



<b>Title</b>	<b>Epigenetic silencing of a long non-coding RNA KIAA0495 in multiple myeloma</b>
<b>Author(s)</b>	<b>Wong, KY; Li, Z; Zhang, X; Leung, GKK; Chan, GCF; Chim, JCS</b>
<b>Citation</b>	<b>Molecular Cancer, 2015, v. 14, article no. 175</b>
<b>Issued Date</b>	<b>2015</b>
<b>URL</b>	<b><a href="http://hdl.handle.net/10722/223219">http://hdl.handle.net/10722/223219</a></b>
<b>Rights</b>	<b>Molecular Cancer. Copyright © BioMed Central Ltd.; This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License.</b>

SHORT COMMUNICATION

Open Access



# Epigenetic silencing of a long non-coding RNA *KIAA0495* in multiple myeloma

Kwan Yeung Wong<sup>1</sup>, Zhenhai Li<sup>1</sup>, Xiaoqin Zhang<sup>2</sup>, Gilberto Ka Kit Leung<sup>2</sup>, Godfrey Chi-fung Chan<sup>3</sup> and Chor Sang Chim<sup>1\*</sup>

## Abstract

In multiple myeloma, a long non-coding RNA, *KIAA0495* (alias *PDAM/TP73-AS1*), had been found progressively downregulated from normal plasma cell to benign monoclonal gammopathy of undetermined significance to symptomatic myeloma. Herein, by methylation-specific PCR, the putative *KIAA0495* promoter was found unmethylated in all healthy controls ( $N = 14$ ) but methylated in 50 % of myeloma cell lines ( $N = 10$ ). *KIAA0495* methylation was shown inversely correlated with *KIAA0495* expression. However, *KIAA0495* methylation was detected in none of both primary myeloma samples at diagnosis ( $N = 61$ ) and at relapse/progression ( $N = 16$ ). Collectively, despite frequently methylated in cell lines, *KIAA0495* methylation appeared unimportant in the pathogenesis or progression of myeloma.

**Keywords:** Long non-coding RNA, *KIAA0495*, DNA methylation, Myeloma, Tumour suppressor

## Introduction

Long non-coding RNA (lncRNA) is a novel class of functional RNA molecules >200 nucleotides with little or no protein-coding capacity [1, 2]. Based on their genomic locations relative to annotated protein-coding genes, lncRNAs are broadly classified into intergenic, intragenic, antisense, pseudogene, or divergent transcripts [3, 4]. Functionally, lncRNAs play a role in, but not limited to, development, differentiation, and carcinogenesis [2, 5]. Moreover, lncRNAs are found dysregulated and therefore potentially to be oncogenic or tumour suppressive in human cancers [6, 7].

Multiple myeloma is a form of haematological cancer originated from malignant transformation of plasma cells [8]. Clinically, this disease begins with benign monoclonal gammopathy of undetermined significance (MGUS), some of which may undergo asymptomatic smouldering multiple myeloma (SMM), and progresses to symptomatic myeloma [9]. Genetically, despite D-type cyclins are apparently upregulated in all patients, myeloma is a heterogeneous disease characterized by specific gains or

losses of chromosomes, or reciprocal translocations, such as  $t(4;14)(p16.3;q32)$  and  $t(14;16)(q32;q23)$  [10].

DNA methylation refers to the catalytic addition of a methyl ( $-CH_3$ ) group to the cytosine ring of a CpG dinucleotide [11]. Human cancers are characterized by loss of global DNA methylation but gain of methylation at promoter-associated CpG islands (a cluster of CpG dinucleotides) and hence transcriptional silencing of specific tumour suppressor genes or miRNAs [12]. In myeloma, DNA methylation has been shown to mediate silencing of multiple tumour suppressor genes and miRNAs, and has been implicated in the pathogenesis and prognosis of the disease [13, 14].

Interestingly, by gene expression profiling, *KIAA0495*, a lncRNA transcribed from chromosome 1p36, has been shown to be progressively downregulated from normal plasma cell to MGUS to symptomatic myeloma [15]. Moreover, methylation-mediated silencing of *KIAA0495* has been demonstrated in oligodendroglial tumours, leading to enhanced cisplatin resistance via upregulation of anti-apoptotic B-cell CLL/lymphoma 2-like 1 (*BCL2L1*) [16]. Hence, we hypothesized that DNA methylation may account for the progressive downregulation of *KIAA0495* in the pathogenesis of myeloma. Herein, we have studied and reported methylation of *KIAA0495* in myeloma cell

\* Correspondence: jcschim@hku.hk

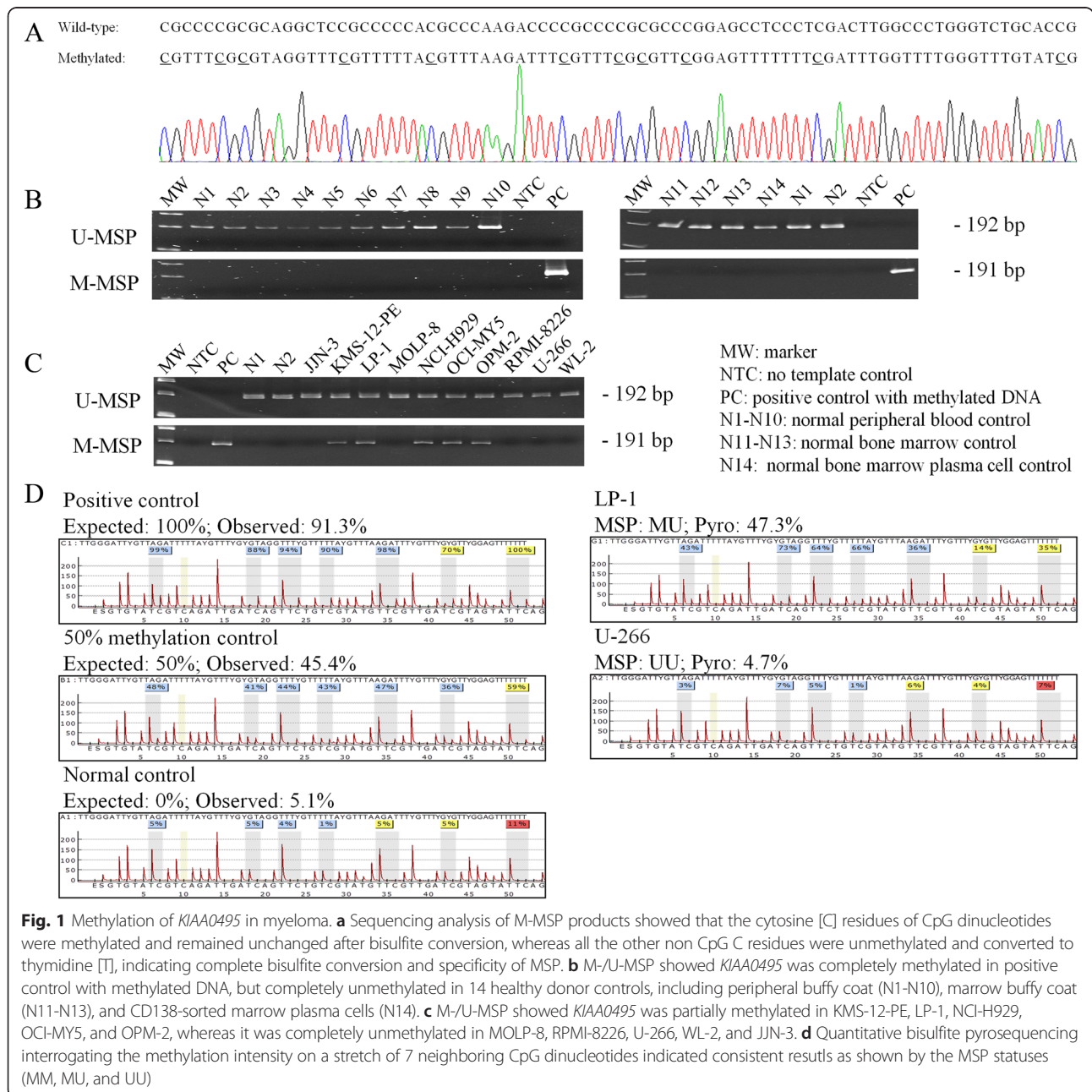
<sup>1</sup>Department of Medicine, Queen Mary Hospital, The University of Hong Kong, Pokfulam Road, Pokfulam, Hong Kong

Full list of author information is available at the end of the article

**Table 1** Primer sequences and PCR reaction conditions

Primer set	Forward primer (5' – 3')	Reverse primer (5' – 3')	Product size (bp)	MgCl <sub>2</sub> /Tm/cycles	Reference
(I) Methylation-specific polymerase chain reaction (MSP)					
<i>KIAA0495</i>					
M-MSP	TGG AGA TAA CGG GTT TAG AGA AAT C	AAC GAA AAC AAA AAT AAA ACC TTC G	191	1.5 mM/58 °C/40X	–
U-MSP	TGG AGA TAA TGG GTT TAG AGA AAT T	AAA CAA AAA CAA AAA TAA AAC CTT CA	192	2.0 mM/58 °C/40X	–
(II) Reverse transcription-polymerase chain reaction (RT-PCR)					
<i>KIAA0495</i>					
	GCT GCT TGC TGT ACG TGG TG	CGT GGC TGA CAC AAA CTT GC	178	1.5 mM/60 °C/35X	[16]
<i>GAPDH</i>	ACC ACA GTC CAT GCC ATC ACT	TCC ACC ACC CTG TTG CTG TA	452	1.5 mM/60 °C/24X	

Keys: Tm, annealing temperature; M-MSP, MSP for the methylated allele; U-MSP, MSP for the unmethylated allele



**Fig. 1** Methylation of *KIAA0495* in myeloma. **a** Sequencing analysis of M-MSP products showed that the cytosine [C] residues of CpG dinucleotides were methylated and remained unchanged after bisulfite conversion, whereas all the other non CpG C residues were unmethylated and converted to thymidine [T], indicating complete bisulfite conversion and specificity of MSP. **b** M-/U-MSP showed *KIAA0495* was completely methylated in positive control with methylated DNA, but completely unmethylated in 14 healthy donor controls, including peripheral buffy coat (N1-N10), marrow buffy coat (N11-N13), and CD138-sorted marrow plasma cells (N14). **c** M-/U-MSP showed *KIAA0495* was partially methylated in KMS-12-PE, LP-1, NCI-H929, OCI-MY5, and OPM-2, whereas it was completely unmethylated in MOLP-8, RPMI-8226, U-266, WL-2, and JLN-3. **d** Quantitative bisulfite pyrosequencing interrogating the methylation intensity on a stretch of 7 neighboring CpG dinucleotides indicated consistent results as shown by the MSP statuses (MM, MU, and UU)

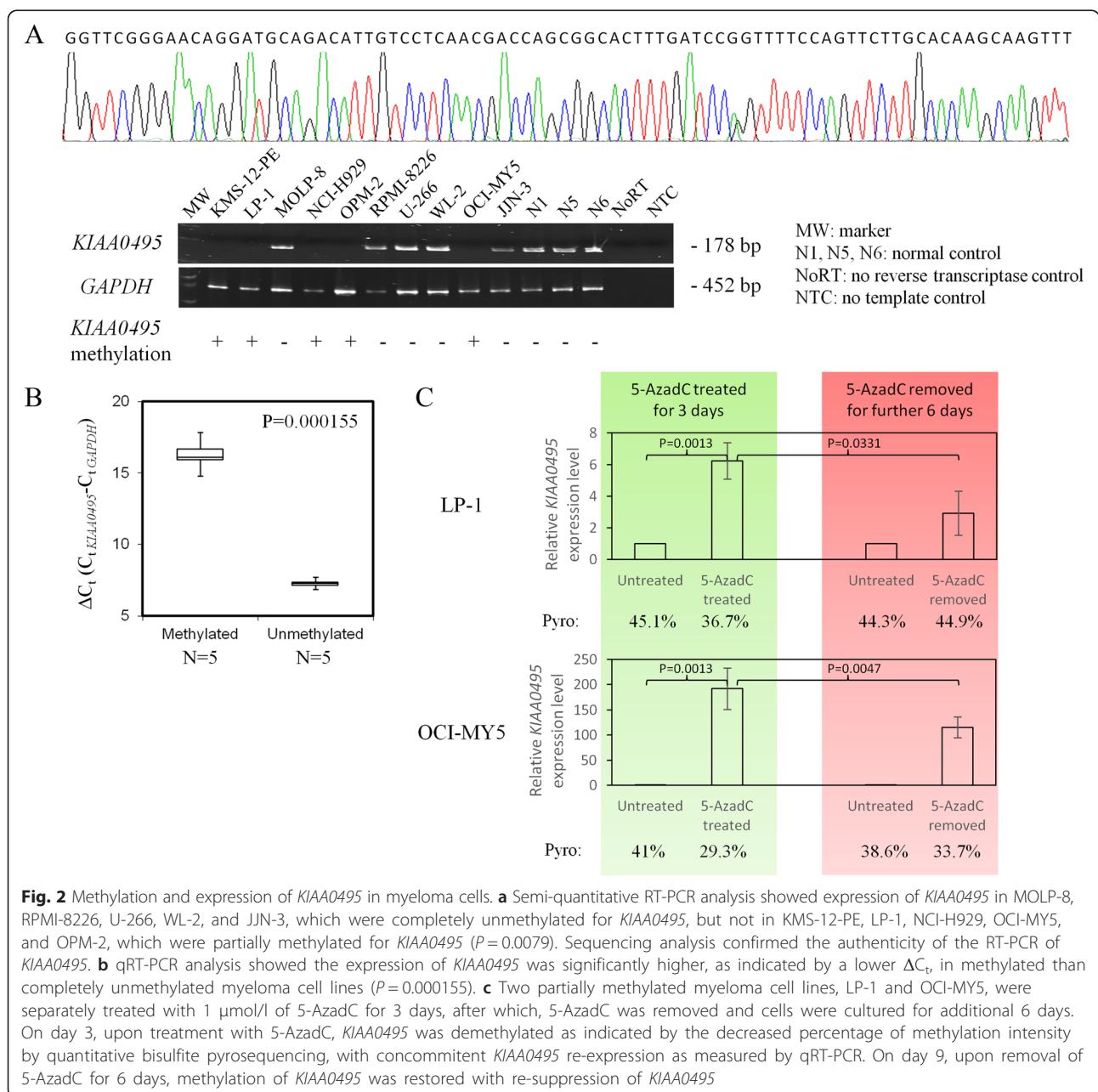
lines, primary myeloma marrow samples at diagnosis, and at relapse/progression. Materials and methods have been incorporated as Additional file 1.

**Findings**

**Methylation-specific PCR: *KIAA0495* methylation in healthy controls and myeloma cell lines**

While the putative *KIAA0495* promoter region was found embedded in a CpG island (Additional file 2: Figure S1), MSP primers were designed to study methylation of this CpG island in a panel of healthy

controls [peripheral (*N* = 10) and marrow (*N* = 3) buffy coat, CD138-sorted healthy plasma cell (*N* = 1)], and myeloma cell lines (*N* = 10) (Table 1). Direct sequencing analysis of M-MSP products from bisulfite-treated positive control showed expected conversion of unmethylated cytosine into uracil (turned to thymidine after PCR), with methylated cytosine remained unchanged, indicating complete bisulfite conversion and MSP specificity (Fig. 1a). None of 14 healthy controls showed *KIAA0495* methylation (Fig. 1b). On the other hand, in myeloma cell lines, KMS-12-PE, LP-1, NCI-H929,



**Fig. 2** Methylation and expression of *KIAA0495* in myeloma cells. **a** Semi-quantitative RT-PCR analysis showed expression of *KIAA0495* in MOLP-8, RPMI-8226, U-266, WL-2, and JIN-3, which were completely unmethylated for *KIAA0495*, but not in KMS-12-PE, LP-1, NCI-H929, OCI-MY5, and OPM-2, which were partially methylated for *KIAA0495* (*P* = 0.0079). Sequencing analysis confirmed the authenticity of the RT-PCR of *KIAA0495*. **b** qRT-PCR analysis showed the expression of *KIAA0495* was significantly higher, as indicated by a lower  $\Delta C_t$ , in methylated than completely unmethylated myeloma cell lines (*P* = 0.000155). **c** Two partially methylated myeloma cell lines, LP-1 and OCI-MY5, were separately treated with 1  $\mu$ mol/l of 5-AzadC for 3 days, after which, 5-AzadC was removed and cells were cultured for additional 6 days. On day 3, upon treatment with 5-AzadC, *KIAA0495* was demethylated as indicated by the decreased percentage of methylation intensity by quantitative bisulfite pyrosequencing, with concomitant *KIAA0495* re-expression as measured by qRT-PCR. On day 9, upon removal of 5-AzadC for 6 days, methylation of *KIAA0495* was restored with re-suppression of *KIAA0495*

OPM-2, and OCI-MY5 were partially methylated, whereas MOLP-8, RPMI-8226, U-266, WL-2, and JIN-3 were completely unmethylated for *KIAA0495* (Fig. 1c). Moreover, the MSP methylation statuses of controls and cell lines were confirmed by quantitative bisulfite pyrosequencing (Fig. 1d). These data suggested that methylation of *KIAA0495* was tumour-specific, consistent with other tumour suppressor protein-coding genes and non-coding miRNAs in myeloma [17–20].

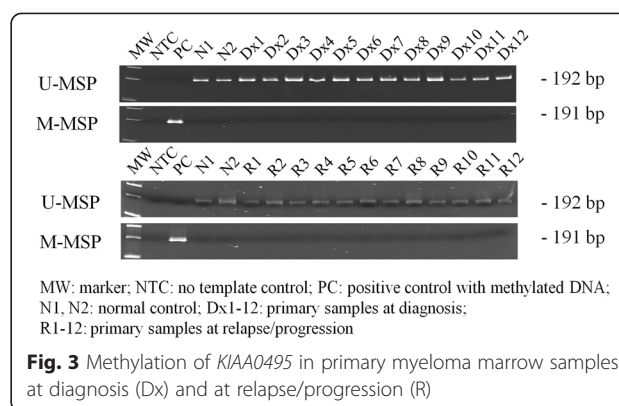
#### *KIAA0495* methylation and expression in myeloma cell lines

To determine whether *KIAA0495* methylation was associated with silencing of *KIAA0495* expression, the expression level of *KIAA0495* was measured by semi-quantitative RT-PCR and quantitative real-time PCR (qPCR), and correlated with the *KIAA0495* methylation status as detected by MSP in myeloma cell lines. Of the 10 myeloma cell lines, *KIAA0495* methylation was associated with lower expression as detected by both semi-quantitative RT-PCR ( $P = 0.0079$ ; Fig. 2a) and quantitative real-time PCR ( $P = 0.000155$ ; Fig. 2b).

Moreover, myeloma cell lines with *KIAA0495* methylation were treated with a hypomethylating agent, 5-AzadC, followed by pyrosequencing and qPCR analyses. Upon 5-AzadC treatment, LP-1 and OCI-MY5 cells, which were partially methylated for *KIAA0495*, showed progressive demethylation of the *KIAA0495* promoter, as evidenced by the decreased methylation percentage on a stretch of seven consecutive CpG dinucleotides, together with concomitant re-expression of the *KIAA0495* transcript, as illustrated by the increased expression relative to the untreated control (LP-1:  $P = 0.0013$ ; OCI-MY5:  $P = 0.0013$ ; Fig. 2c). Nevertheless, when 5-AzadC-treated cells were continually cultured with fresh medium in the absence of 5-AzadC for a further 6 days, methylation of the *KIAA0495* promoter was restored, with simultaneous suppression of the *KIAA0495* expression (LP-1:  $P = 0.0331$ ; OCI-MY5:  $P = 0.0447$ ; Fig. 2c). Taken together, the methylation of *KIAA0495* was inversely correlated with *KIAA0495* expression level in myeloma cell lines, similar to the methylation-mediated silencing of *KIAA0495* demonstrated in glioma cell lines and primary oligodendroglial tumour cells [16], suggesting that methylation of the promoter-associated CpG island emerged to be one of the mechanisms resulting in the regulation of lncRNAs in cancer cells.

#### Methylation-specific PCR: *KIAA0495* methylation in myeloma primary samples at diagnosis and at relapse

To examine if methylation of *KIAA0495* was also detected in primary samples, methylation of *KIAA0495* was studied in 61 primary samples at diagnosis and 16 primary samples at relapse by MSP. However, none of



these samples showed methylation of *KIAA0495* (Fig. 3), indicating methylation of *KIAA0495* was rarely detected in primary myeloma samples. Hence, similar to the studies of *miR-9-1*, *miR-9-3*, and *miR-124-1* [21, 22], these data suggested that methylation of certain tumour suppressor non-coding miRNAs or lncRNAs was acquired *in vitro* during continuous culture of myeloma cells, hence not pathogenic. Therefore, methylation was not the mechanism leading to the progressive downregulation of *KIAA0495* from normal plasma cell to MGUS to symptomatic myeloma [15], suggestive of other mechanisms, such as histone modification and miRNA-mediated repression, may come into play. For instance, in gastric cancer cells, histone deacetylation was found to be associated in the downregulation of a tumour suppressor lncRNA, *FENRR* [23]. Moreover, in breast cancer cells, miR-21 was shown to target and hence downregulate the expression of a tumour suppressor lncRNA, *GASS* [24]. Furthermore, while *KIAA0495* resides on 1p36, which has been shown frequently deleted in newly diagnosed patients with myeloma [25], the loss of expression of *KIAA0495* may be associated with chromosome deletion. Last but not least, inactivation of *KIAA0495* may also be accounted by haploinsufficiency, which has been demonstrated in the *TP53* in myeloma [26].

#### Conclusion

Methylation of *KIAA0495* is tumour-specific, leading to reversible silencing of *KIAA0495* expression in myeloma cell lines. On the other hand, it is rarely detected in primary samples of myeloma at diagnosis or at relapse, and hence not responsible for the progressive downregulation of *KIAA0495* from normal plasma cell to MGUS to symptomatic myeloma, as shown by the gene expression profiling, and remains as a possible *in vitro* event acquired during continuous culture of myeloma cells.



## Additional files

**Additional file 1: Materials and methods.** (DOCX 20 kb)

**Additional file 2: Figure S1.** Schematic diagram showing the transcription start sites (TSS) of *KIAA0495*, distribution of CpG dinucleotides (red lines), and amplicons of methylation-specific PCR (green box) and pyrosequencing (purple box). (TIFF 339 kb)

### Abbreviations

lncRNA: Long non-coding RNA; MGUS: Monoclonal gammopathy of undetermined significance; miRNA: microRNA; MSP: Methylation-specific PCR; M-MSP: Methylated MSP; U-MSP: Unmethylated MSP; PCR: Polymerase chain reaction; RT-PCR: Reverse transcription-PCR; SMM: Smouldering multiple myeloma.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

KYW and ZZ carried out the experiments. KYW, ZZ and CSC drafted the manuscript. KYW and CSC conceived of the study. All authors participated in the design of the study. All authors read and approved the final manuscript.

### Author details

<sup>1</sup>Department of Medicine, Queen Mary Hospital, The University of Hong Kong, Pokfulam Road, Pokfulam, Hong Kong. <sup>2</sup>Department of Surgery, Queen Mary Hospital, The University of Hong Kong, Pokfulam Road, Pokfulam, Hong Kong. <sup>3</sup>Department of Paediatrics and Adolescent Medicine, Queen Mary Hospital, The University of Hong Kong, Pokfulam Road, Pokfulam, Hong Kong.

Received: 8 June 2015 Accepted: 10 September 2015

Published online: 26 September 2015

## References

- Mercer TR, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nat Rev Genet.* 2009;10:155–9.
- Spizzo R, Almeida MI, Colombatti A, Calin GA. Long non-coding RNAs and cancer: a new frontier of translational research. *Oncogene.* 2012;31:4577–87.
- Derrien T, Johnson R, Bussotti G, Tanzer A, Djebali S, Tilgner H, et al. The GENCODE v7 catalog of human long noncoding RNAs: analysis of their gene structure, evolution, and expression. *Genome Res.* 2012;22:1775–89.
- Kung JTY, Colognori D, Lee JT. Long Noncoding RNAs: Past, Present, and Future. *Genetics.* 2013;193:651–69.
- Fatica A, Bozzoni I. Long non-coding RNAs: new players in cell differentiation and development. *Nat Rev Genet.* 2014;15:7–21.
- Gibb E, Brown C, Lam W. The functional role of long non-coding RNA in human carcinomas. *Mol Cancer.* 2011;10:38.
- Gutschner T, Diederichs S. The Hallmarks of Cancer: A long non-coding RNA point of view. *RNA Biol.* 2012;9:703–19.
- Morgan GJ, Walker BA, Davies FE. The genetic architecture of multiple myeloma. *Nat Rev Cancer.* 2012;12:335–48.
- Kyle RA, Durie BGM, Rajkumar SV, Landgren O, Blade J, Merlini G, et al. Monoclonal gammopathy of undetermined significance (MGUS) and smoldering (asymptomatic) multiple myeloma: IMWG consensus perspectives risk factors for progression and guidelines for monitoring and management. *Leukemia.* 2010;24:1121–7.
- Fonseca R, Bergsagel PL, Drach J, Shaughnessy J, Gutierrez N, Stewart AK, et al. International Myeloma Working Group molecular classification of multiple myeloma: spotlight review. *Leukemia.* 2009;23:2210–21.
- Jones PA. Functions of DNA methylation: islands, start sites, gene bodies and beyond. *Nat Rev Genet.* 2012;13:484–92.
- Esteller M. Epigenetics in Cancer. *N Engl J Med.* 2008;358:1148–59.
- Chim CS, Kwong YL, Liang R. Gene hypermethylation in multiple myeloma: lessons from a cancer pathway approach. *Clin Lymphoma Myeloma.* 2008;8:331–9.
- Wong KY, Huang X, Chim CS. DNA methylation of microRNA genes in multiple myeloma. *Carcinogenesis.* 2012;33:1629–38.
- Zhan F, Barlogie B, Arzoumanian V, Huang Y, Williams DR, Hollmig K, et al. Gene-expression signature of benign monoclonal gammopathy evident in multiple myeloma is linked to good prognosis. *Blood.* 2007;109:1692–700.
- Pang JCS, Li KKW, Lau KM, Ng YL, Wong J, Chung NYF, et al. *KIAA0495/PDAM* Is Frequently Downregulated in Oligodendroglial Tumors and Its Knockdown by siRNA Induces Cisplatin Resistance in Glioma Cells. *Brain Pathol.* 2010;20:1021–32.
- Chim CS, Fung TK, Cheung WC, Liang R, Kwong YL. *SOCS1* and *SHP1* hypermethylation in multiple myeloma: implications for epigenetic activation of the *Jak/STAT* pathway. *Blood.* 2004;103:4630–5.
- Chim CS, Pang R, Fung TK, Choi CL, Liang R. Epigenetic dysregulation of *Wnt* signaling pathway in multiple myeloma. *Leukemia.* 2007;21:2527–36.
- Wong KY, Liang R, So CC, Jin DY, Costello JF, Chim CS. Epigenetic silencing of *MIR203* in multiple myeloma. *Br J Haematol.* 2011;154:569–78.
- Wong KY, Yim RLH, So CC, Jin D-Y, Liang R, Chim CS. Epigenetic inactivation of the *MIR34B/C* in multiple myeloma. *Blood.* 2011;118:5901–4.
- Wong KY, So CC, Loong F, Chung LP, Lam WW, Liang R, et al. Epigenetic inactivation of the *miR-124-1* in haematological malignancies. *PLoS one.* 2011;6, e19027.
- Zhang Q, Wang LQ, Wong KY, Li ZY, Chim CS. Infrequent DNA methylation of *miR-9-1* and *miR-9-3* in multiple myeloma. *J Clin Pathol.* 2015;68:557–61.
- Xu TP, Huang MD, Xia R, Liu XX, Sun M, Yin L, et al. Decreased expression of the long non-coding RNA *FENDRR* is associated with poor prognosis in gastric cancer and *FENDRR* regulates gastric cancer cell metastasis by affecting fibronectin1 expression. *J Hematol Oncol.* 2014;7:63.
- Zhang Z, Zhu Z, Watabe K, Zhang X, Bai C, Xu M, et al. Negative regulation of lncRNA *GAS5* by *miR-21*. *Cell Death Differ.* 2013;20:1558–68.
- Walker BA, Leone PE, Chiecchio L, Dickens NJ, Jenner MW, Boyd KD, et al. A compendium of myeloma-associated chromosomal copy number abnormalities and their prognostic value. *Blood.* 2010;116:e56–65.
- Teoh PJ, Chung TH, Sebastian S, Choo SN, Yan J, Ng SB, et al. *p53* haploinsufficiency and functional abnormalities in multiple myeloma. *Leukemia.* 2014;28:2066–74.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

