



Haworth, J., Jenkinson, H., Kerrigan, S., Brittan, J., & Nobbs, A. (2017). Cooperativity of streptococcal surface proteins in binding platelets and extracellular matrix: Spring Meeting for Clinician Scientists in Training 2017. *Lancet*, 389, Supplement 1, S45. https://doi.org/10.1016/S0140-6736(17)30441-5

Peer reviewed version

License (if available): CC BY-NC-ND

Link to published version (if available): 10.1016/S0140-6736(17)30441-5

Link to publication record in Explore Bristol Research PDF-document

This is the author accepted manuscript (AAM). The final published version (version of record) is available online via Elsevier at http://www.sciencedirect.com/science/article/pii/S0140673617304415. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: http://www.bristol.ac.uk/pure/about/ebr-terms

Cooperativity of streptococcal surface proteins in binding platelets and extracellular matrix

Jennifer Haworth, Howard Jenkinson, Steve Kerrigan, Jane Brittan, Angela Nobbs

Poster 34

University of Bristol, Bristol UK (J Haworth PhD, H Jenkinson PhD, J Brittan PhD, A Nobbs PhD); and Royal College of Surgeons in Ireland, Dublin, Ireland (S Kerrigan PhD)

Correspondence to:

Dr Jennifer Haworth, Oral and Dental Sciences, University of Bristol, Bristol BS1 2LY, UK

Jennifer.Haworth@bristol.ac.uk

Abstract

Background The ability of the oral bacterium *Streptococcus gordonii* to bind platelets and extracellular matrix (ECM) contributes to its virulence in infective endocarditis. Surface protein PadA has recently been found to be crucial for platelet activation. The hypothesis is that PadA is dependent upon another surface protein (Hsa) for *S gordonii* to activate platelets and adhere to ECM. We aimed to determine the respective roles of Hsa and PadA in platelet adhesion, and ascertain PadA function in ECM binding.

Methods *S gordonii* DL1 Δ*padA* and Δ*padA*Δ*hsa* knockout mutants were generated by allelic replacement. Mutants were complemented using PadA or Hsa expression plasmids under the control of a nisin-inducible promoter. PadA expression by knock-out and knock-in strains was confirmed by western immunoblot of cell-wall protein extracts. Platelet adhesion to bacteria was measured under static conditions in a p-nitrophenol assay. Bacterial adhesion to ECM proteins was determined by crystal violet assay.

Findings Static platelet adhesion by *S gordonii* $\Delta padA$ mutant was reduced by 30% compared with wild-type. $\Delta padA\Delta hsa$ was more than 80% reduced in binding platelets. Expression of padA in $\Delta padA\Delta hsa$ failed to restore any platelet adhesion, whereas expression of hsa in $\Delta padA\Delta hsa$ mutant restored binding to 70% of wild-type levels. The $\Delta padA$ mutant cells were reduced in binding cellular fibronectin by 25% and vitronectin by 60%. Deletion of hsa abrogated vitronectin binding. Complementation of $\Delta padA\Delta hsa$ with either hsa or padA alone did not restore vitronectin binding.

Interpretation PadA requires the presence of Hsa to interact with platelets. PadA has a minor role in binding cellular fibronectin alongside other surface adhesins. In vitronectin binding, Hsa requires the presence of functional PadA for efficient binding. These results suggest that the *S gordonii* surfaceanchored proteins Hsa and PadA work in concert to mediate processes relevant to host colonisation and pathogenesis.

Funding Wellcome Trust (grant WT097285MA awarded to JH).

Contributors

HJ, AN, and SK conceptualised the study. HJ, AN, and SK devised methods. JH, JB, SK, and AN conducted investigations. JH drafted the abstract. HJ and AN reviewed and edited the abstract. HJ, AN, and SK supervised the study.

Declaration of interests

We declare no competing interests.