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1	Quantitative Spectroscopic Analysis of Heterogeneous
2	Mixtures: the Correction of Multiplicative Effects
3	Caused by Variations in Physical Properties of Samples
4	
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18 **ABSTRACT**: Spectral measurements of complex heterogeneous types of mixture samples are often 19 affected by significant multiplicative effects resulting from light scattering, due to physical variations 20 (e.g. particle size and shape, sample packing and sample surface, etc.) inherent within the individual 21 samples. Therefore, the separation of the spectral contributions due to variations in chemical 22 compositions from those caused by physical variations is crucial to accurate quantitative spectroscopic 23 analysis of heterogeneous samples. In this work, an improved strategy has been proposed to estimate the 24 multiplicative parameters accounting for multiplicative effects in each measured spectrum, and hence 25 mitigate the detrimental influence of multiplicative effects on the quantitative spectroscopic analysis of 26 heterogeneous samples. The basic assumption of the proposed method is that light scattering due to 27 physical variations has the same effects on the spectral contributions of each of the spectroscopically 28 active chemical component in the same sample mixture. Based on this underlying assumption, the 29 proposed method realizes the efficient estimation of the multiplicative parameters by solving a simple 30 quadratic programming problem. The performance of the proposed method has been tested on two 31 publicly available benchmark data sets (i.e. near-infrared total diffuse transmittance spectra of 32 four-component suspension samples and near infrared spectral data of meat samples) and compared 33 with some empirical approaches designed for the same purpose. It was found that the proposed method 34 provided appreciable improvement in quantitative spectroscopic analysis of heterogeneous mixture 35 samples. The study indicates that accurate quantitative spectroscopic analysis of heterogeneous mixture 36 samples can be achieved through the combination of spectroscopic techniques with smart modeling 37 methodology.

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Keywords: Heterogeneous mixture samples, Multiplicative light scattering effects, Modified optical
path-length estimation and correction, Dual calibration strategy, Spectroscopic quantitative analysis

43 **1. Introduction**

44 The quantitative analysis of heterogeneous mixture samples using conventional instruments such as 45 HPLC generally involves troublesome and time-consuming sample preparations. Due to their high measuring speed, multiplicity of analysis, non-destructivity, flexibility and especially requirement of 46 less or even no sample preparations, spectroscopic technologies such as near infrared (NIR), mid 47 infrared (MIR) and Fourier-transform Raman spectroscopy (FT-Raman) have been increasingly applied 48 49 to the analysis of complex systems in areas of chemicals, food processing, agriculture and pharmaceuticals, etc¹⁻⁶. However, when analyzing complex heterogeneous mixture samples that exhibit 50 spectroscopic instrumentation, the 51 sample-to-sample variability in physical properties using 52 multiplicative light scattering effects caused by the uncontrolled variations in optical path length due to 53 the physical differences between samples (e.g. particle size and shape, sample packing, and sample surface, etc) would 'scale' the entire spectral measurement and hence mask the spectral variations 54 relating to the content differences of chemical compounds in the samples ⁷. The presence of dominant 55 multiplicative effects in spectral data could invalidate the underlying assumption of commonly used 56 multivariate linear calibration methods such as PCR⁸ and PLS⁹ which postulates a linear relationship 57 58 between spectral measurements and the contents of chemical components, and hence significantly 59 deteriorate the predictive performance of calibration models built by multivariate linear calibration 60 methods. The separation of the spectral contributions due to variations in chemical compositions from 61 those caused by multiplicative effects is therefore crucial to the accurate quantitative analysis of messy spectral data with multiplicative effects. 62

A number of chemometric pre-processing methods, e.g., Multiplicative Signal Correction (MSC)⁷. 63 Standard Normal Variate (SNV)¹⁰, Inverted Signal Correction (ISC)¹¹, Extended Inverted Signal 64 Correction (EISC)¹², Extended MSC (EMSC)¹³ and Modified EMSC¹⁴ have been proposed to remove 65 66 the multiplicative effects caused by variations in physical properties of samples. However MSC, ISC and EISC could only be applied to a spectrum that has wavelength regions containing no chemical 67 information, i.e. influenced only by the multiplicative effects. Otherwise, they could result in 68 69 dramatically poor results. The applicability of EMSC and the modified EMSC is limited due to the 70 requirement of the pure spectra for all spectroscopically active chemical components present in the 71 samples which is difficult to satisfy in practice.

72 Recently, Thennadil et al. proposed an interesting approach for the correction of multiple light scattering effects by making use of radiative transfer theory ¹⁵⁻¹⁶. Though this approach can to some 73 74 extent improve the predictive performance of multivariate calibration models, its implementation 75 complexity and the requirement of three measurements for each mixture sample (i.e. total diffuse 76 transmittance, total diffuse reflectance and collimated transmittance) make it difficult to use in practice. More recently in a review of pharmaceutical applications of separation of absorption and scattering in 77 near-infrared spectroscopy, similar concepts to the approach mentioned above are discussed ¹⁷. Another 78 79 similar approach to compensate for the scattering effects in reflectance spectroscopy was developed by Kessler et al. by integrating Kubelka–Munk equation with multivariate curve resolution (MCR)¹⁸. Like 80 81 the method based on radiative transfer theory, the application of hard model constrained MCR-ALS 82 algorithm is dependent on the availability of two measurements for each mixture sample (i.e. the diffuse reflectance spectra of a sample with an optically infinite thickness and a sample of finite thickness).
Hence the scope of its applicability is also limited.

85 To overcome these limitations, one of the present authors developed a novel multiplicative effect correction approach, Optical Path-Length Estimation and Correction (OPLEC) ^{19, 20}. OPLEC adopted 86 87 the following two-step procedure for the correction of multiplicative effects in spectral measurements. 88 First of all, the multiplicative parameters accounting for multiplicative effects in the spectral 89 measurements of the calibration samples are estimated by a unique method deduced solely from the 90 linear transformation of the calibration spectral measurements. And then the multiplicative effects in the 91 spectral measurements of the test samples are efficiently removed by a dual-calibration strategy. 92 Without placing any requirement on the spectral measurements, OPLEC can efficiently separate the 93 multiplicative effects of samples' physical properties from the spectral variations related to the chemical 94 compositions, and hence has much wider applicability than other methods reported in the literature. The 95 development of OPLEC provided an important contribution to the solution of multiplicative light 96 scattering issues. Whereas the first step of OPLEC, i.e. the estimation of the multiplicative parameters for the calibration samples, involves the determination of the number of spectroscopically active 97 98 chemical components in the systems under study. A poor estimation of the number of chemical 99 components would result in suboptimal performance of OPLEC. For complex systems, the estimation of 100 the number of chemical components is not a trivial task. Therefore, the OPLEC method needs to be 101 refined to realize its full potential for spectroscopic quantitative analysis of heterogeneous mixtures.

102 The objectives of this study were (1) to redesign the method in OPLEC for the estimation of the 103 multiplicative parameters for the spectral measurements of the calibration samples, (2) to develop a simple but effective approach for determining the optimal model parameter (i.e. the number of spectroscopically active chemical components) in OPLEC, (3) to improve the robustness of OPLEC when being applied to complex systems, and finally (4) to evaluate the performance of the modified OPLEC method on two publicly available benchmark data sets.

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110 **2. Theory**

111 2.1 The dual calibration strategy adopted by OPLEC to correct multiplicative effects

For spectral measurements with multiplicative effects caused by changes in the optical path-length due to the physical variations of the samples, the measured spectrum (\mathbf{x}_i , row vector) of sample *i* composed of *J* chemical components can be approximated by the following model ^{6, 7, 21}:

$$\mathbf{x}_{i} = p_{i} \sum_{j=1}^{J} c_{i,j} \mathbf{s}_{j}, \quad i = 1, 2, ..., I$$

115 Where $c_{i,j}$ is the concentration of the *j*-th chemical component in the *i*-th mixture sample; \mathbf{s}_j represents 116 the pure spectrum of *j*-th chemical component in the mixtures. The coefficient p_i accounts for the 117 multiplicative effects in the spectral measurements of the *i*-th sample caused by changes in the optical 118 path-length due to the physical variations of the sample; *I* denotes the number of calibration samples. 119 Assume the first component is the target constituent in the mixtures and $\sum_{j=1}^{J} c_{i,j} \mathbf{s}_j = 1$ (which strictly 120 hold for $c_{i,j}$ representing unit-free concentration such as weight fraction and mole fraction), then eq.1 121 can also be expressed as:

$$\mathbf{x}_{i} = p_{i}c_{i,1}\Delta\mathbf{s}_{1} + p_{i}\mathbf{s}_{2} + \sum_{j=3}^{J} p_{i}c_{i,j}\Delta\mathbf{s}_{j}, \quad \Delta\mathbf{s}_{j} = \mathbf{s}_{j} - \mathbf{s}_{2}$$
(2)

It is obvious that a linear relationship exists between \mathbf{x}_i and p_i , and also between \mathbf{x}_i and $p_i c_{i,1}$. It should 122 123 be noted that this conclusion would also hold when the content of one constituent (or matrix substances) does not vary over mixture samples. Provided the multiplicative parameter vector \mathbf{p} ($\mathbf{p} = [p_1; p_2; ...; p_I]$) 124 125 for the calibration samples is available (actually it can be estimated from the calibration spectra by the 126 multiplicative parameter estimation method outlined in section 2.2), two following calibration models 127 can therefore be built by multivariate linear calibration methods such as PLS. The first model is between **X** (**X** = $[x_1; x_2; ...; x_n]$) and **p**, and the other is 128 between Х and $diag(c_1)p$ $(diag(\mathbf{c}_1)\mathbf{p} = [p_1 \times c_{11}; p_2 \times c_{21}; \dots; p_I \times c_{I1}])$. For simplicity, the same number of latent components is 129 130 generally used in the above two PLS calibration models. Once the spectrum of a test sample has been 131 recorded, the content of the target constituent in the test sample can then be obtained by dividing the 132 prediction of the second calibration model by the corresponding prediction of the first calibration model.

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134 2.2 Multiplicative parameter estimation

Obviously, the estimation of the multiplicative parameter vector **p** for the calibration samples is the key to the correction of the multiplicative effects by the above dual calibration strategy. The performance of the multiplicative parameter estimation method in the original OPLEC method ¹⁹ relies on the accurate estimation of the number of spectroscopically active chemical components in the systems under study. Poor estimation of the number of chemical components could significantly affect the performance of OPLEC. With a view to improve the robustness of OPLEC, the following refined method for the
estimation of multiplicative parameter vector **p** for the calibration samples was proposed in this work.

142 Suppose the singular value decomposition of **X** ($\mathbf{X} = [\mathbf{x}_1; \mathbf{x}_2; ...; \mathbf{x}_I]$) can be expressed as follows:

$$\mathbf{X} = [\mathbf{U}_{s}, \mathbf{U}_{n}] \begin{bmatrix} \sum_{s} & 0\\ 0 & \sum_{n} \end{bmatrix} [\mathbf{V}_{s}, \mathbf{V}_{n}]^{\mathrm{T}} = \mathbf{U}_{s} \sum_{s} \mathbf{V}_{s}^{\mathrm{T}} + \mathbf{E}$$
((

143 Where, $\mathbf{E} = \mathbf{U}_n \sum_n \mathbf{V}_n^{\mathrm{T}}$; superscript 'T' denotes the transpose; subscripts 's' and 'n' signify that the 144 corresponding factors represent spectral information and noise, respectively. Suppose the actual number 145 of spectroscopically active chemical components in the system studied is *r*, then both \mathbf{U}_s and \mathbf{V}_s consist 146 of *r* columns. According to eq.2, both vectors **p** and diag(\mathbf{c}_1)**p** are in the column space of \mathbf{U}_s , so the 147 following equations hold:

$$\mathbf{U}_{s}\mathbf{U}_{s}^{\mathrm{T}}\mathbf{p}=\mathbf{p}$$

$$\mathbf{U}_{s}\mathbf{U}_{s}^{\mathrm{T}}diag(\mathbf{c}_{1})\mathbf{p} = diag(\mathbf{c}_{1})\mathbf{p}$$
(5)

)

(4

Since there is no requirement to know the absolute value of p_i , p_i can be assumed to be no less than unity ($\mathbf{p} \ge \mathbf{1}$). Therefore, the vector \mathbf{p} satisfying equations 4 and 5 can be obtained by solving the following constrained optimization problem:

$$\min_{\mathbf{p}} \frac{1}{2} \left(\left\| \mathbf{U}_{s} \mathbf{U}_{s}^{\mathrm{T}} \mathbf{p} - \mathbf{p} \right\|_{2}^{2} + \frac{1}{w^{2}} \left\| \mathbf{U}_{s} \mathbf{U}_{s}^{\mathrm{T}} diag(\mathbf{c}_{1}) \mathbf{p} - diag(\mathbf{c}_{1}) \mathbf{p} \right\|_{2}^{2} \right), \text{ subject to } \mathbf{p} \ge 1$$
(6)

151 Where, $\|\|_2$ denotes l^2 norm; *w* is a weight to balance the two parts in the above optimization function. 152 It can be simply set to be the maximum element of \mathbf{c}_1 . The above constrained optimization problem can be transformed into an equivalent quadratic programming problem (which can be resolved by the *quadprog* function in MATLAB. The MATLAB code for the multiplicative parameter estimation method is available in Supporting Information):

$$\min_{\mathbf{p}} f(\mathbf{p}) = \frac{1}{2} \mathbf{p}^{\mathrm{T}} ((\mathbf{I} - \mathbf{U}_{s} \mathbf{U}_{s}^{\mathrm{T}}) + diag(\mathbf{c}_{1} / w)(\mathbf{I} - \mathbf{U}_{s} \mathbf{U}_{s}^{\mathrm{T}}) diag(\mathbf{c}_{1} / w))\mathbf{p}, \text{ such that } -\mathbf{p} \leq -1$$
(7)

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157 2.3 Determination of the number of columns in U_s

158 Theoretically, the number of columns in U_s (i.e. parameter r) should equal to the number of 159 spectroscopically active chemical components in the systems under study. It is generally difficult to 160 determine the exact number of spectroscopically active chemical components in a complex system. 161 Moreover, when the spectral data does not strictly obey the model in eq. 1, the optimal number of 162 columns in U_s might not solely depend on the number of spectroscopically active chemical components 163 in the system under study, which would further complicate the situation. Fortunately, a simple 164 mathematical analysis reveals that min $f(\mathbf{p})$ decreases dramatically with the increase of r at the very 165 start, and then tends to be steady when r exceeds certain threshold value. Therefore, the optimal value of 166 r can be determined by locating the turning point in the plot of min $f(\mathbf{p})$ versus r.

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169 **3. Case studies**

The effectiveness of the modified OPLEC method (hereafter referred to $OPLEC_m$) with respect to its ability to estimate multiplicative parameters was first tested on the near-infrared total diffuse transmittance spectra of four-component suspension system consisting of water, deuterium, ethanol, and polystyrene (hereafter referred to four-component suspension data). To further explore the potential of
OPLEC_m, another real-world near-infrared transmittance spectra of meat samples recorded on a Tecator
Infratec Food and Feed Analyzer (hereafter referred to tecator data) is employed. This spectral data set
is publicly available and hence ensures that the interested reader can repeat the analysis.

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178 *3.1 Four-component suspension data*¹⁶

179 The four-component suspension system is composed of three fully miscible absorbing species of water, 180 deuterium oxide and ethanol and a species that both absorbs and scatters light (i.e., a particulate species 181 of polystyrene). Specifically, the range of particle size and concentration were chosen to be $100 \sim 500$ nm 182 and 1~5 wt%, respectively, such that the following conditions were satisfied: stable suspension, multiple 183 scattering, and sufficient signals in measurement. A total of 42 samples were prepared using various 184 combinations of the concentrations of the four components and particle sizes of which the total diffuse 185 transmittance (T_d) spectra were recorded on a scanning spectrophotometer (CARY 5000) fitted with a 186 diffuse reflectance accessory (DRA-2500). The spectral data were collected in the wavelength region of 187 1500-1880 nm with an interval of 2nm, resulting in measurements at 191 discrete wavelengths per 188 spectrum. Twenty-two suspension samples' spectra were randomly selected to construct the calibration 189 data set. The remaining twenty spectra from the other suspension samples made up the test data set. The 190 absorbing-only species of deuterium oxide with concentration range between 20% and 58 wt% was 191 taken as the analyte of interest in the present analysis and all the total diffuse transmittance spectra were 192 transformed into absorbance spectra prior to the analysis. More experimental details can be found in the 193 original paper of Steponavicius and Thennadil¹⁶.

195 *3.2 Tecator data*²²

196 This benchmark spectral data set consists of the near-infrared absorbance spectra of 240 meat samples 197 recorded on a Tecator Infratec Food and Feed Analyzer working in the wavelength range 850-1050 nm 198 with an interval of 2nm by the Near Infrared Transmission principle. Each sample contains finely 199 chopped pure meat with different moisture, fat and protein contents. A Soxhlet method was used as the 200 laboratory reference for fat determination. The Soxhlet values ranged from 2% to 59% fat. The 240 201 spectra were divided into 5 data sets for the purpose of model validation and extrapolation studies 202 (calibration set: 129; validation set: 43; test set: 43; extrapolation set for fat: 8; extrapolation set for 203 protein: 7). The task in the present work is restricted to predict the fat content (%) of a meat sample on 204 the basis of its near infrared absorbance spectrum, the extrapolation set for protein is therefore excluded. 205 The tecator data is available at http://lib.stat.cmu.edu/datasets/tecator.

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207 *3.3 Data pre-treatment*

For the aforementioned two data sets, the possible additive baseline effects and wavelength dependent spectral variations were firstly removed by projecting the measured spectra onto the orthogonal complement of the space spanned by the row vectors of $\mathbf{M} = [\mathbf{1}; \boldsymbol{\lambda}; \boldsymbol{\lambda}^2]^{19}$. The pre-processed spectra were then used to calculate the multiplicative parameter vector \mathbf{p} for the calibration samples. The dual calibration models in OPLEC_m were built on the pre-processed spectra by using PLS method. The predictive performance of OPLEC_m was compared with those of PLS calibration models with and without the application of data preprocessing methods such as MSC, SNV, EISC and EMSC as long as 215 they are applicable. The root-mean-square error of prediction (RMSEP) was used to assess the 216 performance of the calibration models.

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219 **4. Results and discussion**

220 4.1 Four-component suspension data

221 The raw total transmittance spectra of the four-component suspension samples are presented in Figure 1. 222 It can be observed that the variations in polystyrene particle size and concentration across samples 223 resulted in significant additive baseline shift as well as multiplicative effects in the spectral data. 224 Though the additive baseline effects and possible wavelength dependent spectral variations can be 225 readily removed by orthogonal projection pre-processing, the multiplicative effects as a consequence of 226 the changes in sample's effective optical path-length are rather difficult to correct. Such multiplicative 227 effects can not be effectively modeled by multivariate linear calibration models either. Without being properly corrected or modeled, they can significantly deteriorate the predictive performance of 228 multivariate linear calibration models ^{13, 19}. 229



Wavelength (nm)Figure 1: The raw spectra of the four component suspension system.

232 As stated in the theory section, OPLEC_m can effectively correct the multiplicative effects in spectral measurements. OPLEC_m consists of two main steps. The first step is to estimate the multiplicative 233 234 parameter vector **p** for the calibration samples from the orthogonal projection pre-processed spectra. 235 The estimation of the multiplicative parameter vector \mathbf{p} for the calibration samples requires the 236 determination of the actual number of spectral variation sources (r) in the calibration spectra, which can be achieved by scrutinizing the plot of $\min_{\mathbf{p}} f(\mathbf{p})$ versus r (Figure 2). From Figure 2, it can be seen that 237 min $f(\mathbf{p})$ decreases obviously when the number of columns of U_s increases from one to three and 238 including more components in U_s leads to no significant changes in $\min_{\mathbf{p}} f(\mathbf{p})$, which means the most 239 240 spectral information relevant to \mathbf{p} and diag(\mathbf{c}_1) \mathbf{p} was included in the first three principal components of 241 U_s . Therefore, the optimal value of *r* was then set to three.

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Figure 2: The relationship between $\min_{\mathbf{p}} f(\mathbf{p})$ and the number of columns of \mathbf{U}_{s} (i.e. *r*) for the four component suspension data.

247 After the estimation of the multiplicative parameter vector \mathbf{p} for the calibration samples, one can 248 assess the applicability of OPLEC_m to the spectral data set by examining the two plots of **p** vs $\mathbf{U}_{s}\mathbf{U}_{s}^{T}\mathbf{p}$ 249 and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^T diag(\mathbf{c}_1)\mathbf{p}$, respectively (supporting information, Figure S-1). As shown in 250 Figure S-1, both **p** and $diag(\mathbf{c}_1)\mathbf{p}$ are in good agreement with $\mathbf{U}_{\mathbf{c}}\mathbf{U}_{\mathbf{c}}^T\mathbf{p}$ and $\mathbf{U}_{\mathbf{c}}\mathbf{U}_{\mathbf{c}}^Tdiag(\mathbf{c}_1)\mathbf{p}$, respectively, which confirms that a linear relationship exists between \mathbf{x}_i and p_i , and also between \mathbf{x}_i and 251 252 $p_i c_{i1}$. The dual calibration strategy of OPLEC_m is therefore applicable to the four component 253 suspension data. Figure S-1 also reveals the presence of significant variations of multiplicative effects 254 $(p_i \text{ varying from 1 to 3.09})$ in the calibration samples. Multiplicative effect correction methods such as OPLEC_m are therefore needed to remove such significant multiplicative effects in the spectral 255 256 measurements.

257 Figure 3a compared the predictive performance of the optimal OPLEC_m calibration model for 258 deuterium oxide and the corresponding optimal PLS models with and without the application of 259 preprocessing methods (e.g. SNV, MSC, EISC and EMSC). Obviously, as a result of the presence of 260 severe multiplicative effects, PLS calibration model built on the raw calibration spectra could not give 261 satisfactory predictions for the deuterium oxide in the test suspension samples. Preprocessing the 262 calibration spectra by MSC, SNV or EISC can, to some extent, improve the predictive performance of 263 PLS calibration models in terms of RMSEP values. However, due to the lack of a wavelength region 264 containing no chemical information in the spectral data, the multiplicative effects can not be fully 265 corrected by MSC, SNV or EISC. Hence, the predictive errors of the PLS calibration models built on 266 the calibration spectra pre-processed by MSC, SNV and EISC are still comparatively high. As expected, 267 OPLEC_m offers the best improvement in terms of the predictive ability among all the pre-processed

268	methods. The OPLEC _m calibration model with five underlying components provided the best predictive
269	results with a $RMSEP_{test}$ value as low as 0.005, while the corresponding best $RMSEP_{test}$ value of the
270	PLS calibration model with nine underlying components on the calibration spectra pre-processed by
271	EISC is 0.009. Furthermore, the performance of the OPLEC _m is robust to the number of columns in U_s
272	(Figure 3b). Considering the fact that $OPLEC_m$ does not place any extra requirement on the spectral
273	measurements as other multiplicative effect correction methods do, such a result is quite encouraging.
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Figure 3: a) The predictive performance of $OPLEC_m$ and the PLS models built on the calibration spectra of the four component suspension system preprocessed by different methods (black circle: the raw spectra; red star: MSC; green triangle down: SNV; pink diamond: EISC; blue square: $OPLEC_m$); b) The predictive performance of the optimal $OPLEC_m$ models when U_s with different number of columns (*r*) were used in the calculation of the multiplicative parameter vector **p** for the calibration spectra.

282 4.2 Tecator data

283 As in four component suspension data, there are significant additive baseline effects in the tecator data 284 (supporting information, Figure S-2). Since the changes in physical properties of samples generally 285 result in both additive baseline effects and multiplicative effects, the presence of significant additive baseline effects strongly suggests the existence of multiplicative effects. OPLEC_m was therefore used to 286 287 estimate the multiplicative parameter vector **p** for the calibration samples from the corresponding 288 orthogonal projection pre-processed calibration spectra as described in section 3.3. During the 289 estimation of the multiplicative parameter vector \mathbf{p} for the calibration samples using OPLEC_m, the optimal number of columns included in U_s (i.e. r) is determined by scrutinizing the plot of min $f(\mathbf{p})$ 290 versus r (Figure 4). It can be seen that $\min_{\mathbf{p}} f(\mathbf{p})$ drops sharply as the r increases from one to six, and 291 292 then decreases slowly along with the further increase of r (Figure 4). One can therefore choose six as the 293 optimal number of columns of U_s .



Figure 4: The plot of $\min_{\mathbf{p}} f(\mathbf{p})$ versus the number of columns in \mathbf{U}_s (i.e. r).

It is worth to point out again that the performance of $OPLEC_m$ is quite robust to the choice of r as long as r is big enough but not too large. As shown in Figure 5, The RMSEP value of $OPLEC_m$ for the test samples shows no significant difference when r taking a value between 6 and 11. In practice, such a feature of $OPLEC_m$ can make it more user-friendly when being applied to complex systems.



Figure 5: The RMSEP values for the test samples in the tecator data obtained by the optimal OPLEC_m calibration models when U_s with different number of columns (i.e. *r*) were used in the calculation of the multiplicative parameter vector **p** for the calibration spectra.

305 After the estimation of the multiplicative parameter vector \mathbf{p} for the calibration samples, the dual 306 calibration strategy of OPLEC_m was adopted to mitigate the detrimental of multiplicative effects on the 307 prediction of the fat content. PLS calibration models with and without the application of MSC, SNV and 308 EISC were also established for comparison purposes. The optimal number of underlying components 309 used in the dual calibration models of OPLEC as well as those PLS calibration models was chosen to be 310 the one with minimal root-mean-square error of prediction (RMSEP) for the validation set. The results of OPLEC_m along with those of the four optimal PLS calibration models with and without the 311 312 application of MSC, SNV and EISC were shown in Figure 6.





314 Figure 6: The RMSEP values for the tecator data obtained by different calibration methods.

315 Figure 6 reveals that although the number of latent components (i.e. fourteen) used is sufficiently 316 large, the optimal PLS calibration model on the raw calibration spectra did not give satisfactory 317 predictions for all the four data sets. The RMSEP values for the calibration, validation, test and extrapolation sets are 1.7%, 2.7%, 2.3% and 8.5%, respectively. The application of the empirical 318 319 multiplicative light scattering correction method, SNV saw no significant changes in the RMSEP values 320 for the four data sets. While preprocessing the spectral data by MSC resulted in a dramatic increase in 321 the RMSEP value for the extrapolation set which clearly demonstrates its limitation in practical 322 applications. The EISC preprocessing method surprisingly succeeded in improving the quality of the 323 predictions of PLS calibration model for the tecator data. Its RMSEP values for the calibration, 324 validation, test and extrapolation sets are 0.7%, 0.9%, 1.0% and 3.3%, respectively. The reasons of its 325 success in this particular data set are unclear. As expected, OPLEC_m outperformed all the other methods 326 with RMSEP values for the calibration, validation, test and extrapolation sets equaling to 0.4%, 0.5%, 0.4% and 1.0%, respectively, This remarkable improvement further confirmed the effectiveness of 327 328 OPLEC_m in mitigating the detrimental influence of multiplicative effects on the spectroscopic 329 quantitative analysis of heterogeneous mixture samples.

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332 5. Conclusion

333 The separation of the spectral contributions due to variations in chemical compositions from 334 multiplicative effects caused by physical variations is crucial to the accurate quantitative analysis of 335 complex heterogeneous mixture samples using spectroscopic instruments. In this work, a modified 336 version of Optical Path-Length Correction and Estimation (OPLEC_m) method has been developed to 337 correct the multiplicative effects in spectral measurements. OPLEC_m differs from the original OPLEC 338 method in the way of estimating the multiplicative parameters for the calibration samples. In OPLEC_m, 339 the multiplicative parameters for the calibration samples were obtained by solving a constrained 340 quadratic programming problem, which is much more efficient than the counterpart in the original 341 OPLEC. Furthermore, a simple but effective method has been proposed for the determination of the 342 model parameter involved (i.e. the number of spectroscopically active chemical components in the 343 system under study). Due to the unique multiplicative parameter estimation strategy, the performance of OPLEC_m is much more robust to the choice of the model parameter involved, which makes OPLEC_m 344 more user-friendly when being applied to complex systems. The performance of OPLEC_m has been 345 346 tested on four-component suspension spectral data set and one publicly available benchmark spectral 347 data set. Experimental results reveal that OPLEC_m can achieve satisfactory quantitative results from the 348 spectroscopic measurements of heterogeneous mixtures. Compared with other existing methods 349 designed for the same purpose, OPLEC_m has features of implementation simplicity, wider applicability 350 as well as better performance in terms of quantitative accuracy, and therefore has great potential in 351 quantitative spectroscopic analysis of complex heterogeneous systems.

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304	Supporting Information Available
365	Supporting Information Available MATLAB code for the modified OPLEC, the plots of \mathbf{p} vs $\mathbf{U}_s \mathbf{U}_s^T \mathbf{p}$ and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^T diag(\mathbf{c}_1)\mathbf{p}$
365366	Supporting Information Available MATLAB code for the modified OPLEC, the plots of \mathbf{p} vs $\mathbf{U}_s \mathbf{U}_s^T \mathbf{p}$ and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^T diag(\mathbf{c}_1)\mathbf{p}$ for the four component suspension data, the 129 raw calibration spectra of the tecator data. This material
364365366367	Supporting Information Available MATLAB code for the modified OPLEC, the plots of \mathbf{p} vs $\mathbf{U}_s \mathbf{U}_s^T \mathbf{p}$ and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^T diag(\mathbf{c}_1)\mathbf{p}$ for the four component suspension data, the 129 raw calibration spectra of the tecator data. This material is available free of charge via the Internet at http://pubs.acs.org.

References :

371	(1)	H.W. Siesler, Y. Ozaki, S. Kawata, H.M. Heise, Near-infrared spectroscopy: principal,
372		instruments, applications, WILEY-VCH, Weinheim, 2002
373	(2)	P. Fayolle, D. Picque, G. Corrieu, Monitoring of fermentation processes producing lactic acid
374		bacteria by mid-infrared spectroscopy, Vib. Spectrosc. 1997, 14, 247-252
375	(3)	Y. Roggo, C. Roeseler, M. Ulmschneider, Near infrared spectroscopy for qualitative comparison
376		of pharmaceutical batches, J. Pharm. Biomed. Anal. 2004, 36, 777-786
377	(4)	A. Nordon, D. Littlejohn, A.S. Dann, P.A. Jeffkins, M.D. Richardson, S.L. Stimpson, In situ
378		monitoring of a seed stage of a fermentation process using non-invasive NIR spectrometry, The
379		Analyst, 2008, 133, 660-666
380	(5)	Z.P. Chen, G. Fevotte, A. Caillet, D. Littlejohn, J. Morris, An advanced calibration strategy for
381		in-situ quantitative monitoring of phase transition processes in suspensions using FT-Raman
382		spectroscopy, Anal. Chem. 2008, 80, 6658-6665
383	(6)	Z.P. Chen, J. Morris, A. Borissova, S. Khan, T. Mahmud, R. Penchev, K.J. Roberts, On-line
384		monitoring of batch cooling crystallization of organic compounds using ATR-FTIR spectroscopy
385		coupled with an advanced calibration method, Chemom. Intell. Lab. Syst. 2009, 96, 49-58
386	(7)	P. Geladi, D. MacDougall, H. Martens, Linearization and Scatter-Correction for Near-Infrared
387		Reflectance Spectra of Meat, Appl. Spectrosc. 1985, 39 (3), 491-500

- 388 (8) I.A. Cowe, J.W. McNicol, The Use of Principal Components in the Analysis of Near-Infrared
 389 Spectra, *Appl. Spectrosc.* 1985, *39 (2)*, 257-266
- 390 (9) H. Martens, M. Martens, Multivariate Analysis of Quality: An Introduction, John Wiley and Sons:
 391 Chichester, 2001
- 392 (10) R.J. Barnes, M.S. Dhanoa, S.J. Lister, Standard Normal Variate Transformation and De-trending
 393 of Near-Infrared Diffuse Reflectance Spectra, *Appl. Spectrosc.* 1989, *43 (5)*, 772-777
- 394 (11) I.S. Helland, T. Næs, T. Isaksson, Related versions of the multiplicative scatter correction method
- for preprocessing spectroscopic data, *Chemom. Intell. Lab. Syst.* **1995**, *29 (2)*, 233-241
- 396 (12) D. Pedersen, H. Martens, J. Nielsen, S. Engelsen, Near-infrared absorption and scattering
 397 separated by extended inverted signal correction (EISC): Analysis of near-infrared transmittance
 398 spectra of single wheat seeds, *Appl. Spectrosc.* 2002, *56 (9)*, 1206-1214
- 399 (13) H. Martens, J.P. Nielsen, S.B. Engelsen, Light Scattering and Light Absorbance Separated by
- 400 Extended Multiplicative Signal Correction. Application to Near-Infrared Transmission Analysis of
- 401 Powder Mixtures, Anal. Chem. 2003, 75 (3), 394-404
- 402 (14) S.N. Thennadil, H. Martens, A. Kohler, Physics-based multiplicative scatter correction approaches
- 403 for improving the performance of calibration models, *Appl. Spectrosc.* **2006**, *60*, 315-321
- 404 (15) R. Steponavicius, S.N. Thennadil, Extraction of chemical information of suspensions using
 405 radiative transfer theory to remove multiple scattering effects: application to a model
 406 two-component system, *Anal. Chem.* 2009, *81*, 7713–7723

408		Radiative transfer theory to remove multiple scattering effects: application to a model
409		multicomponent system, Anal. Chem. 2011, 83, 1931-1937
410	(17)	Z. Shi, C. Andersen, Pharmaceutical applications of separation of absorption and scattering in
411		near-infrared spectroscopy (NIRS), J. Pharm. Sci. 2010, 99, 4766-4783
412	(18)	W. Kessler, D. Oelkrug, R. Kessler, Using scattering and absorption spectra as MCR-hard model
413		constraints for diffuse reflectance measurements of tablets, Anal. Chim. Acta, 2009, 642, 127-134
414	(19)	Z.P.Chen, J. Morris, E. Martin, Extracting Chemical Information from Spectral Data with
415		Multiplicative Light Scattering Effects by Optical Path-Length Estimation and Correction, Anal.
416		Chem. 2006, 78(9), 7674-7681
417	(20)	Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil,
417 418	(20)	Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil,J. Faulkner, J. Morris, Calibration of Multiplexed Fiber-Optic Spectroscopy, <i>Anal. Chem.</i> 2011,
417 418 419	(20)	 Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil, J. Faulkner, J. Morris, Calibration of Multiplexed Fiber-Optic Spectroscopy, <i>Anal. Chem.</i> 2011, 83(7), 2655-2659
417418419420	(20)	 Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil, J. Faulkner, J. Morris, Calibration of Multiplexed Fiber-Optic Spectroscopy, <i>Anal. Chem.</i> 2011, 83(7), 2655-2659 Z.P. Chen, J. Morris, Improving the linearity of spectroscopic data subjected to fluctuations in
 417 418 419 420 421 	(20)	 Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil, J. Faulkner, J. Morris, Calibration of Multiplexed Fiber-Optic Spectroscopy, <i>Anal. Chem.</i> 2011, 83(7), 2655-2659 Z.P. Chen, J. Morris, Improving the linearity of spectroscopic data subjected to fluctuations in external variables by the extended loading space standardization, <i>The Analyst</i>, 2008, 133, 914-922
 417 418 419 420 421 422 	(20) (21) (22)	 Z.P.Chen, L.J. Zhong, A. Nordon, D. Littlejohn, M. Holden, M. Fazenda, L. Harvey, B. McNeil, J. Faulkner, J. Morris, Calibration of Multiplexed Fiber-Optic Spectroscopy, <i>Anal. Chem.</i> 2011, 83(7), 2655-2659 Z.P. Chen, J. Morris, Improving the linearity of spectroscopic data subjected to fluctuations in external variables by the extended loading space standardization, <i>The Analyst</i>, 2008, <i>133</i>, 914-922 C. Borggaard, H.H. Thodberg, Optimal minimal neural interpretation of spectra, <i>Anal. Chem.</i>

(16) R. Steponavicius, S.N. Thennadil, Extraction of chemical information of suspensions using

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1	Supporting Information		
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3	Title of the primary article:		
4	Quantitative Spectroscopic Analysis of Heterogeneous Mixtures: the Correction of Multiplicative		
5	Effects Caused by Variations in Physical Properties of Samples		
6	Authors' names:		
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16	Table of content:		
17	1) The MATLAB code for the modified OPLEC method		
18	2) Figure S-1: The plots of \mathbf{p} vs $\mathbf{U}_s \mathbf{U}_s^{\mathrm{T}} \mathbf{p}$ (a) and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^{\mathrm{T}} diag(\mathbf{c}_1)\mathbf{p}$ (b) for the four		
19	component suspension data.		
20 21	3) Figure S-2: The 129 raw calibration spectra of the tecator data.		

22	The MATLAB code for the modified OPLEC method
23	% $[\mathbf{p}, \text{fval}] = \text{OPLEC}_{m}(\mathbf{X}, \mathbf{c}, CompNumb);$
24	% This is an m-file for the estimation of the multiplicative effect vector \mathbf{p} for calibration samples;
25	% X contains \mathbf{x}_i in its rows; \mathbf{x}_i (<i>i</i> =1,2,, <i>I</i>) are the spectra of <i>I</i> calibration samples.
26	% c is the concentration vector of the target chemical component in the calibration samples;
27	% <i>CompNumb</i> is the number of spectroscopically active chemical components in mixture samples;
28	% p is a vector containing the multiplicative scattering parameters for the calibration samples;
29	% fval is the value of objective function at p ;
30	
31	function [p , fval]=OPLECm(X , c , <i>CompNumb</i>);
32	$[\mathbf{U},\mathbf{S},\mathbf{V}]=$ svd $(\mathbf{X});$
33	Us= U(:,1: <i>CompNumb</i>);
34	<i>n</i> =length(c);
35	$w=\max(\mathbf{c});$
36	H1=eye(n, n)- Us* Us';
37	$\mathbf{H2}=\operatorname{diag}(\mathbf{c}./w)^{*}\mathbf{H1}^{*}\operatorname{diag}(\mathbf{c}./w);$

S2

- 38 **H=H1+H2**; % matrix **H** in min(0.5***p**'***H*****p**+**f*****p**);
- 39 f=zeros(n,1); % vector **f** in min(0.5***p**'***H*****p**+**f*****p**);
- 40 A=-eye(n,n); % matrix A in $A*p \le b;$
- 41 **b**=-ones(n,1); % vector **b** in **A*p<=b**;
- 42 **StartingVect**=ones(*n*,1);
- 43 options=optimset('quadprog');
- 44 options=optimset(options,'LargeScale','off','Display','off');
- 45 [**p**,fval]=quadprog(**H**,**f**,**A**,**b**,[],[],[],[],**StartingVect**,options);

46 % After obtaining the model parameter vector **p** for calibration samples, two calibration models are built 47 using the standard PLS toolbox. One is between the concentration vector (**c**) of the target chemical 48 component and the spectral data **X**; the other is between *diag*(**c**)**p** and **X**. The multiplicative effect on 49 the test sample can then be corrected through dividing the prediction of the second calibration model by 50 the prediction of the first calibration model.

51 1) Figure S-1: The plots of p vs $\mathbf{U}_s \mathbf{U}_s^T \mathbf{p}$ (a) and $diag(\mathbf{c}_1)\mathbf{p}$ vs $\mathbf{U}_s \mathbf{U}_s^T diag(\mathbf{c}_1)\mathbf{p}$ (b) for the four 52 component suspension data. The number of columns in \mathbf{U}_s is three.



55 2) Figure S-2: The 129 raw calibration spectra of the tecator data.

