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Over-expressing the C₃ photosynthesis cycle enzyme Sedoheptulose-1-7 Bisphosphatase improves photosynthetic carbon gain and yield under fully open air CO₂ fumigation (FACE)

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Abstract

Background: Biochemical models predict that photosynthesis in C₃ plants is most frequently limited by the slower of two processes, the maximum capacity of the enzyme Rubisco to carboxylate RuBP ($V_{c,max}$), or the regeneration of RuBP via electron transport (J). At current atmospheric [CO₂] levels Rubisco is not saturated; consequently, elevating [CO₂] increases the velocity of carboxylation and inhibits the competing oxygenation reaction which is also catalyzed by Rubisco. In the future, leaf photosynthesis (A) should be increasingly limited by RuBP regeneration, as [CO₂] is predicted to exceed 550 ppm by 2050. The C₃ cycle enzyme sedoheptulose-1,7 bisphosphatase (SBPase, EC 3.1.3.17) has been shown to exert strong metabolic control over RuBP regeneration at light saturation.

Results: We tested the hypothesis that tobacco transformed to overexpressing SBPase will exhibit greater stimulation of A than wild type (WT) tobacco when grown under field conditions at elevated [CO₂] (585 ppm) under fully open air fumigation. Growth under elevated [CO₂] stimulated instantaneous A and the diurnal photosynthetic integral (A') more in transformants than WT. There was evidence of photosynthetic acclimation to elevated [CO₂] via downregulation of $V_{c,max}$ in both WT and transformants. Nevertheless, greater carbon assimilation and electron transport rates (J and J_{max}) for transformants led to greater yield increases than WT at elevated [CO₂] compared to ambient grown plants.

Conclusion: These results provide proof of concept that increasing content and activity of a single photosynthesis enzyme can enhance carbon assimilation and yield of C₃ crops grown at [CO₂] expected by the middle of the 21st century.

Keywords: climate change, photosynthetic carbon reduction cycle, C₃ plants, RuBP regeneration, electron transport, improving photosynthesis

Background

Biochemical models of C₃ photosynthesis (A) predict that A is limited by the slowest of three processes: the maximum carboxylation capacity of the enzyme Rubisco ($V_{c,max}$), the regeneration of Ribulose-5-phosphate (RuBP) via whole chain electron transport (J or J_{max}), or

the inorganic phosphate release from the utilization of triose phosphates (TPU or Pi limited) [1,2]. At current atmospheric [CO₂], and under non stressed conditions, light saturated A operates at the transition between Rubisco and RuBP regeneration limitation. Globally, [CO₂] is expected to increase from current levels of 390 ppm [3] to over 550 ppm by the middle of this century [4,5]. Elevating [CO₂] stimulates C₃ photosynthesis by increasing the substrate for carboxylation, CO₂, and by reducing photorespiration [6,7]. Therefore, as atmospheric carbon dioxide concentration increases, the

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control of photosynthesis will shift away from Rubisco limitation toward RuBP regeneration limitation.

Although photosynthetic stimulation at 550 ppm [CO₂] could in theory increase production by 34%, the observed increase in field C₃ crops is only 15% [7,8]. Additional future increases in yield potential of the world's major crops through an increase in the proportion of biomass allocated to grain or an increase in the efficiency of light capture will be small, as conventional breeding programs are reaching the theoretical maximum with diminishing returns [9-11]. In contrast, model simulations demonstrate that increasing photosynthetic efficiency under current [CO₂] by optimizing the biochemistry of photosynthesis could increase the energy conversion efficiency of a given crop in less time than conventional breeding programs [10,12]. At current levels of crop productivity, global food requirements may outpace current crop production by the middle of this century [11,13,14]. Taken together, these observations suggest that direct improvements in photosynthetic efficiency will be needed if we are to meet global food needs in the future.

A common acclimation response of plants grown at elevated [CO₂] is to allocate fewer resources to Rubisco, thereby downregulating maximum carboxylation capacity (V_{cmax}). This so called photosynthetic acclimation makes more resources available for other metabolic processes [6,15]. The implication is that plants could reallocate resources in the photosynthetic carbon reduction (PCR) cycle to increase the efficiency of N use in elevated [CO₂] [6,7]. In practice, however, plants' photosynthetic resources are not optimally allocated for current [CO₂] nor is their acclimation response optimal in elevated [CO₂] [12]. Theoretically, and by reference to a biochemical model of photosynthesis [i.e., [1]], a plant with a 15% decrease in Rubisco content and 15% increase in RuBP regeneration capacity could translate to a 40% increase in *A* and photosynthetic efficiency of nitrogen use at elevated [CO₂] [Figure 1 in [7]]. It follows that plants engineered with an increased capacity for RuBP regeneration would have a greater increase in productivity in elevated [CO₂] when compared to wild type plants [16-18].

While 11 enzymes are involved in the PCR cycle, modeling and metabolic control analyses have consistently demonstrated that four enzymes are expected to exert the greatest control of flux in the cycle: ribulose biphosphate carboxylase-oxygenase (Rubisco), sedoheptulose-1,7-bisphosphatase (SBPase), aldolase and transketolase [19-21]. Two enzymes, Rubisco and SBPase, are predicted to have the greatest control over carbon assimilation [21,22]. Rubisco is well known to be highly abundant, containing 25% of leaf nitrogen (N) [23] and may in some cases account for up to half

of leaf N [24]. All attempts to improve photosynthesis by manipulating Rubisco expression, activity, or specificity have yielded poor results, in part because of inherent tradeoffs between activity and specificity of the enzyme and limited capacity to add more of this highly abundant protein [25-27]. An additional hurdle to engineering "better" Rubisco is that the functional enzyme requires the coordinated assembly of eight plastid encoded and eight nuclear encoded subunits to form the large (rbcL) and small (rbcS) units of the hexadecameric enzyme [28,29]. With the exception of Rubisco, the other enzymes exerting the greatest control on photosynthesis all function in the RuBP regeneration portion of the PCR cycle. Thus, near term future improvements in photosynthetic biochemistry in C₃ plants are more likely to be achieved by improving content or activity of enzymes other than Rubisco [e.g., [18,21,30,31]].

Sedoheptulose-1,7-bisphosphatase (SBPase) is positioned at the branch point between regenerative (RuBP regeneration) and assimilatory (starch and sucrose biosynthesis) portions of the PCR cycle. It functions to catalyze the irreversible dephosphorylation of sedoheptulose-1,7-bisphosphate (SBP) to sedoheptulose-7-phosphate (S7P). Transketolase then catalyzes the transfer for a two carbon ketol group from S7P to glyceraldehyde-3-phosphate (G3P) to yield xylulose-5-phosphate (X5P) or ribose-5-phosphate (R5P) [32]. SBPase is therefore critical for maintaining the balance between the carbon needed for RuBP regeneration and that leaving the cycle for biosynthesis [20].

Previous experiments have demonstrated that tobacco transformants overexpressing SBPase accumulated more biomass than WT in controlled environment chambers at ambient CO₂ [16]. Smaller increases in biomass were reported for mature SBPase overexpressing plants grown in greenhouse conditions [16]. Additionally, overexpression of SBPase in rice did not increase biomass relative to WT for plants grown at ambient CO₂ levels in two controlled environments [33,34]. The variance in the realized benefit of SBPase overexpression coupled with the fact that RuBP regeneration is highly sensitive to environmental conditions underscores the need to test the response of plants with this single gene manipulation in agronomically relevant conditions [30]. Moreover, models predict that as atmospheric [CO₂] increases so will the benefit of increasing RuBP regeneration capacity in plants [1,21,35]. Therefore, we compared WT and SBPase overexpressing plants under field conditions at ambient and elevated (ca. 585 ppm) [CO₂], and we tested the prediction that transformants would exhibit greater stimulation of photosynthesis and yield than WT plants when grown under fully open air CO₂ fumigation.

Methods

Plant Material

Wild type tobacco (*Nicotiana tabacum* L. cv. Samsun) and sense tobacco plants (T_5 generation *Nicotiana tabacum* L. cv. Samsun) overexpressing a full length *Arabidopsis thaliana* SBPase cDNA, driven by CaMV 35S promoter and the nopaline synthase termination sequence [16], were germinated in Petri dishes and transferred to soil when true leaves emerged. Sense plants (hereafter referred to as 'transformants') were germinated on hygromycin (30 $\mu\text{g/ml}$) medium. One individual from each of two transgenic lines overexpressing SBPase with varying SBPase levels and several randomly selected wild type (WT) individuals were selected for the experiments. Individuals were subsequently clonally propagated by rooting cuttings in peat pots on misting benches and then planted directly in the field at SoyFACE on July 7 2009.

SoyFACE site

The SoyFACE facility is located in the Experimental Research Station of the University of Illinois at Urbana-Champaign [36]. Soybean (*Glycine max*) is grown in eight plots (rings 18 meters in diameter) located within a typically managed soybean field of ca. 40 hectares (ha). Four rings are fumigated with pure $[\text{CO}_2]$ and four rings are non-fumigated controls. Six cuttings of each SBPase genotype (11 and 30) and six of WT were planted in subplots within each ring.

Ambient atmospheric $[\text{CO}_2]$ at the beginning of the 2009 field season was ca. 385 ppm and the target $[\text{CO}_2]$ for elevated rings in 2009 was 585 ppm $[\text{CO}_2]$. In the fumigated rings, 89% of $[\text{CO}_2]$ values recorded every ten minutes from June 19 to September 24, 2009, were within 10% of the target value of 585 ppm. The mean daily $[\text{CO}_2]$ in elevated rings at Soyface during that time was 586.6 ± 19.4 (sd) ppm. Elevated rings were fumigated using a modification of the method of Miglietta *et al.* [37].

Leaf protein and western blotting

Prior to planting, leaf discs were collected from cuttings and immediately frozen in liquid nitrogen to confirm that sense plants had greater SBPase content than WT. Protein quantifications and western blots were performed following [19]. Sample lanes were loaded on an equal protein basis, separated using 10% (w/v) SDS-PAGE, transferred to polyvinylidene difluoride membrane, and probed using antibodies raised against SBPase and transketolase. Antibody target proteins were detected using horseradish peroxidase conjugated to the secondary antibody and ECL chemiluminescence detection reagent (Amersham, Bucks, UK). Western blots were quantified by densitometry using the molecular

imaging Gel Doc XR system (Bio-Rad, Hercules, CA, USA) and imaging software.

In situ measurements of gas exchange and photosynthetic parameters

The diurnal course of photosynthesis at the SoyFACE site was measured on two young fully expand leaves from each genotype at ambient conditions at both normal (385 ppm) $[\text{CO}_2]$ and elevated (585 ppm) $[\text{CO}_2]$ at five time points on two dates in August, 2009. To ensure that each plant was measured in similar environmental conditions, the LEDs of the controlled environment cuvettes of the gas exchange system (LI-6400, LI-COR, Lincoln, Nebraska) were set to deliver the same ambient light PPF. Temperature and relative humidity were similarly set to ambient conditions and kept constant for the duration of each measurement period in the diurnal course. To estimate the total daily carbon gain (A'), photosynthesis was assumed to increase linearly from 0 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at dawn (sunrise) to the first measured value and decrease linearly from the last measured values to 0 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at dusk (sunset). Sunrise and sunset data were determined using the US Naval Observatory website: http://aa.usno.navy.mil/data/docs/RS_OneYear.php. Dew on the leaves prevented us from measuring photosynthesis until about 10:00 h. We estimated A' for each block by integration using the trapezoidal rule and then performed analyses on the integrals [38].

In vivo values of three photosynthetic parameters: maximum carboxylation capacity ($V_{c,\text{max}}$), maximum linear electron transport through photosystem II (J_{max}) and respiration in the light (R_d) were determined by measuring the response of A to intercellular $[\text{CO}_2]$ (C_i) on August 1 and August 15 2009. A vs. C_i curves were measured in situ on one young fully expanded leaf of each genotype in all blocks of each treatment ($n = 4$) with an open gas exchange system (LI-6400, LI-COR, Lincoln, Nebraska). Initially, plants were allowed to reach steady state photosynthesis at their growth $[\text{CO}_2]$ (i.e., 385 ppm or 585 ppm $[\text{CO}_2]$) at a saturating light level of 1500 $\mu\text{mol m}^{-2} \text{ s}^{-1}$. Mean leaf to air vapor pressure deficit (V_{pdL}) was 1.3 ± 0.26 (s.d.), and mean leaf temperature was $26 \pm 1^\circ\text{C}$ (s.d.). Once steady state was reached, photosynthetic $[\text{CO}_2]$ uptake rate (A) and chlorophyll fluorescence parameters were recorded at the growth $[\text{CO}_2]$; then $[\text{CO}_2]$ was decreased in 4 or 5 uniform steps to 50 ppm, returned to growth $[\text{CO}_2]$, and then increased in 4 or 5 uniform steps to 1500 ppm $[\text{CO}_2]$. A minimum of 11 data points were collected for each plant following the methods outlined by Long and Bernacchi [39]. Curves were measured in the morning to avoid confounding treatment and genotype effects with transient decreases in water potential, decreases in

chloroplast inorganic phosphate concentration or decreases in maximum photosystem II (PSII) efficiency (F_v'/F_m').

Electron transport rate (ETR), the actual flux of photons driving PSII, and F_v'/F_m' were calculated using fluorescence parameters, F_s , F_m' , F_o' , [40,41]. Fluorescence parameters were estimated using a Licor 6400 integrated gas exchange system equipped with a fluorescence and light source accessory (LI-6400, LI-COR, Lincoln, Nebraska). F_s is the steady state light adapted fluorescence, F_m' is the maximal fluorescence of a light adapted leaf following a saturating light pulse, and F_o' is the minimal fluorescence of a light adapted leaf that is darkened.

$$ETR = \left(\frac{F_m' - F_s'}{F_m} \right) f I \alpha_{leaf}$$

Where f , is the fraction of photons absorbed by PSII, assumed be 0.5 for C_3 plants; I is the incident photon flux density ($\mu\text{mol m}^{-2} \text{s}^{-1}$); and α is leaf absorptance which was constant (0.87).

A vs. C_i curves were fitted using a biochemical model of photosynthesis [1] including the temperature response functions determined by Bernacchi et al. [42,43] and were solved for the parameters $V_{c,max}$, J_{max} and R_d . The kinetic constants for Rubisco, K_o , K_c and Γ^* in tobacco are taken from [43]. Data below the inflection point of the curve were used to solve for $V_{c,max}$ and R_d using the equation for Rubisco limited photosynthesis [1] and following the method of [39]. Data above the inflection point of the A vs. C_i curve were similarly used to solve for J_{max} using the equation for RuBP limited photosynthesis [1].

Leaf traits and final biomass

Leaf disks (ca. 1.9 cm^2) were collected from plants on August 15 during the midday gas exchange measurements.

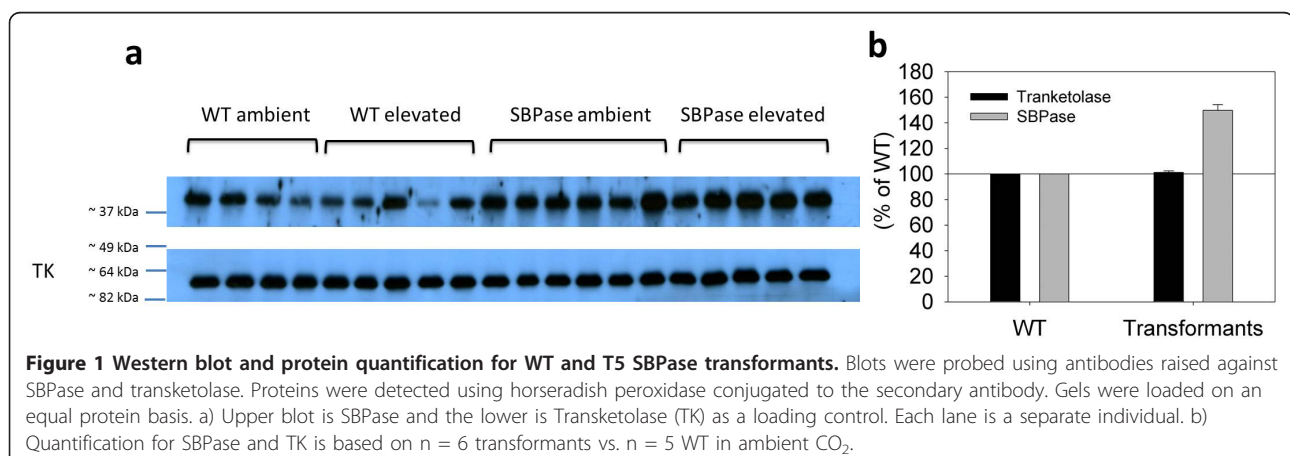
Leaf disks were sealed in pre-cooled vials, placed in coolers and disk fresh weights were determined the same afternoon. Leaf disks were dried at 60°C for 48 hours and then re-weighed. Dry and wet weights were used to determine specific leaf area (SLA) and specific leaf weight (SLW). These same disks were then ground to a fine powder and used to determine leaf carbon (C) and nitrogen (N) content by total combustion (Costech 4010, Valencia, CA, USA).

Statistical analyses were performed using SAS (Version 9.1, SAS institute, Cary, NC) and Jump (Version 4, SAS Institute, Cary NC). Trait and parameter means of SBPase transformant lines were statistically indistinguishable so the lines were pooled for subsequent ANOVAs. Simple effect tests as implemented in SAS (LSMEANS/SLICE) were used to determine if there were significant differences 1) between types within treatments (i.e., WT ambient vs. SBPase ambient) or 2) between treatments within types (i.e., SBPase ambient vs. SBPase elevated). The diurnals at SoyFACE were analyzed as a repeated measures mixed model analysis of variance (PROC MIXED,SAS). As above, SBPase lines were statistically indistinguishable during the time course and were pooled in ANOVAS. Type (SBPase or WT), CO_2 concentration [CO_2] (ambient or elevated), and time of day (time) were fixed factors. Each block contained one ambient and one elevated CO_2 plot and was considered a random factor. As there were only 4 blocks, significant probability was set at $p < 0.1$ a priori to reduce the possibility of type II errors [44,45].

Results

Protein Quantification

SBPase content was 150% (± 4.5) greater in transformants and more uniform relative to WT plants (Figure 1a and 1b). SBPase overexpressing lines did not differ from each other in terms of the SBPase protein content



(Figure 1a). Transketolase content was similar in WT and transformants (Figure 1b).

Diurnal course of gas exchange and electron transport rate

Diurnal trends of photosynthesis and fluorescence parameters were measured at their respective growth [CO₂] (i.e. 380 or 585 ppm) on July 31 and August 15, 2009 (Table 1). On July 31, photosynthetic rate (*A*) was significantly higher in transformants, due to significant differences around midday at elevated (585 ppm) [CO₂] (Figure 2a and 2b). On average, electron transport rate (ETR) (Figure 2c and 2d) was significantly higher for transformants at elevated [CO₂] (simple effect test; $F_{1,12} = 8.43$ $p < 0.05$). Differences in ETR between transformants and WT were driven by significantly lower values for WT plants at midday in elevated [CO₂] on July 31. On August 14, *A* was significantly greater at elevated CO₂ for both WT and transformants (Figure 3a and 3b, Table 1), however, there were no detectable differences in photosynthesis between WT and transformants. ETR was similar for transformants and WT plants in ambient and elevated CO₂ on August 14 (Figure 3c and 3d).

On July 31, elevating [CO₂] increased *A'* for WT and transformants ($F_{1,12} = 15.93$ $p < 0.01$). Transformants had significantly greater *A'* than WT in elevated [CO₂] ($F_{1,12} = 6.89$ $p = 0.01$), but in ambient [CO₂] they were not significantly different (compare Figure 2e and 2f).

Table 1 Repeated measures analysis of variance of diurnal variation of photosynthesis (*A*) and linear electron flux through photosystem II (ETR), for the main effects of plant type (transformants and WT), CO₂ concentration (385 ppm, 585 ppm), and time of day (time)

| 31-Jul | | Photo | | | | ETR | |
|----------------------------|---------|-------|------------------|---------|--------|------------------|--|
| | df | F | P | df | F | P | |
| type | 1, 10.4 | 10.29 | 0.009 | 1, 9.11 | 9.16 | 0.014 | |
| CO ₂ | 1, 10.4 | 28.93 | 0.0003 | 1, 9.11 | 2.04 | 0.187 | |
| type*CO ₂ | 1, 10.4 | 1.99 | 0.188 | 1, 9.11 | 1.99 | 0.191 | |
| time | 4, 73.7 | 21.83 | <.0001 | 4, 79.9 | 16.04 | <.0001 | |
| type*time | 4, 73.7 | 0.41 | 0.804 | 4, 79.9 | 0.35 | 0.846 | |
| CO ₂ *time | 4, 73.7 | 5.75 | 0.000 | 4, 79.9 | 1.58 | 0.189 | |
| type*CO ₂ *time | 4, 73.7 | 0.65 | 0.627 | 4, 79.9 | 0.71 | 0.590 | |
| 14-Aug | | Photo | | | | ETR | |
| | df | F | P | df | F | P | |
| type | 1, 12.4 | 0.98 | 0.342 | 1, 10.9 | 1.54 | 0.240 | |
| CO ₂ | 1, 12.4 | 6.58 | 0.024 | 1, 10.9 | 2.66 | 0.131 | |
| type*CO ₂ | 1, 12.4 | 0.44 | 0.521 | 1, 10.9 | 0 | 0.971 | |
| time | 4, 104 | 29.48 | <.0001 | 4, 102 | 135.52 | <.0001 | |
| type*time | 4, 104 | 0.92 | 0.453 | 4, 102 | 1.16 | 0.333 | |
| CO ₂ *time | 4, 104 | 2.73 | 0.033 | 4, 102 | 1.64 | 0.169 | |
| type*CO ₂ *time | 4, 104 | 0.4 | 0.806 | 4, 102 | 0.45 | 0.775 | |

Diurnal measurements were collected on July 31 and August 14, 2009.

On July 31, *A'* increased 14% for transformants but only 8% for WT. In contrast, on August 15, elevating [CO₂] increased *A'* by 6% for transformants but by 11% for WT ($F_{1,12} = 6.79$ $p < 0.05$). There were no detectable differences in *A'* between transformants and WT in ambient or elevated [CO₂] on August 15 (Figure 3e and 3f).

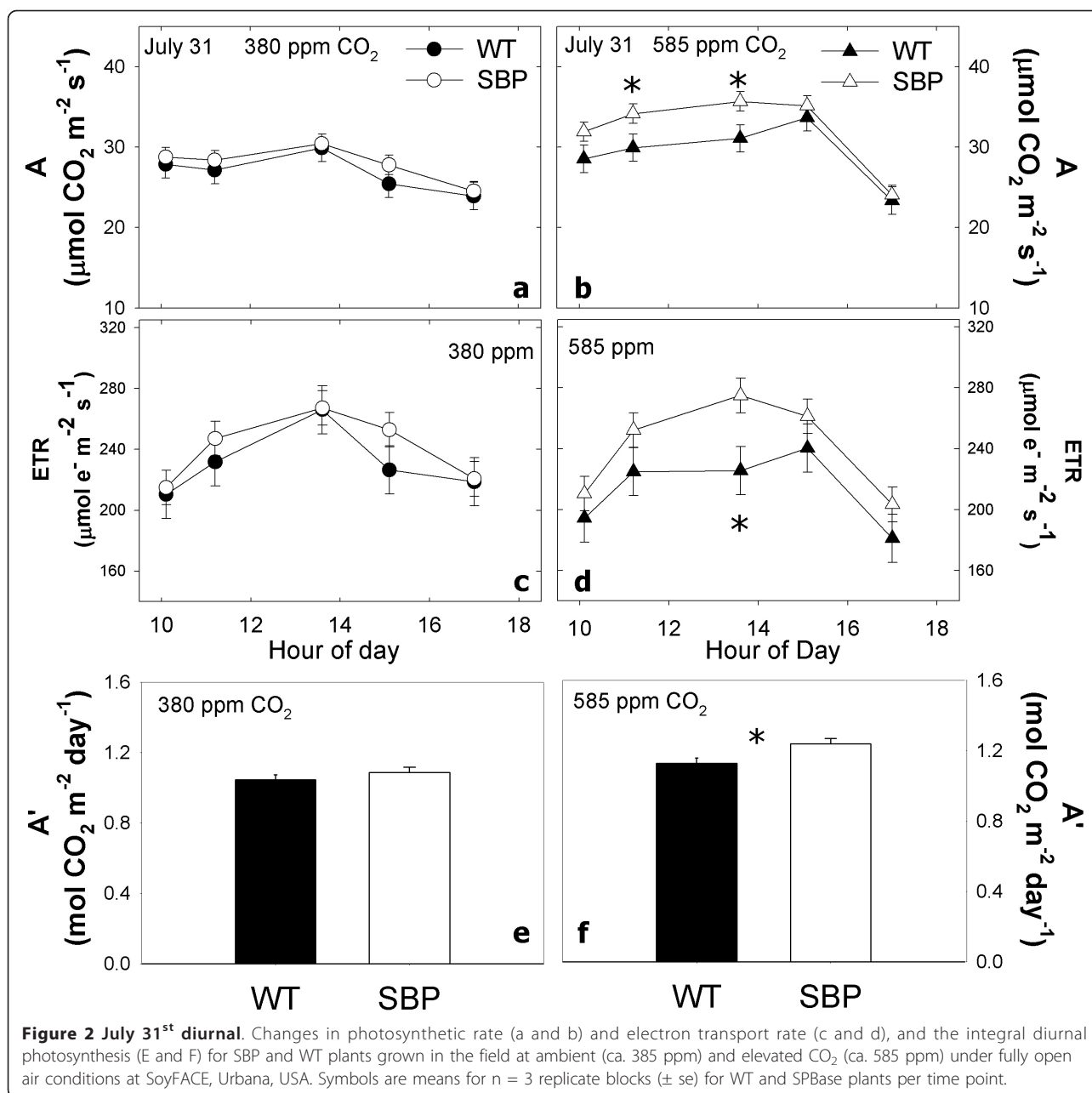
Photosynthetic biochemical parameters

A vs. *C_i* curves were measured in the field the morning following each diurnal (i.e. August 1 and August 15) under similar meteorological conditions as the diurnals. On August 1st *V_{c,max}* tended to be lower in elevated [CO₂] (130.02 ± 5.9) than in ambient [CO₂] (137.13 ± 5.7) but the trend was not significant (Table 2, Figure 4a). There was a type by [CO₂] interaction for the response of *J_{max}* (Table 2). Further analysis revealed that growth at elevated [CO₂] significantly increased *J_{max}* of transformants but not WT ($F_{1,16} = 8.24$ $p < 0.5$) (Figure 4c) on August 1. Consequently, the ratio of *V_{c,max}* to *J_{max}* (*V*/*J*) was similar between WT and transformants at ambient [CO₂]. Elevating [CO₂] significantly reduced *V*/*J* in transformants ($F_{1,14} = 15.56$ $p < 0.01$) but not in WT plants on August 1 (Figure 4e). Growth at elevated [CO₂] significantly increased respiration in the light (*R_d*, Table 2) and transformants had significantly higher *R_d* than WT in both ambient ($F_{1,14} = 7.78$ $p < 0.05$) and elevated [CO₂] ($F_{1,14} = 16.03$ $p < 0.01$) (Figure 4g) on August 1.

On August 15, both *V_{c,max}* and *J_{max}* were significantly lower for plants grown under elevated than ambient [CO₂] (Table 2; Figure 4b and 4d). Transformants had significantly greater *J_{max}* than WT at ambient [CO₂] but not in elevated [CO₂] ($F_{1,20} = 3.87$ $p = 0.06$). Elevating [CO₂] significantly decreased *V*/*J* in transformants and WT (Table 2 Figure 4f). Elevating [CO₂] significantly increased *R_d* for WT and transformants (Figure 4h).

Leaf traits and final biomass

Specific leaf area (SLA) was significantly lower at elevated [CO₂] compared to ambient, and transformants had significantly lower SLA than WT plants (Table 3, Figure 5a). Further analysis revealed that transformant SLA was lower than WT SLA in elevated [CO₂] ($F_{1,15} = 8.75$ $p < 0.01$). Elevating [CO₂] significantly decreased leaf nitrogen content (%N); consequently, the carbon to nitrogen ratio (C:N) of leaves increased significantly in elevated [CO₂] (Table 3, Figure 5b and 5c). Transformant C:N increased more than WT ($F_{1,15} = 9.46$ $p = 0.01$). Above ground biomass (= yield in kg/Ha) was greater for plants grown in elevated [CO₂] and transformant biomass was greater than WT plants (Table 3). Biomass increased more for transformants than WT following growth in elevated [CO₂] (22% vs. 13%) (Figure 5d; $F_{1,15} = 6.37$ $p < 0.05$).



Discussion

The goal of our experiments was to test the hypothesis that tobacco plants transformed to over express the PCR cycle enzyme SBPase would exhibit greater stimulation of carbon assimilation than WT plants when grown at elevated [CO₂] under field conditions [e.g., [17,30,31]].

Transformant biomass increases more than WT at elevated [CO₂]

When grown under fully open air CO₂ fumigation, SBPase overexpressing plants displayed up to 14%

greater light saturated photosynthetic rates (A) and up to 21% more linear electron flux through PSII (ETR) than WT plants. Moreover, after 12 weeks of growth at elevated [CO₂], harvested biomass increased by 13% in WT plants and more than 22% in transformants when compared to plants grown in ambient [CO₂]. In a prior experiment, the same transformants grown in a greenhouse under prevailing light conditions at ambient [CO₂] (ca. 375 ppm) accumulated 12% more biomass than WT plants (Lefebvre et al. 2005)[16]. Here, at ambient [CO₂] (ca. 385 ppm) under field conditions, transformants also yielded 12% more biomass than WT

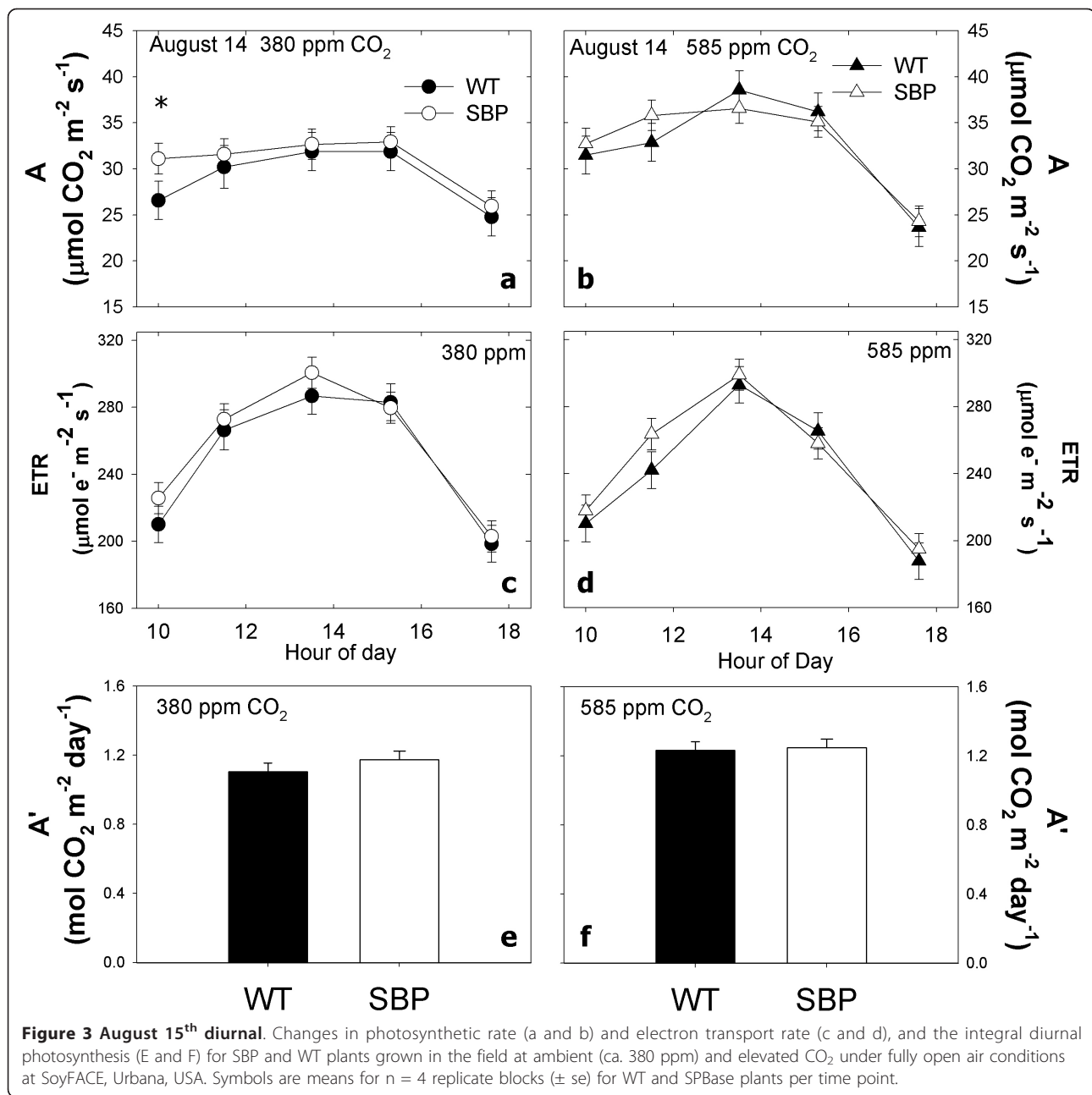


Table 2 ANOVA of photosynthetic parameters $V_{c,max}$ @ 25, potential electron transport rate J_{max} @ 25, $V_{c,max}$ @ 25/ J_{max} @ 25 (V/J), day respiration (R_d), for WT and transformants (Type) at ambient and elevated [CO₂]

| 1-Aug | | V _{c,max} | | | J _{max} | | | V/J | | | R _d | |
|----------------------|---------|--------------------|------------------|-------|------------------|---------------|-------|-------|------------------|-------|----------------|---------------|
| | Df | F | p | df | F | p | df | F | p | df | F | p |
| type | 1, 14.2 | 0.03 | 0.8661 | 1, 16 | 2.58 | 0.1276 | 1, 14 | 1.55 | 0.2329 | 1, 14 | 23.22 | 0.0003 |
| CO ₂ | 1, 14.2 | 0.76 | 0.3979 | 1, 16 | 2.44 | 0.1381 | 1, 14 | 5.86 | 0.0296 | 1, 14 | 17.87 | 0.0008 |
| type*CO ₂ | 1, 14.2 | 0.1 | 0.7524 | 1, 16 | 6.79 | 0.0191 | 1, 14 | 2.81 | 0.116 | 1, 14 | 0.9 | 0.3592 |
| 15-Aug | | V _{c,max} | | | J _{max} | | | V/J | | | R _d | |
| | Df | F | p | df | F | p | df | F | p | df | F | p |
| type | 1, 20 | 2.4 | 0.1371 | 1, 20 | 2.57 | 0.1243 | 1, 20 | 0 | 0.9702 | 1, 20 | 0.03 | 0.8753 |
| CO ₂ | 1, 20 | 73.72 | <.0001 | 1, 20 | 18.18 | 0.0004 | 1, 20 | 40.21 | <.0001 | 1, 20 | 14.98 | 0.001 |
| type*CO ₂ | 1, 20 | 0.3 | 0.5925 | 1, 20 | 1.38 | 0.2531 | 1, 20 | 0.87 | 0.3608 | 1, 20 | 2.5 | 0.1293 |

Parameters were derived from A vs [CO₂] curves measured in the field see methods for details. Only three blocks could be measured on August 1.

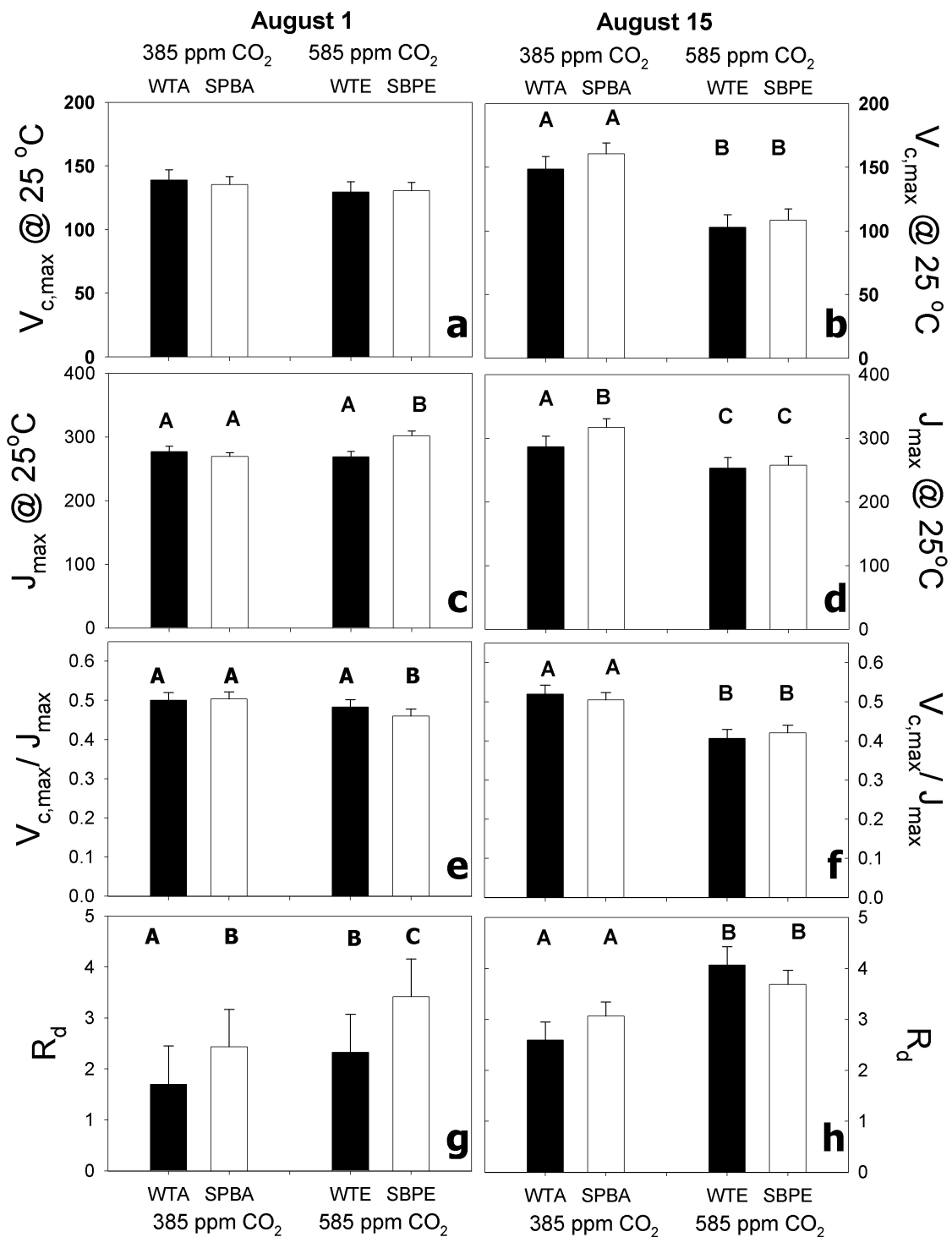


Figure 4 Photosynthetic parameters derived from response of A to [CO₂] using a biochemical model of photosynthesis (see methods). Each day (August 1 and August 15) was analyzed separately with a mixed model ANOVA. Line 11 and line 30 differed only for V/J on aug 1st (*) and were pooled for all other analyses and post hoc tests. Bars are means (± se) (August 1 n = 3) (August 15 n = 4). Bars with different capital letters are significantly different see results for specific p values.

Table 3 Analysis of variance of the effects of [CO₂] and plant type (WT vs. Transformant) on specific leaf area (SLA), leaf nitrogen content (%N), leaf carbon to nitrogen ration (C:N) and final biomass (Kg/ha) for n = 3 blocks

| | df | SLA | | %N | | C:N | | Biomass | |
|----------------------|------|-------|---------------|-------|------------------|-------|---------------|---------|---------------|
| | | F | p | F | p | F | p | F | p |
| type | 1,15 | 6.57 | 0.0217 | 1.22 | 0.2875 | 3.9 | 0.0671 | 4.05 | 0.0625 |
| CO ₂ | 1,15 | 16.69 | 0.001 | 29.65 | <.0001 | 17.36 | 0.0008 | 5.03 | 0.0404 |
| type*CO ₂ | 1,15 | 2.63 | 0.1257 | 3.52 | 0.0809 | 5.65 | 0.0312 | 0.45 | 0.5121 |

plants (see Figure 5) consistent with the Lefebvre et al (2005)[16] greenhouse study. Taken together, these results support our hypothesis and clearly show the benefit of overexpressing SBPase in field grown plants at both current and future levels of atmospheric [CO₂].

WT biomass was 13% greater in elevated [CO₂] when compared to ambient grown WT plants, which is somewhat lower than the average increase in biomass for C₃ crops in FACE experiments [i.e. 19.8% in [46]]. Growth at elevated [CO₂] alters plant insect interaction and increases palatability of crops [47-50]; thus it is possible that yield stimulations were slightly lower because of aphid and hornworm herbivory (pers obs). In tobacco in particular, aphid infestation significantly reduced the stimulatory effect of [CO₂] on biomass [51]. Nevertheless, transformant biomass increased more than WT at elevated [CO₂] (22.7%) and more than the average for C₃ crops in FACE experiments.

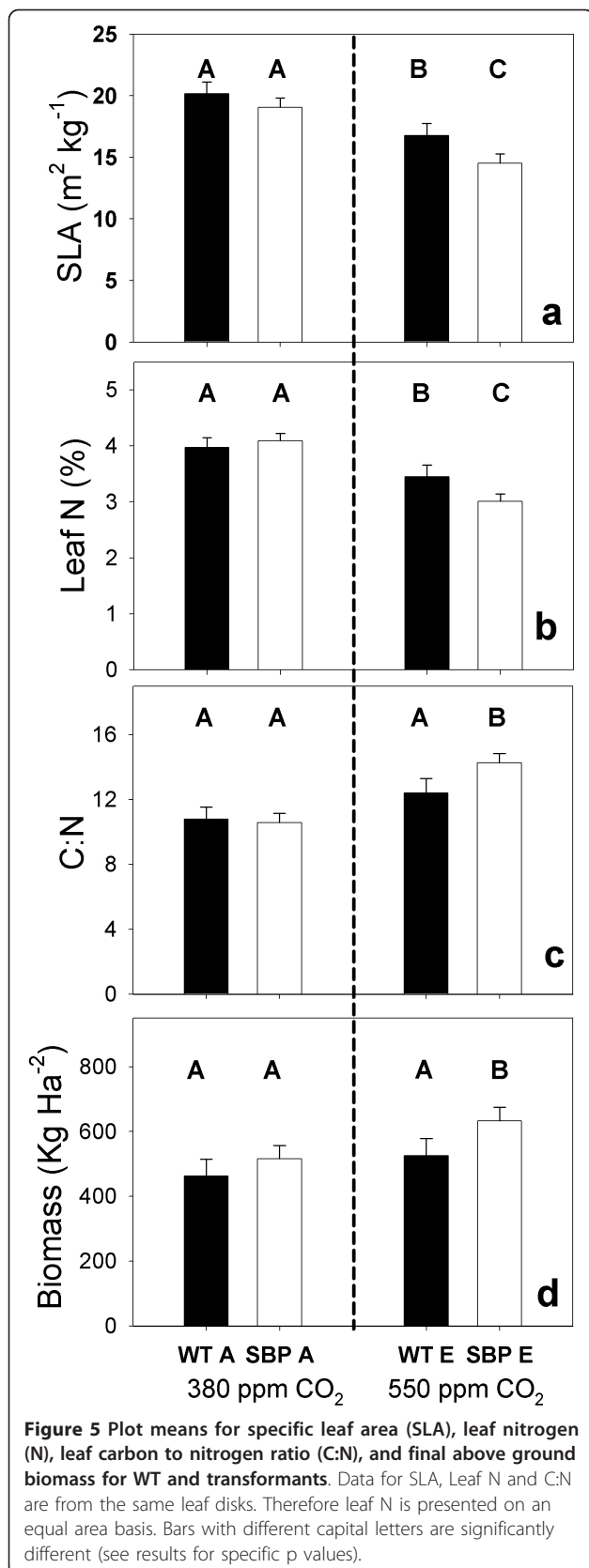
Lefebvre *et al.* (2005)[16] reported that the greatest differences between transformants and WT photosynthetic rates occurred prior to flowering in greenhouse plants and during early development in chamber grown plants. The differences between young expanding and fully expanded leaves could not be accounted for by differential SBPase activity (Lefebvre *et al.* 2005). We show that in ambient and elevated [CO₂] plots, carbon uptake was enhanced more for transformants during the vegetative phase (i.e. July 31) than when plants were starting to flower (August 15). When plants were beginning to flower, differences between transformants and WT were no longer detectable, yet carbon uptake was consistently stimulated for plants growing in elevated [CO₂]. Ultimately, even though the realized increase in *A* and *A'* between WT and transformants falls well short of the theoretical 40% increase in assimilation predicted if plants were to reallocate 15% of photosynthetic resources from Rubisco to RuBP regeneration [e.g., [7]], increases in the carbon uptake of transformants early in growth and prior to flowering were sufficiently large to increase final biomass.

Several studies demonstrate that changing expression and activity level of SBPase directly impacts carbon assimilation, growth, and biomass accumulation in tobacco growing at current ambient [CO₂] (ca. 385 ppm) [16,19,52-55]. While the positive relationship

between SBPase activity and carbon assimilation was clearly shown in WT and transformants [16,19], overexpression of SBPase in rice and tobacco has not always increased biomass for plants grown at ambient [CO₂] levels in controlled environments [16,33,34]. For instance, Lefebvre et al. noted that no increase in photosynthesis or plant yield was evident for tobacco transformants grown in winter when days were shorter and light levels were lower[16] (S. Lefebvre, J.C. Lloyd, and C. Raines unpublished data). The observations of Lefebvre et al. [16] and this study are also consistent with the notion that SBPase exerts control over CO₂ fixation under light saturating conditions. By definition, the amount of SBPase would not affect the light limited rate of photosynthesis which depends on the rate of production of NADPH and ATP on the photosynthetic membrane. Our diurnal measurements are consistent with these expectations, as transformants with increased SBPase activity showed the greatest increases in carbon assimilation relative to wild type plants around midday when light levels were highest. In contrast, there was no difference in assimilation rates between the SBPase overexpressing and wild type plants at the beginning or end of the day (Figure 2).

Acclimation to [CO₂] increases nutrient use efficiency more for transformants than WT

Both WT and transformants showed evidence of a similar decrease in *V_{c,max}* after a month of growth at elevated [CO₂], indicating photosynthetic acclimation via down regulation of in vivo Rubisco capacity. Photosynthetic acclimation to growth in elevated [CO₂] is presumed to be a biochemical adjustment to optimize nitrogen use [6]. As [CO₂] increases so does the catalytic rate of Rubisco, therefore less N needs to be invested in Rubisco to fix carbon. Reallocation of N is then, for instance, available to upregulate respiratory metabolism in response to growth at elevated [CO₂] [56]. SBPase represents less than 1% of the N contained in the enzymes of photosynthetic carbon metabolism [21]. It is therefore remarkable that ca. 50% increase in the amount of this protein in transformants results in detectable increases in CO₂ assimilation. The relatively large increase in CO₂ assimilation at elevated [CO₂] was associated with a significant decrease in leaf N per unit



mass (Figure 5). Thus for a small increase in protein, transformants had a significantly greater increases in nitrogen use efficiency than WT at elevated [CO₂]. The results are consistent with numerous other FACE studies showing that [CO₂] will stimulate growth in spite of photosynthetic acclimation and that growth at elevated [CO₂] increases nitrogen use efficiency [reviewed in [57]].

Transformants and WT plants grown in elevated [CO₂] tended to have higher respiration in the light (R_d) than plants in ambient [CO₂] plots. Leaves of plants grown under elevated [CO₂] accumulate larger concentrations of non-structural carbohydrates (i.e. sugar and starch) [46], and this may underlie higher respiration [58]. Recently, Leakey et al. [56] demonstrated that the acclimation response of respiration to elevated [CO₂] was mediated via transcriptional upregulation of respiratory enzymes. We speculate that the reportedly greater sucrose and starch accumulation in transformants [16] stimulates additional acclimation of respiration to elevated [CO₂] and may therefore also diminish the benefit of overexpressing SBPase. Alternatively, higher R_d in transformants may be a result of the unregulated overexpression of the enzyme. Either way, higher R_d, the requirement for high light, and unmeasured natural stresses all would contribute to a lower realized benefit to overexpressing SBPase in the field.

Conclusion

The data presented in this paper have demonstrated that transgenic tobacco plants with increased SBPase have the potential for greater stimulation of photosynthesis and biomass production relative to wild type tobacco when grown at elevated [CO₂]. Differences between theoretical and realized increases in carbon assimilation are to be expected as studies of PCR cycle antisense plants have demonstrated that the relative importance of any one PCR cycle enzyme is not fixed and will vary according to environmental and developmental conditions [[20], this study, [59]]. Nevertheless, our findings are consistent with the notion that elevating [CO₂] increases the metabolic control of RuBP-regeneration and decreases the control exerted by Rubisco at light saturation [6,7]. Though smaller than theoretically predicted, the increases in photosynthetic stimulation at elevated [CO₂] demonstrated here are indicative that C₃ crop plants can be engineered to meet a rapidly changing environment.

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Authors' contributions

DR Conceived and designed the experiment, acquired and analyzed the data, and wrote the paper. AL aided in data acquisition and analysis, revised the paper, and gave final approval of the manuscript. MK aided in data acquisition, data analysis and gave final approval of the manuscript. CR provided the transformants, provided technical support, revised the paper, and gave final approval of the manuscript. SL and DO conceived and aided in the design of the experiment, revised the manuscript, and gave final approval of the manuscript.

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References

1. Farquhar GD, von Caemmerer S, Berry JA: **A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species.** *Planta* 1980, **149**(1):78-90.
2. Sharkey TD: **O₂ insensitive photosynthesis in C₃ plants -it's occurrence and a possible explanation.** *Plant Physiology* 1985, **78**(1):71-75.
3. Cavagnaro TR, Gleadow RM, Miller RE: **Plant nutrient acquisition and utilisation in a high carbon dioxide world.** *Functional Plant Biology* 2011, **38**(2):87-96.
4. IPCC: **Summary for Policy Makers.** In *Climate Change 2007: The Physical Science Basis Contribution of Working Group 1 to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Edited by: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL. Cambridge, UK and New York, NY: Cambridge University Press; 2007:.
5. Le Quere C, Raupach MR, Canadell JG, Marland G, Bopp L, Ciais P, Conway TJ, Doney SC, Feely RA, Foster P, et al: **Trends in the sources and sinks of carbon dioxide.** *Nature Geoscience* 2009, **2**(12):831-836.
6. Drake BG, Gonzalez-Meler MA, Long SP: **More efficient plants: A consequence of rising atmospheric CO₂?** *Annual Review of Plant Physiology and Plant Molecular Biology* 1997, **48**:609-639.
7. Long SP, Ainsworth EA, Rogers A, Ort DR: **Rising atmospheric carbon dioxide: Plants face the future.** *Annual Review of Plant Biology* 2004, **55**:591-628.
8. Long SP, Ainsworth EA, Leakey ADB, Nosberger J, Ort DR: **Food for thought: Lower-than-expected crop yield stimulation with rising CO₂ concentrations.** *Science* 2006, **312**(5782):1918-1921.
9. Zhu XG, Long SP, Ort DR: **What is the maximum efficiency with which photosynthesis can convert solar energy into biomass?** *Current Opinion in Biotechnology* 2008, **19**(2):153-159.
10. Long SP, Zhu XG, Naidu SL, Ort DR: **Can improvement in photosynthesis increase crop yields?** *Plant Cell and Environment* 2006, **29**(3):315-330.
11. Long SP, Ort DR: **More than taking the heat: crops and global change.** *Current Opinion in Plant Biology* 2010, **13**(3):241-248.
12. Zhu XG, Long SP, Ort DR: **Improving Photosynthetic Efficiency for Greater Yield.** *Annual Review of Plant Biology* 2010, **61**:235-261.
13. Bruinsma J, (ed): **World agriculture: towards 2015/2030: an FAO perspective.** London: Earthscan; 2003.
14. Lobell DB, Cassman KG, Field CB: **Crop Yield Gaps: Their Importance, Magnitudes, and Causes.** *Annual Review of Environment and Resources* 2009, **34**:179-204.
15. Bernacchi CJ, Morgan PB, Ort DR, Long SP: **The growth of soybean under free air CO₂ enrichment (FACE) stimulates photosynthesis while decreasing in vivo Rubisco capacity.** *Planta* 2005, **220**(3):434-446.
16. Lefebvre S, Lawson T, Zakhleniuk OV, Lloyd JC, Raines CA: **Increased sedoheptulose-1,7-bisphosphatase activity in transgenic tobacco plants stimulates photosynthesis and growth from an early stage in development.** (vol 138, pg 451, 2005). *Plant Physiology* 2005, **138**(2):1174-1174.
17. Ainsworth EA, Ort DR: **How do we improve crop production in a warming world?** *Plant Physiology* 2010, **154**(2):526-530.
18. Yamori W, Takahashi S, Makino A, Price GD, Badger MR, von Caemmerer S: **The Roles of ATP Synthase and the Cytochrome b(6)/f Complexes in Limiting Chloroplast Electron Transport and Determining Photosynthetic Capacity.** *Plant Physiology* 2010, **155**(2):956-962.
19. Harrison EP, Willingham NM, Lloyd JC, Raines CA: **Reduced sedoheptulose-1,7-bisphosphatase levels in transgenic tobacco lead to decreased photosynthetic capacity and altered carbohydrate accumulation.** *Planta* 1998, **204**(1):27-36.
20. Raines CA: **The Calvin cycle revisited.** *Photosynthesis Research* 2003, **75**(1):1-10.
21. Zhu XG, de Sturler E, Long SP: **Optimizing the distribution of resources between enzymes of carbon metabolism can dramatically increase photosynthetic rate: A numerical simulation using an evolutionary algorithm.** *Plant Physiology* 2007, **145**(2):513-526.
22. Poolman MG, Fell DA, Thomas S: **Modelling photosynthesis and its control.** *Journal of Experimental Botany* 2000, **51**:319-328.
23. Quick WP, Schurr U, Scheibe R, Schulze ED, Rodermerl SR, Bogorad L, Stitt M: **Decreased Ribulose-1,5-Bisphosphate Carboxylase-Oxygenase in transgenic tobacco transformed with antisense rbcS. 1: impact on photosynthesis in ambient growth conditions.** *Planta* 1991, **183**(4):542-554.
24. Spreitzer RJ, Salvucci ME: **Rubisco: Structure, regulatory interactions, and possibilities for a better enzyme.** *Annual Review of Plant Biology* 2002, **53**:449-475.
25. Suzuki Y, Miyamoto T, Yoshizawa R, Mae T, Makino A: **Rubisco content and photosynthesis of leaves at different positions in transgenic rice with an overexpression of RBCS.** *Plant Cell and Environment* 2009, **32**(4):417-427.
26. Makino A, Sage RF: **Temperature response of photosynthesis in transgenic rice transformed with 'sense' or 'antisense' rbcS.** *Plant and Cell Physiology* 2007, **48**(10):1472-1483.
27. Zhu XG, Portis AR, Long SP: **Would transformation of C-3 crop plants with foreign Rubisco increase productivity? A computational analysis extrapolating from kinetic properties to canopy photosynthesis.** *Plant Cell and Environment* 2004, **27**(2):155-165.
28. Roy H, Andrews TJ: **Rubisco Assembly and Metabolism.** In *Photosynthesis Physiology and Metabolism. Volume 9*. Edited by: Leegood RC, Sharkey TD, von Caemmerer S. Dordrecht: Kluwer; 2000:53-83.
29. Whitney SM, Sharwood RE: **Construction of a tobacco master line to improve Rubisco engineering in chloroplasts.** *Journal of Experimental Botany* 2008, **59**(7):1909-1921.
30. von Caemmerer S, Evans JR: **Enhancing C₃ Photosynthesis.** *Plant Physiology* 2010, **154**(2):589-592.
31. Raines CA: **Transgenic approaches to manipulate the environmental responses of the C₃ carbon fixation cycle.** *Plant Cell and Environment* 2006, **29**(3):331-339.
32. Martin W, Scheibe R, Schnarrenberger C: **The Calvin Cycle and Its Regulation.** In *Photosynthesis: Physiology and Metabolism*. Edited by: Leegood RC, Sharkey TD, von Caemmerer S. Boston: Kluwer Academic Publishers; 2000:9-51.
33. Feng LL, Han YJ, Liu G, An BG, Yang J, Yang GH, Li YS, Zhu YG: **Overexpression of sedoheptulose-1,7-bisphosphatase enhances photosynthesis and growth under salt stress in transgenic rice plants.** *Functional Plant Biology* 2007, **34**(9):822-834.
34. Feng LL, Wang K, Li Y, Tan YP, Kong J, Li H, Li YS, Zhu YG: **Overexpression of SBPase enhances photosynthesis against high temperature stress in transgenic rice plants.** *Plant Cell Reports* 2007, **26**(9):1635-1646.
35. von Caemmerer S: **Biochemical Models of Leaf Photosynthesis.** Collingwood, Victoria, Australia: CSIRO Publishing; 2000.
36. Morgan PB, Bollero GA, Nelson RL, Dohleman FG, Long SP: **Smaller than predicted increase in aboveground net primary production and yield of**

- field-grown soybean under fully open-air [CO₂] elevation. *Global Change Biology* 2005, **11**(10):1856-1865.
37. Miglietta F, Peressotti A, Vaccari FP, Zaldei A, deAngelis P, Scarascia-Mugnozza G: Free-air CO₂ enrichment (FACE) of a poplar plantation: the POPFACE fumigation system. *New Phytologist* 2001, **150**(2):465-476.
38. Bernacchi CJ, Leakey ADB, Heady LE, Morgan PB, Dohleman FG, McGrath JM, Gillespie KM, Wittig VE, Rogers A, Long SP, et al: Hourly and seasonal variation in photosynthesis and stomatal conductance of soybean grown at future CO₂ and ozone concentrations for 3 years under fully open-air field conditions. *Plant Cell and Environment* 2006, **29**(11):2077-2090.
39. Long SP, Bernacchi CJ: Gas exchange measurements, what can they tell us about the underlying limitations to photosynthesis? Procedures and sources of error. *Journal of Experimental Botany* 2003, **54**(392):2393-2401.
40. Baker NR: Chlorophyll fluorescence: A probe of photosynthesis in vivo. *Annual Review of Plant Biology* 2008, **59**:89-113.
41. Genty B, Briantais JM, Baker NR: The relationship between quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica Et Biophysica Acta* 1989, **990**(1):87-92.
42. Bernacchi CJ, Pimentel C, Long SP: In vivo temperature response functions of parameters required to model RuBP-limited photosynthesis. *Plant Cell and Environment* 2003, **26**(9):1419-1430.
43. Bernacchi CJ, Singsaas EL, Pimentel C, Portis AR, Long SP: Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant Cell and Environment* 2001, **24**(2):253-259.
44. Scheiner SM: Theories, Hypotheses and Statistics. In *Design and Analysis of Ecological Experiments*. Edited by: Scheiner SM, Gurevitch J. New York: Oxford University Press; 2001:.
45. Sokal RR, Rohlf FJ: *Estimation and Hypothesis Testing*. *Biometry*. 3 edition. New York: W.H. Freeman; 1995, 127-175.
46. Ainsworth EA, Long SP: What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. *New Phytologist* 2005, **165**(2):351-371.
47. Casteel CL, O'Neill BF, Zavala JA, Bilgin DD, Berenbaum MR, DeLucia EH: Transcriptional profiling reveals elevated CO₂ and elevated O₃ alter resistance of soybean (*Glycine max*) to Japanese beetles (*Popillia japonica*). *Plant Cell and Environment* 2008, **31**(4):419-434.
48. Dermody O, O'Neill BF, Zangerl AR, Berenbaum MR, DeLucia EH: Effects of elevated CO₂ and O₃ on leaf damage and insect abundance in a soybean agroecosystem. *Arthropod-Plant Interactions* 2008, **2**(3):125-135.
49. Lau JA, Tiffin P: Elevated carbon dioxide concentrations indirectly affect plant fitness by altering plant tolerance to herbivory. *Oecologia* 2009, **161**(2):401-410.
50. Zavala JA, Casteel CL, DeLucia EH, Berenbaum MR: Anthropogenic increase in carbon dioxide compromises plant defense against invasive insects. *Proceedings of the National Academy of Sciences of the United States of America* 2008, **105**(13):5129-5133.
51. Fu X, Ye LF, Kang L, Ge F: Elevated CO₂ shifts the focus of tobacco plant defences from cucumber mosaic virus to the green peach aphid. *Plant Cell and Environment* 2010, **33**(12):2056-2064.
52. Olcer H, Lloyd JC, Raines CA: Photosynthetic capacity is differentially affected by reductions in sedoheptulose-1,7-bisphosphatase activity during leaf development in transgenic tobacco plants. *Plant Physiology* 2001, **125**(2):982-989.
53. Harrison EP, Olcer H, Lloyd JC, Long SP, Raines CA: Small decreases in SBPase cause a linear decline in the apparent RuBP regeneration rate, but do not affect Rubisco carboxylation capacity. *Journal of Experimental Botany* 2001, **52**(362):1779-1784.
54. Lawson T, Bryant B, Lefebvre S, Lloyd JC, Raines CA: Decreased SBPase activity alters growth and development in transgenic tobacco plants. *Plant Cell and Environment* 2006, **29**(1):48-58.
55. Miyagawa Y, Tamoi M, Shigeoka S: Overexpression of a cyanobacterial fructose-1,6-/sedoheptulose-1,7-bisphosphatase in tobacco enhances photosynthesis and growth. *Nature Biotechnology* 2001, **19**(10):965-969.
56. Leakey ADB, Xu F, Gillespie KM, McGrath JM, Ainsworth EA, Ort DR: Genomic basis for stimulated respiration by plants growing under elevated carbon dioxide. *Proceedings of the National Academy of Sciences of the United States of America* 2009, **106**(9):3597-3602.
57. Leakey ADB, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP, Ort DR: Elevated CO₂ effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. *Journal of Experimental Botany* 2009, **60**(10):2859-2876.
58. Davey PA, Hunt S, Hymus GJ, DeLucia EH, Drake BG, Karnosky DF, Long SP: Respiratory oxygen uptake is not decreased by an instantaneous elevation of [CO₂], but is increased with long-term growth in the field at elevated [CO₂]. *Plant Physiology* 2004, **134**(1):520-527.
59. Stitt M, Schulze D: Does Rubisco control the rate of photosynthesis and plant growth: an exercise in molecular ecophysiology. *Plant Cell and Environment* 1994, **17**(5):465-487.

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