



University of Dundee

# The Interaction of Hypotaurine and Other Sulfinates with Reactive Oxygen and **Nitrogen Species**

Baseggio Conrado, Alessia; D'Angelantonio, Mila; D'Erme, Maria; Pecci, Laura; Fontana, Mario

Published in: Taurine 10

DOI: 10.1007/978-94-024-1079-2\_45

Publication date: 2017

Document Version Peer reviewed version

Link to publication in Discovery Research Portal

Citation for published version (APA):

Baseggio Conrado, A., D'Angelantónio, M., D'Erme, M., Pecci, L., & Fontana, M. (2017). The Interaction of Hypotaurine and Other Sulfinates with Reactive Oxygen and Nitrogen Species: A Survey of Reaction Mechanisms. In D-H. Lee, S. W. Schaffer, E. Park, & H. W. Kim (Eds.), Taurine 10 (Vol. 975, pp. 573-583). (Advances in Experimental Medicine and Biology ; Vol. 975). Dordrecht: Springer . https://doi.org/10.1007/978-94-024-1079-2 45

#### **General rights**

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.

You may not further distribute the material or use if for any profit-making activity or commercial gain.
You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# The Interaction of Hypotaurine and Other Sulfinates with Reactive Oxygen and Nitrogen Species: A Survey of Reaction Mechanisms

# Alessia BASEGGIO CONRADO<sup>1,2</sup>, Mila D'ANGELANTONIO<sup>3</sup>, Maria D'ERME<sup>1</sup>, Laura PECCI<sup>1</sup>, Mario FONTANA<sup>1\*</sup>

<sup>1</sup>Dipartimento di Scienze Biochimiche, Sapienza Università di Roma –Rome, Italy.

<sup>2</sup>*Photobiology Unit, University of Dundee, Ninewells Hospital & Medical School – UK.* 

<sup>3</sup>ISOF – Istituto per la Sintesi Organica e la Fotoreattività CNR, Bologna – Italy

\*Corresponding author: mario.fontana@uniroma1

Abstract Considerable strides have been made in understanding the oxidative mechanisms involved in the final steps of the cysteine pathway leading to taurine. The oxidation of sulfinates, hypotaurine and cysteine sulfinic acid, to the respective sulfonates, taurine and cysteic acid, has never been associated with any specific enzyme. Conversely, there is strong evidence that in vivo formation of taurine and cysteic acid is the result of sulfinate interaction with a variety of biologically relevant oxidants. In the last decade, many experiments have been performed to understand whether peroxynitrite, nitrogen dioxide and carbonate radical anion could be included in the biologically relevant reactive species capable of oxidizing sulfinates. Thanks to this work, it has been possible to highlight two possible reaction mechanisms (direct and indirect reaction) of sulfinates with reactive oxygen and nitrogen species.

The sulfinates oxidation, mediated by peroxynitrite, is an example of both reaction mechanisms: through a two-electron – direct – reaction with peroxynitrite or through a one-electron – indirect – transfer reaction. In the indirect mechanism, the peroxynitrite homolysis releases hydroxyl and nitrogen dioxide radical and in addition the degradation of short-lived adduct formed by peroxynitrite and  $CO_2$  can generate carbonate radical anion. The reaction of hypotaurine and cysteine sulfinic acid with peroxynitrite-

Taurine 10:

derived radicals is accompanied by extensive oxygen uptake with the generation of transient intermediates, which can begin a reaction by an oxygen-dependent mechanism with the sulfonates, taurine, and cysteic acid as final products. Due to pulse radiolysis studies, it has been shown that transient sulfonyl radicals (RSO<sub>2</sub>) have been produced during the oxidation of both sulfinates by one-electron transfer reaction.

The purpose is to analyze all the aspects of the reactive mechanism in the sulfinic group oxidation of hypotaurine and cysteine sulfinic acid through the results obtained from our laboratory in recent years.

Abbreviations:  $RSO_2^-$ , sulfinates;  $RSO_2^+$ , sulfonyl radicals;  $RSO_3^-$ , sulfonates;  $RSO_2OO^+$ , sulfonyl-peroxyl radicals; HTAU, hypotaurine; CSA, cysteine sulfinic acid; TAU, taurine; CA, cysteic acid; ONOO<sup>-</sup>, peroxynitrite;  $CO_3^-$ , carbonate anion radical; 'NO<sub>2</sub>, nitrogen dioxide; 'NO, nitric oxide

### **1.1 INTRODUCTION**

Hypotaurine (HTAU) and cysteine sulfinic acid (CSA) are the last two metabolic intermediates in the production of taurine in the mammalian cysteine pathway. This pathway is dependent upon three specific enzymes as well as a variety of biologically relevant oxidizing agents (scheme 1) (Huxtable 1992; Stipanuk and Ueki 2011; Ricci et al 1978; Fontana et al 2005; Fellman et al., 1987; Aruoma et al., 1988; Pecci et 1999; Baseggio Conrado et al 2014). It is well known that cysteine dioxygenase is the first enzyme involved in this pathway and its activity leads to CSA. With the addition of molecular oxygen to the cysteine, to its sulfur atom, there is a conversion to the sulfinic acid from the thiol. (Stipanuk and Ueki 2011). At this point, the pathway can go in one of two directions both of which lead to taurine (TAU). In the first, a decarboxylation occurs and HTAU is produced due to the activity of CSA decarboxylase, and the sulfinic group of HTAU is subsequently oxidized to the sulfonic group of TAU. CSA can also undergo oxidation to produce cysteic acid (CA) and, through subsequent decarboxylation, forms TAU (Scheme 1) (Huxtable 1992; Stipanuk and Ueki 2011). This crucial point, the sulfinic group oxidation to the respective sulfonic group, has never been associated with any specific enzyme (Wright et al. 1986; Huxtable 1992). Conversely, evidences that in vivo formation of TAU and CA is the result of sulfinate (RSO<sub>2</sub><sup>-</sup>) interaction with a variety of biologically relevant oxidizing agents, are strong

Adv Exp Med Biol vol. ### | Taurine 10

(Ricci et al. 1978; Fellman et al. 1987; Aruoma et al. 1988; Pecci et al. 1999; Fontana et al. 2005; Baseggio Conrado et al. 2014).



**Scheme 1.** Hypotaurine and cysteine sulfinic acid as intermediates in the mammalian pathway leading from cysteine to taurine

Even though reactive oxygen species (ROS) are known to be capable of oxidizing the sulfinic group of HTAU and CSA, only hydroxyl radicals ('OH) and singlet oxygen have shown high direct reactivity leading to TAU formation (Pecci et al. 1999 and 2000; Arouma et al. 1988). This is in spite of low direct reactivity between hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), super-oxide anion (O<sub>2</sub><sup>-</sup>) and RSO<sub>2</sub><sup>-</sup> (Aruoma et al. 1998).

The oxidation of both HTAU and CSA to TAU and CA, respectively, can be also mediated by peroxynitrite (ONOO<sup>-</sup>) (Fontana et al. 2005 and 2006). The sulfinate oxidation mediated by ONOO<sup>-</sup> is an example of both direct and indirect reactions: via a two-electron mechanism (direct reaction) with ONOO<sup>-</sup> or by a one-electron transfer mechanism (indirect reaction). In the indirect mechanism, the ONOO<sup>-</sup> homolysis releases hydroxyl ('OH) and nitrogen dioxide ('NO<sub>2</sub>) radical and in addition the degradation of short-lived adduct formed by ONOO<sup>-</sup> and CO<sub>2</sub> can generate carbonate radical anion (CO<sub>3</sub><sup>-</sup>) (Fontana et al. 2005 and 2008). Two previous studies, using the peroxidase activity of Cu-Zn superoxide dismutase and horseradish peroxidase, carried out from our group have demonstrated the importance of both carbonate anion and nitrogen dioxide radicals in the oxidation of HTAU (Baseggio Conrado et al. 2014 and 2015). The reaction of HTAU and CSA with peroxynitrite-derived radicals is accompanied by extensive oxygen uptake suggesting the generation of transient sulfonyl

Adv Exp Med Biol vol. ### | Taurine 10

radicals (RSO<sub>2</sub>), which can begin a reaction by an oxygen-dependent mechanism with the sulfonates (RSO<sub>3</sub><sup>-</sup>), TAU and CA, as final products (Fontana et al. 2005 and 2006). Pulse radiolysis studies showed that transient sulfonyl radicals (RSO<sub>2</sub>) have been produced during the oxidation of both sulfinates by one-electron transfer reaction (Baseggio Conrado et al. 2014). The findings that HTAU and CSA efficiently react with hydroxyl radical, singlet oxygen as well as CO3<sup>-</sup> and 'NO2 provided further support for the proposed role of these compounds as antioxidants and free radicaltrapping agents in vivo (Green et al. 1991; Tadolini et al. 1995; Fontana et al. 2004). Of note, in all studies performed on the antioxidant activity of both RSO<sub>2</sub><sup>-</sup>, the scavenger effect of CSA on different targets, such as tyrosine dimerization, is higher than that exerted by HTAU, which can be explained considering the CSA-derived sulforyl radicals. These transient radicals can undergo degradation to sulfite, a secondary product with an efficient protective effect against oxidative reactions. Instead, the RSO<sub>2</sub>. derived from the HTAU oxidation seems to have a reduced tendency to decay.

The mechanisms involved in the interaction of the reactive oxygen (ROS) and nitrogen species (RNS) with  $RSO_2^-$ , HTAU and CSA, have only recently been investigated in depth (Fontana et al. 2005; Baseggio Conrado et al. 2014). In particular, starting from the findings of Cavallini and coworkers (Ricci et al. 1978; Pecci et al. 1999 and 2000) and subsequently, through experiments performed to understand whether peroxynitrite could be included in the biologically relevant reactive species capable of oxidizing  $RSO_2^-$ , it has been possible to highlight the reaction pathway mechanism of  $RSO_2^-$  leading to  $RSO_3^-$  (Fontana et al 2005 and 2006).

# **1.2 Sulfinic group oxidation by one-electron pathway (indirect reaction)**

In recent years, through three different oxidants – ONOO<sup>-</sup>, 'NO<sub>2</sub> and CO<sub>3</sub><sup>-</sup> – that were tested for inclusion as oxidant agents capable of oxidizing the sulfinic group of RSO<sub>2</sub><sup>-</sup>, it has been possible to define the indirect oneelectron pathway in the reaction from RSO<sub>2</sub><sup>-</sup>, HTAU or CSA, to RSO<sub>3</sub><sup>-</sup>, TAU or CA (Baseggio Conrado et al. 2014; Fontana et al. 2005). The extensive ONOO<sup>-</sup> studies were due to both its strong oxidizing and nitrating effect on substrates. It has been suggested that the damage in cellular and tissue in neurodegenerative disorders as well as in inflammatory and autoimmune disease can be related to the oxidizing and nitrating effect of ONOO<sup>-</sup> that acts as a reactive toxic species (Stewart and Heales 2003; Koppenol et al. 1992; Pryor and Squadrito 1995; Huie and Padmaja 1993;

Adv Exp Med Biol vol. ### | Taurine 10

Eiserich et al. 1998;). In vivo, nitric oxide ('NO) and O<sub>2</sub><sup>-</sup> can react through a diffusion-controlled reaction and produce ONOO<sup>-</sup> (scheme 2) (Beckman et al. 1990; Pryor and Squadrito 1995). It has been described that ONOOcould be present with its conjugate acid (ONOOH,  $pK_a = 6.8$ ) which decays rapidly  $(t_{1/2} < 1 \text{ s})$ . ONOOH homolysis can generate two radicals, OH) and NO<sub>2</sub>, capable of oxidizing through an indirect one-electron reaction other target substrates (Radi et al. 2001). Moreover, the decomposition of peroxynitrite-CO<sub>2</sub> adducts, produced under physiological conditions by peroxynitrite predominant reaction with  $CO_2$ , can generate  $CO_3$  and  $NO_2$ (Scheme 2) (Lymar and Hurst 1998; Meli et al. 2002; Augusto et al. 2002). It has been shown that  $CO_3^{-}$  and 'NO<sub>2</sub> stimulate the oxidation, peroxidation, and nitration of several biological targets (Beckman et al. 1990). Furthermore, Bonini and collaborators have suggested that the ONOO<sup>-</sup> production and peroxidase activity of superoxide dismutase (SOD) can play a key role as mediator in the oxidative damage (Bonini et al. 2004a; Bonini et al. 2004b).

 $CO_3^-$  is a strong acid (pK<sub>a</sub> < 0) that through a one-electron mechanism (E° = 1.5 V) can oxidize many biological targets that consequently produces radicals (Augusto et al. 2002; Bonini and Augusto 2001). NO<sub>2</sub> is a neutral radical and shows a greater selectivity towards biomolecules, compared to  $CO_3^-$ . NO<sub>2</sub> can mediate oxidation and nitration of various substrates and it can be generated by many physiological processes, such as 'NO autoxidation, homolysis of ONOO<sup>-</sup> or its addition to  $CO_2$  (Farrel et al. 1992; Torre et al. 1996; Brennan et al. 2002). The relevance of these two radicals in the oxidation of RSO<sub>2</sub><sup>-</sup> has been analysed through the peroxidase activity of two different enzymes, the Cu-Zn SOD and horseradish peroxidase (Baseggio Conrado et al. 2014 and 2015). In particular, we have shown that diffusible  $CO_3^-$ , produced by the SOD/H<sub>2</sub>O<sub>2</sub> system with bicarbonate, is involved in sulfinate oxidation suggesting the generation of transient intermediates in the oxidative step of RSO<sub>2</sub><sup>-</sup> to RSO<sub>3</sub><sup>-</sup> (Baseggio Conrado et al. 2014).

$$^{\bullet}NO + O_2 ^{\bullet} \xrightarrow{}_{k = 3.9 \times 10^9 M^{-1} S^{-1}} ONOO^{-} \xrightarrow{} ONOO^{-} ONOOH \longrightarrow ^{\bullet}NO_2 + ^{\bullet}OH$$

Scheme 2. Formation and fate of peroxynitrite at physiological pH

Adv Exp Med Biol vol. ### | Taurine 10

# **1.3** Sulfonyl radical (RSO<sub>2</sub><sup>•</sup>)/sulfonyl-peroxyl radical (RSO<sub>2</sub>OO<sup>•</sup>) as transient intermediates in the one-electron pathway of sulfinic group oxidation

 $RSO_2^ \xrightarrow{\text{oxidising agents}}$   $RSO_2^\circ$   $\xrightarrow{\text{oxidation}}$   $RSO_2OO^\circ$   $RSO_2-SO_2R$   $RSO_2-SO_2R$ 

Scheme 3. One-electron pathway of sulfinate oxidation

In the indirect one-electron pathway of the  $RSO_2^-$  oxidation, the first step is the reaction between  $RSO_2^-$  and biological oxidizing agents, such as carbonate radical anion, hydroxyl radical and nitrogen dioxide, and is accompanied by the formation of  $RSO_2^+$  (Aruoma et al. 1988; Sehested and Holcman, 1996; Fontana et al 2005 and 2008; Baseggio Conrado et al. 2014 and 2015). Acting as a strong oxidizing species,  $RSO_2^+$  can produce the sulfonyl-peroxyl radical ( $RSO_2OO^+$ ) when oxygen is present (Scheme 3) (Sevilla et al. 1990).

The above fate for the RSO<sub>2</sub> is not the only one, actually, RSO<sub>2</sub> can dimerize and produce the disulfone derivative (RSO<sub>2</sub>-SO<sub>2</sub>R) (Fellman et al. 1987; Ricci et al. 1978; Green and Fellman 1994).

Pulse radiolysis studies carried out by our group, used to evaluate the oneelectron oxidation rate between  $RSO_2^-$ , HTAU and CSA, with 'OH,  $CO_3^-$ , and 'NO<sub>2</sub>, has supported the evidence of the production of  $RSO_2^-$  (Baseggio Conrado et al. 2014). We have observed that during the reaction between HTAU or CSA with  $CO_3^-$  at pH 7.4 maximum absorption spectra around 320 nm (within a region of 300-350 nm), and at 600 nm was also observed a decay of  $CO_3^-$  absorption spectra. (Baseggio Conrado et al. 2014). Taking into account that Sehested and Holcman (1996) observed during the oxidation of methansulfinic acid by 'OH a sulfonyl radical (maximum optical absorption spectra around 325 nm), the spectra obtained in the reaction between  $RSO_2^-$  and oxidizing agents, such as 'OH,  $CO_3^$ and 'NO<sub>2</sub>, can be attributed to  $RSO_2^-$  as a transient intermediate.

$$RSO_2^{\bullet} \xrightarrow{O_2} RSO_2OO^{\bullet} \xrightarrow{RSO_2^{-}} RSO_2OO^{-} + RSO_2^{\bullet}$$

Scheme 4. Formation and fate of sulfonyl-peroxyl radical

In the ONOO<sup>-</sup> experiments, the oxidation of  $RSO_2^-$ , HTAU and CSA has been shown to be associated with an extensive oxygen uptake (Fontana et al. 2005 and 2006). The high  $O_2$  consumption observed is related to the strong oxidizing activity of  $RSO_2^-$  which reacts with oxygen producing the

Adv Exp Med Biol vol. ### | Taurine 10

sulfonyl-peroxyl radical (RSO<sub>2</sub>OO<sup>•</sup>). RSO<sub>2</sub>OO<sup>•</sup> is an intermediate in this mechanism and also a highly reactive species that can react with excess  $RSO_2^-$  and produce peroxysulfonate (RSO<sub>2</sub>OO<sup>-</sup>) (Scheme 4) (Sevilla et al. 1990).

# **1.4 Different fates of HTAU- and CSA-derived sulfonyl radicals** generated by one-electron sulfinate oxidation



Scheme 5. Decomposition pathway of sulfonyl radical to sulfite/sulfate

In all experiments performed to detect the yield in the production of  $RSO_3^-$ , TAU and CA, from RSO2-, HTAU and CSA, a different behaviour has always been reported between the HTAU- and CSA-derived sulfonyl radical (Pecci et al. 2000; Fontana et al. 2005 and 2006; Baseggio Conrado et al. 2014 and 2015). It has been shown that only a fraction of the depleted CSA was oxidized to CA with a yield of around 17%, compared with the almost total oxidation of HTAU in TAU (Baseggio Conrado et al. 2014). Sulfonyl radical derived by CSA has a high tendency to degrade to acetaldehyde and concurrently producing ammonia, CO2, and sulfite (Harman et al. 1984; Pecci et al. 2000). In detail, CSA-derived sulfonyl radicals partly undergo decomposition, producing sulfur dioxide (SO<sub>2</sub>) and a highly carbon-centred radical (R) which can react with  $O_2$  and further degrade to end products, such as ammonia or  $CO_2$  (Scheme 5) (Pecci et al. 2000). In contrast, HTAU-derived sulfonyl and sulfonyl-peroxyl radicals are more stable. It is likely that these highly reactive radicals could also promote the oxidation of suitable target molecules (Fontana et al. 2008).

# **1.5** Two-electron mechanism of sulfinic group oxidation (direct reaction)

 $ONOOH + RSO_2^{-} \longrightarrow RSO_3^{-} + NO_2^{-} + H^+$ 

Scheme 6. Two-electron mechanism of sulfinate oxidation

In the pathway leading from  $RSO_2^-$  to  $RSO_3^-$ , a route in addition to the one-electron mechanism can occur without any oxygen consumption. This route is a two-electron mechanism that can directly produce the  $RSO_3^-$  (Scheme 6) (Fontana et al. 2005 and 2006).

In this reaction, oxygen is transferred directly from  $ONOO^-$  to  $RSO_2^-$  with the production of  $RSO_3^-$  and nitrite in stoichiometric amounts. According to kinetic experiments, the direct reaction between  $ONOO^-$  and HTAU or CSA appears to be a second-order rate with, respectively, 77.4 and 76.4 M<sup>-1</sup>s<sup>-1</sup> constant. Moreover, ONOOH can be considered the reactant in the direct oxidative pathway as a correlation between  $k_{app}$  build-up and pH decrease has been observed (Fontana et al. 2005).

## **1.6 Conclusion**

The oxidation of sulfinates, HTAU and CSA, by reactive oxygen and nitrogen species to form sulfonates, TAU and CA, may occur either through one- or two-electron pathways as shown in scheme 7.



Scheme 7. One and two-electron pathway for the oxidative reactions of sulfinates

To briefly overview the one-electron mechanism: the reaction between sulfinates ( $RSO_2^-$ ) and one-electron oxidants, such as ONOO<sup>-</sup>,  $CO_3^-$  and  $NO_2$ , is accompanied by the generation of intermediate sulfonyl radicals ( $RSO_2^-$ ). Acting as a strong oxidizing species,  $RSO_2^-$  can produce the sul-

Adv Exp Med Biol vol. ### | Taurine 10

fonyl-peroxyl radical (RSO<sub>2</sub>OO<sup>•</sup>) when oxygen is present (① in Scheme 7) (Sevilla et al. 1990; Flyunt et al. 2001). RSO<sub>2</sub>OO<sup>•</sup> is an intermediate in this mechanism and also a highly reactive species that can react with excess RSO<sub>2</sub><sup>-</sup> and produce peroxysulfonate (RSO<sub>2</sub>OO<sup>-</sup>) (② in Scheme 7). Subsequently, the RSO<sub>2</sub>OO<sup>-</sup> formed decomposes to yield sulfonate (RSO<sub>3</sub><sup>-</sup>) and molecular oxygen or oxidizes the excess RSO<sub>2</sub><sup>-</sup> to RSO<sub>3</sub><sup>-</sup> (③ and ④ in Scheme 7).

Moreover, RSO<sub>2</sub> can either proceed through other three routes. It may initiate an oxygen-dependent radical chain propagation with a possible amplification of the oxidative mechanism ( $\$  in scheme 7) (Sevilla et al. 1990; Flyunt et al. 2001) or it can dimerize to form disulfone derivative (RSO<sub>2</sub>–SO<sub>2</sub>R), that subsequently hydrolyzes to yield RSO<sub>3</sub><sup>-</sup> ( $\$  in Scheme 7) (Ricci et al. 1978; Fellman et al. 1987; Green and Fellman 1994). The dimerization route does not require oxygen and can be operative in systems when low oxygen uptake is observed or in anaerobiosis (Ricci et al. 1978; Pecci et al. 1999; Baseggio Conrado et al. 2015).

The last possible route in the indirect pathway is the decomposition of  $RSO_2$ . As reported above, especially CSA-derived sulfonyl radicals can partly undergo degradation with production of sulfite ( $\bigcirc$  in scheme 7) (Harman et al. 1984; Pecci et al. 2000).

In the two-electron process, the direct transfer of oxygen from the oxidant, such as  $ONOO^-$ , to  $RSO_2^-$  generates  $RSO_3^-$  without any oxygen consumption. Which pathway occurs, between the one or two-electron mechanisms, depends mainly on  $RSO_2^-$  concentration. In fact, in the  $ONOO^-$ -mediated oxidation, it has been shown that at low  $RSO_2^-$  concentration,  $RSO_2^+$  formation is associated with high oxygen uptake via the one-electron mechanism. On the contrary, at high  $RSO_2^-$  concentration, the second-order reaction (between  $RSO_2^-$  and  $ONOO^-$ ) becomes more significant, producing a decrease in the oxygen consumption and consequently the direct oxidation of  $RSO_2^-$  without the formation of  $RSO_2^-$  predominates (Fontana et al. 2005 and 2006).

It is noteworthy that the sulfonyl and sulfonyl-peroxyl radicals produced in this mechanism are highly reactive oxidizing agents and could promote oxidative reactions (Pecci et al. 2003; Fontana et al. 2008; Baseggio Conrado et al. 2014). Looking forwards, it would be interesting to investigate the pathophysiological role of sulfonyl or sulfonyl-peroxyl radicals, particularly considering the increased significance of reactive sulfur species (RSS) in cellular oxidative stress. (Schöneich et al. 1992; Giles and Jacob 2002; Jacob 2012).

As outlined in this review, many questions on the oxidative mechanism of sulfinates to sulfonates have been addressed, however, the actual biologi-

cal relevance and the pathophysiological role of generated reactive intermediates deserve to be further investigated.

# **1.7 REFERENCES**

- Aruoma OI, Halliwell B, Hoey BM, Butler J (1988) The antioxidant action of taurine, hypotaurine and their metabolic precursor. Biochem J 256:251-255
- Augusto O, Bonini MG, Amanso AM, Linares E, Santos CCX, De Menezes S (2002) Nitrogen dioxide and carbonate radical anion: two emerging radicals in biology. Free Rad Biol Med 32:841-859
- Baseggio Conrado A, D'Angelantonio M, Torreggiani A, Pecci L, Fontana M (2014) Reactivity of hypotaurine and cysteine sulfinic acid toward carbonate radical anion and nitrogen dioxide as explored by the peroxidase activity of Cu,Zn superoxide dismutase and by pulse radiolysis. Free Radic Res 48:1300-1310
- Baseggio Conrado A, Pecci L, Capuozzo E, Fontana M (2015) Oxidation of Hypotaurine and Cysteine Sulfinic Acid by Peroxidase-generated Reactive Species. Adv Exp Med Biol 803:41-51
- Beckman JS, Beckman TW, Chen J, Marshall PA Freeman BA (1990) Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. Proc Natl Acad Sci USA 87:1620-1624
- Bonini MG, Augusto O (2001) Carbon dioxide stimulates the production of thiyl, sulfinyl and disulfide radical anion from thiol oxidation by peroxynitrite. J Biol Chem 276:9749-9754
- Bonini MG, Fernandes DC, Augusto O (2004a) Albumin oxidation to diverse radicals by the peroxidase activity of Cu,Zn-superoxide dismutase in the presence of bicarbonate or nitrite: diffusible radicals produce cysteinyl and solventexposed and –unexposed tyrosyl radicals. Biochemistry 43:344-351
- Bonini MG, Miyamoto S, Di Mascio P, Augusto O (2004b) Production of carbonate radical anion during xanthine oxidase turnover in the presence of bicarbonate. J Biol Chem 279:51836-51843
- Brennan ML, Wu W, Fu X, Shen Z, Song W, Front H, Vadseth C, Narine L, Lenkiewicz E, Borchers MT, Lusis AJ, Lee JJ, Lee NA, Abu-Soud HM, Ischiropoulos H, Hazen SL (2002) A tale of two controversies. Defining both role of peroxidase in nitrotyrosine formation in vivo using eosinophil peroxidase and myeloperoxidase-deficient mice, and the nature of peroxidase-generated reactive species. J Biol Chem 277:17415-17427
- Eiserich JP, Patel RP, O'Donnel VB (1998) Pathophysiology of nitric oxide and related species: free radical reactions and modification of biomolecules. Molec Aspects Med 19:221-357
- Farrel AJ, Blake DR, Palmer RMJ, Moncada S, (1992) Increased concentration of nitrite in synovial fluid and serum samples suggest increased nitric oxide synthesis in rheumatic diseases. Ann Rheum Dis 51:1219-1222.

Adv Exp Med Biol vol. ### | Taurine 10

- Fellman JH, Green TR, Eicher AL (1987) The oxidation of hypotaurine to taurine: bis-aminoethyl-α-disulfone, a metabolic intermediate in mammalian tissue. Adv Exp Med Biol 217:39-48
- Flyunt R, Makogon O, Schuchmann MN, Asmus KD, von Sonntag C (2001) OHradical-induced oxidation of methanesulfinic acid. The reaction of methanesulfonyl radical in the absence and presence of dioxygen. J Chem Soc Perkin Trans 2:787–792
- Fontana M, Amendola D, Orsini E, Boffi A, Pecci L (2005) Oxidation of hypotaurine and cysteine sulphinic acid by peroxynitrite. Biochem J 389: 233-240
- Fontana M, Duprè S, Pecci L (2006) The reactivity of hypotaurine and cysteine sulfinic acid with peroxynitrite. Adv Exp Med Biol 583:15-24
- Fontana M, Giovannitti F, Pecci L (2008) The protective effect of hypotaurine and cysteine sulphinic acid on peroxynitrite-dependent oxidative reactions. Free Radic Res 42: 320-330
- Fontana M, Pecci L, Duprè S, Cavallini D (2004) Antioxidant properties of sulfinates: protective effect of hypotaurine on peroxynitrite-dependent damage. Neurochem Res 29: 111-116
- Giles GI, Jacob C (2002) Reactive sulfur species: an emerging concept in oxidative stress. Biol Chem 383:375–388
- Green TR, Fellman JH (1994) Effect of photolytically generated riboflavin radicals and oxygen on hypotaurine antioxidant free radical scavenging activity. Adv exp med Biol 359:19-29
- Green TR, Fellman JH, Eicher AL, Pratt KL (1991) Antioxidant role and subcellular location of hypotaurine and taurine in human neutrophils. Biochim Biophys Acta 1073:91-97
- Harman LS, Mottley C, Mason RP (1984) Free radicals metabolites of L-cysteine oxidation. J Biol Chem 259:5609-5611
- Huie RE, Padmaja S (1993) The reaction of NO with superoxide. Free Radic Res Commun 18:195-199
- Huxtable RJ (1992) Physiological actions of taurine. Physiol Rev 72:101-163
- Jacob C (2012) Redox signalling via the cellular thiolstat. Biochem Soc Trans 39:1247–1253.
- Koppenol WH, Moreno JJ, Pryor WA, Ischiropoulos H, Beckman JS (1992) Peroxynitrite, a cloaked oxidant formed by nitric oxide and superoxide. Chem Res Toxicol 5:834-842
- Lymar SV, Hurst JK (1995) Rapid reaction between peroxonitrite ion and carbondioxide. Implications for biological activity. J Am Chem Soc 117:8867-8868
- Meli R, Nauser T, Latal P, Koppenol WH (2002) Reaction of peroxynitrite with carbon dioxide: intermediates and determination of the yield of CO<sub>3</sub><sup>--</sup> and NO<sub>2</sub><sup>•</sup>. J Biol Inorg Chem 7:31-36
- Pecci L, Costa M, Montefoschi G, Antonucci A, Cavallini D (1999) Oxidation of hypotaurine to taurine with photochemically generated singlet oxygen: the effect of azide. Biochem Biophys Res Commun 254:661-665
- Pecci L, Costa M, Antonucci A, Montefoschi G, Cavallini D (2000) Methylene blue photosensitized oxidation of cysteine sulfinic and other sulfinates: the in-

volvement of singlet oxygen and the azide paradox. Biochem Biophys Res Commun 270:782-786

- Pecci L, Montefoschi G, Antonucci A, Costa M, Cavallini D (2003) Methylene Blue photosensitized oxidation of hypotaurine in the presence of azide generates reactive nitrogen species: formation of nitrotyrosine. Biochem Biophys Res Commun 301:411-416
- Pryor WA, Squadrito GL (1995) The chemistry of peroxynitrite: a product from the reaction of nitric oxide with superoxide. Am J Physiol 268:699-722
- Radi R, Peluffo G, Alvarez MN, Naviliat M, Cayota A (2001) Unraveling peroxynitrite formation in biological systems. Free Radic Biol Med 30:463-88
- Ricci G, Duprè S, Federici G, Spoto G, Matarese RM, Cavallini D (1978) Oxidation of hypotaurine to taurine by ultraviolet irradiation. Physiol Chem Phys 10:435-441
- Sehested K, Holcman J (1996) A pulse radiolysis study of the OH radical induced autoxidation of methanesulfinic acid. Radiat Phys Chem 47:357-360
- Sevilla MD, Becker D, Yan M (1990) The formation and structure of the sulfoxyl radicals RSO<sup>•</sup>, RSOO<sup>•</sup>, RSO2<sup>•</sup>, and RSO2OO<sup>•</sup> from the reaction of cysteine, glutathione and penicillamine thiyl radicals with molecular oxygen. Int J Radiat Biol 57:65-81
- Schöneich C, Dillinger U, von Brunchhausen F, Asmus KD (1992) Oxidation of polyunsatured fatty acids and lipids through thiyl and sulfonyl radicals: reaction kinetics, and influence of oxygen and structure of thiyl radicals. Arch Biochem Biophys 292:456–467
- Stewart VC, Heales SJ (2003) Nitric oxide-induced mitochondrial dysfunction: implications for neurodegeneration. Free Radic Biol Med 34:287-303
- Stipanuk MH, Ueki I (2011) Dealing with methionine/homocysteine sulfur: cysteine metabolism to taurine and inorganic sulfur. J Inherit Metab Dis 34:17-32
- Tadolini B, Pintus G, Pinna GG, Bennardini F, Franconi F (1995) Effects of taurine and hypotaurine on lipid peroxidation. Biochem Biophys Res Commun 213:820-826
- Torre D, Ferrario G, Speranza F, Orani A, Fiori GP, Zeroli C (1996) Serum concentrations of nitrite in patients with HIV-1 infection. J Clin Pathol 49:574-576
- Wright CE, Tallan HH, Lin YY, Gaull GE (1986) Taurine: biological update. Ann Rev Biochem 55:427-453