

FACULDADE DE FARMÁCIA Liversidade de Lisba



SNS SERVIÇO NACIONAL DE SAÚDE

REPÚBLICA PORTUGUESA

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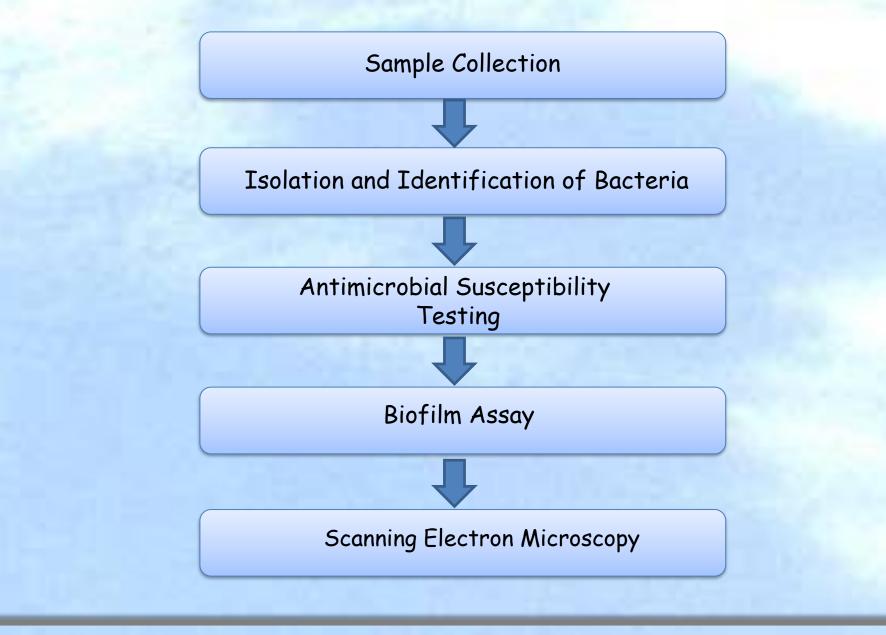
Introduction Artificial lakes, ornamental fountains and other water courses are present in most city parks and green areas, enabling people to practice sports or simply interact with nature and relax. Water is essential for life but, the presence of infectious agents, toxic chemicals, and radiological hazards could represent a threath for human health. Several studies have

Materials and Methods

Water and biofilm samples where collected, in 2 lakes (L1-L2) and ornamental fountains (L3-L4) in February/2015. In May/2015 and monthly during a year (starting March/2016) samples from L4 where collected. The samples were collected within a park with a total area of 26 hectares, located in the northern area of Lisboa, Portugal. Biofilm assembly was monitored by crystal violet assay and SEM and antibiotic susceptibility was performed by conventional methods.

provided evidence for the presence of potential human pathogens in waters containing ornamental fish. The water in these city parks might be dangerous, so this study aims to characterize the microbiota from different ornamental lakes and fountains located in Lisboa, Portugal. A special attention was given to the ability of environmental bacteria to persist within biofilms and the antibiotic susceptibility profiles of clinically relevant members of the ornamental waters microbiota.

Diagram bellow explain the process:



Results:

i doic		microbi						., .				
Actinobacter lwoffii								~				
Aeromonas hydrophila/caviae								×				
Aeromonas sobria	×	*	*	×	*	×	×	~	*	*	*	
Aeromonas spp	*	*	-	•	*	~	-	~	~	~	~	
Aeromonas veronii	×				*						*	
Alcaligenes faecalis spp faecalis	*				*		×				*	
Bacillus spp							•			~		
Bacillus subtilis			×							*		
Bacilo Gneg			•				*					
Comamonas testosteni/ P oleovorans							×		**			
E coli				*	×	×	*	×	×	×		
Enterobacter cloacae complex				*	*	~	•	•	~	*	*	
Enterobacter cloacde complex Escherichia fergusonii				*								
									••		*	
K. oxytoca							•	•	*			
K. pneumoniae				•	*	•	*	×				
K. pneumoniae spp pneumoniae				*	*	*						
Klebsiella oxytoca						×						
Lectercia adecarboxylata									*	-		
Pantoea spp										*		
Pseudomonas fluorescens										×		
Pseudomonas oryzihabitans												
Pseudomonas putida								×		*		
Pseudomonas spp					*							
Raoutela planticola	×											
Raoutella ornithinolytica				×								
Raoutella planticola (K. planticola)									×			
Serraria marcescens					*							
Serratia fonticola					*	×						
Serratia marcescens									×		×	
Serratia plymuthica											*	
Shewanella putrefaciens				×								
Shingomonas paucimobilis		×						×				

Table 2. Antibiotic susceptibility of bacterial isolates identified from ornamental waters inFebruary/2015.

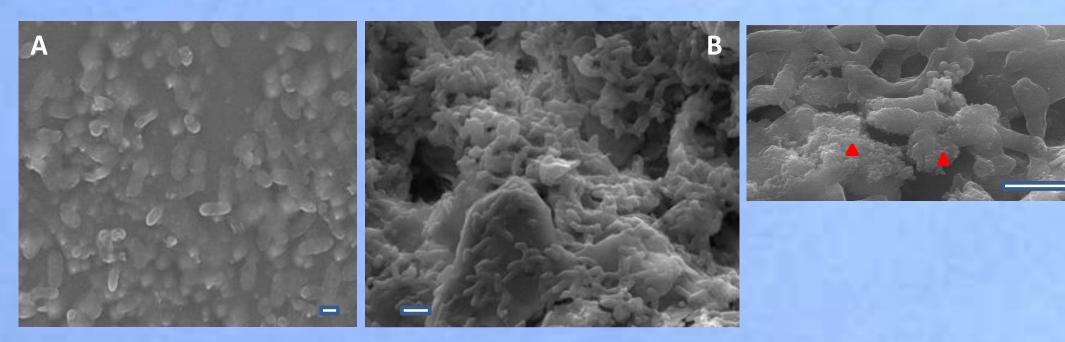
ID	Peaterie		Antibiotic Susceptibility *								
	Bacteria	AMC	FOX	CAZ	CTX	IPM	GM	CIP			
	Klebsiella oxytoca	R	S	S	S	S	S	S			
	Klebsiella pneumoniae	R	S	S	S	S	S	S			
L1	Serratia marcescens	R	S	S	S	S	S	S			
	Serratia odorifera	R	S	S	S	S	S	S			
	Serratia rubidea	S	S	S	S	S	S	S			
	Vibrio metschnikovii	R	R	S	S	S	5	S			

	Elisabethkingia meningoseptica	R	R	5	R	R	5	5
L2	Enterobacter spp.	R	5	5	5	5	S	5
	Stenotrophomonas maltophilia	R	R	R	R	R	5	5
L3	Serratia rubidea	R	S	S	S	S	S	S
	Klebsiella pneumoniae ozonae 1	S	S	S	S	S	S	S
L4	Klebsiella pneumoniae ozonae 2	R	S	S	S	S	S	S
	Pastorella, Shigella	5	S	S	S	S	S	S

* AMC: Amoxicillin/clavulanic acid, FOX: Cefoxitin (FOX), CAZ: Ceftazidime, CTX: Cefotaxime; IMP: Imipenem; GM: Gentamicin; CIP: Ciprofloxacin. R: Resistant, S: Susceptible; I: Intermediate.

Table 3. Bacteria ability to assemble biofilms in vitro at different temperatures.

	OD 570 nm (AU)				
iofilm Recovered Bacteria ID	25 °C	37 ° <i>C</i>			
K. pneumoniae	1.159 <u>+</u> 0.09	0.285 <u>+</u> 0.01			
A. sobria	0.284 <u>+</u> 0.06	0.155 <u>+</u> 0.03			
A. veroni	0.761 <u>+</u> 0.11	0.185 <u>+</u> 0.004			
C. violaceum	0.017 <u>+</u> 0.01	0.096 <u>+</u> 0.05			



25.0

29.0

20.0

19.6

17.0

17.0

17.0

14.0

27.0

Figure 1. Biofilms assembled on cement at 25 °C (A) by A. sobria isolated from biofilms present in L4 (May/2015); (B) and K. pneumoniae isolated from biofilms present in L1 (February/2015); (C) A detail of K. pneumoniae biofilm highlighting the extracellular matrix (red triangles). Scale bar 1 μ m.

Conclusions

- In this study potential human pathogens were identified in samples from a typical urban park.
- These pathogens were found in both planktonic and biofilm forms.
- Waterborne pathogens such as Aeromonas species are a significant cause of acute bacterial gastroenteritis in young children.



Lake (L1)

• The obtained results support a periodic control of ornamental water microbiota as simple preventive

measure to avoid potential health issues.

T(°C)

17.2

19.9



