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1 **Protective effect of phosphates and fluoride on the dissolution of hydroxyapatite**  
2 **and their interactions with saliva**

3

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8

9 **Short title:** Salivary pellicle modulates hydroxyapatite dissolution

10

11

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14

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24

**25 Abstract**

26 This study aimed to investigate the effect of phosphates and fluoride, alone or in  
27 combination, and the influence of salivary pellicle on hydroxyapatite (HA) dissolution.  
28 Baseline dissolution rate of HA discs was measured using a pH-stat system (0.3% citric  
29 acid, pH 3.2). In the first series of experiments, HA discs ( $n=8$ /group) were treated with  
30 a placebo solution (PLA, deionised water); sodium trimetaphosphate (TMP), sodium  
31 tripolyphosphate (TRI) and sodium pyrophosphate (PYRO) at 1% or 8%; 500 ppm F  
32 (500F); 1100 ppm F (1100F); 1100F/1% TMP; 1100F/8% TMP; 1100/1% TRI;  
33 1100F/8% TRI. In the second phase, HA discs were immersed in pooled human saliva  
34 (37° C/ 2h) and treated with PLA; 1100F/1% TMP; 1100F/8% TMP; 1100F/1% TRI;  
35 and 1100F/8% TRI. After treatments, final dissolution rate were measured from three  
36 consecutive 30-min assays. Statistical analysis were performed using 2-way ANOVA  
37 followed by Fisher's test ( $\alpha=0.05$ ). The type and concentration of phosphate tested  
38 significantly influenced HA dissolution; 8% TRI showed the highest reduction (36.9%)  
39 among all treatment solutions. Fluoride alone (1100F) significantly reduced HA  
40 dissolution by 20.7%. When fluoride and phosphates were associated, 1100F/1%TMP,  
41 1100F/8%TMP and 1100F/8% TRI showed the highest percentage reductions of  
42 dissolution (40.3 to 46.1%). Salivary pellicle led to a greater and more sustained  
43 protective effect of the treatment solutions compared to their counterparts without  
44 salivary coating. It was concluded that the association of phosphate and fluoride  
45 enhanced their protective effect against HA dissolution when compared with these  
46 compounds alone, especially in the presence of salivary pellicle.

47

## 48 Introduction

49 Dental erosion is defined as the softening of the tooth surface followed by its  
50 gradual bulk removal, caused by exposure to acids of non-bacterial origin [Lussi et al.,  
51 2011]. Surface softening and enamel loss are most often caused by the consumption of  
52 acidic soft drinks containing citric and/or phosphoric acids [Zero and Lussi, 2005].

53 Conventional fluoride preparations such as toothpastes or mouth rinses have a  
54 limited effect on erosion [Wiegand and Attin, 2003; Magalhães et al., 2011] and  
55 significant inhibition requires either the application in high concentrations or at high  
56 frequency [Ganss et al., 2004], or the use of preparations containing titanium or  
57 stannous ions [Ganss et al., 2010; Wiegand et al., 2010] that might be unsuitable for  
58 routine use because of their low pH and their propensity to stain the teeth. Nonetheless,  
59 promising results have been described for fluoridated products associated with sodium  
60 trimetaphosphate on erosion *in vitro* [Moretto et al., 2010; Manarelli et al., 2011;  
61 Manarelli et al., 2013] and *in situ* [Moretto et al., 2013]. Similarly, sodium  
62 tripolyphosphate and sodium pyrophosphate tetrabasic have also been shown to  
63 promote significant reductions on ~~HA discs the~~ dissolution of HA discs [Barbour et  
64 al., 2005] and ~~on hydroxyapatite crystals dissolution HA crystals~~ [Scaramucci et al.,  
65 2011; 2015] using a pH-stat approach [~~Barbour et al., 2005; Scaramucci et al., 2011;~~  
66 2015], despite these phosphates were not able to reduce enamel and dentine wear *in*  
67 *vitro* [Scaramucci et al., 2011].

68 Saliva contains phosphates, proteins and bicarbonate buffers and is  
69 supersaturated with respect to tooth minerals, such as calcium and phosphate. It is  
70 known that proteins can protect the teeth by the formation of a salivary pellicle when  
71 teeth are exposed to saliva [Siqueira et al., 2007]. This pellicle of adsorbed salivary  
72 proteins might act as a diffusion barrier or a selective permeable membrane reducing  
73 direct contact between acids and tooth surface and thus reducing demineralization of the  
74 surface [Wetton et al., 2007; Jager et al., 2011; White et al., 2012].

75 Based on the above, the aim of the present study was to investigate the effect of  
76 fluoride and phosphates, alone or in combination, on the dissolution of hydroxyapatite  
77 (HA), ~~as well as the~~ ~~The~~ influence of the salivary pellicle ~~on this process was also~~  
78 ~~assessed~~. The study hypothesis was that HA dissolution would be significantly reduced  
79 by the presence of fluoride and phosphates, and that this effect would be influenced by  
80 the presence of salivary acquired pellicle.

81

## 82 **Materials and Methods**

### 83 *Materials*

84 Discs of compressed HA, mean diameter 12.1 mm, mean thickness 1.23 mm,  
85 were purchased from HiMed Inc., Old Bethpage, N.Y., USA. All products were  
86 obtained from Sigma-Aldrich (Poole, Dorset, UK). Solutions were prepared using  
87 deionised water.

88

### 89 *Groups*

90 Treatment solutions included: Placebo (PLA, deionised water); sodium  
91 trimetaphosphate at 1% and 8% (1% TMP and 8% TMP, respectively); sodium  
92 tripolyphosphate at 1% and 8% (1% TRI and 8% TRI, respectively); sodium  
93 pyrophosphate tetrabasic at 1% and 8%) (1% PYRO and 8% PYRO, respectively);  
94 fluoride (500 ppm F and 1100 ppm F, as sodium fluoride); 1100 ppm F with sodium  
95 trimetaphosphate at 1% and 8% (1100 ppm F/1% TMP and 1100 ppm F/8% TMP,  
96 respectively); 1100 ppm F with sodium tripolyphosphate at 1% and 8% (1100 ppm  
97 F/1% TRI and 1100 ppm F/8% TRI). Fluoride concentrations were based on those  
98 found in conventional and low fluoride toothpastes. Fluoride concentrations were chosen  
99 based on those present in conventional formulations (1100 ppm F), as well as in low-  
100 fluoride toothpastes (500 ppm F) available in some countries.

101

### 102 *Measurement of Dissolution Rate*

103 Dissolution in 0.3% citric acid, pH 3.2, was measured using a pH-stat model  
104 (718 Stat Titrino: Metrohm UK, Runcorn, Cheshire, UK) with a 50 ml double-walled  
105 glass reaction vessel fitted with a lid with 3 inlet ports. Water was pumped by a  
106 circulating water bath (Type GD120; Grant Instruments, Cambridge, UK) through the  
107 water jacket to maintain the reaction temperature at 37 °C. Prior to use in the pH-stat,  
108 batches of discs were conditioned to ensure consistency of response. The discs were  
109 first ultra-sonicated in deionized water in a bath with an ultrasonic power of 40 kHz for  
110 15 min to remove loose HA particles. They were then exposed to gently stirred 0.3%  
111 citric acid, pH 3.2, for 30 min at room temperature, then washed in deionised water and  
112 finally air-dried. For use in the pH-stat, the back of the discs were coated with nail

113 varnish to leave an area of 161.4 mm<sup>2</sup> available for reaction. Discs were then fixed with  
114 sticky wax to the tip of glass tubes fitted with a cone suitable for the inlet ports.

115 In each experiment, 30 ml of citric acid solution was introduced into the reaction  
116 vessel and the pH electrode and burette tip fitted. After the system had reached thermal  
117 equilibrium, the pH was adjusted to 3.2 by adding small quantities of concentrated KOH  
118 or HCl solution and finally adjusted using the pH-stat. The reaction was initiated by  
119 introducing the specimen on its holder and addition of titrant (50 mM HCl) was  
120 recorded for 30 min. A control measurement of dissolution rate was made after the  
121 conditioning step and the disc was removed from the holder, washed in deionised water  
122 and dried. After exposing the disc to the chosen treatment, the disc was reattached to the  
123 glass specimen holder and measurement of dissolution rate was made.

124

#### 125 *Saliva Collection*

126 Mixed saliva was collected from a panel of 2 volunteers, who had previously  
127 registered at the saliva bank from the University of Bristol, having received local ethical  
128 approval for this. When saliva was required, each volunteer was provided with a 50 ml  
129 polystyrene Universal tube, marked at the 25 ml level. Each volunteer chewed a square  
130 of Parafilm to stimulate salivary flow and expectorated saliva into the tube until the  
131 mark was reached. These samples were combined and centrifuged using a Centaur 1  
132 (MSE, London, UK) at 4,000 g for 15 min at ambient temperature. The supernatant was  
133 used to treat HA specimens (2 mL/disc).

134

#### 135 *Experiments*

136 *Native HA.* In the first series of experiments, inhibition was tested on non-  
137 saliva-treated HA [Barbour et al., 2005; Jones et al., 2013]. First, a sequence of three  
138 control measurements was performed. Next, the HA discs were individually immersed  
139 in 100 mL of the treatment solutions (PLA; 1% TMP; 8% TMP; 1% TRI; 8% TRI; 1%  
140 PYRO; 8% PYRO; 500 ppm F; 1100 ppm F; 1100 ppm F/1% TMP; 1100 ppm F/8%  
141 TMP; 1100 ppm F/1% TRI; 1100 ppm F/8% TRI) for 100mL/2 min with gentle stirring,  
142 followed by rinsing with deionised water. ~~Fluoride concentrations were chosen based on~~  
143 ~~those present in conventional formulations (1100 ppm F), as well as in low fluoride~~  
144 ~~toothpastes (500 ppm F) available in some countries. Afterwards it was rinsed with~~  
145 ~~deionized water and rinsing in water; Following,~~ the post-treatment dissolution rate was

146 determined in 3 consecutive periods of 30 minutes each. ~~For this, each HA disc were~~  
147 ~~immersed in 100 ml of the respective treatment solutions for 2 min with gentle stirring.~~  
148 ~~Afterwards, it was rinsed with deionized water. For this, Next,~~ 30 mL of citric acid was  
149 added to the reaction vessel and the pH electrode and burette tip were fitted. After the  
150 system ~~had~~ reached equilibrium (pH 3.2 adjusted with KOH or HCl) the reaction was  
151 initiated by introducing the HA disc, and the addition of tritant (50 ~~mMmmol l<sup>-1</sup>~~ HCl)  
152 was recorded for 30 minutes using pH-stat, similarly ~~for the measures did as done~~ for  
153 the control. Fresh citric acid solutions were added to the vessel for each 30-min assay.  
154 This sequence was performed on 8 separate HA discs for each treatment.

155 *Saliva-Coated HA.* After making control measurements of dissolution rate, discs  
156 were immersed in pooled mixed saliva supernatant for 2 h in Petri dishes in an incubator  
157 at 37 ° C. Next, the HA discs were immersed in treatment (PLA; 1100 ppmF/1% TMP;  
158 1100 ppmF/8% TMP; 1100 ppmF/1% TRI; 1100ppmF/8% TRI) 100 ml/2 min with  
159 gentle stirring. After rinsing in water, a series of 3 post-treatment measurements of  
160 dissolution was performed. This sequence was performed on 8 separate HA discs for  
161 each treatment.

### 162 *Statistical Analysis*

164 Analyses were performed using the SigmaPlot (version 12.0) with 5% of  
165 statistical significance level. Data from native and saliva coated discs exhibited a  
166 normal (Kolmogorov-Smirnov) and homogeneous distribution. Treatment solutions  
167 (with or no salivary pellicle) and time (baseline control, post-treatment 1, 2 and 3) were  
168 considered as variation factors. The baseline was determined for each specimen by  
169 averaging the three control runs before treatment. Thus, data were submitted to 2-way  
170 ANOVA, followed by Fisher test ( $p < 0.05$ ).

### 171 **Results**

173 ~~Eleven—Ten~~ of the test solutions without pellicle effected a statistically  
174 significant reduction in the HA dissolution rate in citric acid solutions (Table 1). Among  
175 the phosphates, the highest percentage reduction of dissolution was seen for 8% TRI  
176 (36.9%), while 1100 ppm had the highest percentage reduction of dissolution (20.7%)  
177 among the solutions containing only NaF. For solutions containing NaF and a phosphate  
178 salt in combination, 1100 ppm F/1%TMP, 1100 ppm F/8%TMP e 1100 ppm F/8% TRI

179 showed the highest percentage reductions of dissolution (Table 1), ranging from 40.3 to  
180 46.1%.

181         Regarding the presence of salivary pellicle, all test solutions effected a  
182 statistically significant reduction in the HA dissolution rate in citric acid solution in  
183 specimens covered with salivary pellicle in comparison with their counterparts not  
184 treated with saliva (Figure 1). Salivary coating promoted a more sustained protective  
185 effect of the treatment solutions when compared to their counterparts not treated with  
186 saliva ( $p < 0.001$ ). Also, no significant differences were seen for the treatment groups in  
187 the presence of saliva regarding the reduction of dissolution rate and persistence of  
188 effect over time.

189

## 190 **Discussion**

191         In this study, the pH stat method was employed to investigate the immediate and  
192 sustained effect of sodium trimetaphosphate, sodium tripolyphosphate and sodium  
193 pyrophosphate tetrabasic with or without fluoride on hydroxyapatite dissolution. These  
194 phosphates have shown promising results against dental erosion, but most studies only  
195 evaluated their immediate effects. The present study showed that the reduction of  
196 hydroxyapatite dissolution as well as the duration of this effect was significantly  
197 influenced by type of phosphate and concentration tested, as well as by the salivary  
198 coating. Therefore, the study's hypothesis was accepted.

199         In the first set of experiments, all test solutions were evaluated without the  
200 presence of a salivary coating. It has been shown that fluoride can offer a protection  
201 against dental erosion and this effect is given by a formation of a layer of KOH-soluble  
202 calcium fluoride [Magalhães et al., 2011]. In the present study, a relationship between  
203 fluoride concentration and the protective effect was observed, since reductions in HA  
204 dissolution were 13% and 21% respectively for solutions containing 500 and 1100 ppm  
205 F. The results are in line with previous data showing a reduction of 12% in dissolution  
206 rate of native HA when 300 ppm F was administered [Jones et al., 2013]. However,  
207 these reductions did not persist beyond the first post-treatment run, confirming that  
208 fluoride alone might have a limited action against enamel erosion.

209         Regarding the phosphates evaluated, the highest reduction of HA dissolution rate  
210 (~37.4%) was observed for the solution containing 8% of TRI, and its effects were  
211 persistent up to the third post-treatment run (90 min). Despite the present results are



212 higher than data reported in previous investigations using the same methodology  
213 [Barbour *et al.*, 2005; Scaramucci *et al.*, 2011; 2015], it is noteworthy that the studies  
214 cited above used lower concentrations of this phosphate (ranging from 0.02 to 2%),  
215 what might help to explain the different results. This assumption is reinforced by the  
216 fact that 1% TRI promoted a much lower reduction in HA dissolution (~13%) than at  
217 8%, and its effects were only significant for the first post-treatment run (30 min). As  
218 for the other phosphates tested, the reductions in HA dissolution were less pronounced  
219 for TMP and PYRO at 8%, and their effects did not persist over the 3 post-treatment  
220 runs. The modest effect of TMP is in agreement with previous findings showing that  
221 TMP alone produce negligible effects on bovine enamel de-/remineralisation when  
222 added to different topically applied dental products [Danelon *et al.*, 2014; Manarelli *et al.*,  
223 2014]. The association of TMP and TRI with fluoride further reduced the rate of  
224 hydroxyapatite dissolution compared to these phosphates or fluoride alone. It is  
225 noticeable that while 1% TMP did not produce any significant effect on HA dissolution  
226 (1.2%), its association with 1100 ppm F led to maximum inhibition of 40.3%. Given  
227 that this value was 2-fold higher than that observed for 1100 ppm F alone, the results  
228 indicate that TMP and fluoride have a synergistic effect against HA dissolution,  
229 confirming previous observations using bovine enamel in a pH-cycling model [Castro *et al.*,  
230 2015]. In this sense, despite TMP has been shown to act like a partial barrier to CaF<sub>2</sub>  
231 deposition on enamel surface [Manarelli *et al.*, 2014], CaF<sub>2</sub> and CaF<sup>+</sup> compounds are  
232 believed to retain on TMP molecules (adhered to HA) and to be released to saliva upon  
233 acidification of the oral environment, further reacting with salivary phosphates leading  
234 to the formation of more reactive calcium phosphates [Manarelli *et al.*, 2014]. It is also  
235 noteworthy that the association with fluoride led to a significant increase in the  
236 protective effect of TMP at both concentrations tested (1 and 8%) tested, while for TRI  
237 this effect was only seen at 1%, indicating that each phosphate has an ideal molar ratio  
238 with fluoride in order to achieve the maximum additive or synergistic effect.

239 In the second set of experiments a salivary coating on hydroxyapatite was  
240 introduced, as it is known that saliva plays an important role on dental erosion. The  
241 acquired pellicle is composed of proteins and glycoproteins that act as a protective  
242 barrier, preventing the direct contact between the acid and the tooth surface [Buzalaf *et al.*,  
243 2012]. When compounds were tested with a salivary coating, a greater reduction of  
244 hydroxyapatite dissolution (~65% for all solutions together) was observed compared

245 with discs not previously treated with saliva (~40% for all solutions together) in the first  
246 post-treatment run, what is also in line with previous data using a similar research  
247 protocol [Jones et al., 2013]. It is noteworthy that while the protective effect for native  
248 HA decreased by 63% between the first and the second post treatment run, and by 66%  
249 between the second and the third post-treatment, these decreases were much lower for  
250 HA previously exposed to saliva (15 and 20%, respectively). This indicates that the  
251 protective layer formed by salivary components, fluoride and TMP or TRI was not  
252 totally dissolved from the HA surface after the first exposure to the acid media, but  
253 occurred gradually from its external to the basal components [Joiner et al., 2008].

254 Although the present in vitro model does not fully reproduce acid challenges  
255 occurring in vivo, our data can be helpful in further investigations. In this sense, the  
256 results indicate that fluoride associated with phosphates as TRI and TMP could be an  
257 alternative to the development of oral products against dental erosion. It is important to  
258 highlight that an optimal ratio between phosphate and fluoride should be used in order  
259 to achieve optimum results, based on the present results and also on previous data from  
260 experiments performed with bovine enamel specimens [Takeshita et al., 2009; Castro et  
261 al., 2015]. Another important point is related to the presence of saliva in future studies,  
262 as the effect of the treatments was shown to be highly dependent on the salivary coating  
263 of the specimens [Buzalaf et al., 2012]. Thus, the screening of such agents should  
264 always include specimens that have been pre-treated with saliva as well as native  
265 surfaces.

266 To sum up, it can be concluded that TMP and TRI provided reduction of  
267 hydroxyapatite dissolution when an erosion-like model was used. In addition~~to~~, the  
268 association of these phosphates with fluoride enhanced the~~ir~~ effectiveness compared to  
269 fluoride or phosphates alone. Thus, these associations could be a potential alternative in  
270 future investigations in order to prevent dental erosion.

271

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277

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- 349

350 **Table 1.** Mean (SD) percentage reduction of hydroxyapatite dissolution for each  
 351 treatment solution after exposure to citric acid according to time following exposure  
 352 to treatment solutions

Groups	Post-treatment 1 (30 min)	Post-treatment 2 (60 min)	Post-treatment 3 (90 min)
<b>PLA</b>	0.7 <sup>a</sup> (2.1)	0.6 <sup>a</sup> (2.1)	0.0 <sup>a</sup> (1.3)
<b>1% TMP</b>	1.2 <sup>a</sup> (5.8)	0.4 <sup>a</sup> (4.6)	0.0 <sup>a,e</sup> (3.8)
<b>8% TMP</b>	16.2 <sup>b,c</sup> (4.4)*	10.3 <sup>b,f</sup> (3.6)	5.2 <sup>b</sup> (5.4)
<b>1% TRI</b>	12.6 <sup>b</sup> (4.2)*	8.4 <sup>b,f</sup> (3.7)	2.0 <sup>b,e</sup> (3.8)
<b>8% TRI</b>	36.9 <sup>c,d</sup> (5.4)*	25.7 <sup>c</sup> (3.6)*	17.1 <sup>c</sup> (7.5)*
<b>1% PYRO</b>	7.7 <sup>a,b</sup> (6.7)	3.6 <sup>b</sup> (5.8)	0.0 <sup>b</sup> (5.2)
<b>8% PYRO</b>	18.2 <sup>b,c</sup> (4.9)*	10.3 <sup>b,d</sup> (2.8)*	4.6 <sup>b</sup> (2.5)
<b>500 ppm F</b>	13.1 <sup>b</sup> (3.3)*	9.0 <sup>d,f</sup> (4.2)*	4.8 <sup>b</sup> (6.1)
<b>1100 ppm F</b>	20.7 <sup>c</sup> (5.7)*	12.3 <sup>b</sup> (1.7)*	8.5 <sup>b</sup> (4.0)
<b>1100 ppm F/1%TMP</b>	40.3 <sup>d</sup> (5.5)*	17.2 <sup>e</sup> (4.7)*	2.6 <sup>d</sup> (5.8)
<b>1100 ppm F/8%TMP</b>	42.2 <sup>d</sup> (4.8)*	13.1 <sup>e</sup> (6.5)*	4.6 <sup>d</sup> (5.8)
<b>1100 ppm F/1%TRI</b>	32.3 <sup>c,d</sup> (7.8)*	11.5 <sup>b,d</sup> (8.1)*	7.0 <sup>b</sup> (7.5)
<b>1100 ppm F/8%TRI</b>	46.1 <sup>d</sup> (3.3)*	16.9 <sup>f</sup> (6.8)*	5.4 <sup>b</sup> (6.2)

353 Different superscript letters within each column show statistical difference among treatment  
 354 solutions. Asterisks indicate significantly difference from mean baseline control (Fisher's test,  
 355  $p < 0.05$ ,  $n = 8$ /group).

356 **PLA:** placebo solution (deionised water); **TMP:** sodium trimetaphosphate; **TRI:** sodium  
 357 tripolyphosphate; **PYRO:** sodium pyrophosphate.

358

359 Figure captions

360

361 **Figure 1.** Mean inhibition of hydroxyapatite dissolution for each treatment solution  
362 after exposure to citric acid in specimens with or without salivary coating. Bars indicate  
363 SD. Asterisks indicate significant differences between each post-treatment run and the  
364 control (baseline) measurement. Different letters indicate significant differences among  
365 the groups (Fisher test,  $p < 0.05$ ,  $n = 8/\text{group}$ ).