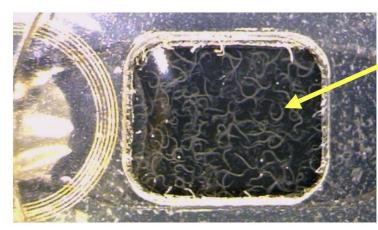
Note: A = amber, B = blue, W = white







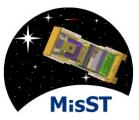
After pumping worms into imaging chamber

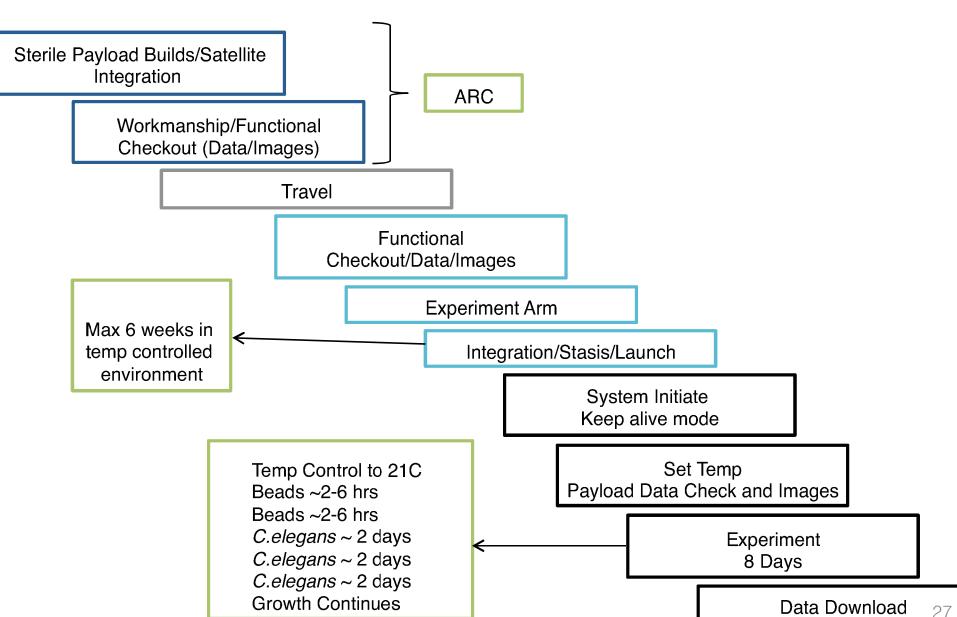


 C. elegans growth after 1 week



Payload ConOps







Payload 1 (Imager): Fluidics materials



Direct contact with C. elegans

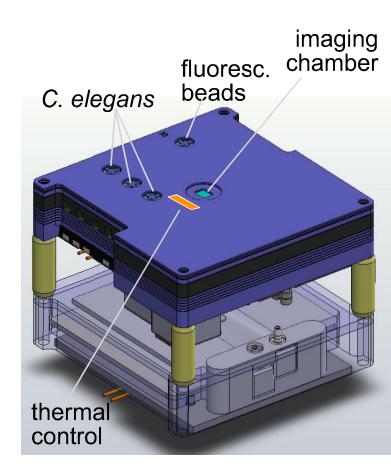
- 1. Polycarb. (machined)
- 2. Polycarb. porous membrane
- 3. Pressure-sensitive adhesive (PSA) (cut edges only)
 - acrylic adhesive, polyester carrier
 - laser-cut edges
- 4. Polystyrene gas-permeable cover film

Contact with growth medium only (reservoir/tubing)

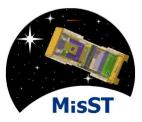
- 1. Poly(ethylene vinylacetate) [EVA] bag
- 2. Polysulfone barbed ports
- 3. Polyurethane tubing

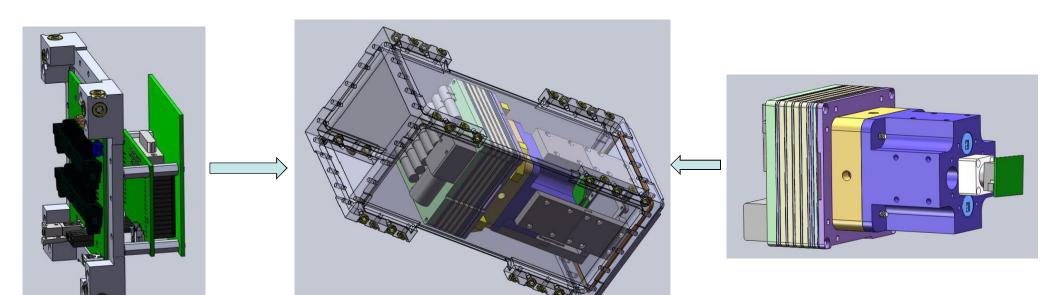
Direct contact with C. elegans and growth medium

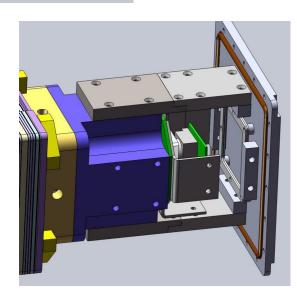
- 1. Internal wetted valve materials
 - 1.PPS = Polyphenylene sulfide ("Fortron")2.PBT = Polybutylene terephthalate ("Valox")3.316 SS
 - 4.FeCr Alloy
 - 5.Silicone Rubber

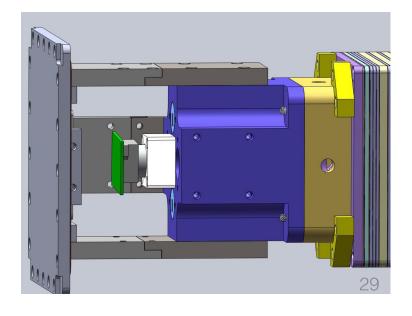




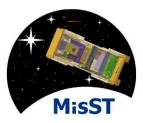




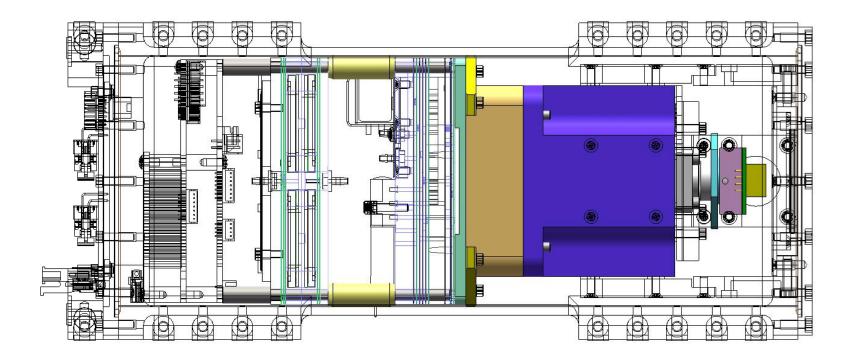




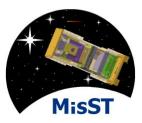




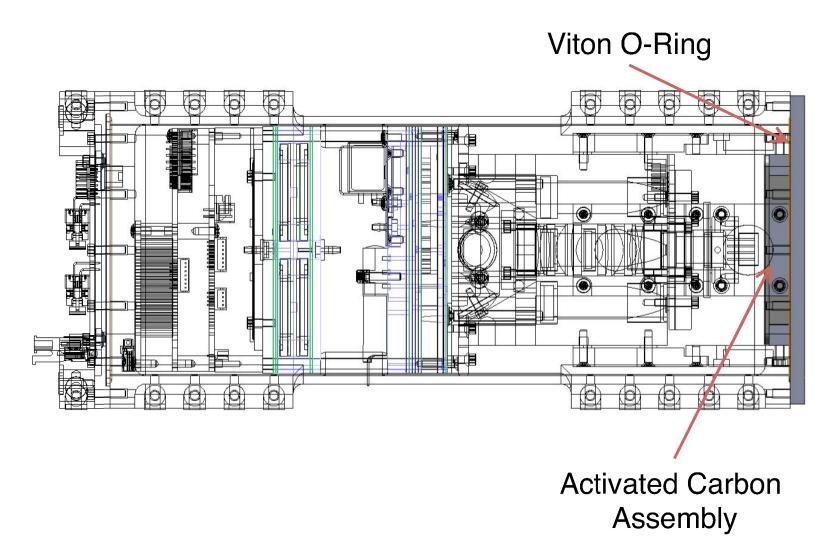
Fluidics Stack Pins - Detail -



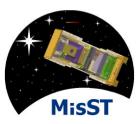




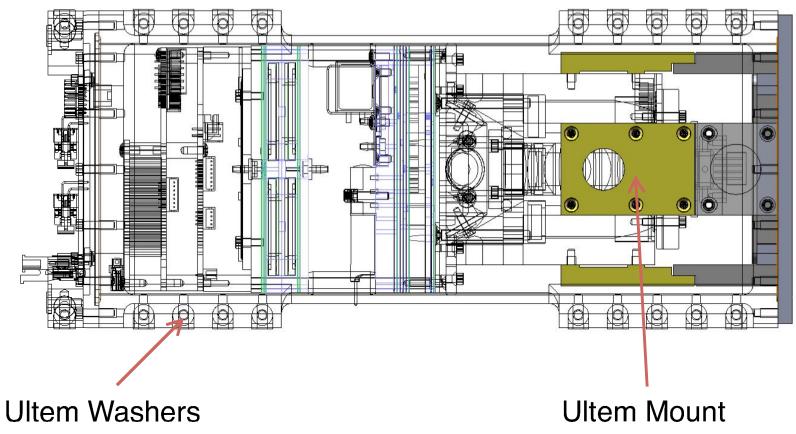
Back-Cover Assembly







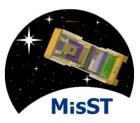
Camera Mount - Mated to Back Panel -



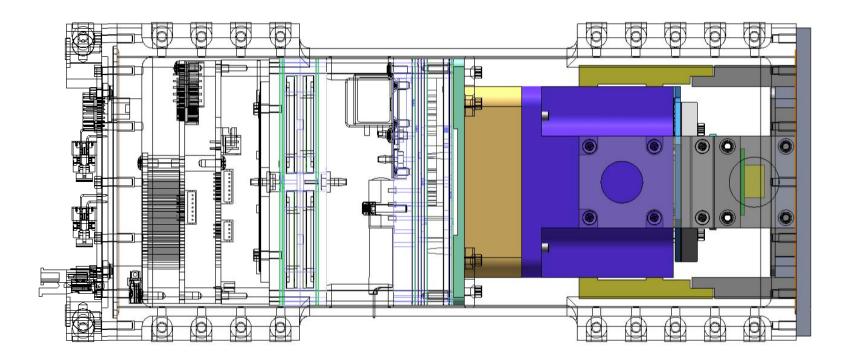
Enclosure Thermal Isolation

Payload Thermal Isolation

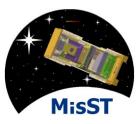




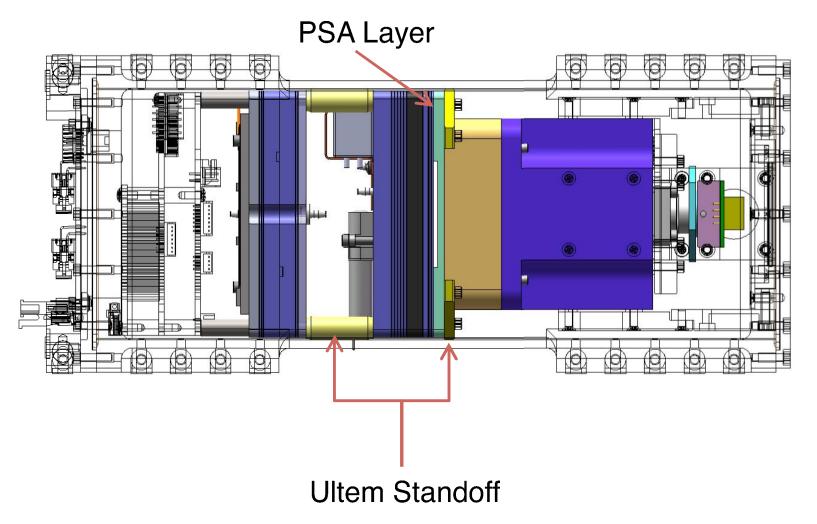
Camera Assembly - Mated to Camera Mount -



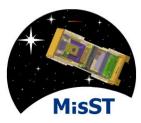




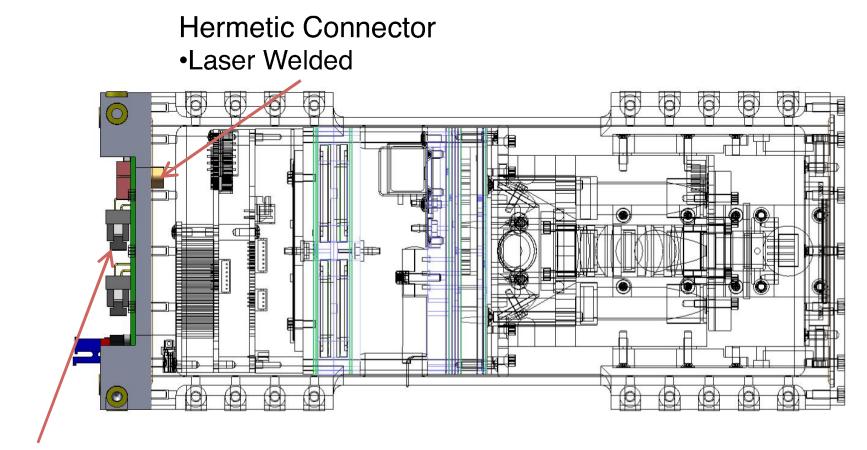
Fluidics Assembly - Mated to Camera Assembly -





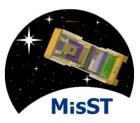


Front Panel Assembly

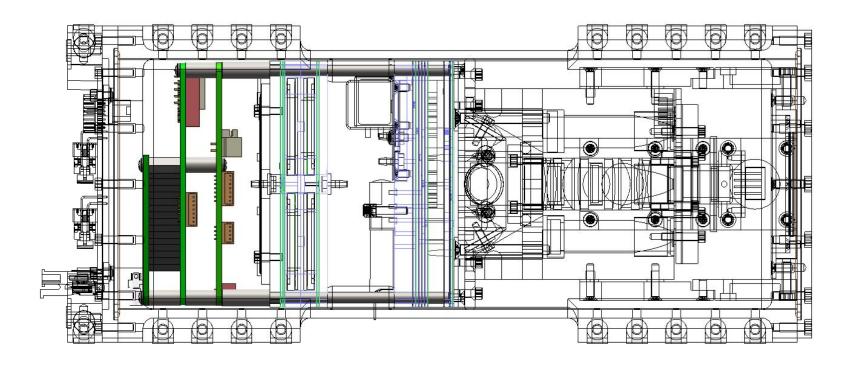


Imager Interface Board



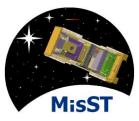


Electronics Assembly - Mated to Fluidics Stack Pins -





Payload Electrical Overview



M410 Imager Interface Board	Translates the bus 50pin cable to the hermetic connector on the payload can. Also has connection for the remove before flight "kill switch"
M420 Imager Payload Analog	Contains all analog payload circuits. Temperature sensor circuits, LED current drivers, RH sensor, pressure sensor and circuitry, Heater circuitry
M422 Imager Payload Digital	Contains all digital and inductive load circuits. Valve H- bridges, motor switches, memory chips, camera interface.
M430 Imager LED	Contains the 4 high power imager illumination led's Attached to a heat sink and devices are properly thermally sunk to allow for continuous use.
M450 Imager Processor	Contains the payload microprocessor and associated circuitry, uC LDO, RTC, FRAM, ADC reference