

Editor-in-Chief

PROFESSOR O. O KEHINDE PHILLIPS

Editor

PROFESSOR N.A. AMUSA
EDITORIAL BOARD

Professor L. O. Adebajo
Department of Microbiology
Olabisi Onabanjo University, Ago-Iwoye

Professor B. M. Ogunsanwo
Department of Chemical Sciences
Olabisi Onabanjo University, Ago-Iwoye

Professor A.W Gbolagade
Department of Mathematical Sciences
Osun State University, Osogbo

Professor S.A Bankole
Department of Microbiology
Olabisi Onabanjo University, Ago-Iwoye

Dr. R. K. Odunaike
Department of Physics
Olabisi Onabanjo University, Ago-Iwoye

Professor V. A. Awoderu
Department of Plant Science and Applied Zoology
Olabisi Onabanjo University, Ago-Iwoye

Professor O. Otolorin
Department of Mathematical Sciences
Olabisi Onabanjo University, Ago-Iwoye

Professor O. O. Kehinde Phillips
Department of Earth Science
Olabisi Onabanjo University, Ago-Iwoye

Professor Oladele Osibanjo
Department of Chemistry
University of Ibadan, Ibadan.

NOTES TO CONTRIBUTORS

INTRODUCTION

The Olabisi Onabanjo University African Journal of Pure and Applied Sciences is directed towards publication of original scientific research and review papers related to Pure Applied Sciences in Africa. These include Physical, Chemical, Biology, Environmental, Horticultural, Agricultural, and Medical Sciences as well as Forestry.

PROCEDURE FOR SUBMISSION:

Manuscripts must be prepared in clean and concise English typed on A4 type with double spacing and organized as indicated below:

1. **Title:**
Must be informative but concise
2. **Author:**
Surname followed by initials with their addresses below the names
3. **Summary:**
To present the objectives and significant findings of the research work
4. **Key words:**
Maximum of four key words indicated below the summary

Other sections of the papers should include **Introduction; Materials and Methods; Results; Discussions Acknowledgements** if need be and **References.**

References: References in the text should be cited as (Berks and Bode, 1969, (OU, *et al.*, 1971). References cited in the text must be listed under bibliography. The list of references should follow patterns below.

Journals: Awoderu V.A and Esuruoso, O. F. (1974) Morphological cultural characteristics and pathogenicity of three isolates of *Pyricularia* in Nigeria. Afr. J. Pl. Protec. 2 (2). 61 – 66.

Books: Hoffmann, G.M.F, Niethaus, F., and Schonbeck, H. (1976). Compendium of wheat diseases. 2nd Edition. APS Press. St. Paul Minnesota. pp 48 – 88 or 300pp.

Proceedings: Atanda, O. A. Awoderu, V. A. and Dina, S. O. (1977) Plant protection in the Sahelian Zone of Nigeria. In Proc. Of the Government Consultation on Crop and Post-Harvest protection needs in the Sahel, FAO Headquarters, Rome 13 – 17 December, 1976 AGP/CPS/76/WP55pp 20 – 25.

Table and Figures: The number of tables and figures should be restricted. It is either the table or the figure and not both in illustrating your data.

Submission of Manuscripts:

3 copies should be forwarded to: The Editor-in-Chief O.O.U African Journal of Pure and Applied Sciences

Attention:
Dean, Faculty of Science, Olabisi Onabanjo University P.M.B. 2002, Ago Iwoye, Ogun State, Nigeria.

A processing fee of N2000 or \$15 in cheque only addressed to the Dean of Science should accompany the Manuscripts.

Abundances of Nitrates and Coliform with Regards to Water Quality In Some Parts of Ibadan, Southwest, Nigeria.

** ODUKOYA, AM, * LANIYAN T.A AND * ARIYO, S.O.

***Department Of Geosciences, University Of Lagos, Lagos, Nigeria.
Sesanbiodun @ Yahoo.com*

* *Department Of Earth Sciences, Olabisi Onabanjo University, P.M.B 2002 Ago Iwoye, Ogun State*

ABSTRACT

Water pollution has been reported throughout the world and Nigeria is not an exception. A very wide range of pollutants have been recognized, including Nitrogen species and bacteria. Nitrate (NO_3) is the main form of Nitrogen which occurs in water and is becoming increasingly widespread because of agriculture activities, the disposal of sewage and wastes. This study thus focused on the assessment of the water samples in Ibadan, Southwest, Nigeria based on these two pollutants (NO_3 and Coliform) and possibility of natural de-nitrification in the study area. The surface and groundwater in Ibadan have been polluted by municipal, industrial wastewater, agricultural activities among others. The nitrate and coliform concentrations at thirty sampling points within Ibadan ranged from 5.89 to 250mg/l and 74-1000cfu/100mg/l respectively. The concentration of nitrate and coliform exceeded the WHO limit of 10mg/l and 0cfu/100mg/l in 93% and 100% of the sampled locations respectively. High nitrate and coliform levels were observed in water from wells from the undeveloped part of the study area. The decrease in nitrate concentration viz a viz an increase in both HCO_3^- and pH along the groundwater flow direction confirmed the possibility of nitrate removal or natural de-nitrification in the study area.

Key words: Water; Nitrate; Pollution; Coliform; De-nitrification.

Introduction

Groundwater pollution in a number of aquifers throughout the world has been reported, and a very wide range of pollutants including N species, heavy metals, chlorinated hydrocarbons, phenols, cyanides, pesticides, major inorganic species and bacteria has been recognized. Nitrate (NO_3) is the main form of N which occurs in groundwater while Coliforms which is a form of bacteria is the

commonest in water. The concentration of coliform bacteria is commonly used as a general bacteriological indicator of water quality standards. The coliforms are a functional group of intestinal bacterial from warm blooded animals. It is used here as an indicator for faecal contamination and a tracer for more highly pathogenic bacteria which occur at lower concentration. Both are becoming increasingly widespread because of agricultural activities, disposal of sewage and domestic wastes. High solubility of nitrate and its anionic form increase its mobility in groundwater. In strongly oxidizing groundwater, nitrate is the stable form of dissolved nitrogen. It moves in groundwater with no transformation and little or no retardation (Freeze and Cherry, 1979).

Ibadan is one of the fast-growing industrialized cities in Africa (Fig1). The population of the city has also increased tremendously in recent times. In conjunction with the fast development, water use has also increased. This region became heavily populated and large industrial centres and intense agricultural developed. Most of the municipal and industrial wastes are disposed directly into the rivers. Neither sewage nor solid waste is controlled. The groundwater of Ibadan, Southwest Nigeria has been polluted by municipal and industrial wastewater disposal into the River and irrigation channels, storage of the domestic wastewater in septic tanks and agricultural activities.

STUDY LOCATION AND GEO-ENVIRONMENTAL SETTING

The study area falls within the Northern part of Ibadan metropolis and located on longitude $3^{\circ}45' - 4^{\circ}03'$ and latitude $6^{\circ}45' - 7^{\circ}30'$ (Fig 1). The study area falls within the humid and subhumid tropical climate of Southwestern Nigeria with two distinct season: the wet season which occurs between March and October with an average annual rainfall of about 1.250mm and dry season from November to February characterized by dry, dusty and relatively cold NE-SW trade winds. Ibadan metropolis is

characterized by high population density. However, like many urban centres in developing countries, Ibadan metropolis is characterized by poor environment planning such as lack of proper sewage and waste disposal systems, lack of adequate water supply and sanitary conditions and direct discharge of domestic sewage water and dumping of domestic refuse into the drainage channels are common practices especially within the congested central portion of the city. Geologically, the study area is characterized by Precambrian Basement Complex composed of quartzites, gneisses and migmatite as the major rock types which are intruded by pegmatites, quartz veins, apiltes and dolerite dykes (Olayinka *et al.*, 1999 Tijani *et al.*, 2004). These intrusions occur as ridges and inselbergs around the adjoining plains and valleys. The overburden is dominated by weathered saprolite units and the thickness depends on the underlying bedrock. Groundwater occurrences are essentially unconfined to semi-confined which depends on the phreatic weathered regolith and presence of joints and fractures in the fresh rock. Morphologically, the unmodified drainage channels form dendritic patterns and run in a southerly direction through much of the city centre (Fig.2). Water flows are irregular during the dry seasons, while population pressure had resulted in built-up areas closer to the stream banks and attendant human activities with increasing run-off constituting constant danger in terms of flooding during the peak of the rainy season

Methodology

Water samples were collected from groundwater and streams within the study area to determine the abundances of nitrates and coliform with regards to their quality (Fig.1). In situ measurements and laboratory analyses were carried out. Groundwater samples were collected from hand dug wells and streams. Temperature (*T*), pH, electrical conductivity (EC) and dissolved oxygen (DO) were measured in the field. Nitrate analyses of the water samples were carried out using an automated hydrazine reduction method performed on a continuous flow analyzer system (ALPHA/AWWA/WEF 1995). Nitrate is reduced to nitrite by hydrazine sulphate in the presence of Copper sulphate. The resulting nitrite plus any nitrite originally present in the sample is diazotized

with Sulphanilamide and forms a colored complex with N- (1-naphthyl) - ethylenedia - mine dichloride. The colored complex is measured colorimetrically at 520nm. This method is applicable to samples having nitrate concentrations between 1 to 50mg/L.

Water samples having nitrate concentrations greater than 50mg/L were diluted to fall within the range of the calibration curve.

30 samples were collected from groundwater. Sampling wells were selected to represent the entire study area. The water samples were collected in sterilized 1.0ml bottles and transported immediately to the laboratory to determine concentration of coliform bacteria. The filtration processes for bacteriological analyses were made within 2 hours from collection. Indicators were determined using classic methods of water filtration (1.0ml of samples and/or its dilutions) on sterile membrane filters (GN-6 Mertical, pore size 0.45 μ m, Pall), with incubation on Endo- Agar LES (BBL) for 24 hours at 35°C for total coliform. All counted colonies were adjusted to 100- ml volume of water.

Results and Discussions

30 groundwater samples were analysed to determine the concentration of Nitrate, coliform and other cations and anions around Ibadan Southwest Nigeria. Other parameters like pH, dissolved oxygen, temperature and total dissolved solids were also determined. Table 1 shows that nitrate ranges between 5.89 to 250mg/l with the mean of 72.61mg/l. coliform ranges between 74 and 1000cfu/100ml with average of 323.3mg/l, TDS ranges between 50mg/l with an average of 399.5mg/l. pH, dissolved oxygen and temperature show mean values of 7.01, 4.53 and 26.43°C respectively. The Dissolved Oxygen concentration was 3.3 to 7.3mg/l. TDS ranges between 300 and 1500mg/l. Cations and Anions follow the following order; Na⁺ > Ca²⁺ > K⁺ > Mg²⁺ and Cl⁻ > NO₃⁻ > HCO₃⁻ > SO₄²⁻ respectively. The summary is shown in Table 2.

Pattern of Nitrate pollution

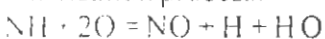
The concentrations of nitrates found in thirty groundwater samples were listed in Table 1. Nitrate and coliform were detected at concentrations exceeding WHO (2004) and EPA (2005) standards (10mg/l and 0Cfu/100mg/l) in 93% and 100% of water samples respectively. Based on the percentile

analysis of the nitrate and coliform concentrations (Table 3) 36.67% and 33.3% of the water samples contained concentrations between 11-45mg/l and 250-500cfu/100mg/l respectively and represent the highest percentage. The lowest percentage which is 6.67% fall within <10mg/l for nitrate and 13.33% which represent 101-150cfu/mg/l for coliform. (Fig2). The average nitrate concentration for the study area was 75mg/l, and 93% of all the samples exceeded the upper limit of 10mg/l (WHO @2004, EPA 2005). Examination of the current pattern of nitrate pollution clearly showed that high nitrate and coliform concentration were found in shallow wells and those sited close to the septic tanks and other municipal wastes as well as agricultural areas (Table 3). Generally, low nitrate concentrations tended to be from those wells from the developed part of the city. Improper construction, sitting, installation, and maintenance of the septic tanks, as well as factors such as depth to groundwater, climate, geology of the site, and septic tank density influence the potential of septic tanks to pollute groundwater. The improper location of wells with respect to septic tanks will increase the potential for the leaching of effluents to the well and groundwater system (Piskin 1973).

Correlation analysis of major constituents

From the correlation analysis in Table 4, three main observations can be summarized as follows;

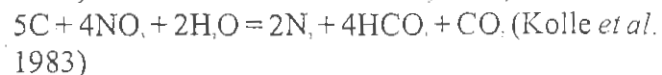
1. The strong positive relationship between NO₃, SO₄, CL and coliform. All of these constituents indicated pollution and their sources were probably from septic tanks and applications of the chemical fertilizer (NH₄)₂SO₄ in the farm around the study area. NH₄ within a nitrogen fertilizer is transformed in the soil into NO₃ and H₂O by the following nitrification process:



2. The negative relationship between NO₃ on one side and other parameters such as TDS, Na⁺, K⁺, Mg²⁺ and Ca²⁺. This confirmed the anthropogenic source of nitrate which is different from other ions whose sources are likely from the bedrock (geology) of the study area.

3. The strong negative correlation (-0.61) between NO₃ and HCO₃⁻ compared to the insignificant relationship with Na⁺, K⁺, Ca²⁺ and Mg²⁺ (-0.18, -0.02, -0.18 and 0.18 respectively). This revealed that the presence of HCO₃⁻ in the groundwater of the study area is closely linked to the removal of NO₃ from the system as a bi-product of

the heterotrophic de-nitrification process and not to natural dissolution from soil or rock (Trudell et al., 1986; Van Beek and Van Puffelen, 1987).



Evidence of De-nitrification in the groundwater system

In order to determine whether the de-nitrification process is already taking place in the groundwater aquifer of the study area some of the water parameters such as HCO₃⁻, pH and SO₄²⁻ were plotted along the groundwater flow directions. To ensure existence of de-nitrification, there must be presence of electron donor and acceptor and organic carbon (OC) as an electron donor. SO₄²⁻ was found to decrease along the direction of groundwater flow (Fig 4) contrary to what should be the case if the electron donor was sulfide (Korom, 1992). Organic carbon (OC) can be supplied to groundwater in two ways. One is where organic carbon is artificially injected into the aquifer while the other is when groundwater interacts with surface water or the root of vegetation. Generally, the majority of all groundwater have dissolved organic carbon (DOC) < 2mg/l (Thurman, 1985).

Change of NO₃ and HCO₃⁻

There was negative correlation between HCO₃⁻ which is the main product of the heterotrophic de-nitrification reaction and NO₃⁻. The HCO₃⁻ concentration increases (produced) while NO₃⁻ concentration decreases (removed) along the flow direction of groundwater (Fig 4). This can be properly explained by the equation above.

Change of pH

The pH increases along the flow of groundwater direction while the concentration of NO₃⁻ decreases along the same direction and this also could be attributed to the removal of nitrate by de-nitrification (Fig 4).

Conclusion

All the parameters analyzed in the groundwater around Ibadan were within both WHO and EPA standards for drinking water except coliform and nitrate. Nitrate and Coliform were detected at concentrations exceeding WHO (2004) and EPA (2005) standards (10mg/l and 0cfu/100mg/l) in 93% and 100% of water samples respectively.

The decreases in nitrate concentrations viz a viz an increase in both HCO₃⁻ and pH along the groundwater flow direction confirmed nitrate removal or natural de-nitrification in the study area

groundwater aquifer. High nitrate concentrations in water supplies are potential hazards to infant health. The consumption of waters with high nitrate concentrations decreases the oxygen carrying capacity of the blood while coliform is responsible for most of the water borne diseases.

References

- APHA-AWWA-WPCF (1981): Standard methods for the examinations of water and wastewater, 15th edition. American Public Health Association, Washington, DC
- Freeze RA, Chery J.A (1979): Groundwater. Prentice Hall, New Jersey.
- Kolle W, Warner P, Strebel O, Bottcher J. (1983): De-nitrification by pyrite in a reducing aquifer. *Vom Wasser*; 61(1): 125-147.
- Olayinka, A., Abimbola A.F., Isibor, R.A., and Rafiu, A.R (1999): A geoelectrical hydrogeochemical investigation of shallow groundwater occurrence in Ibadan, Southwestern Nigeria: *Environment Geology*. Vol.37, pp 31-39.
- Korom. S.F. (1992): Natural de-nitrification in the saturated zone-a review. *Water resources. Res.* 28, 1657-1668.
- Thurman E.M (1985): Organic geochemistry of natural waters. Netherlands, Dordrecht Martinus, Nijhoff. 497p.
- Tijani, M.N., Jinno, K and Horoshiro, Y. (2004): Environment Impact of heavy metals distribution in water and sediments of Ogunpa River, Ibadan area Southwestern Nigeria. *Journal of Mining and Geology*. Vol. 40, (1), pp 73.
- Trudell MR, Gillham Rw, Cherry JA (1986): An in-situ study of the occurrence and rate of denitrification in a shallow unconfined sand aquifer. *J Hydrol* 83 (3/4): 251-268.
- Piskin .R (1973): Evaluation of nitrate content of groundwater in Hall Country, Nebraska. *Groundwater* 11:4-13.
- United States Environment Protection Agency (2005): Quality criteria for water. U.S Govt. Printing Office, Washington D.C.
- Van Beek CGEM, Van Puffelen J(1987): Changes in the chemical composition of drinking water after well infiltration in an unconsolidated sandy aquifer. *Water Resources*
- WHO (2004): Guideline values for chemicals that are of health significance in drinking water quality. World Health Organization, Geneva. pp 491-493.

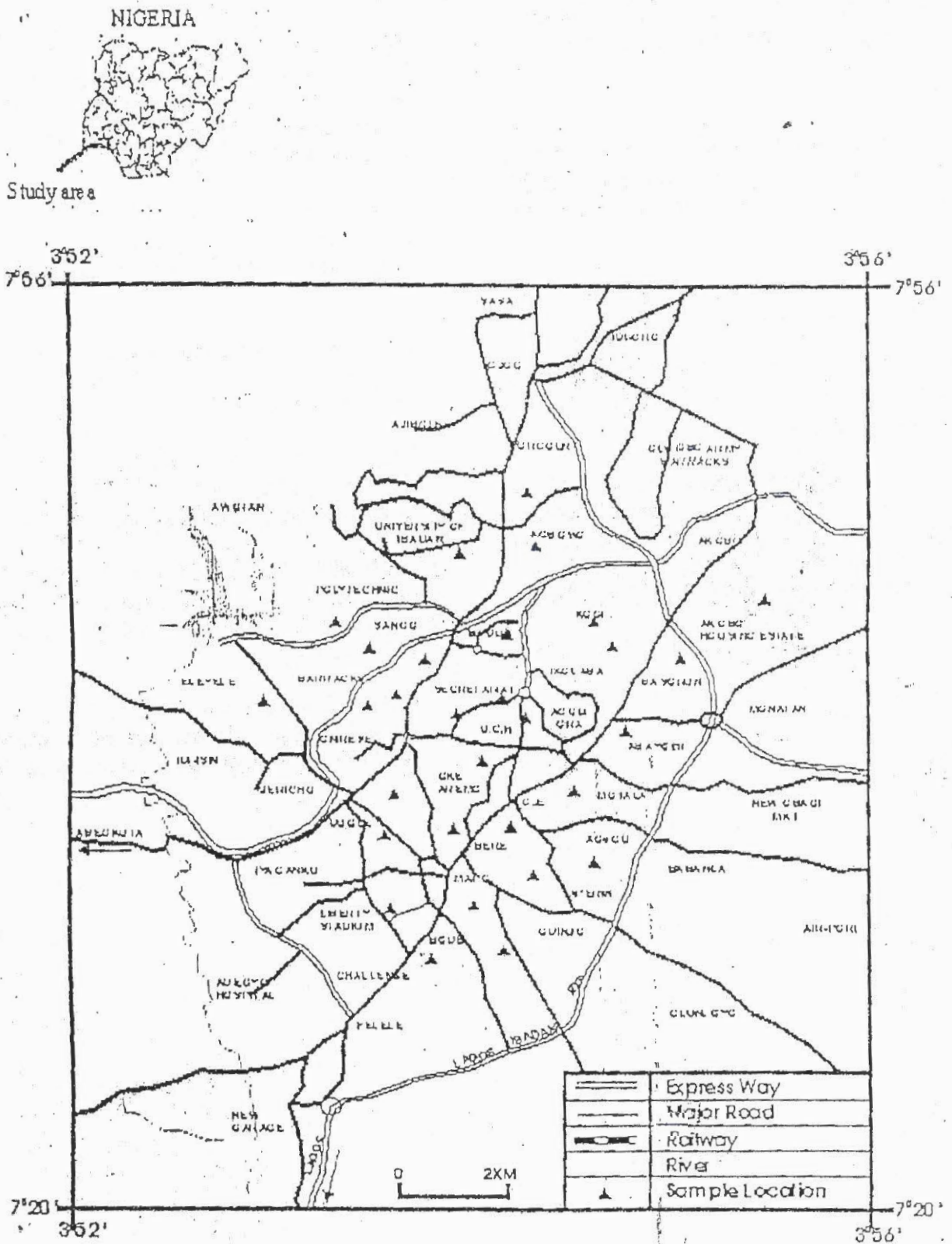


Fig 1: Map of the study area showing sample points

Table 1. Results of Physico-chemical analysis in water samples

mpk	Coliform	Nitrate	TDS	DO	pH	Temp °C	HCO ₃	Cl	SO ₄	Ca	K	Na	Mg
1	80	32	300	3.7	6.2	25	28	127	33	9.6	10.9	18	4.2
2	120	74	800	3.6	6.4	26	77	72	34	12.2	11.8	23.8	3.5
3	100	65	850	3.5	7.4	27	54	181	19	12.5	12.6	28	5.3
4	160	52	500	3.9	7	25	53	85	18	8.4	11.4	38	4.4
5	150	28	900	4.2	6.8	29	54	101	34	12.2	10.8	25	4.6
6	80	20	600	4.8	7.2	28	22	82	24	12.5	9.3	27	2.6
7	280	90	280	3.9	6.9	25	64	70	10	8.4	6.4	38	4.2
8	74	5.89	1000	4.8	6.8	25	75	127	12	12.2	8.9	23.6	4.4
9	280	23.4	200	3.8	6.5	26	74	28	19	5.2	10.1	55	5.4
10	440	28.82	1200	3.3	7.2	25	50	106	25	12.4	11.2	46	4.8
11	120	11.85	120	4.6	7.4	28	51	95.5	10	19.6	10.9	25	3.8
12	280	37.24	280	3.4	7.8	27	62	136	10	12.6	6.3	50	4.2
13	80	7.84	50	3.6	7.6	22	36	54.5	26	10.8	5.68	66	3.6
14	160	43.22	160	4.8	6.9	27	78	127	14	17.6	8.74	65	4.0
15	120	17.51	120	4.4	6.8	26	29	37.5	18	18.5	6.49	55	3.0
16	90	12.41	90	5.2	7.1	25	52	69.5	14	27.9	7.8	36	3.8
17	180	35	180	6.6	6.9	28	30	85	10	20.5	10.9	35	3.6
18	550	120	550	6.8	6.8	30	28	212	22	18.6	11.6	21	3.4
19	155	89	155	7.2	7.3	28	48	132.517		47.4	11.4	11	2.5
20	450	87	450	3.5	7.3	25	62	172.525		48.4	8.5	53.4	4.5
21	300	98	80	3.6	6.9	26	21	54.5	33	36.4	17.5	34	3.4
22	800	150	120	4.6	6.8	27	20.6	120.545		24.4	12.4	42	4.3
23	850	220	100	4.8	7.2	26	37.2	75.5	16	26.8	11	32	2.8
24	500	56	160	5.6	7.4	30	20	82.5	25	14.4	10.5	38	2.4
25	900	213	150	7.3	6.5	25	11.5	77.2	15	9.3	12.5	42	1.8
26	600	125	280	4.5	6.8	26	12	53	34	16.3	17.6	53.4	2.2
27	280	65	70	3.8	7.3	25	28	137	25	26.2	36.3	21	3.2
28	1000	250	440	3.3	7.4	26	43	73.0	18	8.5	16.4	34	6.4
29	200	56	300	4.6	6.9	27	56	23.7	23	8.6	24.4	54	4.6
30	320	65	1500	4.2	6.8	28	98	78.6	32	6.8	14.6	42	3.4

*All parameters in mg/l except coliform that is in cfu/100ml

Table 2: Summary of Physico-chemical results of water samples

Parameters	Min	Max	Mean	ST.Dev.	WHO (2004)	EPA (2005)
pH	6.2	7.8	7.01	0.37	6.5-7.5	6.5-7.5
TDS	50	1000	399.5	373.4	1000	500
DO	3.3	7.3	4.53	1.14		
Temp	25	30	26.43	1.72		
Ca	5.2	48.4	17.57	10.99	200	75
K	5.68	36.3	12.16	5.98		
Na	11	66	37.7	14.18	200	200
Mg	1.8	5.4	3.81	1.02	200	50
HCO ₃	11.5	98	45.8	22.03		
Cl	23.7	212	95.88	44.53	250	250
SO ₄	10	45	22	8.98	400	250
NO ₃	5.89	250	72.61	64.25	10	10
Coliform	74	1000	323.3	269.6	0	0

Table 3: Percentile Analysis of Nitrate and Coliform in water samples

Concentration (mg/l)	Nitrate		Concentration Cfu/100ml	Coliform	
	No of samples	Percentile		No of samples	Percentile
<10	2	6.67%	0-100	06	20%
11-45	11	36.67%	101-150	04	13.33%
46-70	05	16.67%	151-500	14	46.66%
71-100	05	16.67%	501-1000	06	20%
101-200	04	13.33%			
201-250	03	10%			
Total	30	100	Total	30	100

Table 1. Results of Physico-chemical analysis in water samples

mpk	Coliform	Nitrate	TDS	DO	pH	Temp °C	HCO ₃	Cl	SO ₄	Ca	K	Na	Mg
1	80	32	300	3.7	6.2	25	28	127	33	9.6	10.9	18	4.2
2	120	74	800	3.6	6.4	26	77	72	34	12.2	11.8	23.8	3.5
3	100	65	850	3.5	7.4	27	54	181	19	12.5	12.6	28	5.3
4	160	52	500	3.9	7	25	53	85	18	8.4	11.4	38	4.4
5	150	28	900	4.2	6.8	29	54	101	34	12.2	10.8	25	4.6
6	80	20	600	4.8	7.2	28	22	82	24	12.5	9.3	27	2.6
7	280	90	280	3.9	6.9	25	64	70	10	8.4	6.4	38	4.2
8	74	5.89	1000	4.8	6.8	25	75	127	12	12.2	8.9	23.6	4.4
9	280	23.43	200	3.8	6.5	26	74	28	19	5.2	10.1	55	5.4
10	440	28.82	1200	3.3	7.2	25	50	106	25	12.4	11.2	46	4.8
11	120	11.85	120	4.6	7.4	28	51	95.5	10	19.6	10.9	25	3.8
12	280	37.24	280	3.4	7.8	27	62	136	10	12.6	6.3	50	4.2
13	80	7.84	50	3.6	7.6	22	36	54.5	26	10.8	5.68	66	3.6
14	160	43.22	160	4.8	6.9	27	78	127	14	17.6	8.74	65	4.0
15	120	17.51	120	4.4	6.8	26	29	37.5	18	18.5	6.49	55	3.0
16	90	12.41	90	5.2	7.1	25	52	69.5	14	27.9	7.8	36	3.8
17	180	35	180	6.6	6.9	28	30	85	10	20.5	10.9	35	3.6
18	550	120	550	6.8	6.8	30	28	212	22	18.6	11.6	21	3.4
19	155	89	155	7.2	7.3	28	48	132.517		47.4	11.4	11	2.5
20	450	87	450	3.5	7.3	25	62	172.525		48.4	8.5	53.4	4.5
21	300	98	80	3.6	6.9	26	21	54.5	33	36.4	17.5	34	3.4
22	800	150	120	4.6	6.8	27	20.6	120.545		24.4	12.4	42	4.3
23	850	220	100	4.8	7.2	26	37.2	75.5	16	26.8	11	32	2.8
24	500	56	160	5.6	7.4	30	20	82.5	25	14.4	10.5	38	2.4
25	900	213	150	7.3	6.5	25	11.5	77.2	15	9.3	12.5	42	1.8
26	600	125	280	4.5	6.8	26	12	53	34	16.3	17.6	53.4	2.2
27	280	65	70	3.8	7.3	25	28	137	25	26.2	36.3	21	3.2
28	1000	250	440	3.3	7.4	26	43	73.0	18	8.5	16.4	34	6.4
29	200	56	300	4.6	6.9	27	56	23.7	23	8.6	24.4	54	4.6
30	320	65	1500	4.2	6.8	28	98	78.6	32	6.8	14.6	42	3.4

*All parameters in mg/l except coliform that is in cfu/100ml

Table 2: Summary of Physico-chemical results of water samples

Parameters	Min	Max	Mean	ST.Dev.	WHO (2004)	EPA (2005)
pH	6.2	7.8	7.01	0.37	6.5-7.5	6.5-7.5
TDS	50	1000	399.5	373.4	1000	500
DO	3.3	7.3	4.53	1.14		
Temp	25	30	26.43	1.72		
Ca	5.2	48.4	17.57	10.99	200	75
K	5.68	36.3	12.16	5.98		
Na	11	66	37.7	14.18	200	200
Mg	1.8	5.4	3.81	1.02	200	50
HCO ₃	11.5	98	45.8	22.03		
Cl	23.7	212	95.88	44.53	250	250
SO ₄	10	45	22	8.98	400	250
NO ₃	5.89	250	72.61	64.25	10	10
Coliform	74	1000	323.3	269.6	0	0

Table 3: Percentile Analysis of Nitrate and Coliform in water samples

Concentration (mg/l)	Nitrate		Concentration Cfu/100ml	Coliform	
	No of samples	Percentile		No of samples	Percentile
<10	2	6.67%	0-100	06	20%
11-45	11	36.67%	101-150	04	13.33%
46-70	05	16.67%	151-500	14	46.66%
71-100	05	16.67%	501-1000	06	20%
101-200	04	13.33%			
201-250	03	10%			
Total	30	100	Total	30	100

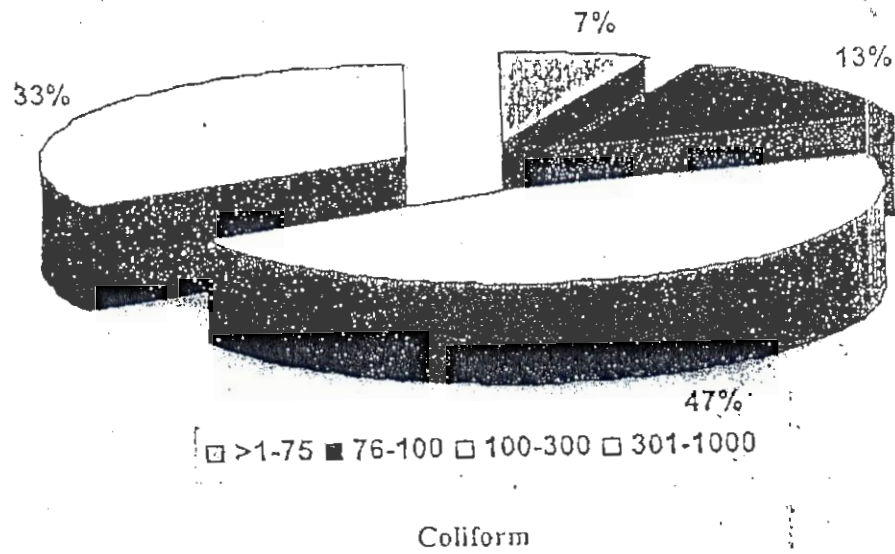
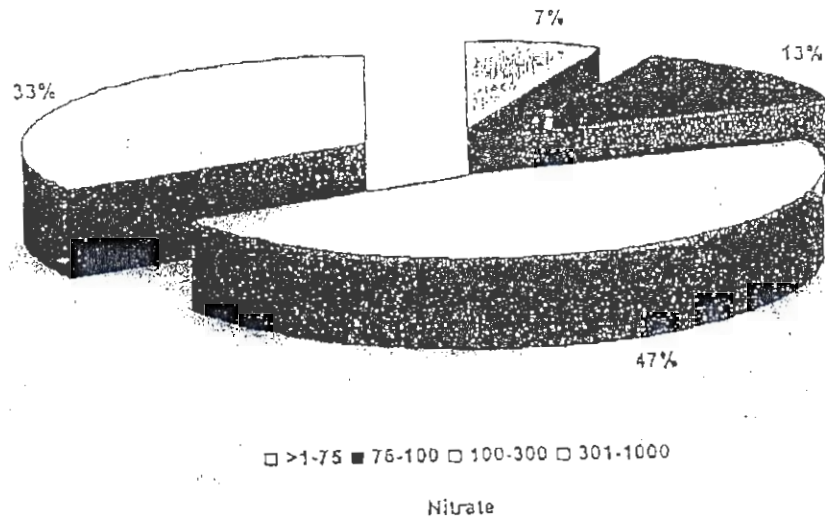


Fig.3: Distribution of Nitrate and Coliform in different percentages. Nitrate in mg/l, coliform in cfu/100ml

