



Mare Incognitum: A Glimpse into Future Plankton Diversity and Ecology Research

Guillem Chust^{1*}, Meike Vogt², Fabio Benedetti³, Teofil Nakov⁴, Sébastien Villéger⁵, Anaïs Aubert^{3,6}, Sergio M. Vallina⁷, Damiano Righetti², Fabrice Not⁸, Tristan Biard^{3,8}, Lucie Bittner⁹, Anne-Sophie Benoiston⁹, Lionel Guidi³, Ernesto Villarino¹, Charlie Gaborit⁷, Astrid Cornils¹⁰, Lucie Buttay¹¹, Jean-Olivier Irisson³, Marlène Chiarello⁵, Alessandra L. Vallim^{12,13}, Leocadio Blanco-Bercial¹⁴, Laura Basconi¹⁵ and Sakina-Dorothee Ayata³

OPEN ACCESS

Edited by:

Cosimo Solidoro,
National Institute of Oceanography
and Experimental Geophysics, Italy

Reviewed by:

Jan Marcin Weslawski,
Institute of Oceanology (PAN), Poland
Jose M. Riascos,
Universidad del Valle, Colombia
Maurizio Ribera D'Alcala',
Stazione Zoologica Anton Dohrn, Italy

*Correspondence:

Guillem Chust
gchust@azti.es

Specialty section:

This article was submitted to
Marine Ecosystem Ecology,
a section of the journal
Frontiers in Marine Science

Received: 30 September 2016

Accepted: 24 February 2017

Published: 10 March 2017

Citation:

Chust G, Vogt M, Benedetti F,
Nakov T, Villéger S, Aubert A,
Vallina SM, Righetti D, Not F, Biard T,
Bittner L, Benoiston A-S, Guidi L,
Villarino E, Gaborit C, Cornils A,
Buttay L, Irisson J-O, Chiarello M,
Vallim AL, Blanco-Bercial L, Basconi L
and Ayata S-D (2017) Mare
Incognitum: A Glimpse into Future
Plankton Diversity and Ecology
Research. *Front. Mar. Sci.* 4:68.
doi: 10.3389/fmars.2017.00068

¹ Marine Research Division, AZTI, Sukarrieta, Spain, ² Environmental Physics Group, Institute for Biogeochemistry and Pollutant Dynamics, ETH Zurich, Zurich, Switzerland, ³ Laboratoire d'Océanographie de Villefranche, Centre National de la Recherche Scientifique, Sorbonne Universités, UPMC Université Paris 06, Villefranche-sur-Mer, France, ⁴ Department of Biological Sciences, University of Arkansas, Fayetteville, AR, USA, ⁵ Laboratoire Biodiversité Marine et ses Usages (MARBEC), UMR 9190 Centre National de la Recherche Scientifique-IRD-UM-IFREMER, Université de Montpellier, Montpellier, France, ⁶ Service des Stations Marines du Muséum National d'Histoire Naturelle, CRESCO, Dinard, France, ⁷ Institute of Marine Sciences (CSIC), Barcelona, Spain, ⁸ Laboratoire Adaptation et Diversité en Milieu Marin UMR7144, Station Biologique de Roscoff, Centre National de la Recherche Scientifique, Sorbonne Universités, UPMC Université Paris 06, Roscoff, France, ⁹ Centre National de la Recherche Scientifique, Institut de Biologie Paris-Seine, Evolution Paris Seine, Sorbonne Universités, UPMC Université Paris 06, Paris, France, ¹⁰ Polar Biological Oceanography, Alfred Wegener Institute for Polar and Marine Research, Bremerhaven, Germany, ¹¹ Centro Oceanográfico de Gijón, Instituto Español de Oceanografía, Gijón, Spain, ¹² Instituto de Biociências, Universidade Estadual Júlio de Mesquita Filho, São Vicente, Brazil, ¹³ Laboratório de Evolução e Diversidade Aquática, Universidade Estadual Júlio de Mesquita Filho, Assis, Brazil, ¹⁴ Bermuda Institute of Ocean Sciences, St. George's, Bermuda, ¹⁵ Università del Salento, CONISMA, Lecce, Italy

With global climate change altering marine ecosystems, research on plankton ecology is likely to navigate uncharted seas. Yet, a staggering wealth of new plankton observations, integrated with recent advances in marine ecosystem modeling, may shed light on marine ecosystem structure and functioning. A EuroMarine foresight workshop on the “Impact of climate change on the distribution of plankton functional and phylogenetic diversity” (PlankDiv) identified five grand challenges for future plankton diversity and macroecology research: (1) What can we learn about plankton communities from the new wealth of high-throughput “omics” data? (2) What is the link between plankton diversity and ecosystem function? (3) How can species distribution models be adapted to represent plankton biogeography? (4) How will plankton biogeography be altered due to anthropogenic climate change? and (5) Can a new unifying theory of macroecology be developed based on plankton ecology studies? In this review, we discuss potential future avenues to address these questions, and challenges that need to be tackled along the way.

Keywords: plankton, macroecology, species distribution, functional diversity, climate change, habitat modeling

INTRODUCTION

Marine ecosystems are altered by anthropogenic climate change and ocean acidification at an unprecedented rate (Waters et al., 2016). In recent years, observational studies have documented shifts in plankton biogeography and community structure in several ocean basins associated to sea warming, with changes that rank among the fastest and largest documented (Beaugrand et al., 2002; Poloczanska et al., 2013; Rivero-Calle et al., 2015). How changes in plankton distribution, phenology, and biomass may impact fisheries and other ecosystem services is poorly quantified (Cheung et al., 2013), with large uncertainties in the magnitude of potential cascading effects caused by trophic mismatch (Edwards and Richardson, 2004), trophic amplification (Chust et al., 2014a), and on global biogeochemical cycles (Doney et al., 2012). In consequence, current management policies suffer from a lack of understanding of marine systems (Borja et al., 2010), and biases arise in the perception of potential ocean calamities in the absence of robust evidence (Duarte et al., 2015).

While recent oceanographic efforts such as Tara Oceans (Pesant et al., 2015) and Malaspina (Duarte, 2015) expeditions have generated a staggering wealth of novel observational data on plankton distribution and diversity (Figure 1), these same data have revealed the extent of our ignorance of marine ecosystem structure and function. A large fraction of plankton diversity recorded in recent surveys cannot be assigned to known taxonomic groups (de Vargas et al., 2015), highlighting how profoundly our knowledge of the planktonic world is biased toward the taxa sampled or cultured. Not only the identity of major players, but also the drivers of community structure and interactions between organisms remain a “*mare incognitum*.” In the surface ocean, plankton composed of prokaryotes (viruses, bacteria, and archaea) and eukaryotes (protists and metazoans; Figure 1) have been shown to form complex interaction networks driven by multiple biotic and abiotic factors (Lima-Mendez et al., 2015), and despite their key role as resource for higher trophic levels, mesopelagic plankton communities are some of the least studied on Earth (St. John et al., 2016).

Despite these gaps in our understanding, the existing data reveal the importance of community composition for marine ecosystem function. For instance, an investigation of planktonic communities at the global scale using high-throughput metagenomic sampling techniques has recently linked carbon export patterns to specific plankton interaction networks (Guidi et al., 2016), suggesting that the who's who in the plankton world is of paramount importance for the carbon cycle. Integrated with revised estimates in species abundance and biomass (Buitenhuis et al., 2013), and combined with advances in statistical (Robinson et al., 2011) and mechanistic modeling techniques (Follows et al., 2007), novel high-throughput metagenomic data may allow us to relate biogeographic patterns of plankton distribution and diversity to further ecosystem processes.

Marine plankton ecology research is thus at a crossroads: At a time where marine ecosystems reveal their nature for the first time, these transient ecosystems have already adapted to environmental changes and are continuing to do so (Waters et al.,

2016), with unknown consequences for ecosystem function, and ecosystem service provision. In this context, a close collaboration between researchers belonging to various fields of plankton ecology appears timely to identify the most pressing questions, and to accelerate progress in our understanding of marine ecosystem structure and function. Recently, a EuroMarine foresight workshop on the “*Impact of climate change on the distribution of plankton functional and phylogenetic diversity*” (PlankDiv), held in March 2016 in Villefranche-sur-Mer, France, gathered experts in climate change ecology, species distribution modeling, plankton biology, as well as genomics and evolution. They identified five fundamental questions in future plankton diversity and macroecology research: (1) What can we learn about plankton communities from the new wealth of high-throughput “omics” data? (2) What is the link between plankton diversity and ecosystem function? (3) How can species distribution models be adapted to represent plankton biogeography? (4) How will plankton biogeography be altered due to anthropogenic climate change? and (5) Can a new unifying theory of macroecology be developed based on plankton ecology studies? These questions, along with their associated challenges, are the subject of this review.

THE NEW WEALTH OF PLANKTON DATA

Several recent circumpolar missions have ushered in a new era of plankton biogeography research at the planetary scale. This recent explosion of biological data is perhaps best exemplified by the output of the *Tara Oceans* expedition (Karsenti et al., 2011). While still only offering a temporal snapshot of marine communities, the 7.2 Terabites of metagenomic data gathered are a 1,000 times that generated by the previous largest marine data project, the *Sorcerer II* Global Ocean Sampling (Rusch et al., 2007). High-throughput omics data offer great potential to reveal the global structure of transient marine planktonic ecosystems, since genetic methods compare favorably to traditional observational methods such as microscopy or flow cytometry in terms of the time expenditure, expert knowledge required to identify organisms, and the cost of equipment and analysis. The growing spatial coverage of data enables researchers to estimate global-scale taxonomic diversity of unicellular eukaryotes (de Vargas et al., 2015), to identify the main environmental drivers of community structure in marine prokaryotes (Sunagawa et al., 2015), and to delve into the complexity of biotic interactions between plankton species spanning multiple domains of life, as well as their link to global biogeochemical cycling (Lima-Mendez et al., 2015; Guidi et al., 2016). Complementary to a “bulk” screening of marine biodiversity, single-cell genomics approaches allow matching of phenotype and genotype, and have been used to investigate the phylogenetic affinities of microbial dark matter (i.e., currently unculturable microbial organisms; Rinke et al., 2013; Hug et al., 2016) and to uncover niche partitioning within globally distributed lineages of marine microbes (Kashtan et al., 2014). In combination, bulk and targeted approaches could unravel the taxonomic composition of planktonic organisms, as well as aspects of their ecological function (Thrash et al., 2014; Louca



© Christian Sardet/CNRS/Tara Expéditions

FIGURE 1 | The staggering wealth of plankton species. Diverse assemblages consist of uni- and multicellular organisms with different sizes, morphologies, feeding strategies, ecological functions, life cycle characteristics, and environmental sensitivities. Courtesy of Christian Sardet, from "Plankton—Wonders of the Drifting World" Univ Chicago Press 2015.

et al., 2016) and genome evolution to new environments (Mock et al., 2017).

Both approaches are challenged by the lack of high quality reference databases (Sunagawa et al., 2015). This highlights the need for comprehensive reference databases to guide the validation and integration of the streams of new data, and their comparison with taxonomic information (e.g., Buitenhuis et al., 2013). In addition, genomic sampling often results in temporal snapshots of one particular aspect of biodiversity [e.g., ribosomal-RNA based Operational Taxonomic Unit (OTU) richness]. Applying this approach to marine plankton communities at similarly broad geographic scales is difficult and expensive, but necessary to improve the assessments of the temporal variability of plankton diversity (Lewandowska et al., 2014). Currently, high-resolution time-series datasets are often restricted to easily-accessible, mostly coastal locations, making extrapolation to the expanses of the open ocean difficult. Therefore, the use of these data for ecological purposes may not be straightforward, especially when trying to estimate abundances of planktonic organisms from metabarcoding (e.g., Decelle et al., 2014).

While the genomic quantification of species composition has become more and more common (Bik et al., 2012; Bik, 2014), and harbors potential for marine ecosystem monitoring in times of rapid environmental and ecosystem change, the link between the identity and the functional role of species remains obscure. Genomic approaches can provide thousands of OTUs, whose metabolic state, morphology, and environmental tolerances are

largely unknown. Supplementary measurements of functional traits in laboratory experiments and the quantification of spatio-temporal variability across populations is severely limited by our success in culturing the large diversity of plankton *in vitro*. Estimates that <30% of plankton are cultivable highlight the daunting task of obtaining such data across the heterogeneous plankton lineages and put alternatives, such as single-cell screens, metatranscriptomic approaches, or *in silico* method developments, to the forefront for the characterization of at least some aspects of plankton diversity.

ASSESSING FUNCTIONAL AND PHYLOGENETIC FACETS OF PLANKTON BIODIVERSITY

Traditional approaches have determined marine biodiversity using species occurrence or abundance information at the regional to global scale (e.g., Tittensor et al., 2010). However, there is a growing consensus about the need to assess other facets of biodiversity such as functional diversity, which accounts for biological traits, and phylogenetic diversity to link environmental changes, ecosystem composition and ecosystem function (Naeem et al., 2012; Mouillot et al., 2013). These two promising concepts developed for macro-organisms should be increasingly used within the marine and climate change contexts to further improve our understanding of the link between plankton

diversity, ecosystem productivity, or additional functions related to global biogeochemical cycles.

Functional diversity uses a set of complementary indices (Mouillot et al., 2013) combining measures of species abundance with selected physiological and ecological traits suggested to reflect the fitness of an organism, and which may influence ecosystem function (Violle et al., 2007). Since certain traits may occur across species pertaining to different taxa, estimates of functional diversity allow for the comparison of assemblages with little (no) taxonomic or phylogenetic overlap, but with similar responses to their environment. This metric can account for the intraspecific variability of ecological strategies (e.g., the trophic status of mixotrophic species), and it can include a diverse range of trait variables (e.g., size, feeding strategy, nutrient uptake kinetics). Although much progress has been made in understanding which characteristics of plankton determine their growth, reproduction, and survival (Litchman and Klausmeier, 2008; Litchman et al., 2013; Benedetti et al., 2016), information on traits is restricted to a few well-studied species (Barton et al., 2013). Consequently, trait choice often depends on subjective criteria such as the availability of data (Petchey and Gaston, 2006), therefore open access trait databases should be developed for marine species (Costello et al., 2015). In addition, it is challenging to measure multiple functional traits of thousands of species. Although omics data could allow identifying traits at the community level (Louca et al., 2016), more research is still needed to assign functional traits to sequences, especially for eukaryotic plankton. Despite these methodological issues, trait-based approach of marine communities opens new opportunities for a better understanding of ecosystem functioning and for the development of ecological indicators (Beauchard et al., 2017).

An alternative approach relies on the interspecific phylogenetic differences as a proxy for the overall diversity of a system, assuming that biological characteristics linked to individual fitness and ecological roles show phylogenetic conservatism, i.e., that communities consisting of species with a lower degree of relatedness differ more in their respective trait values, and are thus more diverse (Mouquet et al., 2012). Phylogenetic diversity indices (Tucker et al., 2016) measure the breadth and distribution of evolutionary history present in an assemblage (Mouquet et al., 2012; Cadotte et al., 2013), using DNA sequences to assess the phylogenetic distances between species, by aligning sequences to a reference tree, or by *de-novo* building of phylogenetic trees (Hinchliff et al., 2015).

With the advent of metagenomic data, these promising approaches need to be further explored in terms of their applicability to and relevance for the description of marine ecosystem function. However, the use of phylogenetic diversity critically depends on methodological advances: a substantial fraction of high-throughput sequences obtained by second generation sequencing for microbial communities may still lack sufficient phylogenetic information to provide a reliable phylogenetic placement. In the near future, the popularization of third generation sequencing (e.g., PacBio, Nanopore), which sequences single molecules of DNA in real time, may circumvent

this problem, and will provide full opportunities to use phylogenetic diversity estimates to study present and future ecosystem function.

SPECIES DISTRIBUTION MODELING—RUNNING BEFORE WE CAN WALK?

Species Distribution Models (SDMs) are statistical tools that model a species realized niche, i.e., the environmental conditions under which the species can maintain a viable population (Hutchinson, 1957), by relating their occurrence or abundance to environmental conditions (Guisan and Zimmermann, 2000). Several key ecological attributes make planktonic species particularly well-suited for SDMs (Robinson et al., 2011): (i) their distribution reflects their environmental preferences, since plankton are short-lived organisms, with population dynamics tightly connected to climate (Sunday et al., 2012); (ii) plankton are less commercially exploited than other marine species, and thus, their spatial patterns are less biased by captures as in the case of many fish and shellfish species. These attributes make them a key group for monitoring the impacts of climate change on biodiversity and ecosystem functioning (Richardson, 2008). So far, SDMs have seldom been applied to study plankton biogeography, with only a handful of studies on phytoplankton (Irwin et al., 2012; Pinkernell and Beszteri, 2014; Brun et al., 2015; Rivero-Calle et al., 2015; Barton et al., 2016) and some more on zooplankton (e.g., Reygondeau and Beaugrand, 2011; Chust et al., 2014b; Villarino et al., 2015; Brun et al., 2016; Benedetti et al., in press). This is due not only to the limited data availability for model development, but also due to several unaddressed methodological issues.

In plankton, a major problem with SDMs is the scarcity of occurrence data, which can lead to an incomplete niche description and/or biased models. A major challenge is therefore to discern biological distribution patterns from patterns of sampling effort, especially in traditional taxonomy-based plankton data sets where reliable absences data are usually unavailable and large regions, such as the South Pacific, are chronically undersampled. Using one of the most extensive plankton data sets to date, the North Atlantic Continuous Plankton Recorder data, Brun et al. (2016) found that a suite of commonly used SDMs are unable to predict and hindcast the distribution of zooplankton and phytoplankton example-species on the decadal scale. One way to improve SDMs is either through careful methodological adjustments, such as a targeted selection of the background (Phillips et al., 2009), the reduction of environmental predictors, and model complexity (Merow et al., 2014). Another approach could be to merge existing data archives and to combine genomic data with traditional approaches in order to reduce the sampling bias. However, since SDMs apply at the species level, this will require specific identifications, either from microscopy, imaging, or sequencing, which would necessitate to keep taxonomic expertise in our laboratories and, in parallel, to develop specific tools for automatic identification.

In their basic form and most common use, classical SDMs do not account generally for three major ecological processes that may be crucial for plankton distribution: (i) the role of dispersal and its limitation, (ii) biotic interactions, and (iii) intraspecific variability, which we discuss below. The relative importance of these processes in shaping planktonic species' ranges is still being under debate (Cermeño and Falkowski, 2009; Chust et al., 2013).

Plankton dispersal is controlled by ocean currents and can impact diversity and community structure (Lévy et al., 2014). Although barriers to dispersal are fewer in the marine realm compared to the terrestrial one (Steele, 1991), coupling ocean connectivity patterns (Tremblay et al., 2008; Foltête et al., 2012) with niche models is likely important. Source-sink dynamics may arise frequently because of the advection of water masses (e.g., Beaugrand et al., 2007; Villar et al., 2015) that can introduce species to unsuitable regions (Pulliam, 2000), potentially biasing SDMs. Future developments for plankton could ensue from graph-based techniques (Dale and Fortin, 2010) and from SDMs coupling with simple dispersal models (Foltête et al., 2012; Zurell et al., 2016).

Furthermore, the need to account for biotic interactions when predicting species distributions has been advocated (Boulangé et al., 2012; Wisz et al., 2013). Recently, the exploration of the plankton “*interactome*” (Lima-Mendez et al., 2015) allowed to describe how biotic interactions occur across trophic levels and relate to environmental conditions and ecosystem functioning, with many new symbiotic interactions identified (Guidi et al., 2016). When prior knowledge is too limited, food-web models could be inferred from simple size-based, or multi-traits assumptions (Albouy et al., 2014), or based on ecosystem models (e.g., Follows et al., 2007; Le Quéré et al., 2016) in combination with satellite estimates of (phyto)plankton community composition (e.g., Hirata et al., 2011).

Finally, SDMs do not consider intraspecific variability, thus assuming that genetic adaptation is negligible. However, many planktonic species exhibit local adaptation (Peijnenburg and Goetze, 2013; Sjöqvist et al., 2015) or consist of several ecotypes with different environmental preferences, and phenotypic plasticity, dispersal, and evolutionary changes could mitigate climate change impacts as they could help species to adapt to changing conditions (O'Connor et al., 2012). One possibility to account for both local adaptation and phenotypic plasticity is to include a population-dependent component in mixed effect models (e.g., Valladares et al., 2014). Furthermore, the joint use of genomic and taxonomic information may help to constrain the differences between subpopulations or ecotypes of a species, and to identify so-called cryptic species.

ADRIFT IN AN OCEAN OF CHANGE

In contrast to works on higher trophic levels (e.g., Cheung et al., 2009), the investigation of the response of plankton to future climate changes has mostly focused more on bulk variables (e.g., biomass, production), with large uncertainties associated with the simulated response of primary and secondary production (e.g., Bopp et al., 2013; Laufkötter et al., 2015). Yet,

observational evidence of changes in planktonic ecosystems has been accumulating over the past decades, with ongoing efforts to attribute these changes to specific environmental drivers (e.g., Beaugrand et al., 2008; Rivero-Calle et al., 2015).

SDMs have been used to support observations of poleward plankton distribution range shifts in response to global warming in the North Atlantic (Beaugrand et al., 2002; Richardson, 2008), as well as changes in the relative abundance of certain groups (Rivero-Calle et al., 2015). However, range shifts and in particular phenological changes can vary according to region and species, leading to unexpected emergent patterns (Richardson et al., 2012; Poloczanska et al., 2013; Burrows et al., 2014; Barton et al., 2015). In fact, multiple non-exclusive and interlinked adaptation strategies at the organismal level may all operate in concert, or, alternatively, the selection of one strategy may reduce the necessity to employ another. For example, shifts in spatial distribution may preclude the necessity for phenological adjustments in a given species attempting to maintain its thermal niche. Other adaptation strategies involve species plasticity and genetic modification in order to face changing conditions (Lavergne et al., 2010; Dam, 2013), which have been documented for spatially isolated zooplankton (Peijnenburg et al., 2006; Yebra et al., 2011), but could not be confirmed for other species (Provan et al., 2009). Another alternative adaptation strategy is the change in depth-distribution, i.e., the migration to deeper waters in search for cooler temperatures carried out by fishes (Perry et al., 2005).

Given the multitude of adaptation options, future projections of ecosystem change are prone to large uncertainties. Moreover, disentangling the effects of anthropogenic climate change on plankton distribution and phenology shifts from other drivers (e.g., climate variability, population dynamics) is equally challenging (Chust et al., 2014b). In particular, the combination of controlling factors, together with systematic biases in sampling effort can lead to biases in estimated trends. The decomposition of factors using different SDMs can detect the so-called “niche tracking,” which is the shift of a species distribution to follow the displacement of their habitat, e.g., poleward shifts (Monahan and Tingley, 2012; Bruge et al., 2016). At the community level, thermal biases between the average thermal affinity of assemblages and local temperature (Stuart-Smith et al., 2015) have to be considered to improve our understanding of the sensitivity of plankton reorganization with warming.

TOWARD A UNIFIED THEORY OF MACROECOLOGY

Predicting how species will respond to global environmental change requires an understanding of the processes generating their current large-scale spatio-temporal patterns of diversity and distribution, which is the essence of macroecology. One such predominant pattern on Earth is the decline in biodiversity of terrestrial and marine macroorganisms from tropical to polar areas (e.g., Tittensor et al., 2010). Hypotheses explaining this pattern often call upon evolutionary history (Mittelbach et al., 2007), diversity-area relations (Rosenzweig,

1995), temperature effects (Allen et al., 2002), or climatic stability (Fraser and Currie, 1996). Although these premises often find empirical support, their testing in the open oceans has been limited. Whereas, zooplankton likely reflect the general latitudinal trend (Beaugrand et al., 2013), bacterioplankton may form seasonal diversity peaks at high (Ladau et al., 2013) and mid (Sunagawa et al., 2015) latitudes, and for phytoplankton the validity of the global pattern itself and the processes that may explain it are still ambiguous (Rodríguez-Ramos et al., 2015; O'Brien et al., 2016). To alleviate data scarcity, which may have contributed to uncertainty, we suggest the implementation of SDMs as strategic tools to integrate novel with traditional data and to depict aspects of global diversity variation across major taxa and spatio-temporal scales.

The validity of the concept of SDM in plankton and its specific adaptation warrant further testing of the processes that determine plankton distribution, abundance, community assembly, and the maintenance of diversity at local to global scales. More than a decade after the appearance of the unified neutral theory of biodiversity (Hubbell, 2001), there is still an active debate on the relative contribution of demographic stochasticity, dispersal, and niche processes on plankton communities (Pueyo, 2006a,b; Cermeño and Falkowski, 2009; Chust et al., 2013), which promoted the revisiting of the “*Paradox of the Plankton*” (Hutchinson, 1961). Recent studies have tried to reconcile neutral and niche theories (Adler et al., 2007) and suggest that neutral combined with metabolic theory can explain macroecological patterns (Tittensor and Worm, 2016). Furthermore, neutral processes might similarly shape both population genetics and community patterns in plankton (Chust et al., 2016). The combination of data from time-series, global *in situ* observations and experiments on marine plankton provides a unique opportunity to characterize the niches of species (Brun et al., 2015) and to explore the relations between ecological niche characteristics (e.g., niche dissimilarity) and local species richness.

Thus, important open questions include: Is plankton community assembly mainly driven by niche assembly or neutral processes? Does this depend on the spatio-temporal scale of observation? Which method(s) can be used to disentangle the dominating process in community assembly and ecosystem structure? What will be the effect of the removal of geographical barriers that have long separated the Earth's biogeographical provinces on marine plankton diversity (“homogocene,” Rosenzweig, 2001)? How does the evolution of microorganism dependency based on gene loss shape the structure and dynamics of communities (Mas et al., 2016)? Due to their fast duplication rates and rapid response to environmental conditions, planktonic communities assemble, dismantle, and re-assemble constantly in natural environments, thus tracking environmental disturbances. Therefore, they are optimally suited to test classical ecological theories established for terrestrial

ecosystems, and to answer questions related to diversity-stability relationships, the area-diversity hypothesis, or food web interactions.

CONCLUSION

Plankton ecology research stands at a crossroads. The staggering increase in the wealth of plankton observation data coincides with a time of significant advances in marine ecosystem modeling, which allow, for the first time, the testing of important theories of macroecology in the marine realm. These achievements offer great promise to shed light on marine ecosystem functioning and ecosystem service provision within the context of global climate change. To unlock their potential, we identified a strong need for concomitant developments in the field of bioinformatics and biostatistics, ecological niche modeling, and genetic reference database assembly, thus allowing for a successful integration of these novel with traditional observations, including taxonomic expertise. Paired with the rigorous verification of new and existing macro-ecological theories in the marine realm, and the testing and application of novel biodiversity metrics that better link ecosystem composition to ecosystem function and ecosystem service provision, these theoretical and empirical advances may allow for the urgently needed quantification of potential impacts of climate change on marine ecosystems and feedbacks to higher trophic levels. Due to the complexity of the task, and the scarcity of observational evidence of these transient ecosystems, we conclude that interdisciplinary, collaborative efforts between experts focussing on different aspects of plankton ecology will be critical in mediating this process.

AUTHOR CONTRIBUTIONS

GC, MV, FB, TN, SV, AA, SMV, DR, JI, and SA conceived and wrote the main manuscript text. All authors reviewed the manuscript.

ACKNOWLEDGMENTS

This research was funded by the EuroMarine Network (<http://www.euromarinenetwork.eu>), through the organization of the PlankDiv EuroMarine Foresight workshop, held at the Observatoire Océanographique de Villefranche-sur-mer, Villefranche-sur-mer, France, in March 2016, and cofounded by the Basque Government (Department Deputy of Agriculture, Fishing and Food Policy). The PlankDiv workshop was also supported by the Laboratoire d'Océanographie de Villefranche-sur-mer (LOV, UPMC/CNRS), the PlankMed action of WP5 MERMEX/MISTRAL, and by the French national programme EC2CO-LEFE (FunOmics project). This is contribution 810 from AZTI Marine Research Division. We thank the editor and three reviewers for their insightful comments, which greatly improved the manuscript.

REFERENCES

- Adler, P. B., Hillerislambers, J., and Levine, J. M. (2007). A niche for neutrality. *Ecol. Lett.* 10, 95–104. doi: 10.1111/j.1461-0248.2006.00996.x
- Albouy, C., Velez, L., Coll, M., Colloca, F., Le Loc'h, F., Mouillot, D., et al. (2014). From projected species distribution to food-web structure under climate change. *Glob. Change Biol.* 20, 730–741. doi: 10.1111/gcb.12467
- Allen, A. P., Brown, J. H., and Gillooly, J. F. (2002). Global biodiversity, biochemical kinetics, and the energetic-equivalence rule. *Science* 297, 1545–1548. doi: 10.1126/science.1072380
- Barton, A. D., Irwin, A. J., Finkel, Z. V., and Stock, C. A. (2016). Anthropogenic climate change drives shift and shuffle in North Atlantic phytoplankton communities. *Proc. Natl. Acad. Sci.* 113, 2964–2969. doi: 10.1073/pnas.1519080113
- Barton, A. D., Lozier, M. S., and Williams, R. G. (2015). Physical controls of variability in North Atlantic phytoplankton communities. *Limnol. Oceanogr.* 60, 181–197. doi: 10.1002/lno.10011
- Barton, A. D., Pershing, A. J., Litchman, E., Record, N. R., Edwards, K. F., Finkel, Z. V., et al. (2013). The biogeography of marine plankton traits. *Ecol. Lett.* 16, 522–534. doi: 10.1111/ele.12063
- Beauchard, O., Veríssimo, H., Queirós, A., and Herman, P. (2017). The use of multiple biological traits in marine community ecology and its potential in ecological indicator development. *Ecol. Indic.* 76, 81–96. doi: 10.1016/j.ecolind.2017.01.011
- Beaugrand, G., Edwards, M., Brander, K., Luczak, C., and Ibanez, F. (2008). Causes and projections of abrupt climate-driven ecosystem shifts in the North Atlantic. *Ecol. Lett.* 11, 1157–1168. doi: 10.1111/j.1461-0248.2008.01218.x
- Beaugrand, G., Lindley, J. A., Helaouet, P., and Bonnet, D. (2007). Macroecological study of *Centropages typicus* in the North Atlantic Ocean. *Prog. Oceanogr.* 72, 259–273. doi: 10.1016/j.pocean.2007.01.002
- Beaugrand, G., Mackas, D., and Goberville, E. (2013). Applying the concept of the ecological niche and a macroecological approach to understand how climate influences zooplankton: advantages, assumptions, limitations and requirements. *Prog. Oceanogr.* 111, 75–90. doi: 10.1016/j.pocean.2012.11.002
- Beaugrand, G., Reid, P. C., Ibañez, F., Lindley, J. A., and Edwards, M. (2002). Reorganisation of North Atlantic marine copepod biodiversity and climate. *Science* 296, 1692–1694. doi: 10.1126/science.1071329
- Benedetti, F., Gasparini, S., and Ayata, S.-D. (2016). Identifying copepod functional groups from species functional traits. *J. Plankton Res.* 38, 159–166. doi: 10.1093/plankt/fbv096
- Benedetti, F., Guilhaumon, F., Adloff, F., and Ayata, S.-D. (in press). Investigating uncertainties in zooplankton composition shifts under climate change scenarios in the Mediterranean Sea. *Ecography*. doi: 10.1111/ecog.02434
- Bopp, L., Resplandy, L., Orr, J. C., Doney, S. C., Dunne, J. P., Gehlen, M., et al. (2013). Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models. *Biogeosciences* 10, 6225–6245. doi: 10.5194/bg-10-6225-2013
- Bik, H. M. (2014). Deciphering diversity and ecological function from marine metagenomes. *Biol. Bull.* 227, 107–116. doi: 10.1086/BBLv227n2p107
- Bik, H. M., Porazinska, D. L., Creer, S., Caporaso, J. G., Knight, R., and Thomas, W. K. (2012). Sequencing our way towards understanding global eukaryotic biodiversity. *Trends Ecol. Evol.* 27, 233–243. doi: 10.1016/j.tree.2011.11.010
- Borja, A., Elliott, M., Carstensen, J., Heiskanen, A. S., and van de Bund, W. (2010). Marine management—towards an integrated implementation of the European marine strategy framework and the water framework directives. *Mar. Pollut. Bull.* 60, 2175–2186. doi: 10.1016/j.marpolbul.2010.09.026
- Boulangeat, I., Gravel, D., and Thuiller, W. (2012). Accounting for dispersal and biotic interactions to disentangle the drivers of species distributions and their abundances. *Ecol. Lett.* 15, 584–593. doi: 10.1111/j.1461-0248.2012.01772.x
- Bruge, A., Alvarez, P., Fontán, A., Cotano, U., and Chust, G. (2016). Thermal niche tracking and future distribution of Atlantic mackerel spawning in response to ocean warming. *Front. Mar. Sci.* 3:86. doi: 10.3389/fmars.2016.00086
- Brun, P., Kiørboe, T., Licandro, P., and Payne, M. R. (2016). The predictive skill of species distribution models for plankton in a changing climate. *Global Change Biol.* 22, 3170–3181. doi: 10.1111/gcb.13274
- Brun, P., Vogt, M., Payne, M. R., Gruber, N., O'Brien, C., Buitenhuis, E. T., et al. (2015). Ecological niches of open ocean phytoplankton. *Limnol. Oceanogr.* 60, 1020–1038. doi: 10.1002/lno.10074
- Buitenhuis, E. T., Hashioka, T., and Quéré, C. L. (2013). Combined constraints on global ocean primary production using observations and models. *Glob. Biogeochem. Cycles* 27, 847–858. doi: 10.1002/gbc.20074
- Burrows, M. T., Schoeman, D. S., Richardson, A. J., Molinos, J. G., Hoffmann, A., Buckley, L. B., et al. (2014). Geographical limits to species-range shifts are suggested by climate velocity. *Nature* 507, 492–495. doi: 10.1038/nature12976
- Cadotte, M., Albert, C. H., and Walker, S. C. (2013). The ecology of differences: assessing community assembly with trait and evolutionary distances. *Ecol. Lett.* 16, 1234–1244. doi: 10.1111/ele.12161
- Cermeño, P., and Falkowski, P. G. (2009). Controls on diatom biogeography in the ocean. *Science* 325, 1539–1541. doi: 10.1126/science.1174159
- Cheung, W. W., Lam, V. W., Sarmiento, J. L., Kearney, K., Watson, R., and Pauly, D. (2009). Projecting global marine biodiversity impacts under climate change scenarios. *Fish Fish.* 10, 235–251. doi: 10.1111/j.1467-2979.2008.00315.x
- Cheung, W. W., Sarmiento, J. L., Dunne, J., Frölicher, T. L., Lam, V. W., Palomares, M. D., et al. (2013). Shrinking of fishes exacerbates impacts of global ocean changes on marine ecosystems. *Nat. Clim. Change* 3, 254–258. doi: 10.1038/nclimate1691
- Chust, G., Allen, J. L., Bopp, L., Schrum, C., Holt, J., Tsiaras, K., et al. (2014a). Biomass changes and trophic amplification of plankton in a warmer ocean. *Glob. Change Biol.* 20, 2124–2139. doi: 10.1111/gcb.12562
- Chust, G., Castellani, C., Licandro, P., Ibaibarriaga, L., Sagarminaga, Y., and Irigoien, X. (2014b). Are Calanus spp. shifting poleward in the North Atlantic? a habitat modelling approach. *ICES J. Mar. Sci.* 71, 241–253. doi: 10.1093/icesjms/fst147
- Chust, G., Villarino, E., Chenuil, A., Irigoien, X., Bizsel, N., Bode, A., et al. (2016). Dispersal similarly shapes both population genetics and community patterns in the marine realm. *Sci. Rep.* 6:28730. doi: 10.1038/srep28730
- Chust, G., Irigoien, X., Chave, J., and Harris, R. P. (2013). Latitudinal phytoplankton distribution and the neutral theory of biodiversity. *Glob. Ecol. Biogeogr.* 22, 531–543. doi: 10.1111/geb.12016
- Costello, M. J., Claus, S., Dekeyser, S., Vandepitte, L., Tuama, É. Ó., Lear, D., et al. (2015). Biological and ecological traits of marine species. *Peer J* 3:e1201. doi: 10.7717/peerj.1201
- Dale, M., and Fortin, M.-J. (2010). From graphs to spatial graphs. *Annu. Rev. Ecol. Syst.* 41, 21–38. doi: 10.1146/annurev-ecolsys-102209-144718
- Dam, H. G. (2013). Evolutionary adaptation of marine zooplankton to global change. *Annu. Rev. Mar. Sci.* 5, 349–370. doi: 10.1146/annurev-marine-121211-172229
- de Vargas, C., Audic, S., Henry, N., Decelle, J., Mahé, F., Logares, R., et al. (2015). Eukaryotic plankton diversity in the sunlit ocean. *Science* 348:1261605. doi: 10.1126/science.1261605
- Decelle, J., Romac, S., Sasaki, E., Not, F., and Mahé, F. (2014). Intracellular Diversity of the V4 and V9 Regions of the 18S rRNA in marine protists (Radiolarians) assessed by high-throughput sequencing. *PLoS ONE* 9, 1–8. doi: 10.1371/journal.pone.0104297
- Doney, S. C., Ruckelshaus, M., Duffy, J. E., Barry, J. P., Chan, F., English, C. A., et al. (2012). Climate change impacts on marine ecosystems. *Annu. Rev. Mar. Sci.* 4, 11–37. doi: 10.1146/annurev-marine-041911-111611
- Duarte, C. M. (2015). Seafaring in the 21st Century: The malaspina 2010 circumnavigation expedition. *Limnol. Oceanogr. Bull.* 24, 11–14. doi: 10.1002/lob.10008
- Duarte, C. M. R. W., Fulweiler, C. E., Lovelock, J. M., Pandolfi, P., Martinetto, M. I., Saunders, M. I., et al. (2015). Ocean calamities: delineating the boundaries between scientific evidence and belief. *Bioscience* 65, 746–747. doi: 10.1093/biosci/biv088
- Edwards, M., and Richardson, A. J. (2004). Impact of climate change on marine pelagic phenology and trophic mismatch. *Nature* 430, 881–884. doi: 10.1038/nature02808
- Follows, M. J., Dutkiewicz, S., Grant, S., and Chisholm, S. W. (2007). Emergent biogeography of microbial communities in a model ocean. *Science* 315, 1843–1846. doi: 10.1126/science.1138544
- Foltête, J. C., Clauzel, C., Vuidel, G., and Tournant, P. (2012). Integrating graph-based connectivity metrics into species distribution models. *Landscape Ecol.* 4, 557–569. doi: 10.1007/s10980-012-9709-4
- Fraser, R. H., and Currie, D. J. (1996). The species richness-energy hypothesis in a system where historical factors are thought to prevail: coral reefs. *Am. Nat.* 148, 138–159. doi: 10.1086/285915

- Guidi, L., Chaffron, S., Bittner, L., Eveillard, D., Larhlimi, A., Roux, S., et al. (2016). Plankton networks driving carbon export in the oligotrophic ocean. *Nature* 532, 465–470. doi: 10.1038/nature16942
- Guisan, A., and Zimmermann, N. (2000). Predictive habitat distribution models in ecology. *Ecol. Model.* 135, 147–186. doi: 10.1016/S0304-3800(00)00354-9
- Hinchliff, C. E., Smith, S. A., Allman, J. F., Burleigh, J. G., Chaudhary, R., Coghill, L. M., et al. (2015). Synthesis of phylogeny and taxonomy into a comprehensive tree of life. *Proc. Natl. Acad. Sci. U.S.A.* 112, 12764–12769. doi: 10.1073/pnas.1423041112
- Hirata, T., Hardman-Mountford, N. J., Brewin, R. J. W., Aiken, J., Barlow, R., Suzuki, K., et al. (2011). Synoptic relationships between surface Chlorophyll-*a* and diagnostic pigments specific to phytoplankton functional types. *Biogeosciences* 8, 311–327. doi: 10.5194/bg-8-311-2011
- Hubbell, S. P. (2001). *The Unified Neutral Theory of Biodiversity and Biogeography*. Princeton, NJ: Princeton University Press.
- Hug, L. A., Baker, B. J., Anantharaman, K., Brown, C. T., Probst, A. J., Castelle, C. J., et al. (2016). A new view of the tree of life. *Nat. Microbiol.* 1:16048. doi: 10.1038/nmicrobiol.2016.48
- Hutchinson, G. E. (1957). Concluding remarks. *Cold Spring Harbor Symp. Quant. Biol.* 22, 415–427. doi: 10.1101/SQB.1957.022.01.039
- Hutchinson, G. E. (1961). The paradox of the plankton. *Am. Nat.* 95, 137–146. doi: 10.1086/282171
- Irwin, A. J., Nelles, A. M., and Finkel, Z. V. (2012). Phytoplankton niches estimated from field data. *Limnol. Oceanogr.* 57, 787–797. doi: 10.4319/lo.2012.57.3.0787
- Karsenti, E., Acinas, S. G., Bork, P., Bowler, C., de Vargas, C., Raes, J., et al. (2011). A holistic approach to marine eco-systems biology. *PLoS Biol.* 9:e1001177. doi: 10.1371/journal.pbio.1001177
- Kashtan, N., Roggensack, S. E., Rodrigue, S., Thompson, J. W., Biller, S. J., Coe, A., et al. (2014). Single-cell genomics reveals hundreds of coexisting subpopulations in wild *Prochlorococcus*. *Science* 344, 416–420. doi: 10.1126/science.1248575
- Ladau, J., Sharpton, T. J., Finucane, M. M., Jospin, G., Kembel, S. W., O'Dwyer, J., et al. (2013). Global marine bacterial diversity peaks at high latitudes in winter. *ISME J.* 7, 1669–1677. doi: 10.1038/ismej.2013.37
- Laufkötter, C., Vogt, M., Gruber, N., Aita-Noguchi, M., Aumont, O., Bopp, L., et al. (2015). Drivers and uncertainties of future global marine primary production in marine ecosystem models. *Biogeosciences* 12, 6955–6984. doi: 10.5194/bg-12-6955-2015
- Lavergne, S., Mouquet, N., Thuiller, W., and Ronce, O. (2010). Biodiversity and climate change: integrating evolutionary and ecological responses of species and communities. *Annu. Rev. Ecol. Evol. Syst.* 41, 321–350. doi: 10.1146/annurev-ecolsys-102209-144628
- Le Quéré, C., Andrew, R. M., Canadell, J. G., Sitch, S., Korsbakken, J. I., Peters, G. P., et al. (2016). Global carbon budget 2016. *Earth Syst. Sci. Data* 8, 605–649. doi: 10.5194/essd-8-605-2016
- Lévy, M., Jahn, O., Dutkiewicz, S., and Follows, M. J. (2014). Phytoplankton diversity and community structure affected by oceanic dispersal and mesoscale turbulence. *Limnol. Oceanogr. Fluids Environ.* 4, 67–84. doi: 10.1215/21573689-2768549
- Lewandowska, A. M., Boyce, D. G., Hofmann, M., Matthiessen, B., Sommer, U., and Worm, B. (2014). Effects of sea surface warming on marine plankton. *Ecol. Lett.* 17, 614–623. doi: 10.1111/ele.12265
- Lima-Mendez, G., Faust, K., Henry, N., Decelle, J., Colin, S., Carcillo, F., et al. (2015). Determinants of community structure in the global plankton interactome. *Science* 348:1262073. doi: 10.1126/science.1262073
- Litchman, E., and Klausmeier, C. A. (2008). Trait-based community ecology of phytoplankton. *Annu. Rev. Ecol. Evol. Syst.* 39, 615–639. doi: 10.1146/annurev.ecolsys.39.110707.173549
- Litchman, E., Ohman, M. D., and Kjørboe, T. (2013). Trait-based approaches to zooplankton communities. *J. Plankton Res.* 35, 473–484. doi: 10.1093/plankt/fbt019
- Louca, S., Parfrey, L. W., and Doebeli, M. (2016). Decoupling function and taxonomy in the global ocean microbiome. *Science* 353, 1272–1277. doi: 10.1126/science.aaf4507
- Mas, A., Jamshidi, S., Lagadeuc, Y., Eveillard, D., and Vandenkoornhuyse, P., (2016). Beyond the black queen hypothesis. *ISME J.* 10, 2085–2091. doi: 10.1038/ismej.2016.22
- Merow, C., Smith, M. J., Edwards, T. C., Guisan, A., McMahon, S. M., Normand, S., et al. (2014). What do we gain from simplicity versus complexity in species distribution models? *Ecography* 37, 1267–1281. doi: 10.1111/ecog.00845
- Mittelbach, G. G., Schemske, D. W., Cornell, H. V., Allen, A. P., Brown, J. M., Bush, M. B., et al. (2007). Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. *Ecol. Lett.* 10, 315–331. doi: 10.1111/j.1461-0248.2007.01020.x
- Mock, T., Otilar, R. P., Strauss, J., McMullan, M., Paajanen, P., Schmutz, J., et al. (2017). Evolutionary genomics of the cold-adapted diatom *Fragilariopsis cylindrus*. *Nature* 541, 536–540. doi: 10.1038/nature20803
- Monahan, W. B., and Tingley, M. W. (2012). Niche tracking and rapid establishment of distributional equilibrium in the house sparrow show potential responsiveness of species to climate change. *PLoS ONE* 7:e42097. doi: 10.1371/journal.pone.0042097
- Mouillot, D., Graham, N. A., Villéger, S., Mason, N. W., and Bellwood, D. R. (2013). A functional approach reveals community responses to disturbances. *Trends Ecol. Evol.* 28, 167–177. doi: 10.1016/j.tree.2012.10.004
- Mouquet, N., Devictor, V., Meynard, C. N., Munoz, F., Bersier, L.-F., Chave, J., et al. (2012). Ecophylogenetics: advances and perspectives. *Biol. Rev.* 87, 769–785. doi: 10.1111/j.1469-185X.2012.00224.x
- Naem, S., Duffy, J. E., and Zavaleta, E. (2012). The functions of biological diversity in an age of extinction. *Science* 336, 1401–1406. doi: 10.1126/science.1215855
- O'Brien, C. J., Vogt, M., and Gruber, N. (2016). Global coccolithophore diversity: drivers and future change. *Prog. Oceanogr.* 140, 27–42. doi: 10.1016/j.pocean.2015.10.003
- O'Connor, M. I., Selig, E. R., Pinsky, M. L., and Altermatt, F. (2012). Toward a conceptual synthesis for climate change responses. *Glob. Ecol. Biogeogr.* 21, 693–703. doi: 10.1111/j.1466-8238.2011.00713.x
- Peijnenburg, K. T., and Goetze, E. (2013). High evolutionary potential of marine zooplankton. *Ecol. Evol.* 3, 2765–2781. doi: 10.1002/ece3.644
- Peijnenburg, K. T., Fauvelot, C., Breeuwer, J. A., and Menken, S. B. (2006). Spatial and temporal genetic structure of the planktonic *Sagittia setosa* (Chaetognatha) in European seas as revealed by mitochondrial and nuclear DNA markers. *Mol. Ecol.* 15, 3319–3338. doi: 10.1111/j.1365-294X.2006.03002.x
- Perry, A. L., Low, P. J., Ellis, J. R., and Reynolds, J. D. (2005). Climate change and distribution shifts in marine fishes. *Science* 308, 1912–1915. doi: 10.1126/science.1111322
- Pesant, S., Not, F., Picheral, M., Kandels-Lewis, S., Le Bescot, N., Gorsky, G., et al. (2015). Open science resources for the discovery and analysis of Tara Oceans data. *Sci. Data* 2:150023. doi: 10.1038/sdata.2015.23
- Petchey, O. L., and Gaston, K. J. (2006). Functional diversity: back to basics and looking forward. *Ecol. Lett.* 9, 741–758. doi: 10.1111/j.1461-0248.2006.00924.x
- Phillips, S. J., Dudík, M., Elith, J., Graham, C. H., Lehmann, A., Leathwick, J., et al. (2009). Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. *Ecol. Appl.* 19, 181–197. doi: 10.1890/07-2153.1
- Pinkernell, S., and Beszteri, B. (2014). Potential effects of climate change on the distribution range of the main silicate sinker of the Southern Ocean. *Ecol. Evol.* 4, 3147–3161. doi: 10.1002/ece3.1138
- Poloczanska, E. S., Brown, C. J., Sydeman, W. J., Kiessling, W., Schoeman, D. S., Moore, P., et al. (2013). Global imprint of climate change on marine life. *Nat. Clim. Change* 3, 919–925. doi: 10.1038/nclimate1958
- Provan, J., Beatty, G. E., Keating, S. L., Maggs, C. A., and Savidge, G. (2009). High dispersal potential has maintained long-term population stability in the North Atlantic copepod *Calanus finmarchicus*. *Proc. R. Soc. Lond. B Biol. Sci.* 276, 301–307. doi: 10.1098/rspb.2008.1062
- Pueyo, S. (2006a). Diversity: between neutrality and structure. *Oikos* 112, 392–405. doi: 10.1111/j.0030-1299.2006.14188.x
- Pueyo, S. (2006b). Self-similarity in species–area relationship and in species abundance distribution. *Oikos* 112, 156–162. doi: 10.1111/j.0030-1299.2006.14184.x
- Pulliam, H. R. (2000). On the relationship between niche and distribution. *Ecol. Lett.* 3, 349–361. doi: 10.1046/j.1461-0248.2000.00143.x
- Reygondeau, G., and Beaugrand, G. (2011). Future climate-driven shifts in distribution of *Calanus finmarchicus*. *Glob. Change Biol.* 17, 756–766. doi: 10.1111/j.1365-2486.2010.02310.x
- Richardson, A. J. (2008). In hot water: zooplankton and climate change. *ICES J. Mar. Sci.* 65, 279–295. doi: 10.1093/icesjms/fsn028

- Richardson, A. J., Brown, C. J., Brander, K., Bruno, J. F., Buckley, L., Burrows, M. T., et al. (2012). Climate change and marine life. *Biol. Lett.* 8, 907–909. doi: 10.1098/rsbl.2012.0530
- Rinke, C., Schwientek, P., Sczyrba, A., Ivanova, N. N., Anderson, I. J., Cheng, J.-F., et al. (2013). Insights into the phylogeny and coding potential of microbial dark matter. *Nature* 499, 431–437. doi: 10.1038/nature12352
- Rivero-Calle, S., Gnanadesikan, A., Del Castillo, C. E., Balch, W. M., and Guikema, S. D. (2015). Multidecadal increase in North Atlantic coccolithophores and the potential role of rising CO₂. *Science* 350, 1533–1537. doi: 10.1126/science.aaa8026
- Robinson, L. M., Elith, J., Hobday, A. J., Pearson, R. G., Kendall, B. E., Possingham, H. P., et al. (2011). Pushing the limits in marine species distribution modelling: lessons from the land present challenges and opportunities. *Glob. Ecol. Biogeogr.* 20, 789–802. doi: 10.1111/j.1466-8238.2010.00636.x
- Rodríguez-Ramos, T., Marañón, E., and Cermeño, P. (2015). Marine nano- and microphytoplankton diversity: redrawing global patterns from sampling-standardized data. *Glob. Ecol. Biogeogr.* 24, 527–538. doi: 10.1111/geb.12274
- Rosenzweig, M. L. (1995). *Species Diversity in Space and Time*. Cambridge: Cambridge University Press.
- Rosenzweig, M. L. (2001). Four questions: what does the introduction of exotic species do to diversity? *Evol. Ecol. Res.* 3, 361–367.
- Rusch, D. B., Halpern, A. L., Sutton, G., Heidelberg, K. B., Williamson, S., Yooseph, S., et al. (2007). The Sorcerer II global ocean sampling expedition: northwest atlantic through eastern tropical pacific. *PLoS Biol.* 5:e77. doi: 10.1371/journal.pbio.0050077
- Sjöqvist, C., Godhe, A., Jonsson, P. R., Sundqvist, L., and Kremp, A. (2015). Local adaptation and oceanographic connectivity patterns explain genetic differentiation of a marine diatom across the North Sea–Baltic Sea salinity gradient. *Mol. Ecol.* 24, 2871–2885. doi: 10.1111/mec.13208
- St. John, M. A., Borja, A., Chust, G., Heath, M., Grigorov, I., Mariani, P., et al. (2016). A dark hole in our understanding of marine ecosystems and their services: perspectives from the mesopelagic community. *Front. Mar. Sci.* 3:31. doi: 10.3389/fmars.2016.00031
- Steele, J. H. (1991). Can ecological theory cross the land-sea boundary? *J. Theor. Biol.* 153, 425–436. doi: 10.1016/S0022-5193(05)80579-X
- Stuart-Smith, R. D., Edgar, G. J., Barrett, N. S., Kininmonth, S. J., and Bates, A. E. (2015). Thermal biases and vulnerability to warming in the world's marine fauna. *Nature* 528, 88–92. doi: 10.1038/nature16144
- Sunagawa, S., Coelho, L. P., Chaffron, S., Kultima, J. R., Labadie, K., Salazar, G., et al. (2015). Structure and function of the global ocean microbiome. *Science* 348:1261359. doi: 10.1126/science.1261359
- Sunday, J. M., Bates, A. E., and Dulvy, N. K. (2012). Thermal tolerance and the global redistribution of animals. *Nat. Clim. Change* 2, 686–690. doi: 10.1038/nclimate1539
- Thrash, J. C., Temperton, B., Swan, B. K., Landry, Z. C., Woyke, T., DeLong, E. F., et al. (2014). Single-cell enabled comparative genomics of a deep ocean SAR11 bathytype. *ISME J.* 8, 1440–1451. doi: 10.1038/ismej.2013.243
- Tittensor, D. P., and Worm, B. (2016). A neutral-metabolic theory of latitudinal biodiversity. *Glob. Ecol. Biogeogr.* 25, 630–641. doi: 10.1111/geb.12451
- Tittensor, D. P., Mora, C., Jetz, W., Lotze, H. K., Ricard, D., Berghe E.V., et al. (2010). Global patterns and predictors of marine biodiversity across taxa. *Nature* 466, 1098–1101. doi: 10.1038/nature09329
- Treml, E. A., Halpin, P. N., Urban, D. L., and Pratson, L. F. (2008). Modeling population connectivity by ocean currents, a graph-theoretic approach for marine conservation. *Landscape Ecol.* 23, 19–36. doi: 10.1007/s10980-007-9138-y
- Tucker, C. M., Cadotte, M. W., Carvalho, S. B., Davies, T. J., Ferrier, S., Fritz, S. A., et al. (2016). A guide to phylogenetic metrics for conservation, community ecology and macroecology. *Biol. Rev. Camb. Philos. Soc.* doi: 10.1111/brv.12252. [Epub ahead of print].
- Valladares, F., Matesanz, S., Guilhaumon, F., Araújo, M. B., Balaguer, L., Benito-Garazón, M., et al. (2014). The effects of phenotypic plasticity and local adaptation on forecasts of species range shifts under climate change. *Ecol. Lett.* 17, 1351–1364. doi: 10.1111/ele.12348
- Villar, E., Farrant, G. K., Follows, M., Garczarek, L., Speich, S., Audic, S., et al. (2015). Environmental characteristics of Agulhas rings affect interocean plankton transport. *Science* 348:1261447. doi: 10.1126/science.1261447
- Villarino, E., Chust, G., Licandro, P., Butenschön, M., Ibaibarriaga, L., Larrañaga, A., et al. (2015). Modelling the future biogeography of North Atlantic zooplankton communities in response to climate change. *Mar. Ecol. Prog. Ser.* 531, 121–142. doi: 10.3354/meps11299
- Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., et al. (2007). Let the concept of trait be functional! *Oikos* 116, 882–892. doi: 10.1111/j.0030-1299.2007.15559.x
- Waters, C. N., Zalasiewicz, J., Summerhayes, C., Barnosky, A. D., Poirier, C., Gałuszka, A., et al. (2016). The Anthropocene is functionally and stratigraphically distinct from the Holocene. *Science* 351, 6269. doi: 10.1126/science.aad2622
- Wis, M. S., Pottier, J., Kissling, W. D., Pellissier, L., Lenoir, J., Damgaard, C. F., et al. (2013). The role of biotic interactions in shaping distributions and realised assemblages of species: implications for species distribution modelling. *Biol. Rev.* 88, 15–30. doi: 10.1111/j.1469-185X.2012.00235.x
- Yebra, L., Bonnet, D., Harris, R. P., Lindeque, P. K., and Peijnenburg, K. T. C. A. (2011). Barriers in the pelagic: population structuring of *Calanus helgolandicus* and *C. euxinus* in European waters. *Mar. Ecol. Prog. Ser.* 428, 135–149. doi: 10.3354/meps09056
- Zurell, D., Thuiller, W., Pagel, J., Cabral, J.S., Münkemüller, T., Gravel, D., et al. (2016). Benchmarking novel approaches for modelling species range dynamics. *Glob. Change Biol.* 22, 2651–2664. doi: 10.1111/gcb.13251

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Chust, Vogt, Benedetti, Nakov, Villéger, Aubert, Vallina, Righetti, Not, Biard, Bittner, Benoiston, Guidi, Villarino, Gaborit, Cornils, Buttay, Irisson, Chiarello, Vallim, Blanco-Bercial, Basconi and Ayata. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.