Possibilities for Misinterpretation in ASV-Speciation Studies of Natural Waters*

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Mögliche Mißdeutungen bei ASV-Speciesuntersuchungen von natürlichen Wässern

Zusammenfassung. Die Probleme, die bei Speciesuntersuchungen von Spurenelementen mit Hilfe der Differentialpuls Anodic Stripping Voltammetrie auftreten, werden zusammenfassend dargestellt. Neben den bei der eigentlichen Bestimmung auftretenden Problemen, werden auch solche erwähnt, die mit der Problemenhme sowie der Lagerung und Vorbehandlung der Proben in Zusammenhang stehen.

Summary. A summary is given of possible pitfalls identified in the speciation studies of trace elements by Differential Pulse Anodic Stripping Voltammetry including sampling-, storageand sample treatment procedures.

Introduction

Speciation studies of trace elements are essential for understanding their bioavailability, adsorption/desorption characteristics and transport mechanisms (involving e.g. ionic forms, (in)organic complexes, colloids). The problems associated with the use of special techniques required for speciation studies at extremely low concentration levels, are more severe than when dealing with mere total concentration determinations. Some problems in speciation studies will be summarized here. They refer to sampling-, storage- and sample treatment procedures, in relation to analytical determination by Anodic Stripping Voltammetry (ASV).

Problems Associated with Sampling-, Storageand Sample Treatment Procedures

Sampling. Extreme precautions and special equipment are required for total trace element concentration determination in natural waters [1, 2]. In addition, distinction of different species may cause problems, e.g. due to the release of organic compounds [3], some of which may change speciation. Clean room or at least clean bench facilities are essential for minimizing or preferably preventing sample contamination [4]. *Filtration*. Filtration results in operationally defined 'dissolved' and 'particulate suspended' fractions. Colloids can play an important role in complexation processes. They are able to pass a filter (usually $0.45 \,\mu\text{m}$ Nuclepore or Millipore). However, they can be removed by ultrafiltration, following $0.45 \,\mu\text{m}$ pore size filtration. Thus, a better estimate is obtained of 'truly dissolved' (< $0.001 \,\mu\text{m}$) forms [5]. All steps in these procedures are likely sources of contamination, however.

Storage. During storage, existing equilibria may be affected. This risk is minimized by analysis of the sample as soon as possible after sampling. If necessary, samples should be stored at 4° C in the dark. This is less likely to change the sample irreversibly than deepfreezing [6].

Pretreatment of Samples. The aim of speciation studies is to gain insight into the qualitative and quantitative aspects related to the different species present in a sample. Addition of chemicals needed for analytical determinations will, in principle, change these aspects.

Thus, the sample should not be acidified, nor should buffer, supporting electrolyte (for ASV measurements), mercury ions (for MFE formation) or any other chemical be added.

DPASV Analysis in Speciation Studies

Problems are also met in the final step of the analytical procedure, i.e. the determination of total concentration or the distinction and separate analysis of different species e.g. by (Differential Pulse) Anodic Stripping Voltammetry. Some of them will be discussed in the following section.

Determination of low trace metal concentrations in natural samples requires a technique with extreme sensitivity, such as Differential Pulse Anodic Stripping Voltammetry (DPASV), which has the additional advantage that distinction of different species even at extremely low concentration levels in natural waters is possible for some trace elements. The HMDE has the advantage that the electrode surface is renewed for each measurement, but it suffers from relatively low sensitivity. Thin mercury film electrodes have strongly increased sensitivities. They can be applied with a preformed film or with an in-situ formed film. Although the latter type has a higher sensitivity, it cannot be used in speciation studies because Hg(II) ions added to the solution to be analysed, change existing equilibria. The preformed jet-stream electrode (JMFE) as described in [7] proves to be more sensitive than the RMFE (Table 1).

^{*} Dedicated to Prof. Dr. W. Fresenius on the occasion of his 70th birthday

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Table 1. Comparison of hanging mercury drop electrode (HMDE), rotating mercury film electrodes (RMFE, in-situ and preformed) and the jet-stream mercury film electrode (JMFE) with respect to copper peak potential (E_p in mV), peak width at half-height ($b_{1/2}$ in mV) and sensitivity in A mol⁻¹dm³ Cu(II). All deposition times were 180 s: the film electrodes (6 mm diameter) were used under optimum conditions

Electrode type	<i>E</i> _p (mV)	<i>b</i> _{1/2} (mV)	Sensitivity (A mol ⁻¹ dm ³) for Cu(II)
HMDE	-110	90	2
RMFE in-situ	-260	45	108
RMFE preformed	-270	70	57
JMFE preformed	-290	70	254

DPASV is often used in speciation studies for the distinction, at the natural pH, of so-called labile and nonlabile species and also for the determination of the complexation capacity (CC) for a specific element. This parameter indicates the amount of ionic metal that can be taken up into non-labile complexes by (organic) ligands present in the natural sample. It can be determined by titration of the ligands with ionic copper, and careful analysis of the data [5, 8, 9].

Several problems can be identified in these procedures.

Adsorption to the Cell. In the absence of possible ligands which are able to form non-labile complexes, the relation between ASV response and concentration of metal added to the solution, is expected to be linear. However, the presence of ligands may result in deviations from linearity. It has been found that such observations can also result from adsorption to the cell wall [10].

Adsorption onto Electrodes. The adsorption of organic matter on the mercury electrode surface is a common problem and is often discussed in the literature. It may result in a shift in the peak potential, broadening of the peaks, in additional and usually sharp peaks at more positive potentials, decreased sensitivity or in non-linear calibration relations.

Organic matter may also influence the behaviour of the platinum counter electrode. Especially in low salinity waters with a high organic matter content, organic matter is easily adsorbed onto the Pt surface. As a consequence, the electrode is shielded and sensitivity decreases. The effects get more serious in consecutive measurements of the CC determination. The adsorbed organics can be observed usually as a yellow film covering the platinum coil.

Adsorption During Equilibration. If the CC measurements are carried out in the equilibration method, where several individual sample aliquots are spiked with different concentrations of copper for equilibration during e.g. 24 h prior to analysis [5], adsorption to the walls of the flasks may influence the CC determinations.

Formation of the Mercury Film. The preformed mercury film electrode may show non-linear response, if inappropriate film formation procedures have been applied.

Experimental conditions during film formation [related to e.g. Hg(II) concentration and deposition time] determine the characteristics of the MFE. An optimum film has a linear calibration curve and has maximum sensitivity. A film that is too thin has decreased sensitivity; a film that is too thick (resulting from high mercury concentrations or long deposition times) adversily affects the calibration curve [11]. The resulting curve may then suggest complexation of added trace elements, in cases where the similarity to complexing capacity properties is in fact due to erroneous electrode characteristics. It can be checked and cured by simple experiments [11].

Conditioning of the MFE. The parameters controlling conditioning of the MFE before and between measurements should also be optimized, in order to avoid memory effects or non-constant electrode behaviour.

pH Control. Purging a water sample with nitrogen results in a pH increase. Existing equilibria and speciation measurements are then likely to be affected. The use of a pH-stat using CO_2 as reagent is recommended [12], rather than the use of buffers for reasons mentioned before.

Complexation Kinetics. Slow reactions may result in slow approach of equilibrium in the complexation reactions after spiking the sample with copper. Then, CC will be underestimated. These problems can be eliminated by application of Ruzic's method [13].

Reaction with Colloids. Inorganic and/or organic colloidal material that passes a $0.45 \,\mu m$ filter, or flocculates/ precipitates formed after filtration, may react with the Cu²⁺ (adsorption, complexation) influencing the determination of complexing capacity.

Intermetallic Compounds. The formation of Cu-Zn intermetallic compounds in the mercury electrode has been reported as cause for non-linearity in CC titration curves [14]. At the low concentrations, used in the CC determinations of natural samples for trace metals, no intermetallic compounds are expected to occur. The suggestion of these authors is probably based on incorrect observations as the applied deposition potential (-700 mV) was not sufficiently negative to reduce zinc.

Most effects mentioned above might be avoided by carefully establishing optimum conditions of the measurements, e.g. choice of materials, cleaning procedures, electrode handling, equilibration, pH control and electrochemical parameters.

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