

KillerRed protein based *in vivo* photodynamic therapy and corresponding tumor metabolic imaging

Qiaoya Lin[†], Shuang Sha[†], Fei Yang, Honglin Jin and Zhihong Zhang^{*}

*Britton Chance Center for Biomedical Photonics
Wuhan National Laboratory for Optoelectronics
Huazhong University of Science and Technology
Wuhan 430074, P. R. China*

*MoE Key Laboratory for Biomedical Photonics
Department of Biomedical Engineering
Huazhong University of Science and Technology
Wuhan 430074, P. R. China
czyzzh@mail.hust.edu.cn

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Photodynamic therapy (PDT) gains wide attention as a useful therapeutic method for cancer. It is mediated by the oxygen and photosensitizer under the specific light irradiation to produce the reactive oxygen species (ROS), which induce cellular toxicity and regulate the redox potential in tumor cells. Nowadays, genetic photosensitizers of low toxicity and easy production are required to be developed. KillerRed, a unique red fluorescent protein exhibiting excellent phototoxic properties, has the potential to act as a photosensitizer in the application of tumor PDT. Meantime, the course of tumor redox metabolism during this treatment was rarely investigated so far. Thus here, we investigated the effects of KillerRed-based PDT on tumor growth *in vivo* and examined the subsequent tumor metabolic states including the changes of nicotinamide adenine dinucleotide hydrogen (NADH) and flavoprotein (Fp), two important metabolic coenzymes of tumor cells. Results showed the tumor growth had been significantly inhibited by KillerRed-based PDT treatment compared to control groups. A home-made cryo-imaging redox scanner was used to measure intrinsic fluorescence and exogenous KillerRed fluorescence signals in tumors. The Fp signal was elevated by nearly 4.5-fold, while the NADH signal decreased by 66% after light irradiation, indicating that Fp and NADH were oxidized in the course of KillerRed-based PDT. Furthermore, we also observed correlation between the fluorescence distribution of

^{*}Corresponding author.

[†]These authors contributed equally to this work.

KillerRed and NADH. It suggests that the KillerRed protein based PDT might provide a new approach for tumor therapy accompanied by altering tumor metabolism.

Keywords: Reactive oxygen species; redox; metabolism; flavoprotein; nicotinamide adenine dinucleotide hydrogen.

1. Introduction

PDT is a useful therapeutic method in the treatment of the head and neck,¹ lung, brain,² skin, breast³ and pancreatic carcinomas in clinic.⁴ It is mediated by photosensitizer and oxygen under the specific light irradiation to produce the reactive oxygen species (ROS),^{5,6} which induce cellular toxicity and regulate the redox potential in tumor cells.^{7,8} Recently, many researchers focus on the development of the photosensitizers to improve the efficacy of PDT method.^{9,10} Nowadays, most of photosensitizers are hydrophobic chemical materials and difficult to be cleared in bodies. Compared with the chemical photosensitizer, such as porphyrin related products, the genetically encoded protein has many advantages. For example, it is of low toxicity to organism and easy to obtain. Thus, developing a photosensitizer based on gene-encoded protein is of great value.¹¹

KillerRed is a unique genetically encoded far red fluorescent protein mutated from the Hydrozoa jellyfish chromoprotein anm2CP¹² and has a phototoxicity 1000-fold more than that of green fluorescent protein (GFP).¹³ It has the potential to act as a photosensitizer. Some groups reported that this KillerRed protein could produce ROS which has toxicity to the tumor cells^{13,15} and its different sub-cellular component localization also had been found, such as membrane^{11,16} or mitochondria¹⁷ which play a major role in photodynamic therapy induced cell death.^{18–20} During the treatment of photodynamic therapy, the genetically encoded photosensitizer will alter the tumor cell redox state and metabolism, however, the course of tumor redox metabolism during PDT treatment was rarely investigated so far.

Tumor metabolism plays an important role in the tumor growth²¹ and differentiation, metastasis²² and apoptosis.²³ The metabolism of tumor cells is different from most normal cells, which is called Warburg effect and is not an effective ATP generation way.²⁴ Investigating tumor metabolism is critical because it can provide an open window for tumor therapeutics.²⁵ There are two autofluorescent indicators,

nicotinamide adenine dinucleotide hydrogen (NADH) and flavoprotein (Fp), as a redox pair to indicate the tumor redox metabolism state.^{26–29} They are both coenzymes in the mitochondrial respiration chain of all tissues. The fluorescence intensity ratio of NADH/(NADH+Fp) can be as a standard for detecting tissue metabolism state which has been utilized for several years.^{30,31}

In this paper, we investigated the antitumor efficacy of KillerRed protein-based PDT in human fibrocarcinoma bearing mice. Meanwhile, the tumor redox state was monitored during the treatment, including NADH and Fp signal collected in tumor cells. We found that the KillerRed protein based PDT inhibited the tumor growth and the tumor was oxidized during the PDT treatment evidenced by increased Fp signal and decreased NADH signal. Furthermore, we also investigated the relationship between exogenous KillerRed protein and NADH, and found that the fluorescence distribution of KillerRed and NADH had correlation evidenced by their high linear coefficient.

2. Materials and Methods

2.1. Construction of plenti-hiko-KillerRed plasmid

To obtain virus transfection plasmid, the virus plasmid vector of interest plenti-hiko-KillerRed was constructed as follows:

KillerRed-F: 5'AAAGGGTCTAGATAAGCAGAGCTGGTTTAGTGAACCGTCAGATCCGCTAG3';

KillerRed-R:5'CCCGGAATTCTTAATCCTCGTCGCTACCGATGGCGCTGGTGATGCGG3'

The primers were used to amplify the dmito-KillerRed gene. The amplification condition is: 96°C 3 min, 96°C 1 min, 60°C 1 min, 72°C 1.5 min, 40 cycles, ended with 72°C 10 min. After that, the KillerRed fragment and the vector plenti-hiko were digested by XbaI and EcoRI simultaneously and then ligated with T4 Ligase.

2.2. Virus packaging and cell infection

After being constructed successfully, the plasmid was packaged with the third HIV lentivirus package system. 3×10^6 HEK-293T cells were seeded into three cell dishes, respectively, after the dishes were treated by the L-Polylysine (Sigma, USA) solution for 15 min. When the HEK-293T cells were adherent, four plasmids (plenti-hiko-KillerRed, pMD2G, pRSV-REV and pMDLg-pRRE) were transfected into the cells simultaneously, 10 μ g for each plasmid, adding 2 \times HBS (3.375 ml), 2.5 M CaCl₂ (0.377 ml) and ddH₂O (2.998 ml) respectively. Since 24 h post transfection, the supernatant of dishes were collected and replaced with fresh culture medium continually every other day for three days, and all supernatant was stored at 4°C. The supernatant was filtered before the virus was concentrated by ultracentrifuge. HT1080 cells were seeded into 96 well-plate, 1×10^4 cells for each well. When the cells were adherent, the concentrated virus was added into the cultured HT1080 cells slowly. After 24 h, the medium was replaced with fresh RPMI 1640 medium. The fluorescent HT1080 cells were digested into several 96 well-plates and the non-fluorescent cells were stamped by toothpick slowly.

2.3. KillerRed-HT1080 and HT1080 cells confocal imaging

To observe the KillerRed protein and endogenous fluorescent images, 2×10^4 HT1080 cells and KillerRed-HT1080 cells were seeded into the microscopy chamber, respectively. The KillerRed-HT1080 cells were irradiated by the 561 nm laser at the power of 100% for 30 min and then 1 μ l DCFH-DA (Beyotime, China) were added and incubated for 30 min before being washed away twice with PBS. The imaging was performed on a PE spinning disk confocal microscopy (Olympus, Japan). (The Fp fluorescent image was taken with a 520–540 nm emission filter excited by a 488 nm laser at 21% of the maximum power with an exposure time of 1 s. The KillerRed fluorescent image was taken with a 582–700 nm emission filter excited by a 561 nm laser at 21% of the maximum power with an exposure time of 1 s.)

2.4. Cell culture and tumor treatment

KillerRed-HT1080 cells were cultured in RPMI 1640 culture medium, (Gibico, USA) supplemented with

10% fetal bovine serum (Gibico, USA) and 100 IU penicillin, 100 mg/ml streptomycin at 37°C 5% CO₂. The cells were washed twice with PBS before being harvested with trypsin. Nude mice (male, 6–8 weeks) were purchased from Silaike company (Shanghai, China). Mice were maintained under SPF conditions, and all animal experiments were performed according to the animal experiment guidelines of the Animal Experimentation Ethics Committee of HUST. Nude mice were inoculated with 2×10^6 KillerRed-HT1080 cells (in 100 μ l PBS) subcutaneously in the left flanks, respectively. Two weeks after that, before the tumor volume reached 1 cm³, mice were anesthetized with 2% (v/v) isoflurane and irradiated once at the tumor area with a 561 nm laser (Coherent Asia, USA) at 80 mV for 3 min every three days. The remaining normal tissue was protected by silver paper from the light irradiation.

2.5. Redox imaging

After being treated with irradiation, the tumor bearing mice were sacrificed immediately under anesthesia, The tumors were excised, and frozen in liquid nitrogen quickly before being mounted by refrigerant (ethanol: glycerin: water = 3:6:1). In order to observe tumor metabolism directly and conveniently, we utilized a redox scanner, a home-made low temperature imaging system in our lab for monitoring tissue mitochondrial NADH and FAD fluorescent signal *ex vivo*. The fluorescent emission wavelength of NADH and Fp is 420–480 nm³² and 530–560 nm³³ respectively. NADH fluorescence in tumors was excited by 360 nm LED (Edmund optics, USA), and collected using a 450 nm filter. The flavoprotein fluorescence was excited by the 450 nm LED, and the emission using 530 nm filter. KillerRed fluorescence was excited by 590 nm LED, and collected using 650 nm filter. The image data was analyzed by Matlab 7.0 software. The NADH and Fp redox images were performed following these formulas: Ratio_{PN} = FL_{NADH} / (FL_{NADH} + FL_{Fp}) and Ratio_{Fp} = FL_{Fp} / (FL_{NADH} + FL_{Fp}).

3. Results

3.1. Detection of ROS produced by KillerRed protein

To investigate the antitumor efficacy of KillerRed protein based PDT in tumor bearing mice, we first

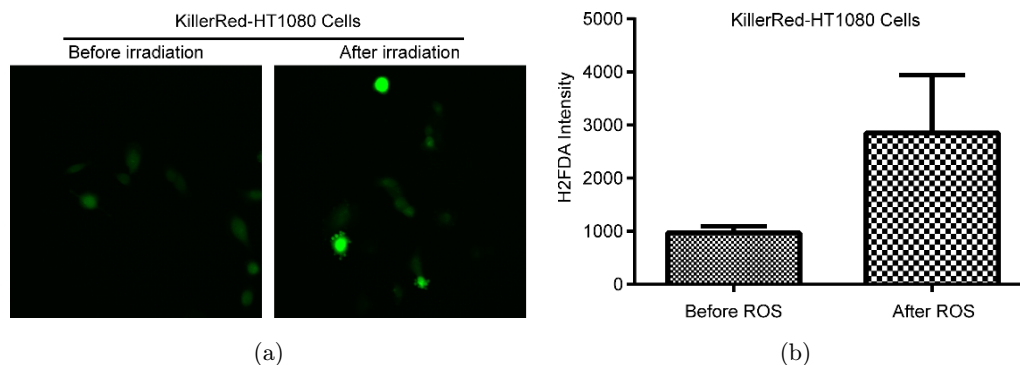


Fig. 1. The image of DCFH-DA stained the HT1080-KillerRed cells under the treatment of the light irradiation. (a) Left panel: The image of DCFH-DA stained the KillerRed-HT1080 cells with no irradiation. Right panel: DCFH-DA stained KillerRed-HT1080 cells under the irradiation for 30 min by 561 nm light. (b) Quantitative analysis from (a).

need to establish the cell line stably expressing KillerRed fluorescent protein. According to previously mentioned transfection method, we used lentivirus to infect the HT1080 cells. Fluorescence of dmito-KillerRed protein was detected in each cell, indicating that we successfully established a HT1080 cell line stably expressing KillerRed.

After that, in order to confirm whether KillerRed protein in HT1080 cell produced ROS, the ROS detection experiment was performed. The fluorescent dye DCFH-DA was used to stain KillerRed-HT1080 cells, and imaged by confocal microscopy. As shown in Fig. 1(a), without irradiation, the fluorescence intensity of green dye DCFH-DA in KillerRed-HT1080 cells was low. After KillerRed-HT1080 cells were treated with the green light irradiation for 30 min, the DCFH-DA dye fluorescence intensity in the KillerRed-HT1080 cells was enhanced [Fig. 1(a)]. Then the fluorescence intensity was quantified, showing that the green fluorescence intensity of DCFH-DA in the irradiated cells was three times stronger than the nonirradiated cells [Fig. 1(b)]. From the results we confirm that the KillerRed protein produced ROS under light irradiation.¹⁶

3.2. Radiation raises cellular autofluorescence in KillerRed expression cells

Since KillerRed protein could produce ROS in the course of green light irradiation, we next monitored the autofluorescence of cells during the irradiation with KillerRed protein. HT1080 cells and mito-KillerRed-HT1080 cells were irradiated with a 561 nm laser at 100% power at different time points (0 min, 15 min, 20 min and 30 min as indicated in Fig. 2),

respectively. The cellular autofluorescent Fp signal was detected using a confocal imaging system [Fig. 2(a)]. The Fp fluorescent signal through 515–550 nm was collected as the flavoprotein signal of cells.^{34–36}

Fig. 2(a) showed Fp and KillerRed protein fluorescent images of KillerRed-HT1080 cells at different time points, the fluorescence intensity of Fp is 318.62 ± 2.01 before irradiation, 397.75 ± 13.67 , 395.46 ± 14.26 and 394.04 ± 14.52 at 15 min, 20 min, 30 min post-irradiation, respectively [Fig. 2(b)]. The data indicated Fp fluorescent signal increased after irradiation. In the meantime, the KillerRed fluorescence was photobleached completely after light irradiation for 15 min. In contrast, no significant difference was found between different time points post-irradiation in control HT1080 cells. As well known, when flavoprotein is oxidized, its fluorescent signal will increase. So we can conclude that the KillerRed fluorescent protein under the light irradiation can induce flavoprotein oxidation in cells *in vitro*.

3.3. *In vivo* antitumor efficacy by KillerRed protein based PDT

Since KillerRed protein under light irradiation could produce ROS efficiently *in vitro*, its antitumor efficacy in tumor bearing mice based photodynamic therapy needs further investigation *in vivo*. Three groups were used in this study: KillerRed-HT1080 with irradiation, KillerRed-HT1080 without irradiation and HT1080 with irradiation. The tumor bearing mice were treated with light irradiation once every three days. The treatment started at 8 days post-tumor inoculation, when the tumor diameter is about 3.5 mm. Meanwhile, the tumor volumes were monitored during the whole treatment. As shown in Fig. 3,

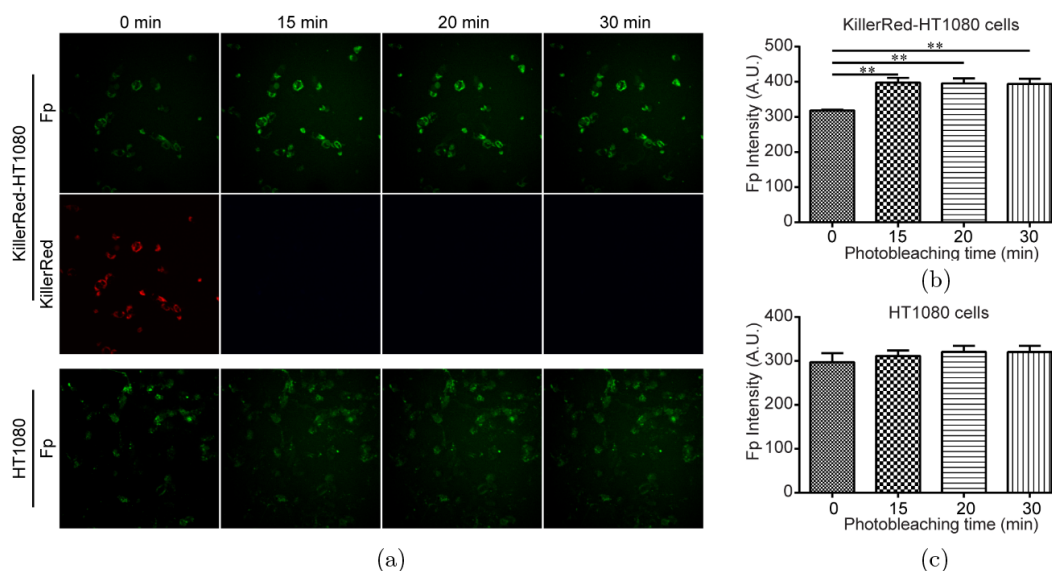


Fig. 2. Autofluorescence and KillerRed fluorescence after green light irradiation. (a) The flavoprotein fluorescence intensities of HT1080 cells and KillerRed-HT1080 cells under the irradiation by 561 nm laser for 0 min, 15 min, 20 min, and 30 min. The first row: the Fp images of KillerRed-HT1080 cells. The middle row: the KillerRed fluorescent images of KillerRed-HT1080 cells. The third row: the Fp images of the HT1080 cells. (b) The quantitative fluorescence intensity analysis of KillerRed-HT1080 cells. (c) The quantitative fluorescence intensity analysis of HT1080 cells. The scale bars represent Mean \pm SEM. (** $p < 0.01$).

the tumor growth of KillerRed-HT1080 tumor bearing mice under laser irradiation had been significantly inhibited compared to the other control groups. While there was no significant difference between KillerRed-HT1080 tumors without irradiation and HT1080 tumors with irradiation. From the results, we can conclude that the KillerRed protein based photodynamic therapy could efficiently inhibit the

human fibrocarcinoma tumor growth and it suggests KillerRed can be utilized as a photosensitizer *in vivo*.

3.4. Cyro-imaging of KillerRed-HT1080 tumors

In order to know the tumor metabolism effect of KillerRed during the course of photodynamic therapy, the cryo-images of KillerRed-HT1080 tumor and HT1080 tumor were collected to monitor the metabolism during PDT treatment. The corresponding NADH, Fp, Ratio_{NADH} and Ratio_{Fp} of tumors are shown in the Fig. 4. The tumors were frozen in liquid nitrogen after a 561 nm laser irradiation (80 mW) for three minutes and imaged using cryo-imaging system. The metabolism images of tumor samples at different depth levels (500 μm , 1000 μm , 1500 μm) were taken. The white circle region of Fp images in Fig. 4(a) (1000 μm) indicates the irradiation region inside tumors. The KillerRed protein in that region was photobleached with the fluorescent signal disappeared. The corresponding NADH fluorescence intensity was decreased and Fp fluorescence intensity was increased obviously at the irradiation region compared to no irradiation region (Fig. 4). The figure below is the histogram of the corresponding fluorescence intensity. The

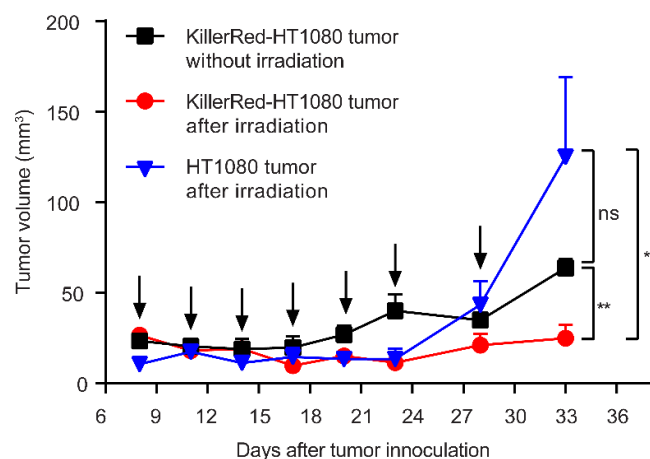


Fig. 3. The tumor volume measurement of the KillerRed-HT1080 tumors and HT1080 tumors. The black arrows indicate the time points at which the mice were treated with light irradiation. The scale bars represent Mean \pm SEM. (* $p < 0.05$, ** $p < 0.01$, $n = 4$).

images taken at other levels (images not shown here) were consistent with images shown in Fig. 4 (a). The fluorescence intensity of Fp and NADH, and corresponding Ratio_{NADH} and Ratio_{Fp} in both groups at the depth of 500 μm, 1000 μm, 1500 μm were quantified respectively [Figs. 4(c)–4(f)]. It showed that the NADH intensity after irradiation decreased by nearly

66% in average at different section level compared to nonirradiation group [Fig. 4(c)], while the Fp intensity increased nearly 4.5 folds [Fig. 4(d)]. Correspondingly, the Ratio_{NADH} after irradiation decreased by nearly 72%, while Ratio_{Fp} increased by 3.8 folds [Figs. 4(e) and 4(f)]. The data indicated that the tumor metabolism was changed under light

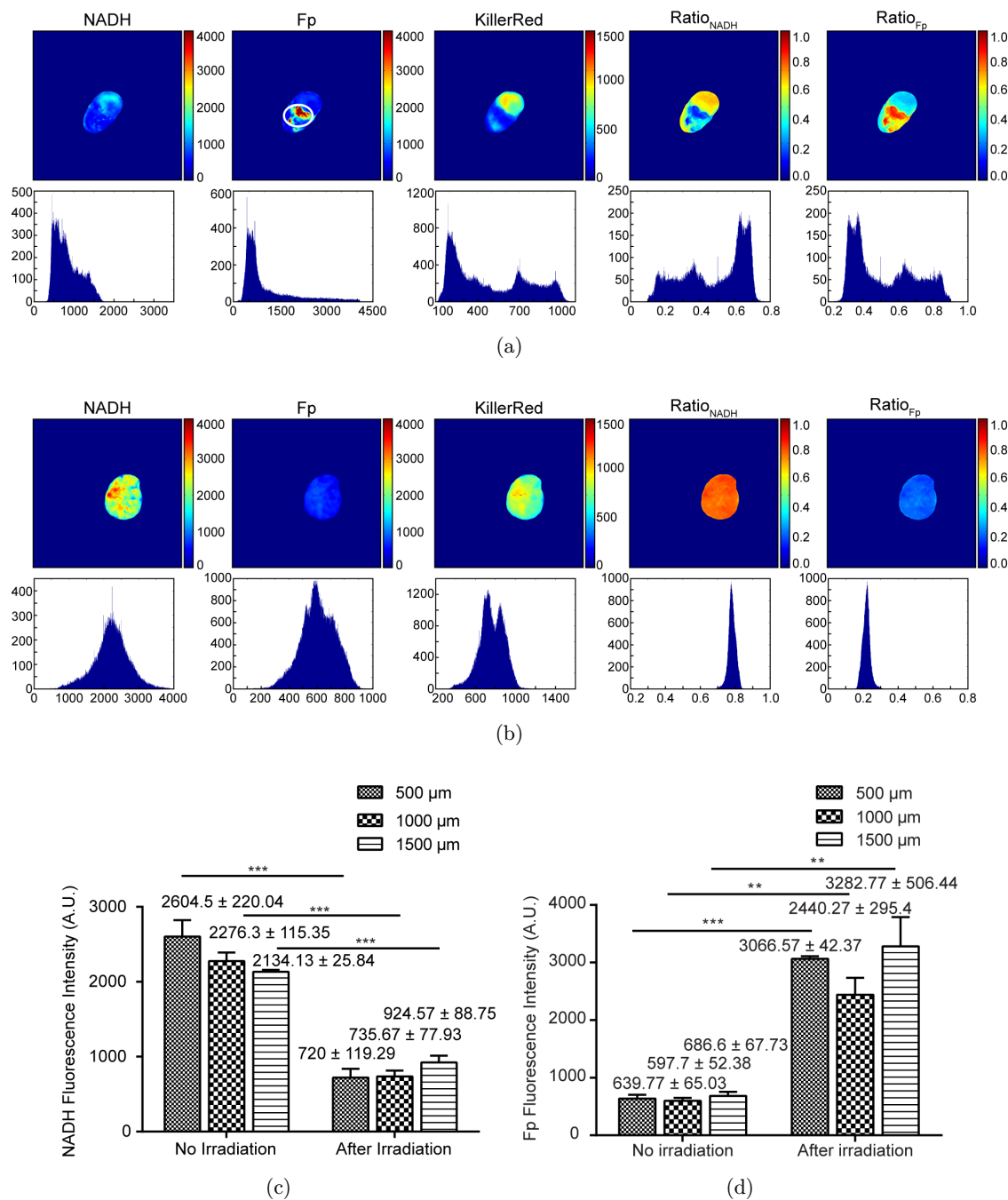


Fig. 4. Cyro-imaging of KillerRed-HT1080 tumor by light irradiation and no irradiation. (a) The cyro-imaging of KillerRed-HT1080 tumor treated by light irradiation. (b) The cyro-imaging of KillerRed-HT1080 tumor with no irradiation. (c)–(f) The quantification data on fluorescence intensity of Fp at different depth, including 500 μm, 1000 μm, 1500 μm.

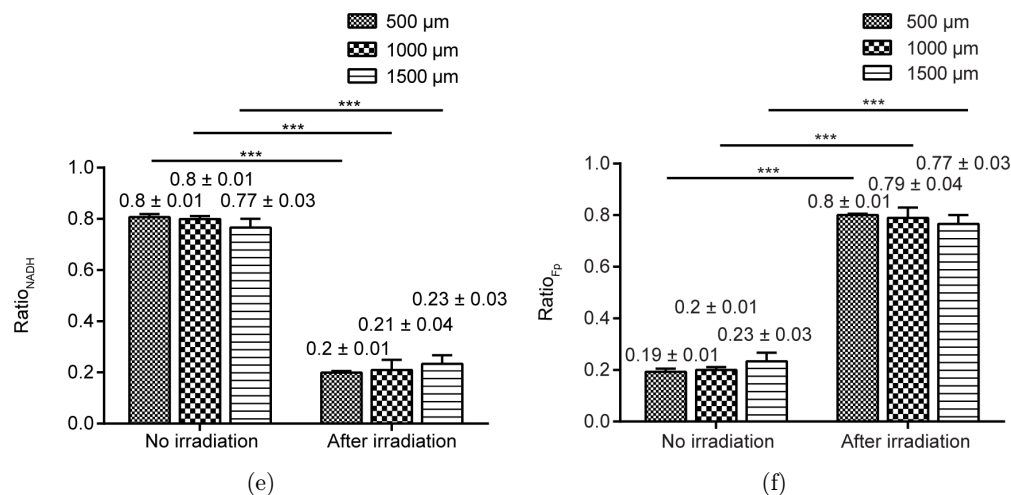


Fig. 4. (Continued)

irradiation in photodynamic therapy while oxidizing the NADH and Fp simultaneously.

3.5. The correlation between KillerRed protein and NADH signal

We also observed that the distribution of KillerRed fluorescent signal in tumors is similar with that of

NADH signal both in irradiation group and non-irradiation group, for which the correlation analysis was performed. The result showed that the coefficient of determination R^2 in KillerRed-HT1080 tumor was 0.859 in irradiation group, and 0.701 in nonirradiation group. High coefficient value (> 0.5) indicated that the distribution of KillerRed had correlation with that of NADH. It suggested that

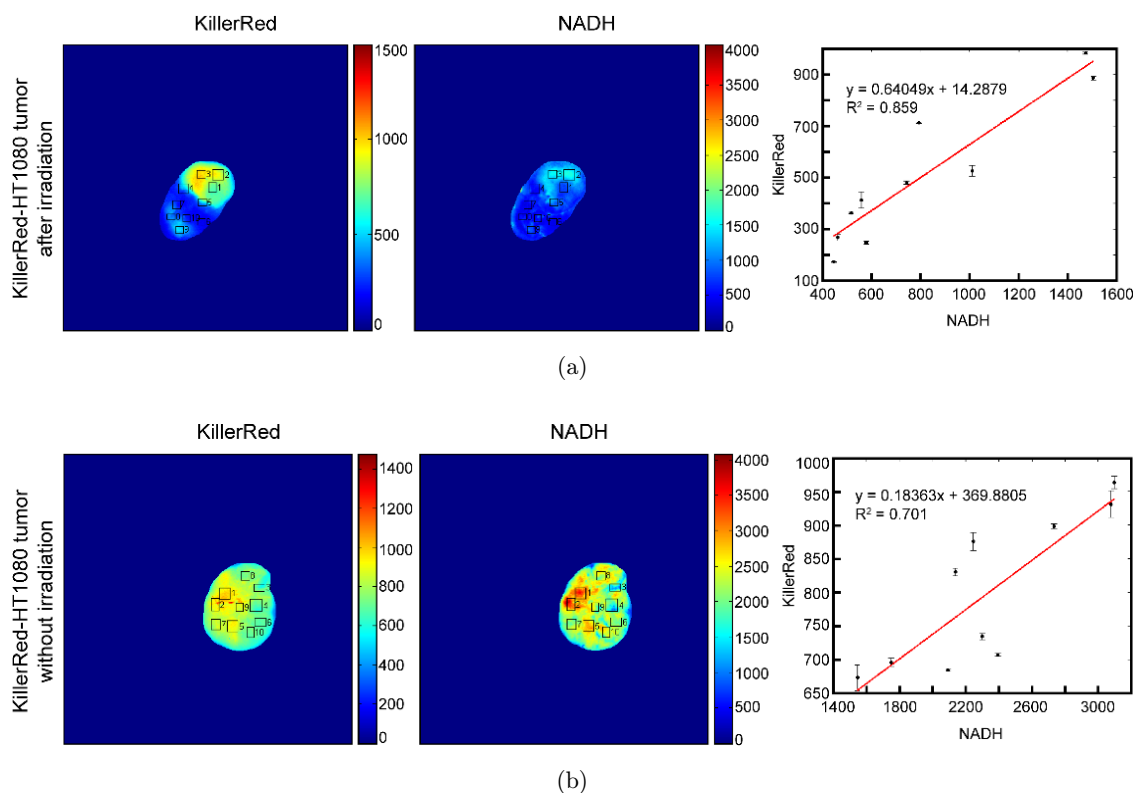


Fig. 5. The correlation between the distribution of KillerRed protein and that of NADH. (a) Tumors after irradiation treatment and (b) Tumors without irradiation.

the tumor redox state, especially the reduction state, had a relationship with the exogenous protein expression.

4. Discussion and Conclusion

KillerRed protein is a potential genetically encoded photosensitizer which can produce ROS by chromophore-assisted light inactivation (CALI) under light irradiation.¹⁴ The ROS accumulated in tumor cells will induce tumor cells apoptosis. Previous study considered that the KillerRed protein can be used as a potential photosensitizer in tumor photodynamic therapy,⁵ however, rarely reports have investigated its antitumor effect *in vivo*. Moreover, the effect of photodynamic therapy on the tumor metabolism is not very clear either. In this study, we investigated the effect on the metabolism of tumor cells *in vitro* and *in vivo* by KillerRed protein when using it as a photosensitizer in a xenograft tumor model.

In our study, it was demonstrated that the KillerRed protein could efficiently produce ROS *in vitro* (Fig. 1). We irradiated KillerRed-HT1080 cells via 561 nm laser at different time points, and detected the endogenous fluorescence intensity. The results showed that the endogenous Fp fluorescence intensity in cells raised obviously compared to HT1080 cells after irradiation for 15 min (Fig. 2). Moreover, the increased Fp signal was maintained at least to 30 min, demonstrating that the flavoprotein was irreversibly oxidized. When utilizing KillerRed protein as a photosensitizer to treat human fibrosarcoma with photodynamic therapy, we observed that the tumor growth was efficiently inhibited (Fig. 3). Under light irradiation, KillerRed protein in tumor region produced ROS, which efficiently killed tumor cells *in vivo*. In the meantime, the tumor endogenous NADH and Fp were oxidized (Fig. 4). It suggests that the KillerRed protein could be used as an efficient photosensitizer in the application of tumor therapy in future.

With tumor progression, the redox metabolism is variable and complicated. Whether the tumor redox state will affect the exogenous gene expression or not is still not clear now. In our study, we also demonstrated that the exogenous protein expression of KillerRed in tumor had relationship with the tumor redox state of NADH. We hypothesized this phenomenon may be induced by KillerRed protein maturation process which requires oxygen,³⁷ since NADH is a common reduced state indicator.

In summary, this study demonstrated the anti-tumor efficacy of KillerRed protein as an alternative PDT photosensitizer and provided useful information regarding the tumor metabolic states following KillerRed-based PDT. It also suggests that KillerRed protein based PDT has broad application for tumor therapy accompanied by altering tumor metabolism in future.

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References

1. M. A. Biel, "Photodynamic therapy and the treatment of head and neck neoplasia," *The Laryngoscope* **108**, 1259–1268 (1998).
2. M. A. Rosenthal, B. Kavar, J. S. Hill, D. J. Morgan, R. L. Nation, S. S. Stylli, R. L. Bassler, S. Uren, H. Geldard, M. D. Green, S. B. Kahl, A. H. Kaye, "Phase I and pharmacokinetic study of photodynamic therapy for high-grade gliomas using a novel boronated porphyrin," *J. Clin. Oncol.* **19**, 519–524 (2001).
3. A. Dimofte, T. C. Zhu, S. M. Hahn, R. A. Lustig, "In vivo light dosimetry for motexafin lutetium-mediated PDT of recurrent breast cancer," *Lasers Surg. Med.* **31**, 305–312 (2002).
4. D. E. Dolmans, D. Fukumura, R. K. Jain, "Photodynamic therapy for cancer," *Nat. Rev. Cancer* **3**, 380–387 (2003).
5. M. V. Shirmanova, E. O. Serebrovskaya, K. A. Lukyanov, L. B. Snopova, M. A. Sirotkina, N. N. Prodanetz, M. L. Bugrova, E. A. Minakova, I. V. Turchin, V. A. Kamensky, S. A. Lukyanov, E. V. Zagaynova, "Phototoxic effects of fluorescent protein KillerRed on tumor cells in mice," *J. Biophotonics* **6**, 283–290 (2013).
6. K. M. S. Russell, B. Vegh, Marina K. Kuimova, "Reactive oxygen species in photochemistry of the

- red fluorescent protein "Killer Red," *Chem. Commun.* **4**, 4887–4889 (2011).
7. V. Adler, Z. Yin, K. D. Tew, Z. Ronai, "Role of redox potential and reactive oxygen species in stress signaling," *Oncogene* **18**, 6104–6111 (1999).
 8. V. Irihimovitch, M. Shapira, "Glutathione redox potential modulated by reactive oxygen species regulates translation of Rubisco large subunit in the chloroplast," *J. Biol. Chem.* **275**, 16289–16295 (2000).
 9. Y. Ye, L. X. Wang, D. P. Zhang, Y. J. Yan, Z. L. Chen, "Studies on photodynamic mechanism of a novel chlorine derivative (TDPC) and its antitumor effect for photodynamic therapy *in vitro* and *in vivo*," *J. Innov. Opt. Health. Sci.* **8**, (2015).
 10. R. W. K. Wu, E. S. M. Chu, Z. Huang, M. C. Olivo, D. C. W. Ip, C. M. N. Yow, "An *in vitro* investigation of photodynamic efficacy of FosPeg (R) on human colon cancer cells," *J. Innov. Opt. Health. Sci.* **8**, (2015).
 11. Z. X. Liao, Y. C. Li, H. M. Lu, H. W. Sung, "A genetically-encoded KillerRed protein as an intrinsically generated photosensitizer for photodynamic therapy," *Biomaterials* **35**, 500–508 (2014).
 12. G. Mueller, "From green to red — To more dead? Autofluorescent proteins as photosensitizers," *J. Photoch. Photobiol. B* **9**, 95–98 (2010).
 13. M. E. Bulina, K. A. Lukyanov, O. V. Britanova, D. Onichtchouk, S. Lukyanov, D. M. Chudakov, "Chromophore-assisted light inactivation (CALI) using the phototoxic fluorescent protein KillerRed," *Nat. Protoc.* **1**, 947–953 (2006).
 14. M. E. Bulina, D. M. Chudakov, O. V. Britanova, Y. G. Yanushevich, D. B. Staroverov, T. V. Cherpurnykh, E. M. Merzlyak, M. A. Shkrob, S. Lukyanov, K. A. Lukyanov, "A genetically encoded photosensitizer," *Nat. Biotechnol.* **24**, 95–99 (2006).
 15. E. O. Serebrovskaya, E. F. Edelweiss, O. A. Stremovskiy, K. A. Lukyanov, D. M. Chudakov, S. M. Deyev, "Targeting cancer cells by using an anti-receptor antibody-photosensitizer fusion protein," *Proc. Natl. Acad. Sci. USA* **106**, 9221–9225 (2009).
 16. W. Waldeck, G. Mueller, M. Wiessler, K. Toth, K. Braun, "Positioning effects of KillerRed inside of cells correlate with DNA strand breaks after activation with visible light," *Int. J. Med. Sci.* **8**, 97–105 (2011).
 17. T. Shibuya, Y. Tsujimoto, "Deleterious effects of mitochondrial ROS generated by KillerRed photodynamic action in human cell lines and *C. elegans*," *J. Photoch. Photobiol.* **117**, 1–12 (2012).
 18. J. Morgan, A. R. Oseroff, "Mitochondria-based photodynamic anti-cancer therapy," *Adv. Drug. Deliv. Rev.* **49**, 71–86 (2001).
 19. X. Wang, "The expanding role of mitochondria in apoptosis," *Genes. Dev.* **15**, 2922–2933 (2001).
 20. S. Orrenius, "Mitochondrial regulation of apoptotic cell death," *Toxicol. Lett.* **149**, 19–23 (2004).
 21. M. Ristow, "Oxidative metabolism in cancer growth," *Curr. Opin. Clin. Nutr.* **9**, 339–345 (2006).
 22. L. Z. Li, "Imaging mitochondrial redox potential and its possible link to tumor metastatic potential," *J. Bioenerg. Biomembr.* **44**, 645–653 (2012).
 23. J. W. Locasale, L. C. Cantley, "Metabolic flux and the regulation of mammalian cell growth," *Cell Metab.* **14**, 443–451 (2011).
 24. P. P. Hsu, D. M. Sabatini, "Cancer cell metabolism: Warburg and beyond," *Cell* **134**, 703–707 (2008).
 25. M. G. Vander Heiden, "Targeting cancer metabolism: A therapeutic window opens," *Nat. Rev. Drug Discov.* **10**, 671–684 (2011).
 26. P. G. Michal Mokry, B. Vidinsky, "*in vivo* monitoring the changes of interstitial pH and FAD/NADH," *Photochem. Photobiol.* **8**, 793–797 (2006).
 27. Z. Zhang, H. Li, Q. Liu, L. Zhou, M. Zhang, Q. Luo, J. Glickson, B. Chance, G. Zheng, "Metabolic imaging of tumors using intrinsic and extrinsic fluorescent markers," *Biosens. Bioelectron.* **20**, 643–650 (2004).
 28. B. Chance, B. Schoener, R. Oshino, F. Itshak, Y. Nakase, "Oxidation-reduction ratio studies of mitochondria in freeze-trapped samples. NADH and flavoprotein fluorescence signals," *J. Biol. Chem.* **254**, 4764–4771 (1979).
 29. L. Z. Li, R. Zhou, H. N. Xu, L. Moon, T. Zhong, E. J. Kim, H. Qiao, R. Reddy, D. Leeper, B. Chance, J. D. Glickson, "Quantitative magnetic resonance and optical imaging biomarkers of melanoma metastatic potential," *Proc. Natl. Acad. Sci. USA* **106**, 6608–6613 (2009).
 30. K. P. Q. Irene Georgakoudi, "Optical imaging using endogenous contrast to assess metabolic state," *Annu. Rev. Biomed. Eng.* **14**, 351–367 (2012).
 31. D. B. Zhihong Zhang, Hui Li, "Redox ratio of mitochondria as an indicator of the response of photodynamic therapy," *J. Biomed. Opt.* **9**, 772–778 (2004).
 32. A. Mayevsky, G. G. Rogatsky, "Mitochondrial function *in vivo* evaluated by NADH fluorescence: from animal models to human studies," *Am. J. Physiol. Cell Ph.* **292**, C615–C640 (2007).
 33. K. D. V. Jayanth Kumar, Ganesan Singaravelu, "In vivo estimation of redox states with autofluorescence spectroscopy in oral submucous fibrosis patients: A Pilot Study," *J. Indian Acad. Oral Med. Radiol.* **24**, 257–260 (2012).
 34. M. S. Islam, M. Honma, T. Nakabayashi, M. Kinjo, N. Ohta, "pH dependence of the fluorescence

- lifetime of FAD in solution and in cells," *Int. J. Mol. Sci.* **14**, 1952–1963 (2013).
35. J. V. Rocheleau, W. S. Head, D. W. Piston, "Quantitative NAD(P)H/fluoroprotein autofluorescence imaging reveals metabolic mechanisms of pancreatic islet pyruvate response," *J. Biol. Chem.* **279**, 31780–31787 (2004).
36. A. V. Kuznetsov, J. Troppmair, R. Sucher, M. Hermann, V. Saks, R. Margreiter, "Mitochondrial subpopulations and heterogeneity revealed by confocal imaging: Possible physiological role?" *Biochim. Biophys. Acta.* **1757**, 686–691 (2006).
37. R. Y. Tsien, "The green fluorescent protein," *Annu. Rev. Biochem.* **67**, 509–544 (1998).