



The University of  
**Nottingham**

UNITED KINGDOM · CHINA · MALAYSIA

Voigt, Jörg-Peter and Fink, Heidrun (2015) Serotonin controlling feeding and satiety. *Behavioural Brain Research*, 277 . pp. 14-31.

**Access from the University of Nottingham repository:**

<http://eprints.nottingham.ac.uk/46066/1/BBR-D-14-00402%20revised.pdf>

**Copyright and reuse:**

The Nottingham ePrints service makes this work by researchers of the University of Nottingham available open access under the following conditions.

This article is made available under the Creative Commons Attribution Non-commercial No Derivatives licence and may be reused according to the conditions of the licence. For more details see: <http://creativecommons.org/licenses/by-nc-nd/2.5/>

**A note on versions:**

The version presented here may differ from the published version or from the version of record. If you wish to cite this item you are advised to consult the publisher's version. Please see the repository url above for details on accessing the published version and note that access may require a subscription.

For more information, please contact [eprints@nottingham.ac.uk](mailto:eprints@nottingham.ac.uk)

1  
2  
3  
4  
5  
6

1

7 **Serotonin controlling feeding and satiety**

8  
9

10  
11  
12 Jörg-Peter Voigt\* and Heidrun Fink<sup>1</sup>

13  
14  
15  
16  
17  
18

19 School of Veterinary Medicine and Science, University of Nottingham, Sutton Bonington,  
20 Loughborough LE12 5RD, UK, <sup>1</sup> Institute of Pharmacology and Toxicology, School of  
21 Veterinary Medicine, Freie Universität Berlin, Koserstr. 20, 14195 Berlin, Germany  
22  
23

24  
25  
26  
27  
28  
29  
30  
31

32 \* Corresponding author:

33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

School of Veterinary Medicine and Science  
University of Nottingham  
Sutton Bonington Campus  
Loughborough  
LE12 5RD  
United Kingdom  
Tel: +44 (0)115 9516408  
Fax: +44 (0)115 9516440  
peter.voigt@nottingham.ac.uk

**Abstract**

Serotonin has been implicated in the control of satiety for almost four decades. Historically, the insight that the appetite suppressant effect of fenfluramine is linked to serotonin has stimulated interest in and research into the role of this neurotransmitter in satiety. Various rodent models, including transgenic models, have been developed to identify the involved 5-HT receptor subtypes. This approach also required the availability of receptor ligands of different selectivity, and behavioural techniques had to be developed simultaneously which allow differentiating between unspecific pharmacological effects of these ligands and 'true' satiation and satiety. Currently, 5-HT<sub>1B</sub>, 5-HT<sub>2C</sub> and 5-HT<sub>6</sub> receptors have been identified to mediate serotonergic satiety in different ways. The recently approved anti-obesity drug lorcaserin is a 5-HT<sub>2C</sub> receptor agonist. In brain, both hypothalamic (arcuate nucleus, paraventricular nucleus) and extrahypothalamic sites (parabrachial nucleus, nucleus of the solitary tract) have been identified to mediate the serotonergic control of satiety. Serotonin interacts within the hypothalamus with endogenous orexigenic (Neuropeptide Y/Agouti related protein) and anorectic ( $\alpha$ -melanocyte stimulating hormone) peptides. In the nucleus of the solitary tract serotonin integrates peripheral satiety signals. Here, the 5-HT<sub>3</sub>, but possibly also the 5-HT<sub>2C</sub> receptor play a role. It has been found that 5-HT acts in concert with such peripheral signals as cholecystinin and leptin. Despite the recent advances of our knowledge, many of the complex interactions between 5-HT and other satiety factors are not fully understood yet. Further progress in research will also advance the development of new serotonergic anti-obesity drugs.

Keywords: 5-HT, 5-HT receptor, hypothalamus, obesity, feeding behaviour, leptin, cholecystinin

## 1. Introduction

What are the behavioural and physiological mechanisms that promote satiety? How is satiety defined? Satiety can be seen as a behavioural state which arises from food consumption and suppresses the initiation of eating for a particular period of time [1]. This description alone suggests a high degree of complexity as peripheral post-ingestive and post-absorptive signals need to be relayed to the brain where they are integrated with other signals to produce (or not) the behavioural state called satiety. The state of satiety is brought about a process called satiation, where sensory, cognitive and early post-ingestive mechanisms bring feeding to a halt and thus stopping a meal. Promoting satiation alone must not necessarily lead to reduced total food intake as the frequency of meals could be increased subsequently. Many peripheral and brain mechanisms have been identified that are involved in the expression satiety and it has been suggested that serotonin accelerates satiation and prolongs satiety [2]. In the following, we will review the role of serotonin in satiety in more detail<sup>1</sup>. The reader will see that, despite immense progress made during the last years, the field is still far from being resolved.

As serotonin (5-Hydroxytryptamine; 5-HT) is a phylogenetically old neurotransmitter, various functions had time to evolve in different phyla, but maybe also in different species. 5-HT receptors exist in animal cells for millions of years and they are as old as adrenoceptors or some peptide receptors, possibly even older [5, 6]. Even in invertebrates such as molluscs (*Aplysia californica*) and annelids (*Hirudo medicinalis*), 5-HT might functionally be related to food intake [7]. 5-HT is involved in feeding even in the honeybee where it has separate effects in the gut and in the insect brain [8]. In general, however, 5-HT seems rather to be involved in appetitive behaviours in invertebrates whereas it has more of a satiating effect in vertebrates [9]. In general, 5-HT neurons seem to be more extensively distributed throughout the body in lower animals than in higher animals including mammals where 5-HT neurones decrease in relative size and are much more clustered, sending axons from these to specific brain areas [10].

---

<sup>1</sup> Although the distinction between satiation and satiety is widely, but not unanimously [3], accepted, we used these terms synonymously. This is for simplification only. The role of serotonin in the structural aspects of feeding behaviour has been reviewed before [4]. As discussed in this review, a reduced food intake is not identical with satiety, but in most cases authors report experimental findings as if changes in food intake stand for changes in satiety.

## 2. Brain 5-HT and satiety

The multitude of 5-HT receptor families and 5-HT receptor subtypes in mammals (Barnes [11-13] and the complex serotonergic innervation of the mammalian brain [14] can possibly explain why 5-HT is involved in so many behaviours [15]. Evidence for an involvement of serotonin in food intake in men accumulated primarily during the 1960s. Thus appetite stimulating properties of the antihistaminergic/antiserotonergic drug cyproheptadine in humans and animals have been reported in the 1960s [16, 17]. During the same decade, fenfluramine (Ponderax) has been introduced as an anti-obesity drug, demonstrating significant weight loss in obese patients [18]. Fenfluramine is an amphetamine analogue and amphetamines' weight reducing effects are known since the 1930s [19-21]. In contrast to the original amphetamines, fenfluramine had no addictive properties allowing its usage as an appetite suppressant on a wider scale. Brain lesions and pharmacological experiments using 5-HT antagonists [22-26] revealed that the hypophagic effect of fenfluramine is indeed based on its serotonergic properties. The brain serotonergic system originates from raphe nuclei in the brainstem [14]. Lesions of these nuclei induce hyperphagia [27] and interfere with the anorectic effect of fenfluramine [28]. The latter finding demonstrates that fenfluramine requires an intact brain serotonergic system to exert its anorectic effect. Later microdialysis experiments, showing a fenfluramine-induced increase in hypothalamic 5-HT-release, could confirm a predominantly central site of action [29, 30].

In 1977, John Blundell [31] summarised the then existing evidence for 5-HT being involved in feeding. As a general rule, increased availability of 5-HT or a direct activation of 5-HT receptors interfered with food intake whereas reduced availability of the transmitter or receptor blockade could induce feeding. Considering an eminent role for brain 5-HT in the control of satiety, one would expect an impact of brain 5-HT synthesis and metabolism on food intake and satiety. Because 5-HT cannot enter the blood brain barrier, the brain needs to synthesize its own 5-HT. The dietary amino acid tryptophan represents the precursor molecule for 5-HT. While entering the brain, tryptophan competes with large neutral amino acids (LNAA) over the transporter at the blood brain barrier. In fact, it is the tryptophan/LNAA ratio which determines the amount of tryptophan that is available to the brain. Therefore, a protein rich diet, providing abundant amino acids would lower the tryptophan/LNAA ratio, less tryptophan can enter the brain, and as a result 5-HT synthesis would decrease. In contrast, carbohydrates promote the release of insulin which facilitates the uptake of LNAA into peripheral tissues, thus improving the tryptophan/LNAA ratio, facilitating tryptophan entry and 5-HT synthesis [32]. In vivo microdialysis has shown that food intake increases hypothalamic 5-HT release [33-35], but a closer investigation into the contribution of

individual macronutrients to this release revealed that the 5-HT increase is actually due to carbohydrates whereas protein has an opposite effect [36]. Administration of the 5-HT precursor amino acid tryptophan itself also reduces food intake [37]. The first step in 5-HT synthesis is the hydroxylation of tryptophan by tryptophan hydroxylase (Tph) forming 5-hydroxytryptophan (5-HTP). There are two isoforms of the enzyme; Tph1 which is predominantly expressed in the periphery, whereas Tph2 is predominantly expressed in the brain [38]. A Tph2 knockout in mice leads to retarded growth and lower body weight in early postnatal development [39, 40]. An independent study found decreased food intake and bone mass in these mice [41] and the effects on body weight could possibly be gender dependent [42]. The lack of brain 5-HT in conjunction with reduced food intake in Tph2 knockout mice seems to be at odds with the concept of 5-HT as satiety factor in the brain, but as this is a constitutional knockout, further research into developmental and aberrations and compensatory effects is required. The upregulation of uncoupling protein 1 (Ucp1) and increased catecholamine levels [41] in Tph2 knockout mice suggest that metabolic effects including a stimulated thermogenesis contribute to the phenotype. In contrast to these genetic studies, irreversible pharmacological inhibition of tryptophan hydroxylase by p-chlorophenylalanine (pCPA) followed by depletion of brain 5-HT, increases food intake [43]. Peripheral administration of 5-hydroxytryptophan (5-HTP), the intermediate product in 5-HT synthesis, reduces food intake [44]. In a final step of 5-HT synthesis, the enzymatic decarboxylation of 5-HTP generates 5-HT. The hypophagic effect of 5-HTP could not be inhibited with a peripherally acting inhibitor of the enzyme, suggesting that the anorectic effect of 5-HTP involves brain mechanisms [45]. Hunger impacts on brain 5-HT synthesis as food deprivation leads to increased brain tryptophan and synthesis and turnover of 5-HT [46, 47]. The major way of terminating the action of synaptic 5-HT is reuptake into the nerve terminal by a specific 5-HT transporter (5-HTT) mechanism. Pharmacological inhibition of this transporter protein would increase the synaptic availability of 5-HT and inhibit appetite and promote satiety as demonstrated for the uptake inhibitor and releaser fenfluramine [48]. Indeed, selective 5-HT reuptake inhibitors (SSRI) increase synaptic 5-HT and reduce food intake [49] in a behaviourally specific manner [50, 51]. However, genetic manipulation of the 5-HTT in mice brings about contrasting effects. Although the absence of the 5-HTT expectedly increases basal extracellular 5-HT [52], these mice show normal food consumption. However, they develop late-onset obesity, possibly due to reduced locomotor activity and hence diminished energy expenditure [53]. Mice overexpressing 5-HTT have lower 5-HT concentrations in various brain regions including the hypothalamus. The potassium stimulated increase in 5-HT was less in these transgenic mice compared with wild-type mice [54]. These mice are smaller and lighter than their wild-type littermates, but

showed no difference in food intake [55]. In a direct comparison, obesity in 5-HTT knockouts and reduced body weight in mice overexpressing 5-HTT were confirmed [56]. Obesity in 5-HTT knockouts and reduced body weight in 5-HTT overexpressing mice are unexpected findings, considering the pharmacological effects of SSRI on feeding. The contribution of compensatory, possibly peripheral mechanism [57], as discussed for Tph2 knockout mice, cannot be ruled out. Indeed, when fenfluramine was administered to 5-HTT knockout mice, they showed a similar behavioural satiety response and the same reduction in food intake as their wild-type littermates, but knockouts were more prone to unspecific (i.e. other than satiating) effects of d-fenfluramine [55].

### 3. 5-HT receptors and feeding

The increasing availability of selective serotonergic agonists and antagonists was a prerequisite to identify the 5-HT receptors which are involved in the control of food intake. One difficulty with an experimental pharmacological approach to satiety is that many pharmacological manipulations can change food intake without actually influencing appetite or satiety. Animals may stop feeding due to locomotor effects of drugs, drug induced nausea, malaise, but also due to stereotypies or sedation. Since animals cannot report 'true' satiety, behavioural parameters had to be identified which are indicative of a 'normal' feeding behaviour in rodents, covering the behavioural sequence from initiating food intake through satiation to satiety and resting. This behavioural satiety sequence (BSS) is an important tool to interpret drug effects on food intake [58-60]. Drugs that promote satiety will facilitate the transition from eating to other behaviours, in particular resting, but will maintain the normal structure of the BSS. By contrast, appetite stimulating drugs or fasting will delay this transition [61, 62]. Behaviourally specific promotion of satiety can be distinguished from unspecific reduction of food intake using this method, as unspecific drug effects appear as disruption of the normal behavioural profile.

Fenfluramine is a 5-HT releaser and re-uptake inhibitor thus increasing the postsynaptic availability of this neurotransmitter. Investigations into fenfluramine and satiety used initially the racemate dl-fenfluramine, whereas recently the more selective d-fenfluramine [63] is being used. As the effect of d-fenfluramine was thought to be based on the increased availability of postsynaptic 5-HT, it was initially a surprising finding that inhibition of brain 5-HT synthesis with pCPA did not interfere with the hypophagic action of d-fenfluramine. Furthermore, low doses of fenfluramine do not impact on 5-HT release but already induce hypophagia [29, 64]. In addition, d-fenfluramine maintains its hypophagic efficacy even following inhibition of 5-HT release [65]. The d-fenfluramine metabolite d-norfenfluramine

1  
2  
3  
4  
5 binds to the 5-HT<sub>2C</sub> receptor and also reduces food intake. This led to the conclusion that  
6 direct 5-HT<sub>2C</sub> receptor activation contributes to the hypophagic effects of d-fenfluramine [66-  
7 68]. Both dl-fenfluramine and d-fenfluramine accelerate the BSS in the majority of studies  
8 conducted in mice or rats [59, 69-73]. By contrast, two studies found a disruption of the BSS  
9 [74, 75], although this is probably due to differences in methodology [59].  
10  
11

12  
13 The hypophagic effect of fenfluramine requires signalling from different 5-HT receptors. The  
14 identification of these receptors advanced our knowledge how serotonergic systems mediate  
15 satiety. Initial experiments already demonstrated that 5-HT<sub>1A/1B</sub> blockade counteracted the  
16 fenfluramine-induced meal size. By contrast, antagonism of the fenfluramine-reduced eating  
17 rate was achieved by ritanserin, a 5-HT<sub>2A/2C</sub> antagonist. These findings suggested that  
18 fenfluramine-induced eating rate and meal size are controlled by different receptor-subtypes  
19 [76].  
20  
21

22  
23 Early pharmacological investigations into the involvement of 5-HT in satiety used  
24 compounds that differ in their affinity to 5-HT receptor subtypes. These included, among  
25 others, the 5-HT<sub>2C/1B</sub> agonist m-chlorophenylpiperazine (mCPP), now a standard molecule  
26 for investigating the mechanisms of 5-HT induced satiety. However, newer compounds,  
27 although less widely available than mCPP, have a greater selectivity for the 5-HT<sub>2C</sub>  
28 receptor. Others molecules that have been used in dissecting the serotonergic mechanisms  
29 of satiety are the 5-HT<sub>1A/1B</sub> agonist RU-24969, the 5-HT<sub>1B/2C</sub> agonist 1-(3-  
30 trifluoromethylphenyl)piperazine (TFMPP) or the 5-HT<sub>2A/2C</sub> agonist 2,5-dimethoxy-4-  
31 iodoamphetamine [77]. Several studies demonstrated hypophagic effects of  
32 intrahypothalamic administration of mCPP, TFMPP, and DOI [77-79]. However, DOI had an  
33 interruptive effect on feeding and the BSS, mainly by inducing hypoactivity, ruling out the 5-  
34 HT-2A receptor as physiological mediator of satiety [51, 80]. The fenfluramine metabolite  
35 mCPP reduced the feeding rate, but not the duration of feeding [51]. Acceleration of the BSS  
36 by mCPP has been reported both for rats and mice [72, 73, 80], but not in a recent rat study  
37 [81]. Activation of the 5-HT<sub>2C</sub> receptor in mice, using the selective agonist VER2379,  
38 accelerated behavioural satiety, but induced also a reduction of non-food reinforced  
39 appetitive responding [82]. Being complementary to, and in combination with  
40 pharmacological approaches, transgenic techniques provided further evidence for the  
41 involvement of 5-HT<sub>1B</sub> and 5-HT<sub>2C</sub> receptors in physiological satiety. A 5-HT<sub>2C</sub> receptor  
42 knockout was accompanied by hyperphagia leading to hyperglycaemia, insulin resistance  
43 and late-onset obesity [83, 84]. The 5-HT<sub>2C</sub> receptor knockout caused a secondary and  
44 age-dependent reduction in the expression of beta 3 receptors in white adipose tissue,  
45 further enhancing obesity [85]. In many rodent models of obesity, including transgenic  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1  
2  
3  
4  
5 models [86], metabolic dysregulations are leading to obesity. In 5-HT2C knockout mice,  
6 there seems to be a strong behavioural component (hyperphagia) to exist that contributes to  
7 the development of obesity. Heisler [87] report that the mCPP failed to suppress food intake  
8 in 5-HT2C knockout mice and the hypophagic effect of fenfluramine is attenuated,  
9 demonstrating a role for the 5-HT2C receptor in mediating d-fenfluramine-induced satiety  
10 [71]. The 5-HT2 knockout model has been suggested as an in vivo screening model for 5-  
11 HT2C receptor ligands [88].  
12  
13  
14  
15

16  
17 The 5-HT1A/1B agonist RU-24969 reduced the time eating, but not the eating rate. Although  
18 this compound has affinity for both the 5-HT1A and the 5-HT1B receptor, the latter mediates  
19 the hypophagic effect [89, 90]. However, conflicting data regarding the effects of the RU-  
20 24969 on the BSS have been reported [51, 91-93]. Using the more selective compound 5-  
21 HT1B agonist CP-94,253, the hypophagic effect as based on a reduction of meal size, could  
22 be confirmed. This more selective 5-HT1B agonist retained the structure of the BSS,  
23 suggesting an involvement of the 5-HT1B receptor in satiety [91-93]. In 5-HT1B receptor  
24 knock-out mice [94] basal food intake and the feeding response to food deprivation remained  
25 unchanged [95]. In contrast, absolute food intake in these mice was increased when intake  
26 measures were not related to body weight. As a consequence, 5-HT1B receptor deletion led  
27 to increased body weight, but leptin levels remained unchanged and despite the increased  
28 body weight these mice were not deemed obese [96]. In line with the aforementioned  
29 pharmacological findings, both fenfluramine and the 5-HT1A/B agonist RU-24969 lost their  
30 hypophagic effects in these mice, further supporting the role of 5-HT1B receptor in satiety  
31 [73, 95]. Lee [97] studied the effect of the selective 5-HT1B agonist CP-94,253 in 5-HT1B  
32 knockout mice, where the satiating effect of the agonist was absent or reduced. In wild-type  
33 mice, the agonist reduced food intake, but not when pre-treated with the selective 5-HT1B  
34 antagonist SB224289. The antagonist itself stimulated food intake, possibly by disinhibition  
35 of satiety. In line with other reports, these findings provide further evidence that the 5-HT1B  
36 receptor is involved in the mediation of tonic satiety.  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48

49 The majority of experimental studies suggest a role for both the 5-HT1B and the 5-HT2C  
50 receptor in mediating endogenous satiety, but the functional relationship between both  
51 receptor subtypes has been studied only more recently by Dalton [98] in 5-HT2C knockout  
52 mice. In wild-type mice, both the 5-HT2C/1B agonist mCPP and the selective 5-HT1B  
53 agonist CP-94,253 advanced post-prandial behavioural satiety, whereas mCPP was  
54 ineffective in the 5-HT2C knockout mice. However, the 5-HT1B agonist was more effective in  
55 5-HT2C knockouts than in wild-type mice, suggesting a compensatory interaction of both  
56 receptors in the mediation of satiety. Analysing the pharmacological effects of 5-HT agonists  
57  
58  
59  
60  
61  
62  
63  
64  
65

and antagonists on the structure of feeding behaviour led to the conclusion that the stimulation of 5-HT<sub>2C</sub> receptors inhibits the rate of eating and that 5-HT<sub>1B</sub> receptors mediate the duration of feeding. The serotonergic control of feeding would be fully expressed, if both receptors are activated [4, 99].

Non-selective 5-HT<sub>2</sub> agonists reduce food intake, but not in a behavioural specific manner [51, 80, 100], making the 5-HT<sub>2A</sub> and 5-HT<sub>2B</sub> receptor less likely candidates for serotonergic satiety [4]. Although there is broad evidence that antagonists at 5-HT<sub>1B</sub> and 5-HT<sub>2C</sub> receptors can induce feeding, this has not been found in all studies. It is likely that baseline level of food intake impact on the effects of 5-HT<sub>1/2</sub> antagonists on feeding [4, 77].

5-HT<sub>3</sub> receptors are widely distributed both in the brain and in the periphery of the body. In the brain, a particular high density of 5-HT<sub>3</sub> receptors has been found in the brain stem. The 5-HT<sub>3</sub> receptor is pharmacologically an exception among the family of 5-HT receptors, as it is a ligand gated ion channel. In preclinical studies 5-HT<sub>3</sub> antagonists induce anxiolysis, improve cognition and mitigate drug withdrawal [101]. 5-HT<sub>3</sub> antagonists like ondansetron are standard antiemetic drugs and reduce secretion and motility in the gut via central and/or peripheral action [101]. Compared to 5-HT<sub>1B</sub> and 5-HT<sub>2C</sub> receptors, relatively little evidence exists that 5-HT<sub>3</sub> receptors are involved in the serotonergic mediation of satiety. Systemic serotonergic activity has been shown to induce an anorectic response due to eating an amino acid imbalanced diet since activation of 5-HT<sub>3</sub> receptors is required to mediate the response [102]. Other studies provide a complex pattern, as the 5-HT<sub>3</sub> antagonist ondansetron increased the intake of sweetened mash, but reduced sucrose intake [103]. Van der Hoek and Cooper [104] revealed a behaviourally specific reduction of palatable food consumption in non-deprived rats following peripheral administration of the selective 5-HT<sub>3</sub> antagonist ondansetron. However, ondansetron did not alter sucrose or chow intake in food deprived rats in a later study, but blunted the anorectic response to a duodenal lipid infusion [105]. The findings of both studies are not necessarily contradictory, as one could assume in both situations a disinhibition of satiety, rather than a satiating effect per se. As ondansetron does not readily cross the blood brain barrier [106], one would assume that these effects on satiety are predominantly peripheral effects. However, antagonism of 5-HT<sub>3</sub> receptors in the nucleus of the solitary tract (NTS) of the brainstem stimulate food intake, indicating also a central site of action [107]. In contrast to these findings in rats, however, no change in food intake has been found in 5-HT<sub>3A</sub> receptor knockout mice. As there is relatively little evidence for an independent role of 5-HT<sub>3</sub> receptors in satiety [108], the prevailing interest in 5-HT<sub>3</sub> receptor antagonists is still the reduction of nausea and vomiting during chemotherapy.

1  
2  
3  
4  
5  
6  
7  
8 The 5-HT6 receptor is almost exclusively expressed in the brain, although it has also been  
9 found in peripheral tissues of various species. The widely expression in the brain includes  
10 the hypothalamus, although the quantification of the expression depends on species and  
11 detection method [109]. Initial experiments did not provide any evidence for the involvement  
12 of the 5-HT6 receptor in satiety [110]. Studies using either intracerebroventricular injections  
13 of 5-HT6 antisense oligonucleotides or intraperitoneal administration of the 5-HT6 receptor  
14 antagonist Ro 04-6790 found decreased feeding behaviour and body weight gain  
15 [111],[112]. The 5-HT6 partial agonist E-6837 induced hypophagia in a rat model of diet-  
16 induced-obesity (DIO) [113], and mice carrying a non-functional 5-HT6 receptor do not  
17 become obese when exposed to a high fat diet [114]. In the latter study, however, there was  
18 no change of habitual feeding on a normal diet in 5-HT6 receptor knockout mice. The  
19 interpretation of these findings should be seen with some caution, as the central distribution  
20 of this receptor is different in mice compared to rats and humans [115]. Whereas 5-HT2C  
21 and 5-HT1B receptor activation induces satiety, it requires inactivation of the 5-HT6 receptor  
22 to reduce food intake, suggesting that different or additional pathways are involved. It has  
23 been hypothesised that 5-HT6 receptors act at GABAergic interneurons in the  
24 hypothalamus. These GABAergic neurones would synapse at pro-opiomelanocortin (POMC)  
25 neurones which release the anorectic peptide  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH).  
26 Antagonists at 5-HT6 receptors could interfere with the GABAergic inhibition and thus  
27 indirectly stimulate  $\alpha$ -MSH, leading to increased satiety [109]. In a mapping study, using  
28 anorectic doses of the 5-HT6 receptor antagonist SB-399855, Garfield [116] detected  
29 increased c-fos immunoreactivity in the hypothalamic paraventricular nucleus (PVN) (but not  
30 in the arcuate nucleus) and the nucleus of the solitary tract (NTS) of the brain stem. This  
31 finding would also exclude a hypothalamic effect similar to 5-HT2C and 5-HT1B agonists  
32 which both have a direct effect in the arcuate nucleus. However, an indirect GABA-mediated  
33 effect as suggested by Woolley et al.[109] would be a possibility. A further mechanism could  
34 be hypothesised that involves the NTS. The increased 5-HT6 receptor expression in the  
35 NTS, a target for peripheral satiety signals, would allow 5-HT6 antagonists to disinhibit  
36 peripheral satiety signals which terminate in the NTS [116].  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52

53  
54 Relatively little is known about the 5-HT4 receptor in satiety, although it could possibly be  
55 involved in stress-induced eating behaviour [117] or reward processing in obese subjects as  
56 an imaging study suggests that 5-HT4 receptor activation occurs in reward circuits (nucleus  
57 accumbens and ventral pallidum). The intensity of signals coming from these two regions  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4  
5 correlated with the body mass index [118]. No evidence exists that that 5-HT5 and 5-HT 7  
6 receptors are involved in satiety.  
7

8  
9 In contrast to the aforementioned 5-HT1B and 5-HT2C agonists and 5-HT6 antagonists, 5-  
10 HT1A agonists do not promote satiety. By acting at somatodendritic autoreceptors in the  
11 raphe nuclei [11, 119-121] the prototypic 5-HT1A agonist 8-hydroxy-2-(di-n-  
12 propylamino)tetralin (8-OH-DPAT) stimulates feeding in satiated rats. The underlying  
13 mechanism is agonism at somatodendritic autoreceptors located on 5-HT neurons in the  
14 dorsal raphe, thus inhibiting 5-HT release [122-128]. This has also been shown for other  
15 agonists at this receptor [77]. Although these effects are behaviourally specific [129], they  
16 follow a circadian pattern [125, 130] and have not been observed in food deprived rats  
17 where 5-HT1A agonists inhibit food intake upon re-feeding [123, 131, 132]. This anorectic  
18 effect of the 5-HT1A effect in food deprived rats is due to stereotypy and various  
19 accompanying signs of the classical 5-HT syndrome [51, 133]. However, when given locally  
20 into the paraventricular nucleus (PVN) of the hypothalamus, 8-OH-DPAT advanced the BBS  
21 [134], possibly by acting at postsynaptic receptors.  
22  
23  
24  
25  
26  
27  
28  
29

30 By facilitating negative feedback, the 5-HT1A agonist 8-OH-DPAT attenuates 5-HT release  
31 also under in-vivo conditions, as measured by microdialysis, but varying across brain regions  
32 [135-137]. In the lateral hypothalamus, peripherally administered 8-OH-DPAT diminished 5-  
33 HT release in rats fed ad libitum, but not in food deprived rats. These latter findings  
34 correspond to the aforementioned effects on food intake, confirming that the impact of 5-  
35 HT1A receptor activation on food intake depends on the nutritional status [132, 138]. A  
36 knockout of this receptor leads to reduced food intake [41]. Very likely, the knockout will  
37 diminish or eliminate the negative somatodendritic feedback and thus increase 5-HT release  
38 that will induce hypophagia via postsynaptic 5-HT2C and 5-HT1B receptor activation,  
39 although no change in habitual feeding in 5-HT1A receptor knockout mice has been  
40 displayed in another study [139].  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50

#### 51 **4. Peripheral 5-HT and satiety**

52

53 The vast majority of the 5-HT in the body is found in the gastrointestinal (GI) tract where it is  
54 stored in enterochromaffin cells. Enterochromaffin cells function as sensory transduction  
55 elements in the gastrointestinal mucosa, responding to chemical and mechanical stimuli by  
56 releasing 5-HT and other potential mediators onto afferent nerve terminals to initiate GI  
57 reflexes and modulate visceral perception [140]. 5-HT will be released after food intake or  
58  
59  
60  
61  
62  
63  
64  
65

intraluminal distension or efferent vagal stimulation. Its primary targets are the mucosal projections of primary afferent neurons including the vagal nerve [141]. 5-HT<sub>4</sub> agonists and 5-HT<sub>3</sub>-antagonists have been used and are being used in a variety of gastrointestinal disorders and chemotherapy-induced nausea [141-144].

However, despite the abundance of 5-HT in the periphery in comparison to the brain, relatively little is known about the involvement of peripheral 5-HT in satiety. A possible explanation could be the fact that the role of brain 5-HT is well established and these mechanisms can be targeted more selectively if the involved receptor subtypes (i.e. 5-HT<sub>2C</sub> or 5-HT<sub>6</sub>) are only or predominantly expressed in the brain. Peripherally administered 5-HT cannot cross the blood brain barrier, but nevertheless induces hypophagia [145, 146]. Peripherally injected 5-HT advanced the BSS, and rats approached satiation quicker and in a behaviourally specific manner [147, 148]. Peripheral 5-HT requires the simultaneous action of gastrointestinal mechanism to elicit a complete behavioural profile of satiety as shown with sham feeding experiments [148]. Peripherally acting 5-HT<sub>1</sub> agonists like 5-carboxamidotryptamine (5-CT) and the 5-HT<sub>2</sub> agonist  $\alpha$ -methyl-5-hydroxytryptamine (5- $\alpha$ -Me-5-HT) also induce hypophagia [149]. The involved 5-HT receptor subtypes need to be identified, as pharmacological studies ruled out 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptors [4, 99]. The sites where exactly peripheral acting 5-HT agonists impact on satiety still require identification, although for 5-HT<sub>2</sub> agonists the pylorus is a candidate [150]. Tph1 deficient mice show normal brain, but low peripheral 5-HT levels [38]. Tph1 knockout showed higher food intake, but unchanged locomotor activity, and gained more body weight. The satiating effect of systemically administered 5-HT was increased in Tph1 knockout mice suggesting adaptive changes in peripheral 5-HT receptors. By contrast, systemic fenfluramine had similar effects both in knockouts and wild-type mice [151]. These data highlight the importance of peripheral 5-HT for the full expression of satiety, but also suggest that peripheral 5-HT depletion does not necessarily lead to a compensatory change in central 5-HT<sub>2C</sub> or 5-HT<sub>1B</sub> mechanism of satiety as shown by the similar effects of fenfluramine in both genotypes.

## 5. Brain mechanisms of serotonergic satiety

The brain mechanisms involved in the control of feeding and satiety are complex, both with regard to the brain structures being involved, but also regarding the participating neurotransmitters and hormones. Serotonergic mechanisms of satiety have been identified in the hypothalamus but also in extrahypothalamic brain structures as for example in the

1  
2  
3  
4  
5 brain stem. Within the hypothalamus, research has been focused lately on the arcuate  
6 nucleus, the PVN and the lateral hypothalamic area (LHA) [152-155].  
7

8  
9 One of the strongest brain stimulators of food intake is Neuropeptide Y (NPY) [156]. NPY  
10 neurons of the hypothalamic arcuate nucleus project directly to the lateral hypothalamus  
11 where they activate both melanin concentrating hormone (MCH) and orexin neurons and this  
12 activation causes feeding, when the arcuate nucleus is stimulated. Projections from the  
13 arcuate NPY neurons to the PVN seem to mediate metabolic functions of NPY [157]. Both 5-  
14 HT1B and 5-HT2C receptors are expressed in the arcuate nucleus [158-160], which receives  
15 input from raphe nucleus neurons [161-163]. Chronic systemic administration of dl-  
16 fenfluramine decreased hypothalamic NPY expression, being consistent with the anorectic  
17 effects of fenfluramine [164]. NPY cell bodies also synthesise agouti-related protein (AgRP)  
18 that increases appetite and decreases metabolism. 5-HT1B agonists hyperpolarise these  
19 neurons via Gi-coupled 5-HT1B receptors [165].  
20  
21  
22  
23  
24  
25  
26

27 The selective 5-HT2C receptor agonist BVT.X reduces acute food intake in both genetic and  
28 diet-induced mice models of obesity following systemic administration. A 7-day infusion of  
29 the agonist via osmotic mini pumps significantly increased POMC mRNA and reduces body  
30 weight in these mice. However, mice lacking the melanocortin (MC) 4 receptor, did not show  
31 the 5-HT2C agonist-induced hypophagia [165]. This latter finding demonstrates that  
32 melanocortins acting on MC4 receptors are a requisite downstream pathway for 5-HT2C  
33 receptor agonists to exert effects on food intake [166]. However, a more recent study,  
34 likewise using MC4 knockout mice, but also MC3 knockout mice, found rather evidence that  
35 MC3 receptors are the more likely candidate [167]. This could be due to differences in study  
36 design, and therefore either receptor could impact on feeding, but under different feeding  
37 conditions. Further studies would be required to solve these discrepancies. Nevertheless,  
38 the role of downstream MC3/MC4 receptors is further supported by pharmacological studies  
39 in rats where MC3/MC4 receptor antagonism attenuates the hypophagic effect of d-  
40 fenfluramine [159]. Hypophagia induced by MC3/MC4 receptor activation is not different  
41 between 5-HT2C knock out and wild-type mice, further suggesting that the serotonergic  
42 effects are upstream of MC3/MC4 receptors [159]. Heisler et al.[168] suggest a model for d-  
43 fenfluramine mediated satiety (or 5-HT2C receptor mediated satiety) which includes the  
44 activation of POMC neurons in the arcuate nucleus via 5-HT2C receptors. POMC is a  
45 precursor for the anorectic protein  $\alpha$ -MSH, and 5-HT also causes a direct release of  $\alpha$ -MSH  
46 from hypothalamic slices [169]. Alpha-MSH is an endogenous MC receptor agonist. The 5-  
47 HT2C receptor mRNA is co-expressed with up to 80% of  $\alpha$ -MSH containing neurons in the  
48 arcuate nucleus and the 5-HT2C/1B agonist mCPP increases Fos-immunoreactivity in this  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4  
5 hypothalamic region [159]. Therefore the model further suggests that the 5-HT<sub>2C</sub> receptor  
6 mediated  $\alpha$ -MSH release leads to a downstream activation of MC<sub>3</sub>/MC<sub>4</sub> receptors [168].  
7 There is recent evidence that 5-HT<sub>1B</sub> receptors inhibit the activity of NPY/AgRP neurons in  
8 the arcuate nucleus [170, 171]. In conclusion, it appears that both 5-HT<sub>2C</sub> and 5-HT<sub>1B</sub>  
9 receptors are involved in arcuate control of feeding, where they act via different, but  
10 complementary mechanisms [170]. In the light of the discussion of possible 5-HT and leptin  
11 interactions in satiety it is important to mention that leptin and 5-HT are activating distinct  
12 POMC neurons in the arcuate nucleus [171], further supporting the view that there is no  
13 direct interaction between 5-HT and leptin at the neuronal level.  
14  
15  
16  
17  
18  
19

20 An important hypothalamic structure in the control of feeding is the PVN where 5-HT and  
21 various agonists reduce food intake by decreased meal size and eating rate [4, 172]. This  
22 pattern is very similar to the effects observed following systemic administration of 5-HT  
23 agonists but different from effects as observed after selective stimulation of peripheral  
24 receptors [4]. The PVN receives input from the arcuate nucleus [173, 174] and also  
25 accommodates MC receptors [175-177]. Local administration of MC<sub>3</sub>/MC<sub>4</sub> agonists into the  
26 PVN reduces food intake whereas antagonists administered into the PVN increase food  
27 intake [178-180]. Although injections of 5-HT<sub>1B</sub> agonists into the PVN reduce food intake,  
28 blockade of the PVN 5-HT<sub>1B</sub> receptors or lesions of this region did not inhibit the satiating  
29 effects of systemic d-fenfluramine [181, 182]. These and other findings suggest that  
30 activation of 5-HT<sub>1B</sub> receptors located in the PVN is sufficient but not necessary to induce  
31 satiety [4]. The selective 5-HT<sub>1B</sub> agonist CP-94,253 stimulated c-fos immunoreactivity in the  
32 PVN and the ventromedial nucleus (VMN) and in various other brain structures including  
33 hindbrain structures after satiating doses, suggesting the involvement of extrahypothalamic  
34 receptors in the 5-HT<sub>1B</sub> mediation of satiety [97].  
35  
36  
37  
38  
39  
40  
41  
42  
43

44 NPY/AgRP and POMC neurons also project from the arcuate nucleus to the LHA where they  
45 control the synthesis of MCH and orexin which both stimulate food intake [183-185]. Orexin  
46 neurons are located specifically in the LHA and the LHA projects to almost all parts of the  
47 brain [186, 187], in particular to serotonergic raphe neurons [187-189]. In the LHA, 5-HT<sub>1A</sub>  
48 receptor immunoreactivity was observed in MCH- and orexin-containing neurons, suggesting  
49 that 5-HT, via postsynaptic 5-HT<sub>1A</sub> receptors, affects the release of these orexigenic  
50 peptides [190]. The 5-HT<sub>2C</sub> agonist mCPP lost its hypophagic potency when both POMC  
51 and orexin, were silenced. Silencing either one only had no effect. Hence, a functional  
52 hypothalamic POMC and orexin activity is a prerequisite for 5-HT<sub>2C</sub> receptor mediated  
53 satiety [191].  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4  
5 Another important appetite stimulant, released from the stomach is ghrelin [192]. As an  
6 important short-term hunger signal, ghrelin is involved in the initiation of a meal [154, 155,  
7 193, 194]. Ghrelin is also released in the brain, in particular in the arcuate nucleus [195,  
8 196]. Peripheral ghrelin targets the arcuate nucleus where ghrelin receptors are expressed  
9 on AgRP neurons [197]; raising the possibility that 5-HT could be a physiological counterpart  
10 of ghrelin or vice versa. Ghrelin mimics the effect of NPY in the PVN [196] and an injection of  
11 5-HT into the PVN inhibits the orexigenic effect of ghrelin, also administered into the PVN  
12 [198]. Recently, a novel heterodimer between the ghrelin receptor (GHS-R1A) and the  
13 unedited 5-HT<sub>2C</sub> receptor has been identified. Dimerization of GHS-R1A receptor with the  
14 unedited 5-HT<sub>2C</sub> receptor reduced the GHS-1RA receptor-mediated calcium influx [199].  
15 This finding not only provides further evidence for interactions between 5-HT ghrelin, but  
16 suggests that these interactions could contribute to the fine-tuning of appetite and satiety.  
17  
18  
19  
20  
21  
22  
23  
24  
25

26 Whereas historically efforts focussed on revealing hypothalamic mechanisms of appetite and  
27 satiety, it became increasingly clear that numerous endocrine and neural factors are  
28 integrated into a complex network of many brain structures [152]. Among those brain  
29 structures hindbrain nuclei, and here the nucleus of the solitary tract (NTS) which is located  
30 in the medulla, play a dominant role. The NTS provides a target in particular for satiety  
31 signals of peripheral origin. The NTS receives afferent input from hypothalamic nuclei, blood  
32 born signals (leptin, ghrelin, glucose) and from the GI tract via the vagus (cholecystokinin;  
33 CCK, peptide YY; PYY and others) [153, 200]. Afferent projections from the rostral NTS  
34 reach the parabrachial nucleus at the junction of the midbrain and pons and at the  
35 hypothalamus, whereas the caudal NTS projects to vagal efferent neurons control  
36 parasympathetic gastrointestinal responses including insulin secretion and gastric emptying  
37 [153]. Decerebrate rats which lack the direct connection between the hindbrain and forebrain  
38 show a reduced feeding response to intra-oral infusion of 12.5% glucose following systemic  
39 administration of fenfluramine or mCPP [201-203]. As Decerebrate rats can only show  
40 behavioural responses controlled by brain stem circuits, these results show that caudal  
41 brainstem receptors are sufficient to produce anorectic effects after systemic administration  
42 of mCPP or fenfluramine. Administration of the 5-HT<sub>2C/2A</sub> antagonist metergoline  
43 completely blocked the anorectic effects of systemic mCPP in this rat model. This  
44 demonstrates that caudal brainstem 5-HT receptors (most likely 5-HT<sub>2C</sub> receptors) are not  
45 only sufficient, but also required to produce anorectic effects of mCPP [203].  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57

58 Food intake (or experimental volume distension) leads to 5-HT secretion from gastric  
59 enterochromaffin cells. This effect is largely relayed by the vagus to the NTS and mediated  
60  
61  
62  
63  
64  
65



1  
2  
3  
4  
5 by 5-HT<sub>3</sub> receptors which are expressed on peripheral dendritic terminals of vagal afferents  
6 innervating the stomach [107, 204-207]. High densities of 5-HT<sub>3</sub> receptor binding sites within  
7 the central nervous system have also been found in the NTS [208, 209] and local  
8 administration of the 5-HT<sub>3</sub> antagonist odansetron increased sucrose intake [107]. After  
9 intestinal anaphylaxis, increased c-fos staining was observed in the NTS, the parabrachial  
10 nucleus, and the hypothalamic PVN [210]. A systemic administration of the 5-HT<sub>3</sub> antagonist  
11 odansetron attenuated this intestinal effect on brain c-fos expression, suggesting a functional  
12 connection of these structures and 5-HT in the efferent control of intestinal disturbances  
13 [210]. Caudal serotonergic neurons of the NTS control the excitability of the parabrachial  
14 nucleus and inhibit feeding [211]. The parabrachial nucleus is connected to the  
15 hypothalamus [212]. Ablation of hypothalamic AgRP neurons leads to aberrant activation of  
16 the parabrachial nucleus and starvation. The parabrachial nucleus has been identified,  
17 therefore, as a functional unit that integrates feeding related signals from several brain  
18 regions [211, 213] and could possibly provide a functional link between hypothalamic and  
19 hindbrain mechanisms of 5-HT mediated satiety. 5-HT<sub>1B</sub> receptor activation in the  
20 parabrachial nucleus reduces food intake [214] and the hypophagic effect of fenfluramine in  
21 the parabrachial nucleus requires 5-HT<sub>B</sub> receptor activation [215].  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34

## 35 **6. 5-HT interactions with CCK and leptin**

36  
37 It is important to emphasize that 5-HT does not work in isolation from other satiety signals.  
38 Considering the neuroanatomy of the serotonergic system, the distribution of 5-HT receptors  
39 and the involvement of brain serotonin in much behaviour, it seems likely that 5-HT, although  
40 having an effect on satiety on its own, could potentially interact with other satiety  
41 mechanisms. Feeding (i.e. energy intake) is essential for survival and this can be secured by  
42 an adaptive control by several mechanisms which would allow the activation of  
43 compensatory responses in case a single mechanism is malfunctioning. Interactions of  
44 satiety mechanisms increase the flexibility and plasticity of the system and should allow  
45 adaptation to different requirements during development (e.g. growth, reproduction, age). In  
46 fact, the probably first candidate for such an interaction that has been investigated is CCK  
47 [216]. Back in 1973, Gibbs et al.[217] already suggested that CCK released from the small  
48 intestine during a meal contributes to termination of the meal and induces postprandial  
49 satiety. Exogenous CCK inhibits food intake in rats in a behaviourally specific manner. Loss  
50 of the CCK-1 receptor in rats due to a spontaneous mutation [218] leads to hyperphagia and  
51 subsequently to obesity [219], although obesity does not develop in CCK-1 receptor knock-  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4  
5 out mice [220]. The prospect of 5-HT-CCK interactions in the control of satiety was primarily  
6 based on pharmacological data. For example, the non-selective 5-HT<sub>1/2</sub> receptor antagonist  
7 metergoline not only attenuated the satiating effect of fenfluramine, but also that of  
8 exogenous CCK. However, neither fenfluramine- nor CCK- induced satiety were affected by  
9 only peripheral 5-HT antagonism with xylamidine [221, 222] *et al.* 1989). Evidence has been  
10 provided that 5-HT<sub>2C</sub> receptor activation would be required for CCK to induce satiety [223].  
11 One could conclude from these results that (exogenous) CCK requires central 5-HT to  
12 induce satiety. Indeed, inhibition of brain 5-HT synthesis attenuates the satiating effect of  
13 CCK [224]. A pharmacologically induced attenuation of central 5-HT release by the 5-HT<sub>1A</sub>  
14 agonist 8-OH-DPAT also counteracts CCK-induced satiety [225, 226], although this has not  
15 been found by Ebenezer and Brooman [227]. Interestingly, antagonism of 5-HT<sub>1A</sub> receptors  
16 with WAY 100135 also attenuated CCK induced satiety, most likely by antagonism at  
17 postsynaptic 5-HT<sub>1A</sub> receptors, an effect that could possibly explained by an increased 5-HT  
18 release following CCK administration [226]. This hypothesis has been confirmed in a later  
19 microdialysis study in food deprived rats which demonstrated that exogenous CCK facilitates  
20 hypothalamic 5-HT release [35]. This would also be in keeping with earlier *in vitro*  
21 experiments demonstrating excitatory effects of CCK on serotonergic neurons in the dorsal  
22 raphe nucleus [228]. When CCK is administered peripherally to 5-HT<sub>2C</sub> receptor knockout-  
23 mice it has no hypophagic effect, adding evidence to previous pharmacological studies  
24 demonstrating that this 5-HT receptor is involved in the mediation of CCK-induced satiety  
25 [229]. Together these indicate that CCK recruits central 5-HT to induce satiety.

26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39 Cooper *et al.* [230] took the opposite approach and investigated if 5-HT induced satiety  
40 would require CCK activity. They and others [231] demonstrated that the CCK<sub>1</sub>-receptor  
41 antagonist devazepide blocked the satiating effect of systemic 5-HT or fenfluramine,  
42 whereas the CCK<sub>2</sub>-receptor antagonist L-365.260 was ineffective. Cooper and Dourish [216,  
43 232] concluded that the CCK<sub>1</sub> receptor is involved in the anorectic effects of fenfluramine as  
44 it facilitates the satiating effect of CCK, resulting in an overall increased satiety. Other  
45 studies, however, did not find an attenuation of 5-HT or fenfluramine induced behavioural  
46 satiety by devazepide [233, 234]. Although devazepide did not impact on fenfluramine-  
47 induced satiety in one of these studies, it notably attenuated fenfluramine-induced  
48 suppression of gastric emptying suggesting a peripheral interaction [234]. In a somewhat  
49 different approach Voigt *et al.* [235] used protease inhibitors to increase the concentration of  
50 endogenous CCK instead of blocking CCK receptors, and hence CCK activity, by  
51 antagonists. Meal-induced CCK release is limited by proteases, and inhibition of these  
52 proteases should therefore induce hypophagia [236]. Although the protease inhibitors and  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

fenfluramine reduced night time feeding, when given separately, no evidence for additive or synergistic effects was found when the compounds were administered in combination.

In the lateral hypothalamus, however, neurons have been identified that respond both to iontophoretic 5-HT and CCK administration, and the response increases with a combined application of both satiety signals, suggesting that the effect of 5-HT and CCK can converge on the same neuron [237]. Both CCK [238] and 5-HT inhibit feeding when injected into the PVN, and lesioning the PVN impairs the hypophagic effect of peripherally administered CCK [239]. Synergistic effects of 5-HT and CCK in the PVN seem also to impact on the motivation to eat [240].

Although there is evidence to support the interactive model for 5-HT and CCK as proposed by Cooper and Dourish [216], more of the data are in favour of CCK requiring 5-HT to mediate satiety rather than 5-HT requiring CCK, although the latter cannot be excluded yet. More recently, further evidence for 5-HT-CCK interactions emerged from studies investigating the involvement of the 5-HT<sub>3</sub> receptor. Compared to 5-HT<sub>1</sub>, 5-HT<sub>2</sub> and 5HT<sub>6</sub> receptors, relatively little data was in favour of 5-HT<sub>3</sub> receptors being specifically involved in satiety. This did not encourage investigations of potential interactions between 5-HT<sub>3</sub> receptors and the CCK system [241]. The first evidence for this arose from a rat study on hypophagia as consequence of eating an amino acid imbalanced diet. This hypophagic response is remediated by the 5-HT<sub>3</sub> antagonist tropisetron. The effect of tropisetron was blunted by the CCK<sub>1</sub> receptor antagonist devazepide, suggesting a possible interaction between CCK and 5-HT in anorexia due to aminoprivic feeding [102]. Inducing satiety by intraduodenal fat infusion could only fully be blocked when the CCK-1 antagonist devazepide and the 5-HT<sub>3</sub> antagonist tropisetron were administered together [242]. Such interactions between CCK and 5-HT<sub>3</sub> antagonists have later been shown for intake of sucrose solution and also of a solid diet [206, 243, 244]. In these experiments, CCK and 5-HT synergistically reduced food intake in a supra-additive manner, suggesting that CCK and 5-HT together bring about a stronger satiety signal than each system alone [245]. Data from the same group shows that the effects of the 5-HT<sub>3</sub> antagonist depends on gastric signals, as gastric distension is required and the CCK antagonist, in contrast to CCK itself, is ineffective in sham-fed rats [206]. The involvement of hindbrain 5-HT<sub>3</sub> receptors in CCK-induced satiation has been demonstrated [107]. These studies, using 'classical approaches' add to the proposed model of interdependent CCK and 5-HT mediated satiety [216], but also emphasise the need not to limit the study of serotonin-mediated satiety to the central nervous system [241].

1  
2  
3  
4  
5 Despite early assumptions of a role of adipose tissue in control of feeding [246] and the  
6 existence of circulating satiety factors [247], only the cloning of the Ob-gene (Ob stands for  
7 obesity) [248] could explain why mice with a deficiency of this gene developed severe  
8 obesity. The Ob-gene codes for a protein that has been later called leptin (Greek; leptos =  
9 thin). Leptin is produced in adipose tissue and blood concentrations of leptin are high in  
10 obese individuals. Leptin crosses the blood brain barrier via a transport mechanism linked to  
11 the leptin receptor [249]. Null-mutation of the Ob-gene causes severe obesity in mice (*ob/ob*  
12 mice) as does the mutation of the leptin receptor (*db/db* mice) [250, 251]. Exogenous leptin  
13 reduces body weight in *ob/ob*-mice down to the level of wild-type control mice [252-255]. By  
14 contrast, leptin administration had no effect in *db/db* mice. The hypothalamic actions of  
15 peripheral leptin are dependent on other hypothalamic signalling systems of hunger and  
16 satiety. Leptin inhibits hunger signals like NPY and AgRP in the hypothalamic arcuate  
17 nucleus but also stimulates POMC which leads to the formation of the satiety signal  $\alpha$ -MSH.  
18 Projections from the arcuate nucleus regulate food intake via MCH and orexins. Synergistic  
19 effects between leptin and CCK in the control of food intake have been described [183-185].  
20 The role of 5-HT-leptin interactions in the control of food intake appears to be less clear  
21 though. Halford and Blundell [256] found little evidence for a direct link between leptin and  
22 5HT in appetite control and have therefore suggested that both leptin and 5-HT represent  
23 separate pathways in the control of food intake. The authors emphasised the concept that  
24 the effects of leptin are rather long lasting (tonic) whereas 5-HT is part of a network for short  
25 acting satiety signals (episodic). The relative independence of leptin and 5-HT is supported  
26 by findings that 5-HT<sub>2C</sub> knockout mice are hyperphagic but their response to exogenous  
27 leptin remains unchanged, although these mice, once being obese, become partially leptin  
28 resistant [84]. However, a later pharmacological study using the 5-HT<sub>2C</sub> receptor antagonist  
29 SB 242084, provided evidence for the involvement of 5-HT<sub>2C</sub> receptors in the mediation of  
30 leptin-induced anorexia [257, 258]. Knockout of 5-HT<sub>2C</sub> receptors in *ob/ob*-mice further  
31 exacerbates obesity [259].

32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48 Peripheral administration of the 5-HT precursor 5-HTP increases serum leptin in mice [260,  
49 261] although it needs to be determined if the involvement of 5-HT in leptin-induced  
50 hypophagia is a direct effect because a further study suggested that hyperleptinemia  
51 following systemic injection of 5-HTP is elicited by 5-HT formed in the peripheral system  
52 [262]. However, immunohistochemical evidence suggests an inverse relationship between 5-  
53 HT and leptin in the dorsal raphe and the hypothalamus. Pharmacological depletion of 5-HT  
54 synthesis and release led to increased leptin immunoreactivity in this brain region [263]. An  
55 impact on feeding behaviour has not been investigated in this study though. Nevertheless  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

one could speculate that the central depletion of 5-HT would reduce (serotonergic) satiety and this could be functionally compensated by increased leptin uptake into the brain. Such an interpretation would require experimental verification as it would be at odds with findings in 5-HTT deficient mice with increased 5-HT in various brain regions where reduced food intake was paralleled by increased leptin levels [264], although the change in leptin concentrations could be a secondary and independent effect.

Whereas all these studies investigated into the impact of serotonergic manipulations on leptin, mostly in the context of food intake, other studies took the reverse approach and looked into serotonergic mechanisms following manipulation of leptin. This approach has some limitations, mainly in that suitable pharmacological antagonists of the leptin receptor have not been widely tested with regard to satiety [265]. Therefore this aspect of putative interactions has been largely studied in leptin deficient mice. In *ob/ob* mice, leptin infusion via osmotic mini pumps reduced food intake and body weight, and increased hypothalamic and brain stem 5-HT concentrations, but not 5-HT concentration in the frontal cortex. Interestingly, this was not observed in lean mice, suggesting enhanced leptin sensitivity in *ob/ob* mice [110]. A recent study by Schellekens [266] analysed the impact of leptin deficiency on 5-HT receptors being involved in the control food intake. The authors found increased hypothalamic 5-HT<sub>1A</sub> receptor expression as well as increased hippocampal 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub>, and 5-HT<sub>6</sub> receptor mRNA expression in obese mice compared to lean control mice. In addition they found decreased hypothalamic and hippocampal 5-HT-turnover, a complementary finding to the earlier observed stimulatory effect of leptin on brain 5-HT turnover [267].

Both leptin and endogenous 5-HT inhibit NPY [170, 183] which could be a common endpoint of their actions. The midbrain raphe projects to the hypothalamic arcuate nucleus where both NPY and POMC neurons are thought to be involved in the serotonergic control of satiety [168, 268]. Cells in the raphe of female pigtailed macaque express 5-HT transporter mRNA, which also serves as a marker of serotonergic neurons, and leptin receptor mRNA, suggesting that leptin may act on serotonergic cells to mediate some of its effects on ingestive behaviour and metabolism [269]. 8-OH-DPAT-induced stimulation of 5-HT<sub>1A</sub> receptors in nucleus raphe pallidus inhibits leptin-induced increases in brown adipose tissue energy expenditure [270]. It has been suggested that serotonergic neurons of the dorsal raphe can uptake leptin following its intracerebroventricular administration [271]. The physiological relevance of this finding needs to be determined, though, as pharmacological studies into the hypophagic effects of centrally administered leptin gave inconsistent results [272-276]. Leptin receptor immunoreactivity has also been identified in ascending

serotonergic neurons [277]. Decreased 5-HT transporter mRNA in neurons of the dorsal raphe nucleus in ob/ob mouse [278] seems to be in line with a rather direct 5-HT-leptin connection. In obese Zucker rats with a mutation in the leptin receptor [279-281] there is hyperexcitability of raphe neurons by leptin in an early developmental stage [282]. In normal rats, intracerebroventricular leptin aggravates the feeding-induced release of 5-HT in the LH [283].

The role of the dorsal raphe in 5-HT interactions has investigated by [41]. Using transgenic techniques they eliminated leptin receptors from serotonergic neurons in the dorsal raphe. This led to increased food intake, body fat, and body weight. This would be in line with most of the studies assuming a stimulatory effect of leptin on 5-HT, or even a recruitment of central 5-HT to induce satiety. However, [41] proposed that 5-HT is largely an orexigenic signal and that leptin-induced hypophagia is mediated by suppressing of activity of 5-HT neurons. Therefore, a lack of inhibition would increase extracellular 5-HT (and therefore appetite) and this effect could be blocked by the 5-HT<sub>1A</sub> antagonist LY426965. Because this antagonist deviates pharmacologically from other 5-HT<sub>1A</sub> antagonists, non-selective effects cannot be fully ruled out without further pharmacological studies [284]. Most importantly, however, there is no evidence so far that LY426965 reduced food intake in a behaviourally specific manner. 5-HT<sub>1A</sub> receptor pharmacology of appetite is complex, as shown with agonists, and net effects on feeding are qualitatively dose dependent and need to take into account effects on both autoreceptors and postsynaptic receptors. Finally, considering 5-HT as an orexigenic factor in mammals would be at odds with preclinical and clinical data.

Regardless of the interpretation of the previous studies [41, 285] and further studies [257, 258, 267], other studies do not provide evidence for direct 5-HT-leptin interactions. In an extensive study, using several mouse lines Lam et al. [286] tried to resolve these discrepancies and aimed to clarify if serotonergic neurons are directly involved in the metabolic effects of leptin. The main outcomes of this study were that, albeit some leptin receptor neurons lie close to 5-HT neurons in the dorsal raphe nucleus, 5-HT neurons do not express these receptors. While leptin hyperpolarizes some non-5-HT dorsal raphe neurons, leptin does not alter the activity of dorsal raphe 5-HT neurons. Furthermore, 5-HT depletion did not impair the anorectic effects of leptin. The serotonin transporter-cre allele (Sert(cre)) is expressed in 5-HT (and developmentally in some non-5-HT) neurons. While Sert(cre) promotes leptin receptor excision in a few leptin receptor neurons in the hypothalamus, it is not active in dorsal raphe receptor neurons, and neuron-specific Sert(cre)-mediated leptin receptor inactivation in mice does not alter body weight or induce adiposity. Thus, leptin does not directly influence 5-HT neurons and does not modulate important appetite-related

determinants via 5-HT neuron function [286]. Because this study could not confirm the previous reports by Yadav [41], and because species differences (primates in [269]) cannot be excluded, further research is required to investigate possible interactions in other brain regions and in the periphery. These studies should integrate models of diet induced obesity at different stages of obesity as possible interactions between leptin and 5-HT may change during development. Even if it remains controversial if 5-HT interacts with leptin at the neuronal level [287], evidence has been provided that 5-HT and leptin act in concert, possibly via a functional synergism at the interface between episodic and tonic satiety.

## **7. 5-HT interactions with other gastrointestinal peptides**

Compared to CCK and leptin, other anorectic peptides of primarily peripheral origin have been studied in less detail with regard to possible interactions with 5-HT. Nevertheless several of these gastrointestinal peptides mediate satiety [288], and are potential candidates for future anti-obesity drugs [289, 290].

Among these gastrointestinal peptides, the hormone insulin is not only involved in the regulation of glucose metabolism, but has also been shown to act as a satiety signal [291, 292]. Although the former is physiologically more significant, central injections of insulin produced hypophagia [293]. Targeted mutation of insulin receptor production in the brain led to obesity in mice, further suggesting a role of brain insulin in the control of feeding [294]. In microdialysis experiments, stimulation of 5-HT release caused activation of insulin in the PVN without affecting insulinemia or glycaemia [295], whereas serum insulin was reduced after administration of the sub-hypophagic doses of the 5-HT<sub>2C</sub> agonist mCPP [296]. This effect was mediated via downstream MC-4 receptors and suggests that pharmacological targeting of 5-HT<sub>2C</sub> receptors may enhance glucose tolerance independently of alterations in body weight [296].

If other peripherally released satiety factors like peptide YY<sub>3-36</sub> (PYY) [297] recruit brain serotonin remains to be investigated, but the behavioural specificity and the hypophagic effect of PYY itself remains a matter of discussion [298]. Glucagon like Peptide-1 (GLP-1) is released in response to a meal from the small intestine [299] and reduces food-intake both in animals and humans [289, 300] for review]. As a central site of action is possible [301-303], it is of interest that both GLP-1 and the GLP agonist exendin-4 stimulate 5-HT release from hypothalamic synaptosomes [304]. However, an icv injection of either one reduced hypothalamic 5-HT level. The primary metabolite of GLP-1, GLP-1 (9-39), had an

antagonistic effect though, as it blocked the effect of GLP-1 on 5-HT release [305]. Similar to the observed loss of CCK-induced satiety, the hypophagic effect of GLP-1 was lost in 5-HT<sub>2C</sub> receptor knockout mice [229, 306]. It cannot be excluded though, that mechanisms downstream to the 5-HT<sub>2C</sub> receptor, rather than the 5-HT<sub>2C</sub> receptor itself, are required to mediate the satiating effect of peripheral GLP-1. The MC4 receptor seems to be a candidate here [306].

C-fos expression pattern suggest that and GLP-1 and CCK use partially independent mechanisms to exert their satiating effects, as, in contrast to CCK, the NTS is probably not required for GLP-1-induced satiation [229]. This interpretation is supported by the finding that the 5-HT<sub>2C</sub> agonist mCPP does not activate GLP-1 neurons in the NTS [307]. It remains to be investigated, if mCPP or other 5-HT agonists activate central GLP-1 neurons at all, as this would be a prerequisite for true interactions between GLP-1 and 5-HT. Due to the short half-life of GLP-1 itself, GLP-1 receptor agonists have been developed which are, due to their longer half-life, pharmacologically more feasible to suppress feeding [300, 308, 309]. Whereas one of them, exendin-4, influences serotonergic neurotransmission [304, 305], another one, liguride, does not seem to require functional 5-HT<sub>2C</sub> receptors to suppress food intake [306]. A recent study [300], however questioned the behavioural specificity of exendin-4-induced anorexia. Taken together, an unambiguous conclusion regarding interactions between GLP-1 and 5-HT in the control of feeding cannot be drawn yet.

## 8. Changes in the brain satiety in different nutritional states

So far the impact of serotonergic manipulations, either pharmacologically or by using transgenic techniques, on feeding and satiety has been discussed. Due to the evidence that brain 5-HT mechanisms are involved in the control of feeding, satiation and satiety, one would expect that these brain systems would undergo changes themselves in situations where overeating or malnutrition occur. In the following we summarise evidence for that without attempting to be comprehensive. Instead we provide data from experimental studies in both genetic models of obesity and models of diet-induced obesity (DIO).

A genetic Zucker rat model of obesity is the obese Zucker rat where brain 5-HT metabolism shows significant abnormalities. However, data differs in detail across studies, as age, gender and brain region impact on 5-HT metabolism. Reduced tryptophan content but unchanged 5-HT content in various brain regions have been reported [310]. An increased 5-HT metabolism, as indicated by the 5-HIAA/5-HT ratio has been found in cortex,



1  
2  
3  
4  
5 hypothalamus and further regions [311]. Orosco et al. [312] investigated into the age  
6 dependency of changes in brain 5-HT and concluded that these changes are secondary to  
7 the development of obesity in female Zucker rats. The authors demonstrated that 5-HT  
8 levels in the medial hypothalamus of female lean and obese Zucker rats, revealed by  
9 chromatography, are not different. However, a decrease in hypothalamic 5-HT turnover has  
10 been seen in male obese Zucker rats at three and six month, but not in ten month of age  
11 [313]. Indeed, in vivo microdialysis showed a lower basal 5-HT release [314], but a,  
12 compared to lean controls, increased serotonergic response to a meal [315]. The latter effect  
13 declines with age [316] which would also be in line with ex vivo 5-HT measures [313]. A  
14 lower hypothalamic baseline concentration, although not being found in some studies, could  
15 be explained with an increased control via raphe somatodendritic 5-HT<sub>1A</sub> autoreceptors,  
16 although the physiological significance of intrinsically hyperexcitable dorsal raphe neurons in  
17 Zucker rats needs to be established [282]. Interestingly, the 5-HT<sub>1A</sub> agonist induced  
18 hypophagia in obese, but not in lean Zucker rats, where it expectedly stimulated feeding  
19 [313]. The aggravated hypothalamic 5-HT release in response to a meal in obese Zucker  
20 rats has been interpreted as a reduced postsynaptic sensitivity to satiety signals [315, 317].  
21 Considering these changes in brain 5-HT metabolism it appears somewhat surprising though  
22 that the hypophagic effects of fenfluramine, the SSRI fluoxetine, and the 5-HT<sub>1B/2C</sub> agonist  
23 TFMPP and 5-HT<sub>2A/2C</sub> agonists were similar in lean and obese Zucker rats [318-322].  
24 However, despite the lack of evidence for postsynaptic 5-HT<sub>1B</sub> or 2C receptors in obese  
25 Zucker rats, the hypophagic effect of 8-OH-DPAT in obese Zucker rats suggests such a pre-  
26 or postsynaptic plasticity, although the localisation of this effect would need to be identified.  
27 Changes in in the excitability of serotonergic raphe neurons in obese Zucker rats occur as  
28 early as between postnatal day 14 and 25 [282]. In addition to presynaptic feedback via  
29 autoreceptors, postsynaptic feedback in the control of 5-HT neurons has been suggested  
30 [323]. Such a postsynaptic feedback could be responsive to changes in 5-HT availability in  
31 projection areas as reported in obese Zucker rats. However, it needs to be considered that  
32 5-HT acts in concert with other neurotransmitters and hormones, and these interactions are  
33 possibly changed as well. Although the primary reason for obesity in Zucker rats is a  
34 dysfunctional leptin receptor [69, 279-281], a dysregulation of the brain serotonergic satiety  
35 system has been demonstrated. This dysregulation, although developmentally secondary in  
36 nature [324], gives an example that 5-HT could possibly contribute to the obese phenotype  
37 and stabilise it.

38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58 Despite sometimes conflicting data, transgenic models, in particular in conjunction with  
59 pharmacological approaches, provide a unique opportunity to study the role of 5-HT in  
60  
61  
62  
63  
64  
65

satiety on a mechanistic and cellular level. Such genetic defects, however, do not account for the obesity epidemic in humans. Rodent models of diet-induced-obesity are taking into account the contribution of environmental factors, e.g. nutritional factors, and their interactions with a given genetic background. Considering the overwhelming evidence for a role of 5-HT in satiety, we should also expect modifications of 5-HT functioning in dietary obese subjects.

A diet-induced-obesity model [325] has been established by exposure of outbred Sprague-Dawley rats to high caloric/high fat diets, where part of the rats become obese (DIO-prone) the other part not (obesity resistant rats; DR-prone rats). When fed on chow, both DIO- and DR-prone showed lower brain 5-HT turnover during the last hour of the light phase, when animals become active and begin foraging for food, as compared to the first hour of the light phase. However, unlike DR-prone rats, DIO-prone rats did not show a significant time-dependent difference in 5-HT turnover in either the arcuate nucleus or the PVN, two hypothalamic brain regions essentially involved in the control of feeding. Upon a 48 hour fast, 5-HT turnover decreased in various hypothalamic and extrahypothalamic brain structures similarly in both cohorts of rats. However, fasted DIO-prone rats showed a much greater reduction in the ventromedial nucleus turnover than fasted DR-prone rats. After feeding an obesogenic diet, DIO rats became obese and the alteration in 5-HT mechanisms disappeared. Whereas the initial abnormalities could possibly predispose the rats to develop obesity upon exposure to a hyperenergetic diet, the normalisation observed in obese DIO-prone rats could possibly contribute to the persistence of obesity [326]. Park et al. [327] fed rats a palatable obesogenic diet for 7 weeks and demonstrated regionally specific changes in binding to at 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub> and 5-HT<sub>2A</sub> receptors being overall consistent with reduced 5-HT release and decreased activity of the 5-HT neurons. The authors suggest that the increased binding may contribute to increased appetite in rats presented with highly palatable diet. In vivo hypothalamic 5-HT release to a meal is already attenuated after one week of feeding a high-fat diet, thus already in a pre-obese state. Continuing feeding for another five weeks led to total abolishment of meal-stimulated hypothalamic 5-HT release [328]. Dietary changes in hypothalamic functioning can obviously occur before the actual onset of obesity.

Alterations in brain 5-HT should also be expected in experimental situations where malnutrition occurs. Tumour bearing rats are used as model of cancer anorexia. These rats show a reduced food intake and upon offering food, and their in-vivo 5-HT release in the VMN of the hypothalamus rose and peaked significantly earlier in tumour bearing rats than in controls. This would be indicative of an earlier occurrence of satiety in these rats. After

surgical removal of the tumour, 24 h food intake had increased to the level of controls and VMN microdialysis showed that 5-HT was normal at baseline, as well as during and after eating [329, 330].

## 9. Serotonin and the pharmacotherapy of obesity

Over the last four decades or so a number of brain mechanisms have been identified that are involved in the control of food intake and satiety [331]. Regarding the recent obesity epidemic, attempts have been made to identify some of these mechanisms as targets for anti-obesity drugs [332-337]. The development of anti-obesity drugs is a complex process, last not least due to interactions between satiety systems, the plasticity of satiety systems and the resulting lack of an 'easy' target. The clinical use of anti-obesity drugs and the recent trends in the development of anti-obesity drugs has been subject of many reviews [332, 336-338] which give also consideration to serotonergic anti-obesity drugs. There are many drugs in different stages of clinical testing, and obviously not all of them recruit the brain 5-HT system for their action. Considering the structural diversity of satiety mechanisms, it is noteworthy that except from the peripherally acting lipase inhibitor orlistat all the other compounds have a, at least primarily, central site of actions. This includes d-fenfluramine and sibutramine; both were withdrawn due to cardio-vascular side effects [337]. A likely cause of fenfluramine-induced valvulopathy is activation of 5-HT<sub>2B</sub> receptors on heart valves by its metabolite norfenfluramine [339, 340]. Sibutramine is a 5-HT and noradrenaline re-uptake inhibitor, very similar to some antidepressant drugs. The potential cardiovascular risk of this compound is probably largely related to its adrenergic properties. The anti-obesity effects of sibutramine, however, are due to its effect on the serotonergic system, but alpha- and beta-adrenoreceptors are also involved [341, 342]. Assuming that only a modest weight loss will be achieved in many patients following a long period of taking the drug, safety becomes the most important issue for anti-obesity drugs. One consequence for optimisation of drug development would therefore be to increase pharmacological selectivity and thus potentially minimise side effects. Regarding 5-HT, one possible way out would be to target 5-HT receptors involved in the regulation of satiety that are predominantly expressed in the brain. The 5-HT<sub>1B</sub> receptor is a less likely target, as this receptor is also located on vascular tissues [343], hence causing potentially vascular side effects. Based on experimental and human studies, the 5-HT<sub>2C</sub> receptor has been identified as a possible target for anti-obesity drugs [338]. Indeed, the 5-HT<sub>2C</sub> agonist lorcaserin (APD356) has been identified in rodents and humans to reduce food intake [344, 345]. Lorcaserin has relatively few side effects (but

could be carcinogenic in rats) and has been approved in 2012 by the FDA for long term treatment of obesity [60, 346].

In the light of multiple actions of 5-HT<sub>2C</sub> ligands in the brain [347] it is somewhat surprising that relatively little experimental data is available which could highlight potential behavioural side effects in humans. Published evidence for behavioural specificity is missing also for lorcaserin [348], although this is also true for many other compounds still undergoing clinical testing [298]. In this context it may be worth considering that translational medicine of disease/obesity between phase 1 and phase 2 of clinical trials could provide early insight in efficacy and safety in humans [349]. Such a translational approach using the 5-HT<sub>2C</sub>/1B agonist mCPP has recently been exemplified in humans [350].

As shown throughout this review, among 5-HT receptor subtypes, the 5-HT<sub>2C</sub> receptor is predominantly involved in mediating satiety. This receptor undergoes mRNA editing that alters the amino-acid coding potential of the predicted second intracellular loop of the receptor and can lead to a 10-15-fold reduction in the efficacy of the interaction between receptors and their G proteins [351, 352]. The extent of editing does not only depend on the medication but also the pathophysiology of the disease [353]. In ob/ob mice, an increase in full-length 5-HT<sub>2C</sub> receptor expression, depending on time of day, as well as differences in 5-HT<sub>2C</sub> receptor editing were found, independent of changes in total 5-HT<sub>2C</sub> receptor mRNA expression [266]. These findings should potentially be considered when experimental data are applied to obese patients, but also when experimental studies using different models of obesity are compared. Finally this could possibly open up a way to a more customised pharmacotherapy.

Another approach to 5-HT<sub>2C</sub> receptor pharmacology in the context of obesity could arise from the dimerization of this G-protein coupled receptor as demonstrated with the ghrelin receptor [199, 354]. Although this concept warrants further research, dimerization of the 5-HT<sub>2C</sub> receptor increases the pharmacological diversity of this receptor and thus the development of new drugs.

Among the other 5-HT receptor subtypes, 5-HT<sub>6</sub> antagonists are undergoing clinical testing at present. 5-HT<sub>6</sub> receptor antagonists are well tolerated but, despite their satiating effects in rodents, are largely tested towards other indications which include dementia [335].

Considering the multiplicity of satiety mechanisms, one could envisage the combination of drugs in a way that they could act in concert to promote satiety, or using single molecules that are targeting different mechanisms. The latter was initially assumed for sibutramine

1  
2  
3  
4  
5 where increased sympathetic activity was hoped to increase energy expenditure. However,  
6 the increased noradrenergic activity had also the potential to cause cardiovascular damage.  
7 Fenfluramine has been combined with amphetamine analogue phentermine in the past and  
8 the recently approved combination of the antiepileptic topiramate and phentermine (Qsymia,  
9 FDA approved in 2012) is a product of this strategy. Further combinations are being clinically  
10 tested [336]. Recent experimental data, however, suggests that this approach does not  
11 immediately lead to the expected results. Whereas sibutramine, for example, advanced the  
12 BSS, thus indicating a satiating effect [355], the combination of sibutramine and the opioid  
13 antagonist naloxone showed rather infra-additive effects [355]. Combining fenfluramine with  
14 rimonabant had additive effects on food intake [356], whereas the combination of mCPP and  
15 naltrexone did not provide any support for a clinically useful combination [81]. In conclusion,  
16 these findings show that each potential combination requires individual testing. A move from  
17 a single-target approach to tackling complex neuronal mechanisms of satiety seems to be  
18 required [357].

19  
20  
21  
22  
23  
24  
25  
26  
27  
28 Another approach to tackle obesity through manipulation of the serotonergic system would  
29 be to influence the motivation to eat [77, 348], maybe in addition to satiety or even  
30 independently. In addition to its satiety promoting effects, fenfluramine also reduces the  
31 motivation to feed [358]. Experimental evidence exists that 5-HT<sub>2C</sub> agonists do not only  
32 inhibit food intake, e.g. the consummatory component of feeding behaviour, but also the  
33 preceding appetitive phase, which is not yet food related [82, 359]. Combining the 5-  
34 HT<sub>2C/1B</sub> agonist mCPP with the CB<sub>1</sub> antagonist/partial agonist rimonabant synergistically  
35 reduced motivated feeding behaviour in an operant paradigm in mice [360]. Testing mCPP in  
36 humans provides clear evidence that this 5-HT<sub>2C/1B</sub> agonist not only promotes satiation  
37 and satiety, but also suppresses appetite [350].

38  
39  
40  
41  
42  
43  
44 Most of the data so far have been implicitly related to a role of 5-HT in the homeostatic  
45 control of satiety. However, there is also a hedonic aspect to feeding and hedonic feeding  
46 will possibly involve dopaminergic mechanisms of reward [152, 361-364]. A functional link  
47 between hypothalamic energy-control mechanisms and the motivational aspect of feeding  
48 has been demonstrated by Helm et al. [240]. Injection of either CCK or 5-HT into the PVN  
49 limits dopamine release in the nucleus accumbens and synergistically activates  
50 acetylcholine release in the accumbens. Highly palatable foods stimulate dopamine release  
51 in the nucleus accumbens [365]. The combined actions of 5-HT and CCK in the PVN may  
52 limit the size of a meal by shifting the animal's motivational state from approach to avoidance  
53 of the food, the latter expressed by either increased accumbal acetylcholine, which controls  
54 dopamine release, or decreased accumbal dopamine [240]. An involvement of the 5-HT<sub>2C</sub>  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4  
5 in brain mechanisms of reward has been suggested [366]. This should be further explored in  
6 the light of the recent obesity epidemic, as the 5-HT<sub>2C</sub> receptor could possibly provide a link  
7 between homeostatic and non-homeostatic (hedonic) eating [165]. Berthoud synthesised  
8 ideas on the regulation of feeding and satiety and pointed out that food intake follows both  
9 homeostatic and ('hedonic') mechanisms. Both are not independent, and while non-  
10 homeostatic eating is frequently attributed to the neurotransmitter dopamine, 5-HT is largely  
11 seen as a neurotransmitter within the homeostatic system, interactions between 5-HT and  
12 dopamine are being discussed, and 5-HT<sub>2C</sub> receptor agonists generally inhibit reward-  
13 related behaviours [366, 367].  
14  
15  
16  
17  
18  
19

20 5-HT has been characterised not only as a satiety signal but also as a developmental signal  
21 [368, 369]. The impact of the nutritional environment during early development is a well-  
22 established fact [370-374]. 5-HT could possibly have a twofold role here, as 5-HT synthesis  
23 depends on tryptophan of nutritional origin, but then the 5-HT satiety system controls feeding  
24 itself. Intrauterine undernutrition leads to resistance to the hypophagic effect of  
25 intracerebroventricular 5-HT and dysregulations of hypothalamic 5-HT<sub>1B</sub> receptor, 5-HT<sub>2C</sub>  
26 receptor and 5-HTT protein expression. Adult offspring of such undernourished rats  
27 developed obesity despite normal habitual food intake suggesting the involvement of further  
28 satiety and metabolic mechanisms [375]. Nutritional programming by overnutrition has also  
29 been demonstrated after feeding a Western-style diet to lactating dams. Offspring from these  
30 rats showed a delayed BSS and a reduced hypothalamic 5-HT turnover although a direct  
31 causal relationship between the two has not been demonstrated in this study [376].  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41

## 42 **10. Serotonin and satiety – What is next?**

43  
44 Despite the vast amount of data on the involvement of serotonin in the control of food intake,  
45 serotonergic mechanisms are sometimes somewhat neglected when brain mechanisms of  
46 satiety are being reviewed [377-379]. This could provoke the question about the importance  
47 of the serotonergic system in satiety in comparison to other neural and endocrine factors.  
48 However, this is rather a rhetorical question, as, based on our current knowledge and  
49 understanding, it is very unlikely that a particular mechanism could be singled out that rules  
50 the “satiety system.” In this context, the fact that a live without (brain) serotonin is possible is  
51 more likely to stimulate further research than giving a definite answer [380]. It does,  
52 however, suggest that brain serotonin, at least with regard to satiety, might have a  
53 modulating function. One could speculate that such a modulating function is required to  
54 allow adaptations to both internal and external changes. Two lines of research are being  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

suggested that could possibly advance our understanding of the role of serotonin and satiety.

Firstly, although the 5-HT receptors that are involved in satiety have been identified, their role in hedonic feeding needs further exploration. 5-HT 2C agonists are still a candidate here and also for the development of anti-obesity drugs. This has been reviewed before, but the future of these compounds depends on their side effects, in particular on the question if these side effects are related to their agonist properties at the 5-HT<sub>2C</sub> receptor. A new and largely unexplored approach to target the 5-HT<sub>2C</sub> receptor has been reported for tackling drug addiction [381]. To minimise side effects, it has been suggested to interfere pharmacologically with intracellular receptor dephosphorylation. 5-HT<sub>2C</sub> receptor activation causes intracellular receptor phosphorylation which prevents desensitisation and enhances resensitisation [382]. Thus an inhibition of dephosphorylation would be functionally similar to extracellular receptor activation. However, it remains to be determined, if such an approach can be used to reduce food intake, and most importantly, if this could lead to the development of new classes of anti-obesity drugs. In addition, direct intracellular effects in the context of insulin secretion have been demonstrated [383], suggesting that intracellular 5-HT functions in various microenvironments act in concert with the known receptor-mediated signalling. If such an intracellular “protein serotonylation” [383, 384] is involved in brain mechanisms of satiety remains to be investigated though. Together with the aforementioned approach to make pharmacological use of receptor dimerization [199], one could expect an increasingly diverse approach to interact with 5-HT<sub>2C</sub> receptors to tackle obesity.

Secondly, as much of the research being reviewed here is driven, either directly or indirectly, by the recent obesity epidemic, it is important to acknowledge further developments in this area. Evidence has accumulated over the last decades that the nutritional environment during early developmental periods has a significant impact on physiology and pathophysiology in adult age [372, 373, 385, 386]. In this context, the aforementioned role of 5-HT in neural development leads to the question how the developing serotonergic system interacts with the early nutritional environment. Placental 5-HT pathways from maternal tryptophan contribute to the fetal programming of the brain. Later in development, there will be a switch to an endogenous brain source of 5-HT [387]. However, as in any case the availability of the 5-HT precursor tryptophan depends on dietary supply [388], an impact of the early nutritional environment on brain development can be expected [389]. There is emerging experimental evidence for a concept of early nutritional programming of hypothalamic function [373]. Caloric undernutrition during late gestation led to increased 5-

HT1A receptor expression in in the VMH and LHA at postnatal day [390].The 5-HT1A receptor has been implicated in neural development [369], and in rodents the hypothalamic satiety system matures postnatally. Projections from the arcuate nucleus, which plays a role in the serotonergic control of feeding, develop during the second week after birth [391]. The hypothalamus remains largely immature until postnatal day 21 [373]. Together these findings give rise to speculations that the brain 5-HT satiety system could be nutritionally programmed. Preliminary evidence for such an assumption comes from experimental studies demonstrating that perinatal protein deficiency attenuates the hypophagic effect of fenfluramine in the offspring [389]. Longitudinal and mechanistic studies should help to identify the critical pre- and postnatal time points when nutritional challenges impact first on brain 5-HT development. Such knowledge could contribute to generate an optimal nutritional environment during early developmental stages. Such an approach would help to develop timed strategies to reduce the risk for eating disorders or obesity in later life as it has already been suggested for psychiatric disorders [392, 393].

## 11. Conclusions

In summary, over the last four decades 5-HT has been identified as an important signal for satiation and satiety, possibly in concert with other satiety signals. Brain mechanisms of 5-HT-induced satiety are being identified, showing an emerging understanding as to which structures are involved and how brain neurotransmitter functioning relates to these structures. The present review could only touch these aspects, but highlighted the 5-HT receptor subtypes that are predominantly involved in 5-HT mediated satiety and, therefore, provide targets for further developments of more selective appetite suppressant drugs. The complex interactions between 5-HT and other endogenous mediators of satiety making 5-HT not an 'easy target' for the development of anti-obesity drugs, as those interactions enable flexible responses and the initiation of compensatory mechanism to respond to nutritional challenges. Looking at a picture that comprises not only homeostatic aspects of feeding, but also accounts for hedonic aspects and developmental aspects of the 5-HT system does not only illustrates the complexity of the topic, but also provides opportunity how to tackle associated health issues.



## References

- [1] Blundell JE. The control of appetite: basic concepts and practical implications. *Schweiz Med Wochenschr.* 1999;129:182-8.
- [2] Blundell JE. Serotonin manipulations and the structure of feeding behaviour. *Appetite.* 1986;7 Suppl:39-56.
- [3] Booth DA, Thibault, L. Macronutrient-specific hungers and satieties and their neural bases, learnt from pre- and postingestional effects of eating particular foodstuffs. In: Berthoud HR, Seeley, R.J., editors. *Neural and etabolic control of macronutrient intake*, Boca Raton: CRC Press; 2000. p. 61-91.
- [4] Simansky KJ. Serotonin and the structure of satiation. In: Smith GP, editor. *Satiation: From gut to brain*, New York: Oxford University Press; 1998. p. 217-62.
- [5] Hen R. Of mice and flies: commonalities among 5-HT receptors. *Trends Pharmacol Sci.* 1992;13:160-5.
- [6] Venter JC, di Porzio U, Robinson DA, Shreeve SM, Lai J, Kerlavage AR, et al. Evolution of neurotransmitter receptor systems. *Prog Neurobiol.* 1988;30:105-69.
- [7] Weiger WA. Serotonergic modulation of behaviour: a phylogenetic overview. *Bi Biol rev Camb Philos Soc.* 1997;72:61-95.
- [8] French AS, Simcock KL, Rolke D, Gartside SE, Blenau W, Wright GA. The role of serotonin in feeding and gut contractions in the honeybee. *J Insect Physiol.* 2014;61:8-15.
- [9] Gillette R. Evolution and function in serotonergic systems. *Integ Comp Biol.* 2006;46:838-46.
- [10] Azmitia EC. Evolution of serotonin: Sunlight to suicide. In: Mueller CP, Jacobs, B.L., editors. *Handbook of the behavioral neurobiology of serotonin*, New York: Academic Press; 2010. p. 3-22.

[11] Barnes NM, Sharp T. A review of central 5-HT receptors and their function. *Neuropharmacology*. 1999;38:1083-152.

[12] Hoyer D, Clarke DE, Fozard JR, Hartig PR, Martin GR, Mylecharane EJ, et al. VII. International union of pharmacology classification of receptors for 5-hydroxytryptamine (serotonin). *Pharmacol Rev*. 1994;46:157-203.

[13] Hoyer D, Hannon JP, Martin GR. Molecular, pharmacological and functional diversity of 5-HT receptors. *Pharmacol Biochem Behav*. 2002;71:533-54.

[14] Hornung J. The neuroanatomy of the serotonergic system. In: Mueller CP, Jacobs, B.L., editors. *Handbook of the behavioral neurobiology of serotonin*, New York: Academic Press; 2010. p. 51-64.

[15] Lucki I. The spectrum of behaviors influenced by serotonin. *Biol Psychiatry*. 1998;44:151-62.

[16] Bergen SS, Jr. Appetite Stimulating Properties of Cyproheptadine. *Am J Dis Child*. 1964;108:270-3.

[17] Stone CA, Wenger HC, Ludden CT, Ross CA, Stavorski JM. Antiserotonin-antihistaminic properties of cyproheptadine. *J Pharmacol Expe Therap*. 1961;131:73-84.

[18] Guy-Grand B. Clinical studies with dexfenfluramine: from past to future. *Obes Res*. 1995;3 Suppl 4:491S-6S.

[19] Lesses MF, Myerson A. Human autonomic pharmacology XVI Benzedrine sulfate as an aid in the treatment of obesity. *New Engl J Med*. 1938;218:119-24.

[20] Nathanson MH. The central action of betaaminopropylbenzene (benzedrine): Clinical observations. *J Pharmacol Expe Therap*. 1937;108:528-31.

[21] Prinzmetal M, Bloomberg W. The use of benzedrine for the treatment of narcolepsy. *J Am Med Assoc*. 1935;105:2051-4.

[22] Barrett AM, Mcsharry L. Inhibition of dug-induced anorexia in rats by methysergide. *J Pharm Pharmacol.* 1975;27:889-95.

[23] Blundell JE, Latham CJ, Leshem MB. Biphasic Action of a 5-Hydroxytryptamine inhibitor on fenfluramine-induced anorexia. *J Pharm Pharmacol.* 1973;25:492-4.

[24] Clineschmidt BV, McGuffin JC, Pflueger AB, Totaro JA. A 5-hydroxytryptamine-like mode of anorectic action for 6-chloro-2-[1-piperazinyl]-pyrazine (MK-212). *Br J Pharmacol.* 1978;62:579-89.

[25] Clineschmidt BV, McGuffin JC, Werner AB. Role of monoamines in the anorexigenic actions of fenfluramine, amphetamine and p-chloromethamphetamine. *Eur J Pharmacol.* 1974;27:313-23.

[26] Jespersen.S, Scheel-Kruger.J. Evidence for a difference in mechanism of action between fenfluramine-Induced and amphetamine-Induced anorexia. *J Pharm Pharmacol.* 1973;25:49-54.

[27] Geyer MA, Puerto A, Menkes DB, Segal DS, Mandell AJ. Behavioral studies following lesions of the mesolimbic and mesostriatal serotonergic pathways. *Brain Res.* 1976;106:257-69.

[28] Samanin R, Ghezzi D, Valzelli L, Garattini S. The effects of selective lesioning of brain serotonin or catecholamine containing neurones on the anorectic activity of fenfluramine and amphetamine. *Eur J Pharmacol.* 1972;19:318-22.

[29] Oluyomi AO, Gibson EL, Barnfield AM, Curzon G. d-Fenfluramine and d-norfenfluramine hypophagias do not require increased hypothalamic 5-hydroxytryptamine release. *Eur J Pharmacol.* 1994;264:111-5.

[30] Schwartz D, Hernandez L, Hoebel BG. Fenfluramine administered systemically or locally increases extracellular serotonin in the lateral hypothalamus as measured by microdialysis. *Brain Res.* 1989;482:261-70.

[31] Blundell JE. Is There a role for serotonin (5-Hydroxytryptamine) in feeding. *Int J Obesity.* 1977;1:15-42.

[32] Wurtman RJ, Hefti F, Melamed E. Precursor control of neurotransmitter synthesis. *Pharmacol Rev.* 1980;32:315-35.

[33] Schwartz DH, McClane S, Hernandez L, Hoebel BG. Feeding increases extracellular serotonin in the lateral hypothalamus of the rat as measured by microdialysis. *Brain Res.* 1989;479:349-54.

[34] Orosco M, Nicolaidis S. Spontaneous feeding-related monoaminergic changes in the rostromedial hypothalamus revealed by microdialysis. *Physiol Behav.* 1992;52:1015-9.

[35] Voigt J-P, Sohr R, Fink H. CCK-8S facilitates 5-HT release in the rat hypothalamus. *Pharmacol Biochem Behav.* 1998;59:179-82.

[36] Rouch C, Nicolaidis S, Orosco M. Determination, using microdialysis, of hypothalamic serotonin variations in response to different macronutrients. *Physiol Behav.* 1999;65:653-7.

[37] Latham CJ, Blundell JE. Evidence for the effect of tryptophan on the pattern of food consumption in free feeding and food deprived rats. *Life Sci.* 1979;24:1971-8.

[38] Walther DJ, Bader M. A unique central tryptophan hydroxylase isoform. *Biochem Pharmacol.* 2003;66:1673-80.

[39] Narboux-Neme N, Angenard G, Mosienko V, Klempin F, Pitychoutis PM, Deneris E, et al. postnatal growth defects in mice with constitutive depletion of central serotonin. *Acs Chem Neurosci.* 2013;4:171-81.

[40] Alenina N, Kikic D, Todiras M, Mosienko V, Qadri F, Plehm R, et al. Growth retardation and altered autonomic control in mice lacking brain serotonin. *P Natl Acad Sci USA.* 2009;106:10332-7.

[41] Yadav VK, Oury F, Suda N, Liu ZW, Gao XB, Confavreux C, et al. A serotonin-dependent mechanism explains the leptin regulation of bone mass, appetite, and energy expenditure. *Cell.* 2009;138:976-89.

[42] Savelieva KV, Zhao SL, Pogorelov VM, Rajan I, Yang Q, Cullinan E, et al. Genetic disruption of both tryptophan hydroxylase genes dramatically reduces serotonin and affects behavior in models sensitive to antidepressants. *Plos One*. 2008;3.

[43] MacKenzie RG, Hoebel BG, Ducret RP, Trulson ME. Hyperphagia following intraventricular p-chlorophenylalanine-, leucine- or tryptophan-methyl esters: lack of correlation with whole brain serotonin levels. *Pharmacol Biochem Behav*. 1979;10:951-5.

[44] Blundell JE, Leshem MB. The effect of 5-hydroxytryptophan on food intake and on the anorexic action of amphetamine and fenfluramine. *J Pharm Pharmacol*. 1975;27:31-7.

[45] Fletcher PJ, Burton MJ. Dissociation of the anorectic actions of 5-HTP and fenfluramine. *Psychopharmacology*. 1986;89:216-20.

[46] Curzon G, Joseph MH, Knott PJ. Effects of immobilization and food deprivation on rat-brain tryptophan metabolism. *J Neurochem*. 1972;19:1967-74.

[47] Katak KM, Wayner MJ, Stein JM. Effects of various periods of food deprivation on serotonin synthesis in the lateral hypothalamus. *Pharmacol Biochem Behav*. 1978;9:535-41.

[48] Blundell JE, Hill AJ. Dexfenfluramine and appetite in humans. *Int J Obesity*. 1992;16:S51-S9.

[49] Goudie AJ, Thornton EW, Wheeler TJ. Effects of Lilly 110140, a Specific Inhibitor of 5-hydroxytryptamine uptake, on food-intake and on 5-Hydroxytryptophan-induced anorexia - evidence for serotonergic inhibition of feeding. *J Pharm Pharmacol*. 1976;28:318-20.

[50] Clifton PG, Barnfield AMC, Philcox L. A Behavioral profile of fluoxetine-induced anorexia. *Psychopharmacology*. 1989;97:89-95.

[51] Simansky KJ, Vaidya AH. Behavioral mechanisms for the anorectic action of the serotonin (5-HT) uptake inhibitor sertraline in rats - comparison with directly acting 5-HT agonists. *Brain Res Bull*. 1990;25:953-60.

[52] Mathews TA, Fedele DE, Coppelli FM, Avila AM, Murphy DL, Andrews AM. Gene dose-dependent alterations in extraneuronal serotonin but not dopamine in mice with reduced serotonin transporter expression. *J Neurosci Meth.* 2004;140:169-81.

[53] Uceyler N, Schutt M, Palm F, Vogel C, Meier M, Schmitt A, et al. Lack of the serotonin transporter in mice reduces locomotor activity and leads to gender-dependent late onset obesity. *Int J Obesity.* 2010;34:701-11.

[54] Jennings KA, Loder MK, Sheward WJ, Pei Q, Deacon RMJ, Benson MA, et al. Increased expression of the 5-HT transporter confers a low-anxiety phenotype linked to decreased 5-HT transmission. *J Neurosci.* 2006;26:8955-64.

[55] Pringle A, Jennings KA, Line S, Bannerman DM, Higgs S, Sharp T. Mice overexpressing the 5-hydroxytryptamine transporter show no alterations in feeding behaviour and increased non-feeding responses to fenfluramine. *Psychopharmacology.* 2008;200:291-300.

[56] Line SJ, Barkus C, Coyle C, Jennings KA, Deacon RM, Lesch KP, et al. Opposing alterations in anxiety and species-typical behaviours in serotonin transporter overexpressor and knockout mice. *Eur Neuropsychopharmacol.* 2011;21:108-16.

[57] Chen JJ, Li Z, Pan H, Murphy DL, Tamir H, Koepsell H, et al. Maintenance of serotonin in the intestinal mucosa and ganglia of mice that lack the high-affinity serotonin transporter: Abnormal intestinal motility and the expression of cation transporters. *J Neurosci.* 2001;21:6348-61.

[58] Antin J, Gibbs J, Holt J, Young RC, Smith GP. Cholecystokinin elicits the complete behavioral sequence of satiety in rats. *J Comp Physiol Psychol.* 1975;89:784-90.

[59] Halford JC, Wanninayake SC, Blundell JE. Behavioral satiety sequence (BSS) for the diagnosis of drug action on food intake. *Pharmacol Biochem Behav.* 1998;61:159-68.

[60] Yanovski SZ, Yanovski JA. Long-term drug treatment for obesity: a systematic and clinical review. *JAMA.* 2014;311:74-86.

[61] Ishii Y, Blundell JE, Halford JC, Rodgers RJ. Palatability, food intake and the behavioural satiety sequence in male rats. *Physiol Behav.* 2003;80:37-47.

[62] Rodgers RJ, Halford JC, Nunes de Souza RL, Canto de Souza AL, Piper DC, Arch JR, et al. Dose-response effects of orexin-A on food intake and the behavioural satiety sequence in rats. *Regul Pept.* 2000;96:71-84.

[63] Garattini S, Mennini T, Bendotti C, Invernizzi R, Samanin R. Neurochemical mechanism of action of drugs which modify feeding via the serotonergic system. *Appetite.* 1986;7 Suppl:15-38.

[64] Gibson EL, Kennedy AJ, Curzon G. d-Fenfluramine- and d-norfenfluramine-induced hypophagia: differential mechanisms and involvement of postsynaptic 5-HT receptors. *EurJPharmacol.* 1993;242:83-90.

[65] Raiteri M, Bonanno G, Vallebuona F. In-vitro and in-vivo effects of d-fenfluramine - no apparent relation between 5-Hydroxytryptamine release and hypophagia. *J Pharmacol Exp Therap.* 1995;273:643-9.

[66] Mennini T, Bizzi A, Caccia S, Codegoni A, Fracasso C, Frittoli E, et al. Comparative-Studies on the Anorectic Activity of D-Fenfluramine in Mice, Rats, and Guinea-Pigs. *N-S Arch Pharmacol.* 1991;343:483-90.

[67] Curzon G, Gibson EL, Oluyomi AO. Appetite suppression by commonly used drugs depends on 5-HT receptors but not on 5-HT availability. *Trends in Pharmacological Sciences.* 1997;18:21-5.

[68] Vickers SP, Dourish CT, Kennett GA. Evidence that hypophagia induced by d-fenfluramine and d-norfenfluramine in the rat is mediated by 5-HT<sub>2C</sub> receptors. *Neuropharmacology.* 2001;41:200-9.

[69] Yamashita T, Murakami T, Iida M, Kuwajima M, Shima K. Leptin receptor of Zucker fatty rat performs reduced signal transduction. *Diabetes.* 1997;46:1077-80.

[70] Vickers SP, Clifton PG, Dourish CT. Behavioural evidence that d-fenfluramine-induced anorexia in the rat is not mediated by the 5-HT<sub>1A</sub> receptor subtype. *Psychopharmacology.* 1996;125:168-75.

[71] Vickers SP, Clifton PG, Dourish CT, Tecott LH. Reduced satiating effect of d-fenfluramine in serotonin 5-HT<sub>2C</sub> receptor mutant mice. *Psychopharmacology*. 1999;143:309-14.

[72] Hewitt KN, Lee MD, Dourish CT, Clifton PG. Serotonin 2C receptor agonists and the behavioural satiety sequence in mice. *Pharmacol Biochem and Behav*. 2002;71:691-700.

[73] Lee MD, Somerville EM, Kennett GA, Dourish CT, Clifton PG. Reduced hypophagic effects of d-fenfluramine and the 5-HT<sub>2C</sub> receptor agonist mCPP in 5-HT<sub>1B</sub> receptor knockout mice. *Psychopharmacology*. 2004;176:39-49.

[74] Montgomery AMJ, Willner P. Fenfluramine disrupts the behavioral satiety sequence in rats. *Psychopharmacology*. 1988;94:397-401.

[75] McGuirk J, Muscat R, Willner P. Effects of Chronically administered fluoxetine and fenfluramine on food-intake, body-weight and the behavioral satiety sequence. *Psychopharmacology*. 1992;106:401-7.

[76] Grignaschi G, Samanin R. Role of 5-HT Receptors in the Effect of d-fenfluramine on feeding patterns in the rat. *Eur J of Pharmacol*. 1992;212:287-9.

[77] De Vry J, Schreiber R. Effects of selected serotonin 5-HT<sub>1</sub> and 5-HT<sub>2</sub> receptor agonists on feeding behavior: possible mechanisms of action. *Neurosci Biobehav Rev*. 2000;24:341-53

[78] Currie PJ, Coscina DV. 5-Hydroxytryptaminergic receptor agonists: effects on neuropeptide Y potentiation of feeding and respiratory quotient. *Brain Res*. 1998;803:212-7.

[79] Hutson PH, Donohoe TP, Curzon G. Infusion of the 5-Hydroxytryptamine Agonists Ru24969 and TFMPP into the paraventricular nucleus of the hypothalamus causes hypophagia. *Psychopharmacology*. 1988;95:550-2.

[80] Kitchener SJ, Dourish CT. An examination of the behavioural specificity of hypophagia induced by 5-HT-1b, 5-HT-1c and 5-HT-2 receptor agonists using the post-prandial satiety sequence in rats. *Psychopharmacology*. 1994;113:369-77.



1  
2  
3  
4  
5 [81] Wright FL, Rodgers RJ. On the behavioural specificity of hypophagia induced in male  
6 rats by mCPP, naltrexone, and their combination. *Psychopharmacology*. 2014;231:787-800.  
7

8  
9  
10 [82] Somerville EM, Horwood JM, Lee MD, Kennett GA, Clifton PG. 5-HT<sub>2C</sub> receptor  
11 activation inhibits appetitive and consummatory components of feeding and increases brain  
12 c-fos immunoreactivity in mice. *Europ J Neurosci*. 2007;25:3115-24.  
13

14  
15  
16 [83] Tecott LH, Sun LM, Akana SF, Strack AM, Lowenstein DH, Dallman MF, et al. Eating  
17 disorder and epilepsy in mice lacking 5-HT<sub>2C</sub> serotonin receptors. *Nature*. 1995;374:542-6.  
18

19  
20  
21 [84] Nonogaki K, Strack AM, Dallman MF, Tecott LH. Leptin-independent hyperphagia and  
22 type 2 diabetes in mice with a mutated serotonin 5-HT<sub>2C</sub> receptor gene. *Nat Med*.  
23 1998;4:1152-6.  
24

25  
26  
27 [85] Nonogaki K, Memon RA, Grunfeld C, Feingold KR, Tecott LH. Altered gene expressions  
28 involved in energy expenditure in 5-HT<sub>2C</sub> receptor mutant mice. *Biochem Biophys Res*  
29 *Commun*. 2002;295:249-54.  
30

31  
32  
33 [86] Inui A. Transgenic approach to the study of body weight regulation. *Pharmacological*  
34 *Reviews*. 2000;52:35-61.  
35

36  
37  
38 [87] Heisler LK, Chu HM, Tecott LH. Epilepsy and obesity in serotonin 5-HT<sub>2C</sub> receptor  
39 mutant mice. *Ann Ny Acad Sci*. 1998;861:74-8.  
40

41  
42  
43 [88] Fletcher PJ, Tampakeras M, Sinyard J, Slassi A, Isaac M, Higgins GA. Characterizing  
44 the effects of 5-HT<sub>2C</sub> receptor ligands on motor activity and feeding behaviour in 5-HT<sub>2C</sub>  
45 receptor knockout mice. *Neuropharmacology*. 2009;57:259-67.  
46

47  
48  
49 [89] Bendotti C, Samanin R. The role of putative 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptors in the  
50 control of feeding in rats. *Life Sci*. 1987;41:635-42.  
51

52  
53  
54 [90] Kennett GA, Curzon G. Evidence that hypophagia induced by mCPP and TFMPP  
55 requires 5-HT<sub>1C</sub> and 5-HT<sub>1B</sub> receptors; hypophagia induced by RU 24969 only requires 5-  
56 HT<sub>1B</sub> receptors. *Psychopharmacology (Berl)*. 1988;96:93-100.  
57  
58  
59  
60  
61  
62  
63  
64  
65

[91] Halford JC, Blundell JE. The 5-HT<sub>1B</sub> receptor agonist CP-94,253 reduces food intake and preserves the behavioural satiety sequence. *Physiol Behav.* 1996;60:933-9.

[92] Lee MD, Simansky KJ. CP-94,253: A selective serotonin(1B) (5-HT<sub>1B</sub>) agonist that promotes satiety. *Psychopharmacology.* 1997;131:264-70.

[93] Lee MD, Kennett GA, Dourish CT, Clifton PG. 5-HT<sub>1B</sub> receptors modulate components of satiety in the rat: behavioural and pharmacological analyses of the selective serotonin(1B) agonist CP-94,253. *Psychopharmacology.* 2002;164:49-60.

[94] Saudou F, Amara DA, Dierich A, Lemeur M, Ramboz S, Segu L, et al. Enhanced aggressive-behavior in mice lacking 5-Ht<sub>1B</sub> receptor. *Science.* 1994;265:1875-8.

[95] Lucas JJ, Yamamoto A, Scearce-Levie K, Saudou F, Hen R. Absence of fenfluramine-induced anorexia and reduced c-fos induction in the hypothalamus and central amygdaloid complex of serotonin 1B receptor knock-out mice. *J Neurosci.* 1998;18:5537-44.

[96] Bouwknecht JA, van der Gugten J, Hijzen TH, Maes RAA, Hen R, Olivier B. Male and female 5-HT<sub>1B</sub> receptor knockout mice have higher body weights than wildtypes. *Physiol Behav.* 2001;74:507-16.

[97] Lee MD, Somerville EM, Kennett GA, Dourish CT, Clifton PG. Tonic regulation of satiety by 5-HT receptors in the mouse: converging evidence from behavioural and c-fos immunoreactivity studies? *Eur J Neurosci.* 2004;19:3017-25.

[98] Dalton GL, Lee MD, Kennett GA, Dourish CT, Clifton PG. Serotonin 1B and 2C receptor interactions in the modulation of feeding behaviour in the mouse. *Psychopharmacology (Berl).* 2006;185:45-57.

[99] Simansky KJ. Serotonergic control of the organization of feeding and satiety. *Behav Brain Res.* 1996;73:37-42.

[100] Halford JCG, Lawton CL, Blundell JE. The 5-HT<sub>2</sub> receptor agonist MK-212 reduces food intake and increases resting but prevents the behavioural satiety sequence. *Pharmacol Biochem Behav.* 1997;56:41-6.

[101] Costall B, Naylor RJ. 5-HT<sub>3</sub> receptors. *Curr Drug Targets CNS Neurol Disord.* 2004;3:27-37

[102] Aja SM, Barrett JA, Gietzen DW. CCK(A) and 5-HT<sub>3</sub> receptors interact in anorectic responses to amino acid deficiency. *Pharmacol Biochem Behav.* 1999;62:487-91.

[103] Cooper SJ, Greenwood SE, Gilbert DB. The selective 5-HT-3 receptor antagonist, ondansetron, augments the anorectic effect of d-amphetamine in nondeprived rats. *Pharmacol Biochem Behav.* 1993;45:589-292.

[104] van der Hoek GA, Cooper SJ. Ondansetron, a selective 5-HT<sub>3</sub> receptor antagonist, reduces palatable food consumption in the nondeprived rat. *Neuropharmacology.* 1994;33:805-11.

[105] Savastano DM, Hayes MR, Covasa M. Serotonin-type 3 receptors mediate intestinal lipid-induced satiation and Fos-like immunoreactivity in the dorsal hindbrain. *Am J Physiol Regul Integr Comp Physiol.* 2007;292:R1063-70.

[106] Simpson KH, Murphy P, Colthup PV, Whelan P. Concentration of ondansetron in cerebrospinal-fluid following oral dosing in volunteers. *Psychopharmacology.* 1992;109:497-8.

[107] Hayes MR, Covasa M. Dorsal hindbrain 5-HT<sub>3</sub> receptors participate in control of meal size and mediate CCK-induced satiation. *Brain Research.* 2006;1103:99-107.

[108] Lee MD, Clifton, P.G. Role of the serotonergic system in appetite and ingestion control. In: Mueller CP, Jacobs, B.L., editors. *Handbook of the behavioural neurobiology of serotonin*, New York: Academic Press; 2010. p. 331-46.

[109] Woolley ML, Marsden CA, Fone KC. 5-HT<sub>6</sub> receptors. *Curr Drug Targets CNS Neurol Disord.* 2004;3:59-79.

[110] Harris RB, Zhou J, Redmann SM, Jr., Smagin GN, Smith SR, Rodgers E, et al. A leptin dose-response study in obese (ob/ob) and lean (+/?) mice. *Endocrinology.* 1998;139:8-19.

[111] Bentley JC. Effect of the 5-HT<sub>6</sub> agonist, Ro 04-6790 on food consumption in rats trained to a fixed feeding regime. *Br J Pharmacol.* 1999;126:66P.

[112] Woolley ML, Bentley JC, Sleight AJ, Marsden CA, Fone KC. A role for 5-HT<sub>6</sub> receptors in retention of spatial learning in the Morris water maze. *Neuropharmacology.* 2001;41:210-9.

[113] Fisas A, Codony X, Romero G, Dordal A, Giraldo J, Merce R, et al. Chronic 5-HT<sub>6</sub> receptor modulation by E-6837 induces hypophagia and sustained weight loss in diet-induced obese rats. *Br J Pharmacol.* 2006;148:973-83.

[114] Frassetto A, Zhang J, Lao JZ, White A, Metzger JM, Fong TM, et al. Reduced sensitivity to diet-induced obesity in mice carrying a mutant 5-HT<sub>6</sub> receptor. *Brain Res.* 2008;1236:140-4.

[115] Hirst WD, Abrahamsen B, Blaney FE, Calver AR, Aloj L, Price GW, et al. Differences in the central nervous system distribution and pharmacology of the mouse 5-hydroxytryptamine-6 receptor compared with rat and human receptors investigated by radioligand binding, site-directed mutagenesis, and molecular modeling. *Mol Pharmacol.* 2003;64:1295-308.

[116] Garfield AS, Burke LK, Shaw J, Evans ML, Heisler LK. Distribution of cells responsive to 5-HT receptor antagonist-induced hypophagia. *Behav Brain Res.* 2014;266C:201-6.

[117] Bockaert J, Claeysen S, Compan V, Dumuis A. 5-HT<sub>4</sub> receptors, a place in the sun: act two. *Curr Opin Pharmacol.* 2011;11:87-93.

[118] Haahr ME, Rasmussen PM, Madsen K, Marnier L, Ratner C, Gillings N, et al. Obesity is associated with high serotonin 4 receptor availability in the brain reward circuitry. *Neuroimage.* 2012;61:884-8.

[119] Hoyer D, Pazos A, Probst A, Palacios JM. Serotonin receptors in the human brain. I. Characterization and autoradiographic localization of 5-HT<sub>1A</sub> recognition sites. Apparent absence of 5-HT<sub>1B</sub> recognition sites. *Brain Res.* 1986;376:85-96.

[120] Kia HK, Brisorgueil M-J, Hamon M, Calas A, Verge D. Ultrastructural localization of 5-hydroxytryptamine-1A receptors in the rat brain. *J Neurosci Res.* 1996;46:697-708.

[121] Pompeiano M, Palacios JM, Mengod G. Distribution and cellular localization of mRNA coding for 5-HT<sub>1A</sub> receptor in the rat brain: correlation with receptor binding. *J Neurosci.* 1992;12:440-53.

[122] Dourish CT, Huston PH, Curzon G. Low doses of the putative serotonin agonist 8-hydroxy-2-(di-n-propylamino) tetralin (8-OH-DPAT) elicit feeding in the rat. *Psychopharmacology.* 1985;86:197-204.

[123] Dourish CT, Hutson PH, Curzon G. Characteristics of feeding induced by the serotonin agonist 8-hydroxy-2-(di-n-propylamino) tetralin (8-OH-DPAT). *Brain Res Bull.* 1985;15:377-84.

[124] Bendotti C, Samanin R. 8-hydroxy-2-(di-n-propylamino)tetralin (8-OH-DPAT) elicits eating in free-feeding rats by acting on central serotonin neurons. *Eur J Pharmacol.* 1986;121:147-50.

[125] Currie PJ, Coscina DV. Diurnal-variations in the feeding response to 8-OH-DPAT injected into the dorsal or median raphe. *Neuroreport.* 1993;4:1105-7

[126] Fletcher PJ, Davies M. Dorsal raphe microinjection of 5-HT and indirect 5-HT agonists induces feeding in rats. *Eur J Pharmacol.* 1990;184:265-71.

[127] Hutson PH, Dourish CT, Curzon G. Evidence that the hyperphagic response to 8-OH-DPAT is mediated by 5-HT<sub>1A</sub> receptors. *Eur J Pharmacol.* 1988;150:361-6.

[128] Hutson PH, Dourish CT, Curzon G. Neurochemical and behavioral evidence for mediation of the hyperphagic action of 8-OH-DPAT by 5-HT cell body autoreceptors. *Eur J Pharmacol.* 1986;129:347-52.

[129] Dourish CT, Clark ML, Iversen SD. 8-OH-DPAT elicits feeding and not chewing - evidence from liquid diet studies and a diet choice test. *Psychopharmacology.* 1988;95:185-8.

[130] Aulakh CS, Wozniak KM, Haas M, Hill JL, Zohar J, Murphy DL. Food intake, neuroendocrine and temperature effects of 8-OHDPAT in the rat. *Eur J Pharmacol.* 1988;146:253-9.

[131] Ebenezer IS. Effects of the 5-HT<sub>1A</sub> agonist 8-OH-DPAT on food intake in food-deprived rats. *Neuroreport.* 1992;3:1019-22.

[132] Voigt J-P, Kienzle A, Sohr R, Rex A, Fink H. Feeding and 8-OH-DPAT-related release of serotonin in the rat lateral hypothalamus. *Pharmacol Biochem Behav.* 2000;65:183-9.

[133] Jacobs BL. An animal behavior model for studying central serotonergic synapses. *Life Sci.* 1976;19:777-85.

[134] Lopez-Alonso VE, Mancilla-Diaz JM, Rito-Domingo M, Gonzalez-Hernandez B, Escartin-Perez RE. The effects of 5-HT<sub>1A</sub> and 5-HT<sub>2C</sub> receptor agonists on behavioral satiety sequence in rats. *Neurosci Lett.* 2007;416:285-8.

[135] Adell A, Carceller A, Artigas F. In vivo brain dialysis study of the somatodendritic release of serotonin in the raphe nuclei of the rat: Effects of 6-hydroxy-2-(di-n-propylamino)teralin. *J Neurochem.* 1993;60:1673-81.

[136] Casanovas JM, Lesourd M, Artigas F. The effect of the selective 5-HT<sub>1A</sub> agonists alnespirone (S-20499) and 8-OH-DPAT on extracellular 5-hydroxytryptamine in different regions of rat brain. *Br J Pharmacol.* 1997;122:733-41.

[137] Hjorth S, Sharp T. Effect of the 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT on the release of 5-HT in dorsal and median raphe-innervated rat brain regions as measured by in vivo microdialysis. *Life Sci.* 1991;48:1779-86.

[138] Voigt JP, Nwaiser B, Rex A, Mayer C, Fink H. Effect of 5-HT<sub>1A</sub> receptor activation on hypothalamic glucose. *Pharmacol Res.* 2004;50:359-65.

[139] Bechtholt AJ, Smith K, Gaughan S, Lucki I. Sucrose intake and fasting glucose levels in 5-HT(1A) and 5-HT(1B) receptor mutant mice. *Physiol Behav.* 2008;93:659-65.

[140] Bertrand PP, Bertrand RL. Serotonin release and uptake in the gastrointestinal tract. *Auton Neurosci*. 2010;153:47-57.

[141] Gershon MD, Tack J. The serotonin signaling system: From basic understanding to drug development-for functional GI disorders. *Gastroenterology*. 2007;132:397-414.

[142] Tonini M. 5-Hydroxytryptamine effects in the gut: the 3, 4, and 7 receptors. *Neurogastroent Motil*. 2005;17:637-42.

[143] Sanger GJ. 5-hydroxytryptamine and the gastrointestinal tract: where next? *Trends in Pharmacological Sciences*. 2008;29:465-71.

[144] Camilleri M. Serotonin in the gastrointestinal tract. *Curr Opin Endocrinol Diabetes Obes*. 2009;16:53-9.

[145] Fletcher PJ, Burton MJ. Effects of manipulations of peripheral serotonin on feeding and drinking in the rat. *Pharmacol Biochem and Behav*. 1984;20:835-40.

[146] Pollock JD, Rowland N. peripherally administered serotonin decreases food-Intake in rats. *Pharmacol Biochem and Behav*. 1981;15:179-83.

[147] Edwards S, Stevens R. Peripherally administered 5-hydroxytryptamine elicits the full behavioural sequence of satiety. *Physiol Behav*. 1991;50:1075-7.

[148] Simansky KJ, Jakubow J, Sisk FC, Vaidya AH, Eberle-Wang K. Peripheral serotonin is an incomplete signal for eliciting satiety in sham-feeding rats. *Pharmacol Biochem Behav*. 1992;43:847-54.

[149] Simansky KJ, Sisk FC, Vaidya AH, Eberle-Wang K. Peripherally administered alpha-methyl-5-hydroxy-tryptamine and 5-carboxamidotryptamine reduce food intake via different mechanisms in rats. *Behav Pharmacol*. 1989;1:241-6.

[150] Eberle-Wang K, Braun BT, Simansky KJ. Serotonin contracts the isolated rat pylorus via a 5-HT<sub>2</sub>-like receptor. *Am J Physiol*. 1994;266:R284-91.

[151] Bert B, Fink H, Walther DJ, Voigt JP. Peripheral serotonin and the control of food intake. *Fund Clin Pharmacol.* 2008;22:122.

[152] Berthoud HR. Multiple neural systems controlling food intake and body weight. *Neurosci Biobehav Rev.* 2002;26:393-428.

[153] Grill HJ, Hayes MR. Hindbrain Neurons as an Essential Hub in the Neuroanatomically Distributed Control of Energy Balance. *Cell Metabolism.* 2012;16:296-309.

[154] Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG, et al. Ghrelin enhances appetite and increases food intake in humans. *J Clin Endocrinol Metab.* 2001;86:5992.

[155] Tschop M, Smiley DL, Heiman ML. Ghrelin induces adiposity in rodents. *Nature.* 2000;407:908-13.

[156] Clark JT, Kalra PS, Crowley WR, Kalra SP. Neuropeptide Y and human pancreatic polypeptide stimulate feeding behavior in rats. *Endocrinology.* 1984;115:427-9.

[157] Currie PJ, Coiro CD, Niyomchai T, Lira A, Farahmand F. Hypothalamic paraventricular 5-hydroxytryptamine: Receptor-specific inhibition of NPY-stimulated eating and energy metabolism. *Pharmacol Biochem and Behav.* 2002;71:709-16.

[158] Clemett DA, Punhani T, Duxon MS, Blackburn TP, Fone KC. Immunohistochemical localisation of the 5-HT<sub>2C</sub> receptor protein in the rat CNS. *Neuropharmacology.* 2000;39:123-32.

[159] Heisler LK, Cowley MA, Tecott LH, Fan W, Low MJ, Smart JL, et al. Activation of central melanocortin pathways by fenfluramine. *Science.* 2002;297:609-11.

[160] Xu Y, Jones JE, Kohno D, Williams KW, Lee CE, Choi MJ, et al. 5-HT<sub>2C</sub>Rs expressed by pro-opiomelanocortin neurons regulate energy homeostasis. *Neuron.* 2008;60:582-9.

[161] Sim LJ, Joseph SA. Arcuate nucleus projections to brainstem regions which modulate nociception. *J Chem Neuroanat.* 1991;4:97-109.



1  
2  
3  
4  
5 [162] Kiss JZ, Cassell MD, Palkovits M. Analysis of the ACTH/beta-End/alpha-MSH-  
6 immunoreactive afferent input to the hypothalamic paraventricular nucleus of rat. *Brain Res.*  
7 1984;324:91-9.  
8  
9

10  
11 [163] Zec N, Filiano JJ, Kinney HC. Anatomic relationships of the human arcuate nucleus of  
12 the medulla: a Dil-labeling study. *Journal of neuropathology and experimental neurology.*  
13 1997;56:509-22.  
14  
15

16  
17 [164] Choi S, Blake V, Cole S, Fernstrom JD. Effects of chronic fenfluramine administration  
18 on hypothalamic neuropeptide mRNA expression. *Brain Research.* 2006;1087:83-6.  
19  
20

21  
22 [165] Berthoud HR. Metabolic and hedonic drives in the neural control of appetite: who is the  
23 boss? *Curr Opin Neurobiol.* 2011;21:888-96.  
24  
25

26  
27 [166] Lam DD, Przydzial MJ, Ridley SH, Yeo GSH, Rochford JJ, O'Rahilly S, et al. Serotonin  
28 5-HT<sub>2C</sub> receptor agonist promotes hypophagia via downstream activation of melanocortin 4  
29 receptors. *Endocrinology.* 2008;149:1323-8.  
30  
31

32  
33 [167] Rowland NE, Fakhari KJ, Robertson KL, Haskell-Luevano C. Effect of serotonergic  
34 anorectics on food intake and induction of Fos in brain of mice with disruption of  
35 melanocortin 3 and/or 4 receptors. *Pharmacol Biochem and Behav.* 2010;97:107-11.  
36  
37

38  
39 [168] Heisler LK, Cowley MA, Kishi T, Tecott LH, Fan W, Low MJ, et al. Central serotonin  
40 and melanocortin pathways regulating energy homeostasis. *Melanocortin System.*  
41 2003;994:169-74.  
42  
43

44  
45 [169] Tiligada E, Wilson JF. Regulation of alpha-melanocyte-stimulating hormone-release  
46 from superfused slices of rat hypothalamus by serotonin and the interaction of serotonin with  
47 the dopaminergic system inhibiting peptide release. *Brain Res.* 1989;503:225-8.  
48  
49

50  
51 [170] Heisler LK, Jobst EE, Sutton GM, Zhou LG, Borok E, Thornton-Jones Z, et al.  
52 Serotonin reciprocally regulates melanocortin neurons to modulate food intake. *Neuron.*  
53 2006;51:239-49.  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

[171] Sohn JW, Xu Y, Jones JE, Wickman K, Williams KW, Elmquist JK. Serotonin 2C receptor activates a distinct population of arcuate pro-opiomelanocortin neurons via TRPC channels. *Neuron*. 2011;71:488-97.

[172] Leibowitz SF, Alexander JT. Hypothalamic serotonin in control of eating behavior, meal size, and body weight. *Biol Psychiatry*. 1998;44:851-64.

[173] Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obesity*. 2001;25:S63-S7.

[174] Atasoy D, Betley JN, Su HH, Sternson SM. Deconstruction of a neural circuit for hunger. *Nature*. 2012;488:172-7.

[175] Kishi T, Aschkenasi CJ, Lee CE, Mountjoy KG, Saper CB, Elmquist JK. Expression of melanocortin 4 receptor mRNA in the central nervous system of the rat. *J Comp Neurol*. 2003;457:213-35.

[176] Siljee JE, Unmehopa UA, Kalsbeek A, Swaab DF, Fliers E, Alkemade A. Melanocortin 4 receptor distribution in the human hypothalamus. *Eur J Endocrinol*. 2013;168:361-9.

[177] Balthasar N, Dalgaard LT, Lee CE, Yu J, Funahashi H, Williams T, et al. Divergence of melanocortin pathways in the control of food intake and energy expenditure. *Cell*. 2005;123:493-505.

[178] Giraudo SQ, Billington CJ, Levine AS. Feeding effects of hypothalamic injection of melanocortin 4 receptor ligands. *Brain Res*. 1998;809:302-6.

[179] Wirth MM, Olszewski PK, Yu C, Levine AS, Giraudo SQ. Paraventricular hypothalamic alpha-melanocyte-stimulating hormone and MTII reduce feeding without causing aversive effects. *Peptides*. 2001;22:129-34.

[180] Kask A, Schiöth HB. Tonic inhibition of food intake during inactive phase is reversed by the injection of the melanocortin receptor antagonist into the paraventricular nucleus of the hypothalamus and central amygdala of the rat. *Brain Research*. 2000;887:460-4.

[181] Grignaschi g, Sironi F, Samanin R. The 5-hT-1B receptor mediates the effect of d-fenfluramine on eating caused by intra-hypothalamic injection of neuropeptide Y. *Eur J Pharmacol.* 1995;274:221-4.

[182] Fletcher PJ, Currie PJ, Chambers JW, Coscina DV. Radiofrequency lesions of the PVN fail to modify the effects of serotonergic drugs on food intake. *Brain Res.* 1993;630:1-9.

[183] Elias CF, Aschkenasi C, Lee C, Kelly J, Ahima RS, Bjorbaek C, et al. Leptin differentially regulates NPY and POMC neurons projecting to the lateral hypothalamic area. *Neuron.* 1999;23:775-86.

[184] Sahu A. Leptin decreases food intake induced by melanin-concentrating hormone (MCH), galanin (GAL) and neuropeptide Y (NPY) in the rat. *Endocrinology.* 1998;139:4739-42.

[185] Smith FJ, Campfield LA, Moschera JA, Bailon PS, Burn P. Brain administration of OB protein (leptin) inhibits neuropeptide-Y-induced feeding in ob/ob mice. *Regul Pept.* 1998;75-76:433-9.

[186] Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, et al. Neurons containing hypocretin (orexin) project to multiple neuronal systems. *Journal of Neuroscience.* 1998;18:9996-10015.

[187] Nambu T, Sakurai T, Mizukami K, Hosoya Y, Yanagisawa M, Goto K. Distribution of orexin neurons in the adult rat brain. *Brain Res.* 1999;827:243-60.

[188] Peyron C, Petit J-M, Rampan C, Jouviet M, Luppi P-H. Forebrain afferents to the rat dorsal raphe nucleus demonstrated by retrograde and anterograde tracing methods. *Neuroscience.* 1998;82:443-68.

[189] Brown RE, Sergeeva OA, Eriksson KS, Haas HL. Convergent excitation of dorsal raphe serotonin neurons by multiple arousal systems (orexin/hypocretin, histamine and noradrenaline). *Journal of Neuroscience.* 2002;22:8850-9.

[190] Collin M, Backberg M, Onnestam K, Meister B. 5-HT1A receptor immunoreactivity in hypothalamic neurons involved in body weight control. *Neuroreport.* 2002;13:945-51.

[191] Nonogaki K, Kaji T. Hypothalamic orexin and pro-opiomelanocortin activities are essential for the anorexic effects of m-chlorophenylpiperazine in mice. *Int J Neuropsychoph.* 2010;13:1261-7.

[192] Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature.* 1999;402:656-60.

[193] Ariyasu H, Takaya K, Tagami T, Ogawa Y, Hosoda K, Akamizu T, et al. Stomach is a major source of circulating ghrelin, and feeding state determines plasma ghrelin-like immunoreactivity levels in humans. *J Clin Endocrinol Metab.* 2001;86:4753-8.

[194] Bagnasco M, Tulipano G, Melis MR, Argiolas A, Cocchi D, Muller EE. Endogenous ghrelin is an orexigenic peptide acting in the arcuate nucleus in response to fasting. *Regul Pept.* 2003;111:161-7.

[195] Lu S, Guan JL, Wang QP, Uehara K, Yamada S, Goto N, et al. Immunocytochemical observation of ghrelin-containing neurons in the rat arcuate nucleus. *Neurosci Lett.* 2002;321:157-60.

[196] Cowley MA, Smith RG, Diano S, Tschop M, Pronchuk N, Grove KL, et al. The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron.* 2003;37:649-61.

[197] Mondal MS, Date Y, Yamaguchi H, Toshinai K, Tsuruta T, Kangawa K, et al. Identification of ghrelin and its receptor in neurons of the rat arcuate nucleus. *Regul Pept.* 2005;126:55-9.

[198] Currie PJ, John CS, Nicholson ML, Chapman CD, Loera KE. Hypothalamic paraventricular 5-hydroxytryptamine inhibits the effects of ghrelin on eating and energy substrate utilization. *Pharmacol Biochem Behav.* 2010;97:152-5.

[199] Schellekens H, van Oeffelen WE, Dinan TG, Cryan JF. Promiscuous dimerization of the growth hormone secretagogue receptor (GHS-R1a) attenuates ghrelin-mediated signaling. *J Biol Chem.* 2013;288:181-91.

[200] Berthoud HR, Sutton GA, Townsend RL, Patterson LM, Zheng HY. Brainstem mechanisms integrating gut-derived satiety signals and descending forebrain information in the control of meal size. *Physiol Behav.* 2006;89:517-24.

[201] Grill HJ, Donahey JCK, King L, Kaplan JM. Contribution of caudal brainstem to d-fenfluramine anorexia. *Psychopharmacology.* 1997;130:375-81.

[202] Kaplan JM, Donahey J, Baird JP, Simansky KJ, Grill HJ. d-fenfluramine anorexia: dissociation of ingestion rate, meal duration, and meal size effects. *Pharmacol Biochem Behav.* 1997;57:223-9.

[203] Kaplan JM, Song S, Grill HJ. Serotonin receptors in the caudal brainstem are necessary and sufficient for the anorectic effect of peripherally administered mCPP. *Psychopharmacology (Berl).* 1998;137:43-9.

[204] Glatzle J, Sternini C, Robin C, Zittel TT, Wong H, Reeve JR, Jr., et al. Expression of 5-HT<sub>3</sub> receptors in the rat gastrointestinal tract. *Gastroenterology.* 2002;123:217-26.

[205] Hayes MR, Covasa M. Gastric distension enhances CCK-induced Fos-like immunoreactivity in the dorsal hindbrain by activating 5-HT<sub>3</sub> receptors. *Brain Res.* 2006;1088:120-30.

[206] Hayes MR, Moore RL, Shah SM, Covasa M. 5-HT<sub>3</sub> receptors participate in CCK-induced suppression of food intake by delaying gastric emptying. *Am J Physiol Regul Integr Comp Physiol.* 2004;287:R817-23.

[207] Mazda T, Yamamoto H, Fujimura M, Fujimiya M. Gastric distension-induced release of 5-HT stimulates c-fos expression in specific brain nuclei via 5-HT<sub>3</sub> receptors in conscious rats. *Am J Physiol Gastrointest Liver Physiol.* 2004;287:G228-35.

[208] Laporte AM, Koscielniak T, Ponchant M, Verge D, Hamon M, Gozlan H. Quantitative autoradiographic mapping of 5-HT<sub>3</sub> receptors in the rat CNS using [<sup>125</sup>I] lodo-zacopride and [<sup>3</sup>H] zacopride as radioligands. *Synapse.* 1992;10:271-81.

[209] Pratt GD, Bowery NG. The 5-HT<sub>3</sub> receptor ligand, [3H]BRL 43694, binds to presynaptic sites in the nucleus tractus solitarius of the rat. *Neuropharmacology*. 1989;28:1367-76.

[210] Castex N, Fioramonti J, Fargeas MJ, Bueno L. c-fos expression in specific rat brain nuclei after intestinal anaphylaxis: involvement of 5-HT<sub>3</sub> receptors and vagal afferent fibers. *Brain Res*. 1995;688:149-60.

[211] Wu Q, Clark MS, Palmiter RD. Deciphering a neuronal circuit that mediates appetite. *Nature*. 2012;483:594-U112.

[212] Fulwiler CE, Saper CB. Subnuclear organization of the efferent connections of the parabrachial nucleus in the rat. *Brain Res Rev*. 1984;7:229-59.

[213] Carter ME, Soden ME, Zweifel LS, Palmiter RD. Genetic identification of a neural circuit that suppresses appetite. *Nature*. 2013;503:111-14

[214] Lee MD, Aloyo VJ, Fluharty SJ, Simansky KJ. Infusion of the serotonin(1B) (5-HT<sub>1B</sub>) agonist CP-93,129 into the parabrachial nucleus potently and selectively reduces food intake in rats. *Psychopharmacology*. 1998;136:304-7.

[215] Simansky KJ, Nicklous DM. Parabrachial infusion of D-fenfluramine reduces food intake - Blockade by the 5-HT<sub>1B</sub> antagonist SB-216641. *Pharmacol Biochem Behav*. 2002;71:681-90.

[216] Cooper SJ, Dourish CT. Multiple cholecystokinin (CCK) receptors and CCK-monoamine interactions are instrumental in the control of feeding. *Physiol Behav*. 1990;48:849-57.

[217] Gibbs J, Young RC, Smith GP. Cholecystokinin decreases food intake in rats. *J Comp Physiol Psychol*. 1973;3:488-95.

[218] Funakoshi A, Miyasaka K, Shinozaki H, Masuda M, Kawanami T, Takata Y, et al. An animal model of congenital defect of gene expression of cholecystokinin (CCK)-A receptor. *Biochem Biophys Res Commun*. 1995;210:787-96.

[219] Moran TH, Katz LF, Plata-Salaman CR, Schwartz GJ. Disordered food intake and obesity in rats lacking cholecystokinin A receptors. *Am J Physiol.* 1998;274:R618-R25.

[220] Kopin AS, Mathes WF, McBride EW, Nguyen M, Al-Haider W, Schmitz F, et al. The cholecystokinin-A receptor mediates inhibition of food intake yet is not essential for the maintenance of body weight. *J Clin Invest.* 1999;103:383-91.

[221] Neill C, Cooper SJ. Evidence that d-fenfluramine anorexia is mediated by 5-HT1 receptors. *Psychopharmacology.* 1989;97:213-8.

[222] Stallone D, Nicolaidis S, Gibbs J. Cholecystokinin-induced anorexia depends on serotonergic function. *AmJPhysiol.* 1989;256:R1138-R41.

[223] Poeschla B, Gibbs J, Simansky KJ, Greenberg D, Smith GP. Cholecystokinin-induced satiety depends on activation of 5-HT1c receptors. *Am J Physiol.* 1993;264:R62-R4.

[224] Esfahani N, Bednar I, Qureshi GA, Sodersten P. Inhibition of serotonin synthesis attenuates inhibition of ingestive behavior by CCK-8. *Pharmacol Biochem Behav.* 1995;51:9-12.

[225] Poeschla B, Gibbs J, Simansky KJ, Smith GP. The 5HT1A agonist 8-OH-DPAT attenuates the satiating action of cholecystokinin. *Pharmacol Biochem Behav.* 1992;42:541-3.

[226] Voigt JP, Fink H, Marsden CA. Evidence for the involvement of the 5-HT1A receptor in CCK induced satiety in rats. *Naunyn Schmiedebergs Arch Pharmacol.* 1995;351:217-20.

[227] Ebenezer IS, Brooman J. Pretreatment with the 5-HT-1A receptor agonists 8-OH-DPAT or gepirone does not attenuate the inhibitory effect of systemically administered cholecystokinin (CCK) on food intake in rats. *Meth Find Exp Clin Pharmacol.* 1994;16:589-95.

[228] Boden PR, Woodruff GN, Pinnock RD. Pharmacology of a cholecystokinin receptor on 5-hydroxytryptamine neurons in the dorsal raphe of the rat. *Br J Pharmacol.* 1991;102:635-8.

[229] Asarian L. Loss of cholecystokinin and glucagon-like peptide-1-induced satiation in mice lacking serotonin 2C receptors. *Am J Physiol-Reg I.* 2009;296:R51-R6.

[230] Cooper SJ, Dourish CT, Barber DJ. Reversal of the anorectic effect of (+)-fenfluramine in the rat by the selective cholecystokinin receptor antagonist MK-329. *Br J Pharmacol.* 1990;99:65-70.

[231] Grignaschi G, Mantelli B, Fracasso C, Anelli M, Caccia S, Samanin R. Reciprocal interaction of 5-hydroxytryptamine and cholecystokinin in the control of feeding patterns in rats. *Br J Pharmacol.* 1993;109:491-4.

[232] Cooper SJ. Cholecystokinin modulation of serotonergic control of feeding behavior. *Ann N Y Acad Sci.* 1996;780:213-22.

[233] Eberle-Wang K, Simansky KJ. The CCK-A receptor antagonist, devazepide, blocks the anorectic action of CCK but not peripheral serotonin in rats. *Pharmacol Biochem Behav.* 1992;43:943-7.

[234] Francis J, Dourish CT, Cooper SJ. Devazepide attenuates dl-fenfluramine-induced suppression of gastric emptying but not food intake in the 17 h food-deprived rat. *Physiol Behav.* 1997;62:545-50.

[235] Voigt JP, Wenz D, Voits M, Fink H. Does increased endogenous CCK interact with serotonin to reduce food intake in rats? *Peptides.* 2000;21:1895-901.

[236] Weller A, Smith GP, Gibbs J. Endogenous Cholecystokinin Reduces Feeding in Young-Rats. *Science.* 1990;247:1589-91.

[237] Zippel U, Heidel E, Davidowa H. Action of cholecystokinin and serotonin on lateral hypothalamic neurons of rats. *Eur J Pharmacol.* 1999;379:135-40.

[238] Crawley JN, Corwin RL. Biological actions of cholecystokinin. *Peptides.* 1994;15:731-55.

[239] Crawley JN, Kiss JZ. Paraventricular nucleus lesions abolish the inhibition of feeding induced by systemic cholecystokinin. *Peptides.* 1985;6:927-35.



[240] Helm KA, Rada P, Hoebel BG. Cholecystokinin combined with serotonin in the hypothalamus limits accumbens dopamine release while increasing acetylcholine: a possible satiation mechanism. *Brain Research*. 2003;963:290-7.

[241] Aja S. Serotonin-3 receptors in gastric mechanisms of cholecystokinin-induced satiety. *Am J Physiol-Reg I*. 2006;291:R112-R4.

[242] Burton-Freeman B, Gietzen DW, Schneeman BO. Cholecystokinin and serotonin receptors in the regulation of fat-induced satiety in rats. *Am J Physiol*. 1999;276:R429-R34.

[243] Daughters RS, Hofbauer RD, Grossman AW, Marshall AM, Brown EM, Hartman BK, et al. Ondansetron attenuates CCK induced satiety and c-fos labeling in the dorsal medulla. *Peptides*. 2001;22:1331-8.

[244] Hayes MR, Savastano DM, Covasa M. Cholecystokinin-induced satiety is mediated through interdependent cooperation of CCK-A and 5-HT<sub>3</sub> receptors. *Physiol Behav*. 2004;82:663-9

[245] Hayes MR, Covasa M. CCK and 5-HT act synergistically to suppress food intake through simultaneous activation of CCK-1 and 5-HT<sub>3</sub> receptors. *Peptides*. 2005;26:2322-30.

[246] Kennedy GC. The role of depot fat in the hypothalamic control of food intake in the rat. *Proc R Soc Lond B Biol Sci*. 1953;140:578-96.

[247] Coleman DL. Effects of parabiosis of obese with diabetes and normal mice. *Diabetologia*. 1973;9:294-8.

[248] Zhang YY, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homolog. *Nature*. 1994;372:425-32.

[249] Friedman JM. Leptin, leptin receptors and the control of body weight. *Eur J Med Res*. 1997;2:7-13.

[250] Ahima RS, Flier JS. Adipose tissue as an endocrine organ. *Trends Endocrin Met*. 2000;11:327-32.

[251] Friedman MJ. Leptin and the neural circuit regulating body weight and metabolism. In: Kordon C, Hanoune J, Danyzer R, Christen Y, editotrs. Brain somatic cross-talk and the central control of metabolism, Berlin Heidelberg: Springer; 2003. p. 15-35.

[252] Pelleymounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T, et al. Effects of the obese gene product on body weight regulation in ob/ob mice. *Science*. 1995;269:540-3.

[253] Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, et al. Weight-reducing effects of the plasma protein encoded by the obese gene. *Science*. 1995;269:543-6.

[254] Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P. Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science*. 1995;269:546-9.

[255] Weigle DS, Bukowski TR, Foster DC, Holderman S, Kramer JM, Lasser G, et al. Recombinant Ob protein reduces feeding and body-weight in the Ob/Ob mouse. *J Clin Invest*. 1995;96:2065-70.

[256] Halford JCG, Blundell JE. Separate systems for serotonin and leptin in appetite control. *Ann Med*. 2000;32:222-32.

[257] von Meyenburg C, Langhans W, Hrupka BJ. Evidence that the anorexia induced by lipopolysaccharide is mediated by the 5-HT<sub>2C</sub> receptor. *Pharmacol Biochem Behav*. 2003;74:505-12.

[258] Yamada J, Sugimoto Y, Hirose H, Kajiwara Y. Role of serotonergic mechanisms in leptin-induced suppression of milk intake in mice. *Neurosci Lett*. 2003;348:195-7.

[259] Wang B, Chehab FF. Deletion of the serotonin 2c receptor from transgenic mice overexpressing leptin does not affect their lipodystrophy but exacerbates their diet-induced obesity. *Biochem Biophys Res Commun*. 2006;351:418-23.

[260] Yamada J, Sugimoto Y, Ujikawa M. The serotonin precursor 5-hydroxytryptophan elevates serum leptin levels in mice. *Eur J Pharmacol.* 1999;383:49-51.

[261] Yamada J, Sugimoto Y, Ujikawa M. Involvement of leptin in hypophagia induced by the serotonin precursor 5-hydroxytryptophan (5-HTP) in mice. *Biol Pharm Bull.* 2006;29:557-9.

[262] Yamada J, Ujikawa M, Sugimoto Y. Serum leptin levels after central and systemic injection of a serotonin precursor, 5-hydroxytryptophan, in mice. *Eur J Pharmacol.* 2000;406:159-62.

[263] Fernandez-Galaz MC, Fernandez-Agullo T, Carrascosa JM, Ros M, Garcia-Segura LM. Leptin accumulation in hypothalamic and dorsal raphe neurons is inversely correlated with brain serotonin content. *Brain Res.* 2010;1329:194-202.

[264] Chen X, Margolis KJ, Gershon MD, Schwartz GJ, Sze JY. Reduced serotonin reuptake transporter (SERT) function causes insulin resistance and hepatic steatosis independent of food intake. *PLoS One.* 2012;7:e32511.

[265] Shpilman M, Niv-Spector L, Katz M, Varol C, Solomon G, Ayalon-Soffer M, et al. Development and characterization of high affinity leptins and leptin antagonists. *J Biol Chem.* 2011;286:4429-42.

[266] Schellekens H, Clarke G, Jeffery IB, Dinan TG, Cryan JF. Dynamic 5-HT<sub>2C</sub> receptor editing in a mouse model of obesity. *PLoS One.* 2012;7:e32266.

[267] Calapai G, Corica F, Corsonello A, Sautebin L, Di Rosa M, Campo GM, et al. Leptin increases serotonin turnover by inhibition of brain nitric oxide synthesis. *J Clin Invest.* 1999;104:975-82.

[268] Sohn JW, Williams KW. Functional heterogeneity of arcuate nucleus pro-opiomelanocortin neurons: implications for diverging melanocortin pathways. *Mol Neurobiol.* 2012;45:225-33.

[269] Finn PD, Cunningham MJ, Rickard DG, Clifton DK, Steiner RA. Serotonergic neurons are targets for leptin in the monkey. *J Clin Endocrinol Metab.* 2001;86:422-6.

[270] Morrison SF. Activation of 5-HT<sub>1A</sub> receptors in raphe pallidus inhibits leptin-evoked increases in brown adipose tissue thermogenesis. *Am J Physiol Regul Integr Comp Physiol*. 2004;286:R832-7.

[271] Fernandez-Galaz MC, Diano S, Horvath TL, Garcia-Segura LM. Leptin uptake by serotonergic neurones of the dorsal raphe. *J Neuroendocrinol*. 2002;14:429-34.

[272] Seeley RJ, van Dijk G, Campfield LA, Smith FJ, Burn P, Nelligan JA, et al. Intraventricular leptin reduces food intake and body weight of lean rats but not obese Zucker rats. *Horm Metab Res*. 1996;28:664-8.

[273] Widdowson PS, Upton R, Buckingham R, Arch J, Williams G. Inhibition of food response to intracerebroventricular injection of leptin is attenuated in rats with diet-induced obesity. *Diabetes*. 1997;46:1782-5.

[274] Hulseley MG, Lu H, Wang T, Martin RJ, Baile CA. Intracerebroventricular (i.c.v.) administration of mouse leptin in rats: behavioral specificity and effects on meal patterns. *Physiol Behav*. 1998;65:445-55.

[275] Cusin I, Rohner-Jeanrenaud F, Stricker-Krongrad A, Jeanrenaud B. The weight-reducing effect of an intracerebroventricular bolus injection of leptin in genetically obese *fa/fa* rats. Reduced sensitivity compared with lean animals. *Diabetes*. 1996;45:1446-50.

[276] Choi S, Sparks R, Clay M, Dallman MF. Rats with hypothalamic obesity are insensitive to central leptin injections. *Endocrinology*. 1999;140:4426-33.

[277] Hay-Schmidt A, Helboe L, Larsen PJ. Leptin receptor immunoreactivity is present in ascending serotonergic and catecholaminergic neurons of the rat. *Neuroendocrinology*. 2001;73:215-26.

[278] Collin M, Hakansson-Ovesjo ML, Misane I, Ogren SO, Meister B. Decreased 5-HT transporter mRNA in neurons of the dorsal raphe nucleus and behavioral depression in the obese leptin-deficient *ob/ob* mouse. *Brain Res Mol Brain Res*. 2000;81:51-61.

[279] Phillips MS, Liu Q, Hammond HA, Dugan V, Hey PJ, Caskey CJ, et al. Leptin receptor missense mutation in the fatty Zucker rat. *Nat Genet.* 1996;13:18-9.

[280] White BD, Martin RJ. Evidence for a central mechanism of obesity in the Zucker rat: role of neuropeptide Y and leptin. *Proc Soc Exp Biol Med.* 1997;214:222-32.

[281] Chua SC, Jr., White DW, Wu-Peng XS, Liu SM, Okada N, Kershaw EE, et al. Phenotype of fatty due to Gln269Pro mutation in the leptin receptor (Lepr). *Diabetes.* 1996;45:1141-3.

[282] Ohliger-Frerking P, Horwitz BA, Horowitz JM. Serotonergic dorsal raphe neurons from obese Zucker rats are hyperexcitable. *Neuroscience.* 2003;120:627-34.

[283] Telles MM, Guimaraes RB, Ribeiro EB. Effect of leptin on the acute feeding-induced hypothalamic serotonergic stimulation in normal rats. *Regul Pept.* 2003;115:11-8.

[284] Tordera R, Pei Q, Newson M, Gray K, Sprakes M, Sharp T. Effect of different 5-HT1A receptor antagonists in combination with paroxetine on expression of the immediate-early gene Arc in rat brain. *Neuropharmacology.* 2003;44:893-902.

[285] Yadav VK, Oury F, Tanaka KF, Thomas T, Wang Y, Cremers S, et al. Leptin-dependent serotonin control of appetite: temporal specificity, transcriptional regulation, and therapeutic implications. *J Exp Med.* 2011;208:41-52.

[286] Lam DD, Leininger GM, Louis GW, Garfield AS, Marston OJ, Leshan RL, et al. Leptin does not directly affect CNS serotonin neurons to influence appetite. *Cell Metab.* 2011;13:584-91.

[287] Donovan MH, Tecott LH. Serotonin and the regulation of mammalian energy balance. *Frontiers in neuroscience.* 2013;7:36.

[288] Ritter RC. Gastrointestinal mechanisms of satiation for food. *Physiol Behav.* 2004;81:249-73.

[289] Halford JC, Cooper GD, Dovey TM. The pharmacology of human appetite expression. *Curr Drug Targets.* 2004;5:221-40.

[290] Rodgers RJ, Tschop MH, Wilding JP. Anti-obesity drugs: past, present and future. *Dis Model Mech.* 2012;5:621-6.

[291] Woods SC, Porte D, Jr. The role of insulin as a satiety factor in the central nervous system. *Adv Metab Disord.* 1983;10:457-68.

[292] Vanderweele DA. Insulin as a satiating signal. In: Smith GP, editor. *Satiation: From gut to brain*, New York: Oxford University Press; 1998. p. 198-216.

[293] Woods SC, Lotter EC, McKay LD, Porte D, Jr. Chronic intracerebroventricular infusion of insulin reduces food intake and body weight of baboons. *Nature.* 1979;282:503-5.

[294] Bruning JC, Gautam D, Burks DJ, Gillette J, Schubert M, Orban PC, et al. Role of brain insulin receptor in control of body weight and reproduction. *Science.* 2000;289:2122-5.

[295] Orosco M, Rouch C, Gerozissis K. Activation of hypothalamic insulin by serotonin is the primary event of the insulin-serotonin interaction involved in the control of feeding. *Brain Res.* 2000;872:64-70.

[296] Zhou L, Sutton GM, Rochford JJ, Semple RK, Lam DD, Oksanen LJ, et al. Serotonin 2C receptor agonists improve type 2 diabetes via melanocortin-4 receptor signaling pathways. *Cell Metab.* 2007;6:398-405.

[297] Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, et al. Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature.* 2002;418:650-4.

[298] Rodgers RJ, Holch P, Tallett AJ. Behavioural satiety sequence (BSS): Separating wheat from chaff in the behavioural pharmacology of appetite. *Pharmacol Biochem Behav.* 2010.

[299] Holst JJ. The physiology of glucagon-like peptide 1. *Physiol Rev.* 2007;87:1409-39.

[300] Wright FL, Rodgers RJ. Behavioural profile of exendin-4/naltrexone dose combinations in male rats during tests of palatable food consumption. *Psychopharmacology (Berl).* 2014.

[301] Abbott CR, Monteiro M, Small CJ, Sajedi A, Smith KL, Parkinson JR, et al. The inhibitory effects of peripheral administration of peptide YY(3-36) and glucagon-like peptide-1 on food intake are attenuated by ablation of the vagal-brainstem-hypothalamic pathway. *Brain Res.* 2005;1044:127-31.

[302] Tang-Christensen M, Larsen PJ, Goke R, Fink-Jensen A, Jessop DS, Moller M, et al. Central administration of GLP-1-(7-36) amide inhibits food and water intake in rats. *Am J Physiol.* 1996;271:R848-56.

[303] Rowland NE, Crews EC, Gentry RM. Comparison of Fos induced in rat brain by GLP-1 and amylin. *Regul Pept.* 1997;71:171-4.

[304] Brunetti L, Orlando G, Recinella L, Leone S, Ferrante C, Chiavaroli A, et al. Glucagon-like peptide 1 (7-36) amide (GLP-1) and exendin-4 stimulate serotonin release in rat hypothalamus. *Peptides.* 2008;29:1377-81.

[305] Owji AA, Khoshdel Z, Sanea F, Panjehshahin MR, Shojaee Fard M, Smith DM, et al. Effects of intracerebroventricular injection of glucagon like peptide-1 and its related peptides on serotonin metabolism and on levels of amino acids in the rat hypothalamus. *Brain Res.* 2002;929:70-5.

[306] Nonogaki K, Suzuki M, Sanuki M, Wakameda M, Tamari T. The contribution of serotonin 5-HT<sub>2C</sub> and melanocortin-4 receptors to the satiety signaling of glucagon-like peptide 1 and liraglutide, a glucagon-like peptide 1 receptor agonist, in mice. *Biochem Biophys Res Commun.* 2011;411:445-8.

[307] Lam DD, Zhou L, Vegge A, Xiu PY, Christensen BT, Osundiji MA, et al. Distribution and neurochemical characterization of neurons within the nucleus of the solitary tract responsive to serotonin agonist-induced hypophagia. *Behav Brain Res.* 2009;196:139-43.

[308] Larsen PJ, Fledelius C, Knudsen LB, Tang-Christensen M. Systemic administration of the long-acting GLP-1 derivative NN2211 induces lasting and reversible weight loss in both normal and obese rats. *Diabetes.* 2001;50:2530-9.

[309] Raun K, von Voss P, Gotfredsen CF, Golozoubova V, Rolin B, Knudsen LB. Liraglutide, a long-acting glucagon-like peptide-1 analog, reduces body weight and food

intake in obese candy-fed rats, whereas a dipeptidyl peptidase-IV inhibitor, vildagliptin, does not. *Diabetes*. 2007;56:8-15.

[310] Finkelstein JA, Chance WT, Fischer JE. Brain serotonergic activity and plasma amino acid levels in genetically obese Zucker rats. *Pharmacol Biochem Behav*. 1982;17:939-44.

[311] Tsujii S, Nakai Y, Fukata J, Nakaishi S, Takahashi H, Usui T, et al. Monoamine metabolism and its responses to food deprivation in the brain of Zucker rats. *Physiol Behav*. 1988;44:495-500.

[312] Orosco M, Trouvin JH, Cohen Y, Jacquot C. Ontogeny of brain monoamines in lean and obese female Zucker rats. *Physiol Behav*. 1986;36:853-6.

[313] Voigt J-P, Schade R, Fink H, Hörtnagl H. Role of 5-HT<sub>1A</sub> receptors in the control of food intake in obese Zucker rats of different ages. *Pharmacol Biochem Behav*. 2002;72:403-9.

[314] Meguid MM, Fetissov SO, Blaha V, Yang ZJ. Dopamine and serotonin VMN release is related to feeding status in obese and lean Zucker rats. *Neuroreport*. 2000;11:2069-72.

[315] Orosco M, Rouch C, Meile MJ, Nicolaidis S. Spontaneous feeding-related monoamine changes in rostromedial hypothalamus of the obese Zucker rat: a microdialysis study. *Physiol Behav*. 1995;57:1103-6.

[316] Lemierre S, Rouch C, Nicolaidis S, Orosco M. Combined effect of obesity and aging on feeding-induced monoamine release in the rostromedial hypothalamus of the Zucker rat. *Int J Obes Relat Metab Disord*. 1998;22:993-9.

[317] De Fanti BA, Hamilton JS, Horwitz BA. Meal-induced changes in extracellular 5-HT in medial hypothalamus of lean (Fa/Fa) and obese (fa/fa) Zucker rats. *Brain Res*. 2001;902:164-70.

[318] Grinker JA, Drewnowski A, Enns M, Kissileff H. Effects of d-amphetamine and fenfluramine on feeding patterns and activity of obese and lean Zucker rats. *Pharmacol Biochem Behav*. 1980;12:265-75.



[319] Orosco M, Bremond J, Jacquot C, Cohen Y. Fenfluramine and brain transmitters in the obese Zucker rat. *Neuropharmacology*. 1984;23:183-8.

[320] Chaouloff F, Coupry I, Baudrie V. Cortical [3H]ketanserin binding and 5-HT<sub>2A</sub> receptor-mediated behavioral responses in obese Zucker rats. *Pharmacol Biochem Behav*. 1995;50:309-12.

[321] Dryden S, Frankish HM, Wang Q, Pickavance L, Williams G. The serotonergic agent fluoxetine reduces neuropeptide Y levels and neuropeptide Y secretion in the hypothalamus of lean and obese rats. *Neuroscience*. 1996;72:557-66.

[322] Koulu M, Huupponen R, Hanninen H, Pesonen U, Rouru J, Seppala T. Hypothalamic neurochemistry and feeding behavioral responses to clonidine, an alpha-2-agonist, and to trifluoromethylphenylpiperazine, a putative 5-hydroxytryptamine-1B agonist, in genetically obese Zucker rats. *Neuroendocrinology*. 1990;52:503-10.

[323] Sharp T, Boothman L, Raley J, Queree P. Important messages in the 'post': recent discoveries in 5-HT neurone feedback control. *Trends Pharmacol Sci*. 2007;28:629-36.

[324] Horwitz BA, Hamilton JS, Routh VH, Green K, Havel P, Chan A. Adiposity and serum leptin increase in fatty (fa/fa) BNZ neonates without decreased VMH serotonergic activity. *AmJPhysiol*. 1998;274:E1009-E17.

[325] Levin BE, Triscari J, Sullivan AC. Altered sympathetic activity during development of diet-induced obesity in rat. *Am J Physiol*. 1983;244:R347-55.

[326] Hassanain M, Levin BE. Dysregulation of hypothalamic serotonin turnover in diet-induced obese rats. *Brain Res*. 2002;929:175-80.

[327] Park S, Harrold JA, Widdowson PS, Williams G. Increased binding at 5-HT(1A), 5-HT(1B), and 5-HT(2A) receptors and 5-HT transporters in diet-induced obese rats. *Brain Res*. 1999;847:90-7.

[328] Banas SM, Rouch C, Kassis N, Markaki EM, Gerozissis K. A dietary fat excess alters metabolic and neuroendocrine responses before the onset of metabolic diseases. *Cell Mol Neurobiol.* 2009;29:157-68.

[329] Blaha V, Yang ZJ, Meguid MM, Chai JK, Oler A, Zadak Z. Ventromedial nucleus of hypothalamus is related to the development of cancer-induced anorexia: in vivo microdialysis study. *Acta Medica (Hradec Kralove).* 1998;41:3-11.

[330] Varma M, Torelli GF, Meguid MM, Chai JK, Blaha V, Laviano A, et al. Potential strategies for ameliorating early cancer anorexia. *The Journal of surgical research.* 1999;81:69-76.

[331] Adan RA. Mechanisms underlying current and future anti-obesity drugs. *Trends Neurosci.* 2013;36:133-40.

[332] Halford JC. Clinical pharmacotherapy for obesity: current drugs and those in advanced development. *Curr Drug Targets.* 2004;5:637-46.

[333] Garfield AS, Heisler LK. Pharmacological targeting of the serotonergic system for the treatment of obesity. *J Physiol.* 2009;587:49-60.

[334] Heal DJ, Gosden J, Smith SL. A review of late-stage CNS drug candidates for the treatment of obesity. *Int J Obes (Lond).* 2012.

[335] Heal DJ, Smith SL, Fisas A, Codony X, Buschmann H. Selective 5-HT<sub>6</sub> receptor ligands: progress in the development of a novel pharmacological approach to the treatment of obesity and related metabolic disorders. *Pharmacol Ther.* 2008;117:207-31.

[336] Heal DJ, Gosden J, Smith SL. A review of late-stage CNS drug candidates for the treatment of obesity. *Int J Obes (Lond).* 2013;37:107-17.

[337] Bello NT, Liang NC. The use of serotonergic drugs to treat obesity--is there any hope? *Drug Des Devel Ther.* 2011;5:95-109.

[338] Bickerdike MJ, Vickers SP, Dourish CT. 5-HT<sub>2C</sub> receptor modulation and the treatment of obesity. *Diabetes Obe Metab.* 1999;1:207-14.

[339] Ni W, Li MW, Thakali K, Fink GD, Watts SW. The fenfluramine metabolite (+)-norfenfluramine is vasoactive. *J Pharmacol Exp Ther.* 2004;309:845-52.

[340] Rothman RB, Baumann MH. Serotonergic drugs and valvular heart disease. *Expert Opin Drug Saf.* 2009;8:317-29.

[341] Higgs S, Cooper AJ, Barnes NM. Reversal of sibutramine-induced anorexia with a selective 5-HT(2C) receptor antagonist. *Psychopharmacology (Berl).* 2011;214:941-7.

[342] Jackson HC, Bearham MC, Hutchins LJ, Mazurkiewicz SE, Needham AM, Heal DJ. Investigation of the mechanisms underlying the hypophagic effects of the 5-HT and noradrenaline reuptake inhibitor, sibutramine, in the rat. *Br J Pharmacol.* 1997;121:1613-8.

[343] Lanfumey L, Hamon M. 5-HT1 receptors. *Curr Drug Targets CNS Neurol Disord.* 2004;3:1-10.

[344] Smith SR, Prosser WA, Donahue DJ, Morgan ME, Anderson CM, Shanahan WR, et al. Lorcaserin (APD356), a selective 5-HT(2C) agonist, reduces body weight in obese men and women. *Obesity (Silver Spring).* 2009;17:494-503.

[345] Thomsen WJ, Grottick AJ, Menzaghi F, Reyes-Saldana H, Espitia S, Yuskin D, et al. Lorcaserin, a novel selective human 5-hydroxytryptamine2C agonist: in vitro and in vivo pharmacological characterization. *J Pharmacol Exp Ther.* 2008;325:577-87.

[346] Bray GA, Ryan DH. Update on obesity pharmacotherapy. *Ann N Y Acad Sci.* 2014;1311:1-13.

[347] Jensen NH, Cremers TI, Sotty F. Therapeutic potential of 5-HT2C receptor ligands. *Scientific World Journal.* 2010;10:1870-85.

[348] Halford JC. Lorcaserin--not a new weapon in the battle with appetite. *Nature reviews Endocrinology.* 2010;6:663-4.

[349] Dawson GR, Craig KJ, Dourish CT. Validation of experimental medicine methods in psychiatry: the P1vital approach and experience. *Biochem Pharmacol.* 2011;81:1435-41.

[350] Thomas JM, Dourish CT, Tomlinson JW, Hassan-Smith Z, Higgs S. Effects of the 5-HT<sub>2C</sub> receptor agonist meta-chlorophenylpiperazine on appetite, food intake and emotional processing in healthy volunteers. *Psychopharmacology (Berl)*. 2014;231:2449-59.

[351] Niswender CM, Sanders-Bush E, Emeson RB. Identification and characterization of RNA editing events within the 5-HT<sub>2C</sub> receptor. *Ann N Y Acad Sci*. 1998;861:38-48.

[352] Burns CM, Chu H, Rueter SM, Hutchinson LK, Canton H, Sanders-Bush E, et al. Regulation of serotonin-2C receptor G-protein coupling by RNA editing. *Nature*. 1997;387:303-8.

[353] Decher N, Netter MF, Streit AK. Putative impact of RNA editing on drug discovery. *Chem Biol Drug Des*. 2013;81:13-21.

[354] Schellekens H, Dinan TG, Cryan JF. Taking two to tango: a role for ghrelin receptor heterodimerization in stress and reward. *Front Neurosci*. 2013;7:148.

[355] Tallett AJ, Blundell JE, Rodgers RJ. Sibutramine & naloxone: infra-additive interaction in the regulation of appetite? *Behav Brain Res*. 2010;207:174-81.

[356] Rowland NE, Mukherjee M, Robertson K. Effects of the cannabinoid receptor antagonist SR 141716, alone and in combination with dexfenfluramine or naloxone, on food intake in rats. *Psychopharmacology (Berl)*. 2001;159:111-6.

[357] Green AR, Marsden CA. How do we re-engage the pharmaceutical industry in research on serotonin and psychiatric disorders? *Acs Chem Neurosci*. 2013;4:9-12.

[358] Thurlby PL, Samanin R. Effects of anorectic drugs and prior feeding on food-rewarded runway behavior. *Pharmacol Biochem Behav*. 1981;14:799-804.

[359] Higgins GA, Silenieks LB, Rossmann A, Rizos Z, Noble K, Soko AD, et al. The 5-HT<sub>2C</sub> receptor agonist lorcaserin reduces nicotine self-administration, discrimination, and reinstatement: relationship to feeding behavior and impulse control. *Neuropsychopharmacology*. 2012;37:1177-91.

[360] Ward SJ, Lefever TW, Jackson C, Tallarida RJ, Walker EA. Effects of a Cannabinoid1 receptor antagonist and Serotonin2C receptor agonist alone and in combination on motivation for palatable food: a dose-addition analysis study in mice. *J Pharmacol Exp Ther.* 2008;325:567-76.

[361] Johnson AW. Eating beyond metabolic need: how environmental cues influence feeding behavior. *Trends Neurosci.* 2013;36:101-9.

[362] Saper CB, Chou TC, Elmquist JK. The need to feed: homeostatic and hedonic control of eating. *Neuron.* 2002;36:199-211.

[363] Figlewicz DP, MacDonald Naleid A, Sipols AJ. Modulation of food reward by adiposity signals. *Physiol Behav.* 2007;91:473-8.

[364] Berthoud HR. The neurobiology of food intake in an obesogenic environment. *Proc Nutr Soc.* 2012;71:478-87.

[365] Hajnal A, Smith GP, Norgren R. Oral sucrose stimulation increases accumbens dopamine in the rat. *Am J Physiol Regul Integr Comp Physiol.* 2004;286:R31-7.

[366] Higgins GA, Sellers EM, Fletcher PJ. From obesity to substance abuse: therapeutic opportunities for 5-HT<sub>2C</sub> receptor agonists. *Trends Pharmacol Sci.* 2013;34:560-70.

[367] Hayes DJ, Greenshaw AJ. 5-HT receptors and reward-related behaviour: a review. *Neurosci Biobehav Rev.* 2011;35:1419-49.

[368] Lauder JM. Ontogeny of the serotonergic system in the rat: serotonin as a developmental signal. *Ann N Y Acad Sci.* 1990;600:297-313.

[369] Whitaker-Azmitia PM. Serotonin and development. In: Mueller CP, Jacobs, B.L., editors. *Handbook of the behavioural neurobiology of serotonin*, New York: Academic Press; 2010. p. 309-24.

[370] Langley-Evans SC. Nutritional programming of disease: unravelling the mechanism. *J Anat.* 2009;215:36-51.

[371] Kirk SL, Samuelsson AM, Argenton M, Dhonye H, Kalamatianos T, Poston L, et al. Maternal obesity induced by diet in rats permanently influences central processes regulating food intake in offspring. *PLoS One*. 2009;4:e5870.

[372] Levin BE. Metabolic imprinting: critical impact of the perinatal environment on the regulation of energy homeostasis. *Philosophical transactions of the Royal Society of London Series B, Biological sciences*. 2006;361:1107-21.

[373] Bouret SG. Role of early hormonal and nutritional experiences in shaping feeding behavior and hypothalamic development. *J Nutr*. 2010;140:653-7.

[374] Alfaradhi MZ, Ozanne SE. Developmental programming in response to maternal overnutrition. *Front Genet*. 2011;2:27.

[375] Porto LC, Sardinha FL, Telles MM, Guimaraes RB, Albuquerque KT, Andrade IS, et al. Impairment of the serotonergic control of feeding in adult female rats exposed to intra-uterine malnutrition. *Br J Nutr*. 2009;101:1255-61.

[376] Wright TM, Fone KC, Langley-Evans SC, Voigt JP. Exposure to maternal consumption of cafeteria diet during the lactation period programmes feeding behaviour in the rat. *Int J Dev Neurosci*. 2011;29:785-93.

[377] Oswal A, Yeo G. Leptin and the control of body weight: a review of its diverse central targets, signaling mechanisms, and role in the pathogenesis of obesity. *Obesity (Silver Spring)*. 2010;18:221-9.

[378] Druce M, Bloom SR. The regulation of appetite. *Arch Dis Child*. 2006;91:183-7.

[379] Valassi E, Scacchi M, Cavagnini F. Neuroendocrine control of food intake. *Nutrition, metabolism, and cardiovascular diseases : NMCD*. 2008;18:158-68.

[380] Mosienko V, Beis D, Pasqualetti M, Waider J, Matthes S, Qadri F, et al. Life without brain serotonin: Reevaluation of serotonin function with mice deficient in brain serotonin synthesis. *Behav Brain Res*. 2014.

[381] Muller CP, Carey RJ. Intracellular 5-HT 2C-receptor dephosphorylation: a new target for treating drug addiction. *Trends Pharmacol Sci.* 2006;27:455-8.

[382] Backstrom JR, Price RD, Reasoner DT, Sanders-Bush E. Deletion of the serotonin 5-HT2C receptor PDZ recognition motif prevents receptor phosphorylation and delays resensitization of receptor responses. *J Biol Chem.* 2000;275:23620-6.

[383] Paulmann N, Grohmann M, Voigt JP, Bert B, Vowinckel J, Bader M, et al. Intracellular serotonin modulates insulin secretion from pancreatic beta-cells by protein serotonylation. *PLoS Biol.* 2009;7:e1000229.

[384] Walther DJ, Stahlberg S, Vowinckel J. Novel roles for biogenic monoamines: from monoamines in transglutaminase-mediated post-translational protein modification to monoaminylation deregulation diseases. *FEBS J.* 2011;278:4740-55.

[385] Langley-Evans SC. Nutrition in early life and the programming of adult disease: a review. *J Hum Nutr Diet* 2014 (in press).

[386] Godfrey KM, Inskip HM, Hanson MA. The long-term effects of prenatal development on growth and metabolism. *Semin Reprod Med.* 2011;29:257-65.

[387] Bonnin A, Levitt P. Fetal, maternal, and placental sources of serotonin and new implications for developmental programming of the brain. *Neuroscience.* 2011;197:1-7.

[388] Fernstrom JD, Wurtman RJ. Brain serotonin content: increase following ingestion of carbohydrate diet. *Science.* 1971;174:1023-5.

[389] Lopes de Souza S, Orozco-Solis R, Grit I, Manhaes de Castro R, Bolanos-Jimenez F. Perinatal protein restriction reduces the inhibitory action of serotonin on food intake. *Eur J Neurosci.* 2008;27:1400-8.

[390] Manuel-Apolinar L, Rocha L, Damasio L, Tesoro-Cruz E, Zarate A. Role of prenatal undernutrition in the expression of serotonin, dopamine and leptin receptors in adult mice: implications of food intake. *Mol Med Rep.* 2014;9:407-12.

[391] Bouret SG, Draper SJ, Simerly RB. Formation of projection pathways from the arcuate nucleus of the hypothalamus to hypothalamic regions implicated in the neural control of feeding behavior in mice. *J Neurosci.* 2004;24:2797-805.

[392] Oberlander TF. Fetal serotonin signaling: setting pathways for early childhood development and behavior. *J Adolesc Health.* 2012;51:S9-16.

[393] Kepser LJ, Homberg JR. The neurodevelopmental effects of serotonin: A behavioural perspective. *Behav Brain Res.* 2014.