



Simon, N., & Dodd, A. (2017). A new link between plant metabolism and circadian rhythms? *Plant, Cell and Environment*, 40(7), 995-996.
<https://doi.org/10.1111/pce.12939>

Peer reviewed version

License (if available):
CC BY-NC

Link to published version (if available):
[10.1111/pce.12939](https://doi.org/10.1111/pce.12939)

[Link to publication record in Explore Bristol Research](#)
PDF-document

This is the author accepted manuscript (AAM). The final published version (version of record) is available online via Wiley at <http://onlinelibrary.wiley.com/doi/10.1111/pce.12939/abstract>. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/pure/about/ebr-terms>

1 **A new link between plant metabolism and circadian rhythms?**

2

3 Noriane M. L. Simon and Antony N. Dodd*

4

5 School of Biological Sciences, University of Bristol, Bristol BS8 1TQ, U.K.

6

7 * Corresponding author, antony.dodd@bristol.ac.uk; +44 (0)117 394 1176.

8

9 The rotation of the Earth around its axis generates 24 h cycles of environmental change,
10 such as daily rhythms of light and temperature. Circadian clocks, cellular biological
11 oscillators that generate 24-hour rhythms of gene expression and metabolism, are thought to
12 synchronize the functioning of organisms with these daily environmental changes. Circadian
13 regulation enables organisms to anticipate environmental changes such as dawn and dusk,
14 and co-ordinate their metabolism, physiology and behaviour with daily changes in the
15 environment. This is particularly important for plants, which cannot move to escape
16 environmental challenges.

17 In the experimental model *Arabidopsis thaliana* (*Arabidopsis*), correct circadian regulation
18 increases photosynthesis, biomass accumulation, survival, seed number and viability (Green
19 *et al.*, 2002; Dodd *et al.*, 2005). It is estimated that almost 90% of *Arabidopsis* transcripts
20 can oscillate in abundance over the 24 h cycle, with about 30% of transcripts being
21 circadian-regulated (Michael *et al.*, 2008). This multitude of genes under circadian control
22 highlights the pervasiveness of circadian regulation in co-ordinating the functioning of plants
23 with their rhythmic environment. Because photosynthetic light harvesting can only occur
24 during the day, and stored carbohydrate reserves require mobilization at night to supply
25 respiration and growth, plant metabolism is intimately associated with cycles of day and

26 night. Building upon extensive underpinning research into the molecular genetics of
27 circadian oscillators, the interactions between metabolism, signalling and circadian
28 regulation have become an important growth area in plant circadian biology. For example,
29 breakthroughs have demonstrated that the rate of nocturnal starch breakdown is intricately
30 timed so that plants do not starve at night (Graf *et al.*, 2010), sugars produced by
31 photosynthesis can entrain the circadian clock (Haydon *et al.*, 2013), and the concentrations
32 of ions such as Ca^{2+} and Mg^{2+} are regulated by, and can regulate, the circadian oscillator
33 (Dodd *et al.*, 2007; Feeney *et al.*, 2016).

34 In this issue of *Plant, Cell and Environment*, Shin *et al.*, 2017 identified another potential
35 connection between metabolism and circadian regulation. The authors established that an
36 energy-sensing protein complex can influence circadian rhythms. AKIN10 (known also as
37 KIN10 or SnRK1.1) is a catalytic α -subunit of Snf1 (sucrose non-fermenting1)-related kinase
38 1 (SnRK1), which is an evolutionarily conserved energy sensor. SnRK1 controls metabolic
39 enzymes through protein phosphorylation (Sugden *et al.*, 1999), and also regulates > 1000
40 transcripts in response to starvation by controlling transcription factor activity (Baena-
41 González *et al.*, 2007; Mair *et al.*, 2015). SnRK1 plays such a fundamental role in energy
42 metabolism that AKIN10 knockouts are lethal (Baena-González *et al.*, 2007). By
43 overexpressing AKIN10 with a chemically-inducible promoter, the authors explored the role
44 of AKIN10 in circadian regulation. They found that inducing very high levels of AKIN10
45 expression caused the circadian clock to assume a long period, of up to 5 h longer than
46 controls, when plants were under conditions of continuous light. Interestingly, the long
47 circadian period caused by AKIN10 overexpression disappeared in experiments performed
48 under continuous darkness, such that AKIN10 overexpressing plants had the same circadian
49 period as the controls. When AKIN10 overexpressors were in constant darkness,
50 supplementing the growth media with sugars did not restore the long circadian period that
51 occurred in the light. The authors interpret this to indicate that starvation does not cause the
52 insensitivity of circadian period to AKIN10 overexpression in darkness. Instead, Shin *et al.*

53 (2017) propose that the influence of AKIN10 upon circadian period forms a response to the
54 light environment.

55 The study also found that under both light/dark cycles and constant light, AKIN10
56 overexpression caused a delay in the peak of expression of transcripts encoding the
57 evening-expressed circadian oscillator component GIGANTEA (GI). This is interesting
58 because *gi-11* mutants are insensitive to a long-term effect of sucrose upon the circadian
59 oscillator (Dalchau *et al.*, 2011). Additionally, the authors found that the period of plants
60 harbouring the *tic-2* mutation in the circadian oscillator gene *TIME FOR COFFEE (TIC)* had
61 reduced sensitivity to the effects of AKIN10 overexpression, suggesting a role for TIC in the
62 regulation of circadian period by AKIN10.

63 It is intriguing that AKIN10, a key player in the regulation of energy metabolism of
64 Arabidopsis, can influence circadian rhythms. The work of Shin *et al.* (2017) builds on
65 studies demonstrating bidirectional regulatory interactions between circadian regulation and
66 metabolism (Fig. 1). For example, the environmental cycles of day and night dictate when
67 photosynthesis can occur, and photosynthesis is also regulated extensively by the circadian
68 oscillator (Dodd *et al.*, 2014). Importantly, the products of photosynthesis can, in turn, entrain
69 the circadian oscillator (Haydon *et al.*, 2013). Each morning, the upregulation of
70 photosynthesis causes an accumulation of sugars, which alters circadian oscillator gene
71 expression and can adjust the circadian phase (Haydon *et al.*, 2013). Similarly, the circadian
72 oscillator controls the rate of nocturnal starch consumption (Graf *et al.*, 2010), with one
73 mathematical model for the regulation of nocturnal starch degradation assuming the
74 presence of a sugar sensing mechanism (Feugier & Satake, 2013). In this way, the
75 environment affects metabolism, metabolism regulates the circadian oscillator, and the
76 circadian oscillator regulates metabolism (Fig. 1).

77 By demonstrating that a subunit of the central energy sensor SnRK1 affects the functioning
78 of the circadian oscillator, Shin *et al.* (2017) have identified a mechanism that has the
79 potential to couple metabolism with circadian regulation. This adds to the evidence that

80 reciprocal regulation between the circadian oscillator and energy metabolism exists across
81 several Kingdoms of life. For example, in mammals there are circadian rhythms of NAD⁺ and
82 ATP synthesis and feeding can reset the circadian oscillator, and in both plants and
83 cyanobacteria, the availability of energy can regulate circadian rhythms (Rust *et al.*, 2011;
84 Bass, 2012; Haydon *et al.*, 2013). Therefore, the long circadian period caused by AKIN10
85 overexpression (Shin *et al.* 2017) could point to a role for AKIN10 in interfacing the circadian
86 oscillator with both metabolism and environment, given the extensive influence of
87 environmental conditions upon the metabolic state of plants. In future, it will be informative to
88 determine the function and position of SnRK1 within the circadian system, to understand
89 how a sensor of cellular energy status contributes to the responses of plants to the daily
90 changes that occur in the environment.

91

92 **Figure legend**

93 **Figure 1.** Reciprocal regulation between the environment, metabolism and circadian
94 rhythms in *Arabidopsis thaliana*. SnRK1 is thought to regulate metabolic enzyme activity and
95 transcription in response to energy availability. In this issue, Shin *et al.* (2017) have identified
96 a new role for the SnRK1 catalytic subunit AKIN10 in regulating circadian period (shown in
97 red).

98

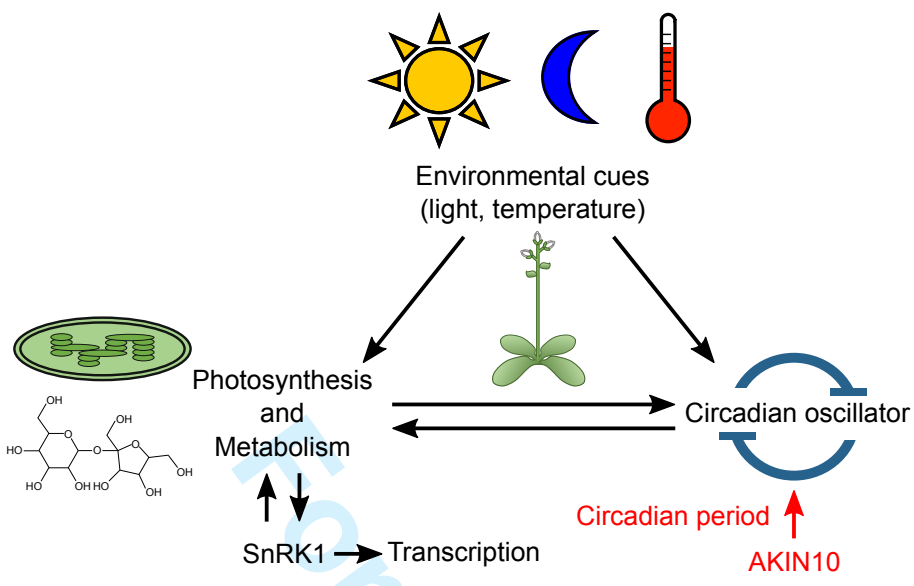
99 **References**

- 100 **Baena-González E, Rolland F, Thevelein JM, Sheen J. 2007.** A central integrator of
101 transcription networks in plant stress and energy signalling. *Nature* **448**(7156): 938-
102 942.
- 103 **Bass J. 2012.** Circadian topology of metabolism. *Nature* **491**(7424): 348-356.
- 104 **Dalchau N, Baek SJ, Briggs HM, Robertson FC, Dodd AN, Gardner MJ, Stancombe**
105 **MA, Haydon MJ, Stan G-B, Gonçalves JM, et al. 2011.** The circadian oscillator
106 gene *GIGANTEA* mediates a long-term response of the *Arabidopsis thaliana*
107 circadian clock to sucrose. *Proceedings of the National Academy of Sciences*
108 **108**(12): 5104-5109.
- 109 **Dodd AN, Gardner MJ, Hotta CT, Hubbard KE, Dalchau N, Love J, Assie J-M,**
110 **Robertson FC, Jakobsen MK, Gonçalves J, et al. 2007.** The *Arabidopsis* circadian
111 clock incorporates a cADPR-based feedback loop. *Science* **318**(5857): 1789.
- 112 **Dodd AN, Kusakina J, Hall A, Gould PD, Hanaoka M. 2014.** The circadian regulation of
113 photosynthesis. *Photosynthesis Research* **119**(1): 181-190.
- 114 **Dodd AN, Salathia N, Hall A, Kévei E, Tóth R, Nagy F, Hibberd JM, Millar AJ, Webb**
115 **AAR. 2005.** Plant circadian clocks increase photosynthesis, growth, survival, and
116 competitive advantage. *Science* **309**(5734): 630.
- 117 **Feeney KA, Hansen LL, Putker M, Olivares-Yañez C, Day J, Eades LJ, Larrondo LF,**
118 **Hoyle NP, O'Neill JS, van Ooijen G. 2016.** Daily magnesium fluxes regulate cellular
119 timekeeping and energy balance. *Nature* **532**(7599): 375-379.
- 120 **Feugier F, Satake A. 2013.** Dynamical feedback between circadian clock and sucrose
121 availability explains adaptive response of starch metabolism to various photoperiods.
122 *Frontiers in Plant Science* **3**: 305.
- 123 **Graf A, Schlereth A, Stitt M, Smith AM. 2010.** Circadian control of carbohydrate availability
124 for growth in *Arabidopsis* plants at night. *Proceedings of the National Academy of*
125 *Sciences* **107**(20): 9458-9463.

- 126 **Green RM, Tingay S, Wang Z-Y, Tobin EM. 2002.** Circadian rhythms confer a higher level
127 of fitness to *Arabidopsis* plants. *Plant Physiology* **129**(2): 576-584.
- 128 **Haydon MJ, Mielczarek O, Robertson FC, Hubbard KE, Webb AAR. 2013.**
129 Photosynthetic entrainment of the *Arabidopsis thaliana* circadian clock. *Nature* **502**:
130 689-692.
- 131 **Mair A, Pedrotti L, Wurzinger B, Anrather D, Simeunovic A, Weiste C, Valerio C,**
132 **Dietrich K, Kirchler T, Nägele T, et al. 2015.** SnRK1-triggered switch of bZIP63
133 dimerization mediates the low-energy response in plants. *eLife* **4**.
- 134 **Michael TP, Mockler TC, Breton G, McEntee C, Byer A, Trout JD, Hazen SP, Shen R,**
135 **Priest HD, Sullivan CM, et al. 2008.** Network discovery pipeline elucidates
136 conserved time-of-day-specific cis-regulatory modules. *PLOS Genetics* **4**(2): e14.
- 137 **Rust MJ, Golden SS, O'Shea EK. 2011.** Light-driven changes in energy metabolism
138 directly entrain the cyanobacterial circadian oscillator. *Science* **331**(6014): 220.
- 139 **Shin J, Sanchez-Villarreal A, Davis A, Du S, Berendzen K, Koncz C, Li C, Davis SJ.**
140 **2017.** The metabolic sensor AKIN10 modulates the *Arabidopsis* circadian clock in a
141 light-dependent manner. *Plant, Cell and Environment*.
- 142 **Sugden C, Donaghy PG, Halford NG, Hardie DG. 1999.** Two SNF1-related protein kinases
143 from spinach leaf phosphorylate and inactivate 3-hydroxy-3-methylglutaryl-coenzyme
144 A reductase, nitrate reductase, and sucrose phosphate synthase *in vitro*. *Plant*
145 *Physiology* **120**(1): 257-274.

146

147



Preprint Review Only