

Microbial growth yield as a new parameter in environmental chemistry and risk assessment

Brock, Andreas Libonati; Kästner, Matthias; Trapp, Stefan

Publication date:
2017

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):

Brock, A. L., Kästner, M., & Trapp, S. (2017). Microbial growth yield as a new parameter in environmental chemistry and risk assessment. Abstract from 14th International Symposium on Persistent Toxic Substances, Nagoya, Japan.

DTU Library

Technical Information Center of Denmark

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Microbial growth yield as a new parameter in environmental chemistry and risk assessment

Andreas Libonati Brock¹, Matthias Kästner², Stefan Trapp¹

¹ DTU Environment, Technical University of Denmark, Kgs. Lyngby, Denmark

² UFZ—Helmholtz-Centre for Environmental Research, Department of Environmental Biotechnology, Leipzig, Germany

Chemicals traded on the European market subject to a chemical safety assessment are assessed for their persistency, bioaccumulation and toxicity (PVB/vPvB) [1]. If the degradation half-life in soil is more than 120 days the chemical is labelled persistent. In soil incubation experiments non-extractable residues (NER) are often formed. NER are defined as residues that are not extractable using extraction methods that do not alter the matrix [2]. Kästner *et al.* [3] differentiated between three types of NER: sequestered NER (type I); covalently bound NER (type II); and biogenic NER (type III). Only type I and II can be considered NER according to the IUPAC definition [2]. A considerable fraction of NER can be of biogenic origin (e.g. [4]). Despite this, ECETOC [5] recommends that NER is considered as a 'separate component' in risk assessment; however, analytical methods are needed to differentiate biogenic NER from other types of NER and currently not part of standard procedures. In REACH [6] incorporation of labelled carbon into biomass should be considered as a potential removal pathway. Consequently, a method to distinguish biogenic NER from type I and II NER is needed. Biogenic NER is the sum of microbial biomass X formed by degradation and its decay products (soil organic matter SOM). The biogenic NER is thus related to the microbial growth yield which is defined as the mass of microorganisms formed per mass of substrate consumed [7,8]:

$$[X_{biogenic\ NER}] = \frac{Y}{1-Y} [CO_2] \quad (1)$$

$$[SOM_{biogenic\ NER}] = \frac{f \times Y}{(1-Y) + (1-f) \times Y} [CO_2] \quad (2)$$

Eqs. 1 and 2 quantify the upper ($X_{biogenic\ NER}$) and lower ($SOM_{biogenic\ NER}$) boundaries of biogenic NER formed due to microbial growth, decay and incorporation into SOM. Since the CO₂ released during degradation is measured, estimation of the yield enables us to elucidate the nature of NER. Recently, we developed a yield estimation method based on the molecule's nutritional value. The method is based on the Gibbs free energy released from mineralisation, the Nernst equation, and structural information [7]. The advantage is that it can be used to all kind of molecules, including xenobiotics and pesticides, without knowledge about microbial metabolic pathways. Using this method, typical carbon yields of xenobiotics range from 0 (chlorobenzenes) to 0.57 g cell carbon (g substrate carbon)⁻¹ (synthetic plant hormones) under aerobic conditions. Under nitrate-reducing conditions, yields are somewhat lower, while under sulfate-reducing conditions the range is from 0 to 0.24 g cell carbon (g substrate carbon)⁻¹. Bacterial growth, and thus high yield, is a prerequisite for the rapid degradation unless other growth substrates can be used [8], which is often the case in co-metabolic degradation processes. Using Eq. (1) and (2) we found that for many pesticides a considerable part of the NER formed in degradation experiment is likely biogenic [9]. The approach outlined provides a method to quantify the biogenic NER fraction of NER observed in soil degradation studies. Beyond NER estimation, the method enables identification of compounds which are non-stable, and allows closing the carbon cycle in turnover models.

References

- [1] REACH (2014) Guidance on Information Requirements and Chemical Safety Assessment, Chapter R.11: PBT/vPvB assessment, European Chemical Agency, Helsinki, Finland.
 - [2] Roberts, T. R. (1984) IUPAC Reports on Pesticides: Non-extractable pesticide residues in soils and plants, *Pure Appl. Chem.*, 56 (7), 945-956.
 - [3] Kästner, M., Nowak, K. M., Miltner, A., Trapp, S., & Schäffer, A. (2014) Classification and Modelling of Nonextractable Residue (NER) Formation of Xenobiotics in Soil – A Synthesis, *Crit. Rev. Env. Sci. Technol.*, 44, 2107–2171.
 - [4] Nowak, K. M., Miltner, A., Gehre, M., Schäffer, A., & Kästner, M. (2011) Formation and fate of bound residues from microbial biomass during 2, 4-D degradation in soil, *Environ. Sci. Technol.*, 45(3), 999–1006.
 - [5] ECETOC (2013). Development of interim guidance for the inclusion of non-extractable residues (NER) in the risk assessment of chemicals, Technical Report No. 118, Brussels, Belgium.
 - [6] REACH (2016) Guidance on Information Requirements and Chemical Safety Assessment, Chapter R.7b: Endpoint specific guidance, European Chemical Agency, Helsinki, Finland.
 - [7] Trapp S., Brock, A. L., Nowak, K. M. & Kästner, M. (*in revision*) Prediction of the formation of biogenic non-extractable residues during degradation of environmental chemicals from biomass yields, *Environ. Sci. Technol.*
 - [8] Kovarova-Kovar, K., & Egli, T. (1998) Growth Kinetics of Suspended Microbial Cells: From Single-Substrate-Controlled Growth to Mixed-Substrate Kinetics. *Microbiol. Mol. Biol. Rev.*, 62 (3), 646–666.
 - [9] Brock, A. L., Kästner, M. & Trapp, S. (*submitted*) Microbial growth yield estimates from thermodynamics and its importance for degradation of pesticides and formation of biogenic non-extractable residues, *SAR QSAR Environ. Res.*
-