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# Accelerated modification of the zona pellucida is the primary cause of decreased fertilizability of oocytes in the 129 inbred mouse strain

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# Accelerated Modification of the Zona Pellucida is the Primary Cause of Decreased Fertilizability of Oocytes in the 129 Inbred Mouse Strain

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**Short title:** Decreased fertilizability in 129 mice

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**Abstract**

We investigated whether the small litter size in 129 inbred mouse strain results from a reduction in oocyte fertilizability. Sensitivity of the zona pellucida to α-chymotrypsin was examined for oocytes collected at 14 (shortly after ovulation), 17, and 20 h after hCG injection. Passage of spermatozoa through the zona pellucida (using an *in vitro* fertilization (IVF) technique) and the density of cortical granules were examined for oocytes collected at 14 and 17 h after hCG injection. The capability of the oolemma to fuse with the sperm plasma membrane was also evaluated by IVF using zona-free eggs. The zona pellucida became markedly resistant to the enzyme 17 h after hCG injection. IVF rates significantly decreased at this time. In addition, there was a significant reduction in the density of cortical granules. When zona-free oocytes were inseminated, high fertilization rates were obtained at both 17 and 14h after hCG injection. These results indicate that accelerated modification of the zona pellucida primarily causes a decreased fertilizability of oocytes in 129 mice, resulting in the low reproductive performance of this strain.

Keywords: 129 mouse, Fertility, Oocyte, Zona pellucida, Cortical granules

## Introduction

The 129 inbred mouse strain is useful as an animal model of testicular teratoma (Stevens, 1967; Matin, 2007), and it greatly contributes to production of genetically engineered mice as a supplier of embryonic stem (ES) cells (Evans & Kaufman, 1981; Martin, 1981). Apart from these useful traits for genetic research, it has been reported that the litter size of 129 mice is considerably smaller than that of other popular inbred strains such as C3H and C57BL/6 (Verley *et al*., 1967; Nagasawa *et al*., 1973; Festing, 1979). The low reproductive performance of 129 mice may be attributable to failure in fertilization rather than embryonic death because the *in vitro* fertilization (IVF) rates are usually low (Sztein *et al*., 2000; Byers *et al*., 2006; Kawai *et al*., 2006). Our previous study found that the *in vivo* fertilization rate was less than 50% even when 129 females were mated with C57BL/6J males with normal fertility (Hino *et al*., 2009). These findings strongly suggest that the low fertilization rate of 129 mice arises from oocytes. One of the possible causative factors is chemical alteration of the zona pellucida enclosing the oocyte; thus, spermatozoa cannot pass through the structure. Another is that the capability of the oolemma to fuse with the sperm plasma membrane is low in this strain.

To determine the exact causative factor(s) of the low fertilization rates of strain 129 mice, the sensitivity of the zona pellucida to protease and the fertilizability of oocytes were examined at different times after ovulation. The capability of the oolemma to fuse with the sperm plasma membrane was evaluated by IVF assay using zona-free oocytes. In addition, development of fertilized eggs was followed up to full term.

## Material and methods

*Animals*

The 129 inbred mouse strain used in the present study were of two 129 substrains; 129+Ter/Sv mice were purchased from CLEA Japan (Tokyo, Japan) and 129/SvEv mice were purchased from Biological Research Laboratories (Füllinsdorf, Switzerland). Inbred C57BL/6J mice with normal reproductive performance were purchased from CLEA Japan (Tokyo, Japan) and used as a standard of comparison. Females from 2 to 4 months of age and males from 3 to 4 months of age (strain 129 and C57BL/6J mice) were used in the experiments. A mature MCH (ICR) hybrid mouse strain from CLEA Japan (Tokyo, Japan) served as recipients of embryo transfer. The 129+Ter/Sv, 129/SvEv, and C57BL/6J were referred to as 129T, 129S and B6/J, respectively. All mice were kept under specific pathogen-free conditions for at least 1 week before use. They were fed ad libitum under controlled lighting conditions (Light: 08:00 to 20:00) at a temperature of 23 ± 1°C and humidity of 55 ± 10%. All experimental procedures were approved by the Animal Care and Use Committee of the Mitsubishi Kagaku Institute of Life Sciences.

*Media*

Organic and inorganic reagents were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan), unless specifically stated. The medium used for oocyte collection and *in vitro* fertilization (IVF) was TYH medium (Toyoda *et al*., 1971a). The culture medium for embryos was modified Whitten medium (mWM) (Nomura & Katsuki, 1987), which consists of 109.51 mM NaCl, 4.78 mM KCl, 1.19 mM $KH_2PO_4$, 1.19 mM $MgSO_4 \cdot 7H_2O$, 22.62 mM $NaHCO_3$, 5.56 mM glucose, 0.23 mM sodium pyruvate (Nacalai

Tesque, Kyoto, Japan), 1.49 mM calcium lactate·5$H_2O$, 75 mg/l penicillin G potassium (Meiji Seika, Tokyo, Japan), 50 mg/l streptomycin sulphate (Meiji Seika), 0.01 mM 2-mercaptoethanol (Nacalai Tesque), 0.05 mM EDTA·2Na (Nacalai Tesque), 1 mg/l phenol red, and 3 g/l bovine serum albumin (BSA) (Yagai Co. Ltd., Yamagata, Japan).

*Time of ovulation after hCG injection*

Females were intraperitoneally injected with 7.5 IU eCG followed 48 h later with 7.5 IU hCG. They were sacrificed by cervical dislocation at 1-h intervals from 10 to 15 h after hCG injection and their oviducts were removed. When cumulus-enclosed oocytes were detected in the ampullary region of the oviducts, they were collected and put in a droplet (50 μl) of TYH medium containing 0.01% hyaluronidase (Sigma-Aldrich, St Louis, MO, USA) under paraffin oil (Fisher Scientific, Fair Lawn, NJ, USA). Five to 10 min later, cumulus-free oocytes were washed twice with TYH medium. The number of oocytes was recorded.

*Dissolution of the zona pellucida by chymotrypsin*

Cumulus-enclosed oocytes were obtained from females at 14, 17, and 20 h after hCG injection and one-cell (1-cell) embryos from females 1 day after mating (24h after hCG injection). After removal of the cumulus cells by treatment with 0.01% hyaluronidase, they were washed thoroughly with TYH medium and transferred to droplets (50 μl; two to four oocytes/embryos per droplet) of TYH medium containing 1.5 IU α-chymotrypsin (Sigma-Aldrich, St Louis, MO, USA) under paraffin oil at 37°C under 5% $CO_2$ in air. They were observed for the absence of the zona pellucida at 10, 30, 60, 120, 180, and 280 min. For each case, 46 to 73 oocytes and 26 to 33 1-cell embryos were examined.

*IVF assay with zona-intact oocytes*

To examine passage of spermatozoa through the zona pellucida, IVF was performed according to the procedure by Toyoda *et al*. (1971a; b). Spermatozoa obtained from the cauda epididymis of 129T, 129S, and B6/J males were introduced into a droplet (300 μl) of TYH medium under paraffin oil and were incubated for 1.5 to 2 h at 37°C under 5% $CO_2$ in air to induce capacitation.

Cumulus-enclosed oocytes were obtained at 14 and 17 h after hCG injection. They were transferred to a droplet (300 μl) of TYH medium, and inseminated by adding the preincubated sperm suspension. The final concentration of spermatozoa at the time of insemination was 150-200 cells/μl. In every IVF experiment, two to three females were used.

Five to 6 h after insemination, the eggs were washed thoroughly with mWM, and the formation of pronuclei and extrusion of a second polar body were microscopically examined. Ova with both male and female pronuclei and a second polar body were recorded as monospermic ova, those with more than two pronuclei and a second polar body as polyspermic ova, and those with one pronucleus as parthenogenetic ova.

*IVF assay with zona-free oocytes*

The cumulus-enclosed oocytes were obtained from 129T and B6/J females at 16 h after hCG injection. Cumulus cells were dispersed by hyaluronidase treatment, and the zona pellucidae were completely dissolved in acidic Tyrode's solution (pH 2.5) followed by three rapid washes in TYH medium. The zona-free eggs were directly transferred to a droplet (300 μl) of TYH medium containing 129T spermatozoa following preincubation for 1.5 to 2 h at a

126 concentration of 3 and 10 cells/μl. Insemination was completed 17 h after hCG injection.
127 Nine hours after insemination, the number of pronuclei was scored.
128
129 *Cortical granule staining and quantification*
130 Zona-free eggs of 129T, 129S, and B6/J females were obtained 14 and 17 h after
131 hCG injection as mentioned above. To examine whether the spontaneous release of cortical
132 granules occurs *in vitro*, some of cumulus-enclosed oocytes recovered 14h after hCG
133 injection were cultured in TYH medium for 3h. The zona-free oocytes were fixed in 3.7%
134 paraformaldehyde in Dulbecco's PBS (D-PBS) for 30 min and blocked in D-PBS containing
135 0.3% BSA (blocking solution). They were washed three times in blocking solution, and then
136 permeabilized in D-PBS containing 0.1% Triton X-100 for 5 min. After washing three times
137 in blocking solution, they were incubated for 30 min in D-PBS containing 100 μg/ml FITC
138 conjugate-lens culinaris agglutinin LCA (Sigma-Aldrich, St Louis, MO, USA) which
139 specifically attaches to the cortical granules (Ducibella *et al*., 1988). They were washed three
140 times in blocking solution and mounted with Vectashield containing DAPI (Vector, RL-1000,
141 Burlingame, CA, USA). All procedures were conducted at room temperature.
142 The cortical granule density in 100 $\mu m^2$ area of the cortex was determined under
143 fluorescence microscope by counting LCA-labeled cortical granules. For each case, 26 to 39
144 oocytes from three to four females were examined.
145
146 *Embryo transfer*
147 Some of monospermic zygotes produced by IVF assay with 129T and 129S
148 zona-intact oocytes recovered at 14 and 17 h after hCG injection were cultured in mWM

medium under 5% $CO_2$ in air at 37°C. On the next day, resultant two-cell embryos were transferred into the oviduct of pseudopregnant ICR females. Two to four females were used as recipients, and 20 embryos were transferred into each recipient. Pregnant females were either sacrificed 19 days after the transfer or allowed to deliver in order to count live pups.

*Statistical analysis*

Fertilization rates of zona-intact oocytes were analyzed by one-way ANOVA after transformation into arcsine values. The average number of ovulated oocytes and density of cortical granules were compared by Student's *t*-test. Results of IVF assay with zona-free oocytes and embryo development were analyzed by $\chi^2$-test. Differences were considered to be significant with $P < 0.05$.

## Results

*Time of ovulation after hCG injection*

Table 1 presents the number of females with oocytes in the oviducts and the number of oocytes recovered at different times after hCG injection in 129T and B6/J strains. In 129T, oocytes were recovered from only one female at 12 h after hCG injection and from all females at and after 13 h after hCG injection. The number of oocytes recovered reached the maximum level at 14 h after hCG injection, indicating that ovulation was completed between 12 and 14 h after hCG injection in this strain. By contrast, all females had ovulated by 12 h after hCG injection in B6/J. Thus, it appears that ovulation after hCG injection in 129T females occurs approximately 2 h later compared to B6/J females. Interestingly, the mean number of oocytes recovered on completion of ovulation was obviously larger in 129T than

172 in B6/J ($P < 0.01$).

173

174 *Change in sensitivity of the zona pellucida to chymotrypsin*

175 Sensitivity of the zona pellucida of oocytes to chymotrypsin was markedly
176 dependent upon both the time after ovulation and the mouse strain (Figure 1). When oocytes
177 were recovered at 14 h after hCG injection (shortly after ovulation) in 129T and 129S, the
178 zona pellucida was usually digested by the enzyme within 10 min. However, the structure
179 became considerably resistant to the enzyme when oocytes were recovered at 17 h
180 (approximately 3 h after ovulation) and 20 h (approximately 6 h after ovulation) after hCG
181 injection. In B6/J oocytes, a high sensitivity of the zona pellucida to chymotrypsin persisted
182 until 17 h after hCG injection (approximately 5 h after ovulation). Even in oocytes recovered
183 at 20 h after hCG injection (approximately 8 h after ovulation), digestion of the zona
184 pellucida was seen in more than 90% of oocytes by 120 min. The zona pellucida of 1-cell
185 embryos of all strains was highly resistant to the enzyme until 120 min.

186

187 *IVF assay with zona-intact oocytes*

188 Results of the IVF assay are presented in Table 2. In the IVF assay with 129T
189 oocytes, B6/J spermatozoa were also used for insemination to check on the results obtained
190 by 129T spermatozoa. When 129T oocytes recovered 14 h after hCG injection were used,
191 fertilization rates were similar to those of B6/J oocytes regardless of donors of spermatozoa.
192 A similar result was found when 129S oocytes were inseminated 14 h after hCG injection.
193 When 129T oocytes were inseminated 17 h after hCG injection, however, the fertilization
194 rates were significantly reduced and the low fertilization rate was inadequately improved by

use of B6/J spermatozoa. Consequently, the fertilization rates of 129T oocytes recovered 17 h after hCG injection were much lower than that of B6/J oocytes recovered at the same time. When 129S and B6/J oocytes were inseminated 17 h after hCG injection, the fertilization rate was significantly lower in 129S oocytes than in B6/J oocytes. In this study, monospermy was seen in more than 90% of fertilized eggs regardless of the time of oocyte recovery after hCG injection and origin of gametes.

*IVF assay with zona-free oocytes*

Immediately after the zona-free oocytes were placed in a droplet of sperm suspension, they were quickly attached by some spermatozoa. In both low and high sperm concentrations, fertilization rates were significantly higher in 129T oocytes than in B6/J oocytes (Table 3), indicating that the capability of oolemma of 129 mouse oocytes to fuse with sperm plasma membrane never deteriorated up to at least 3 h after ovulation.

*Observation of cortical granules*

All of 129T and 129S oocytes recovered at 14 and 17 h after hCG injection showed typical metaphase II configuration and cortical granule domain (Figures 2a and 2c). However, the density of cortical granules in 129T and 129S oocyte recovered 17 h after hCG injection significantly decreased (Figures 2b, 2d and 3).This reduction occurred when oocytes recovered 14h after hCG injection were cultured *in vitro* for further 3h. Although the density of cortical granules in B6/J oocytes recovered 14h after hCG injection was low, there was no such reduction in density of cortical granules. Thus, partial release of cortical granules occurred in a time-dependent manner with 129T and 129S oocytes even when the oocytes

were cultured *in vitro*.

*Embryo development*

In IVF assay with oocytes recovered 14 and 17h after hCG injection, almost all (98-100%) of monospermic zygotes in 129T and 129S developed to two-cell embryos. After embryo transfer, all of the recipient females became pregnant (Table 4). The embryos of both substrains well developed to term and there was no significant difference in percentage of live pups delivered between both oocyte recovery times after hCG injection. These percentages in 129T and 129S were comparable to that obtained in our previous study (Suzuki-Migishima et al., 2009).

**Discussion**

The present study found that the zona pellucida of 129 mouse oocytes became resistant to chymotrypsin approximately 3 h after ovulation (17 h after hCG injection), and concomitantly rates of successful IVF in the 129 mouse oocytes significantly decreased; however, their oolemma maintained the capability to fuse with the sperm plasma membrane. The decrease of IVF rates persisted even when B6/J spermatozoa were used. These findings indicate that the zona pellucida of 129 mouse oocytes primarily hampers fertilization. Because there was a significant reduction in the density of cortical granules, the partial release of cortical granules might have caused the zona pellucida to become resistant to the enzyme.

Usually, the zona pellucida acquires resistance to proteases after penetration of the spermatozoa into the oocyte cytoplasm (Smithberg, 1953; Krzanowska, 1972; Mintz &

241 Gearhart, 1973; Schmell & Gulyas, 1980; Gulyas & Yuan, 1985), ensuring an oocyte
242 monospermy (Barros & Yanagimachi, 1971; Sato, 1979). Xu *et al*. (1997) reported that
243 spontaneous activation, which is accompanied by release of some cortical granules,
244 modification of the zona pellucida, and transition of metaphase to anaphase, occurs in
245 post-ovulatory aged oocytes of CF-1 mice. Compared to freshly-ovulated oocytes, the aged
246 mouse oocytes were reportedly susceptible to artificial parthenogenetic stimuli (Fulton &
247 Whittingham, 1978; Kubiak, 1989; Collas *et al*., 1989; Xu *et al*., 1997). Therefore, partial
248 release of cortical granules found in 129 mouse oocytes approximately 3 h after ovulation
249 might have resulted from spontaneous activation of the oocytes. However, it appeared that
250 the activation was too weak to induce the transition of metaphase to anaphase because there
251 were no oocytes exhibiting the spindle of anaphase configuration.
252 In most mammalian species, the epithelial cells of the oviducts secrete glycoproteins
253 (OGPs), which can interact with the zona pellucida of ovulated oocytes (Buhi, 2002). Coy *et*
254 *al*. (2008) reported that the OGPs made the zona pellucida of bovine and swine oocytes
255 resistant to proteases and thereby contributed to prevent polyspermy. More recently, it has
256 been found that OGPs function as adhesive ligands for mouse spermatozoa (Lyng & Shur,
257 2009). However, it remains unknown whether OGPs can chemically alter the zona pellucida
258 to block polyspermy.
259 Our previous study demonstrated that 129 female mice showed reduction in litter
260 size after natural mating (Hino *et al*., 2009). The time interval between ovulation and sperm
261 penetration *in vivo* has been estimated to be 3 to 5 h in spontaneously ovulated females and 1
262 to 3 h in superovulated females in common mouse strains (Braden & Austin, 1954; Edwards
263 & Gates, 1959; Braden, 1962). If this is the case in 129 mice, the small litter size following

natural mating could be due to poor fertilization via chemical alteration of the zona pellucida before penetration of spermatozoa.

Sztein *et al*. (2000) and Byers *et al*. (2006) reported that 129 female mice had relatively low IVF rates (53% and 24%, respectively) even when oocytes were recovered 13 to 14.5 h after hCG injection; however, in our study, IVF rates with oocytes recovered 14 h after hCG injection were 78% in 129T mice and 54% in 129S mice. According to our results, ovulation would be either in progress or finished at this time; therefore, the oocytes should maintain normal fertilizability. Although it is difficult to directly compare our results with previous results because of different IVF methodologies, the time lag of fertilization following insemination may cause a discrepancy in the IVF rates between studies. In the present study, spermatozoa were adequately capacitated by preincubation for 90 to 120 min; however, the sperm preincubation was short (about 10 min) in the previous studies (Sztein *et al*., 2000; Byers *et al*., 2006). Toyoda *et al*. (1971b) reported that the passage of spermatozoa through the zona pellucida was delayed following the preincubation for less than 15 min compared to the preincubation for 60 to 120 min because of inadequate sperm capacitation. It is therefore possible that the low IVF rates reported in the previous studies may be explained by the alteration of the zona pellucida before the penetration of spermatozoa through it.

In humans, even though production of spermatozoa is normal in number and motility, fertilization failure (0%) and/or low fertilization (<25%) occurs in 4-20% of the couples undergoing IVF (Barlow *et al*., 1990; Molloy *et al*., 1991; Roest *et al*., 1998). Although the causes remain unclear, the possibility exists that spermatozoa fail to pass through the zona pellucida. Olds-Clarke (1996) reported that not only the sperm velocity but also the quality of the zona pellucida influences the success of IVF. Männikkö *et al*. (2005) reported that gene

mutation affecting the structure of the zona pellucida is associated with the IVF failure. If an adverse alteration of the zona pellucida actually occurs in the oocytes of the infertile women, the 129 mice may be useful as a relevant animal model of unexplained fertilization failure.

In conclusion, 129 mouse oocytes exhibit the short fertilizable life span. This is due to accelerated alteration of the zona pellucida, which is probably caused by the spontaneous release of cortical granules. The use of oocytes immediately after ovulation can improve the IVF rate and enhance the reproductive efficiency of 129 inbred mouse strains.

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390 **Figure Legends**

391 **Figure 1.** Difference in dissolution time of the zona pellucida by chymotrypsin among
392 oocytes recovered at 14, 17, and 20 h after hCG injection in 129T, 129S and B6/J mice.
393
394 **Figure 2.** Fluorescence micrographs of cortical granules in oocytes of 129T mice. **a**: oocyte
395 recovered 14 h after hCG injection, **b**: higher magnification of the frame in **a**, **c**: oocyte
396 recovered 17 h after hCG injection, **d**: higher magnification of the frame in **c**. A scale bar
397 represents 30 μm in a and c, and 5μm in b and d, respectively.
398
399 **Figure 3.** Comparison of cortical granule density in oocytes recovered 14 and 17 h after hCG
400 injection in 129T and B6/J mice. White and black bars represent the cortical granule density
401 when oocytes were recovered 14 and 17h after hCG injection, respectively. Gray bars
402 represent the cortical granule density when oocytes recovered 14h after hCG injection were
403 cultured *in vitro* for 3h. *$P<0.05$; **$P<0.01$.

**Table 1** Ovulation in 129T and B6/J females at different intervals after hCG injection

| Time (h) after hCG injection | 129T | | | B6/J | | |
|---|---|---|---|---|---|---|
| | No. of females examined | No. of females with oocytes | No. of oocytes collected (mean) | No. of females examined | No. of females with oocytes | No. of oocytes collected (mean) |
| 10 | - | - | - | 5 | 1 | 2 (0.4) |
| 11 | - | - | - | 5 | 5 | 34 (6.8) |
| 12 | 5 | 1 | 1 (0.2) | 5 | 5 | 163 (32.6) |
| 13 | 5 | 5 | 119 (23.8) | 5 | 5 | 135 (27.0) |
| 14 | 4 | 4 | 222 (55.5) | - | - | - |
| 15 | 6 | 6 | 324 (54.0) | - | - | - |

**Table 2** IVF assay with 129T, 129S and B6/J zona-intact oocytes recovered at 14 and 17 h after hCG injection

| Time (h) after hCG injection | Donors | | No. of exp. | No. of ova examined | No. of fertilized ova | | | No. of parthenogenetic eggs (with one pronucleus) |
|---|---|---|---|---|---|---|---|---|
| | Oocytes | Spermatozoa | | | Total (%) | Monospermic | Polyspermic | |
| 14 | 129T | 129T | 4 | 423 | 329 (77.8)[a] | 321 | 8 | 5 |
| | B6/J | | 4 | 184 | 157 (85.3)[b] | 149 | 8 | 1 |
| | 129T | B6/J | 4 | 335 | 236 (70.4)[c] | 229 | 7 | 3 |
| | B6/J | | 4 | 244 | 214 (87.7)[d] | 203 | 11 | 2 |
| | 129S | 129S | 4 | 224 | 120 (53.6)[e] | 119 | 1 | 10 |
| | B6/J | | 4 | 227 | 155 (68.3)[f] | 152 | 3 | 3 |
| 17 | 129T | 129T | 4 | 407 | 107 (26.3)[g] | 105 | 2 | 3 |
| | B6/J | | 4 | 214 | 142 (66.4)[h] | 137 | 5 | 0 |
| | 129T | B6/J | 4 | 429 | 202 (47.1)[i] | 201 | 1 | 2 |
| | B6/J | | 4 | 200 | 185 (92.5)[j] | 181 | 4 | 0 |
| | 129S | 129S | 4 | 274 | 128 (46.7)[k] | 128 | 0 | 1 |
| | B6/J | | 4 | 175 | 143 (81.7)[l] | 141 | 2 | 1 |

・Statistical significance of B6/J oocytes: g vs. h ($P < 0.05$), i vs. j ($P < 0.01$), and k vs. l ($P < 0.01$).

・Statistical significance between 14 and 17 h post-hCG: a vs g ($P < 0.05$).

**Table 3** IVF rate of 129T and B6/J zona-free oocytes inseminated 17 h after hCG injection

| Sperm concentration (/μl) | Donors | | No. of oocytes examined | No. of fertilized eggs | | |
|---|---|---|---|---|---|---|
| | Oocytes | Spermatozoa | | Total (%) | monospermy | polyspermy |
| 3 | 129T | 129T | 68 | 59 (86.8)[a] | 57 | 2 |
| | B6/J | 129T | 70 | 43 (61.4)[b] | 61 | 6 |
| 10 | 129T | 129T | 69 | 67 (97.1)[c] | 61 | 6 |
| | B6/J | 129T | 71 | 62 (88.7)[d] | 50 | 13 |

・Statistical significance of B6/J oocytes: a vs. b ($P < 0.01$), c vs. d ($P < 0.05$).

**Table 4** Offspring from 129T and 129S embryos produced by IVF assay with oocytes recovered 14 and 17 h after hCG injection

| Time (h) after hCG injection | Donors | | No. of 2-cell embryos transferred | No. of recipients used | No. of pregnant recipients | No. (%) of pups |
|---|---|---|---|---|---|---|
| | Oocytes | Spermatozoa | | | | |
| 14 | 129T | 129T | 80 | 4 | 4 | 53 (66.3) |
| | 129S | 129S | 60 | 3 | 3 | 38 (63.3) |
| 17 | 129T | 129T | 60 | 3 | 3 | 42 (70.0) |
| | 129S | 129S | 40 | 2 | 2 | 23 (57.5) |

Figure 1

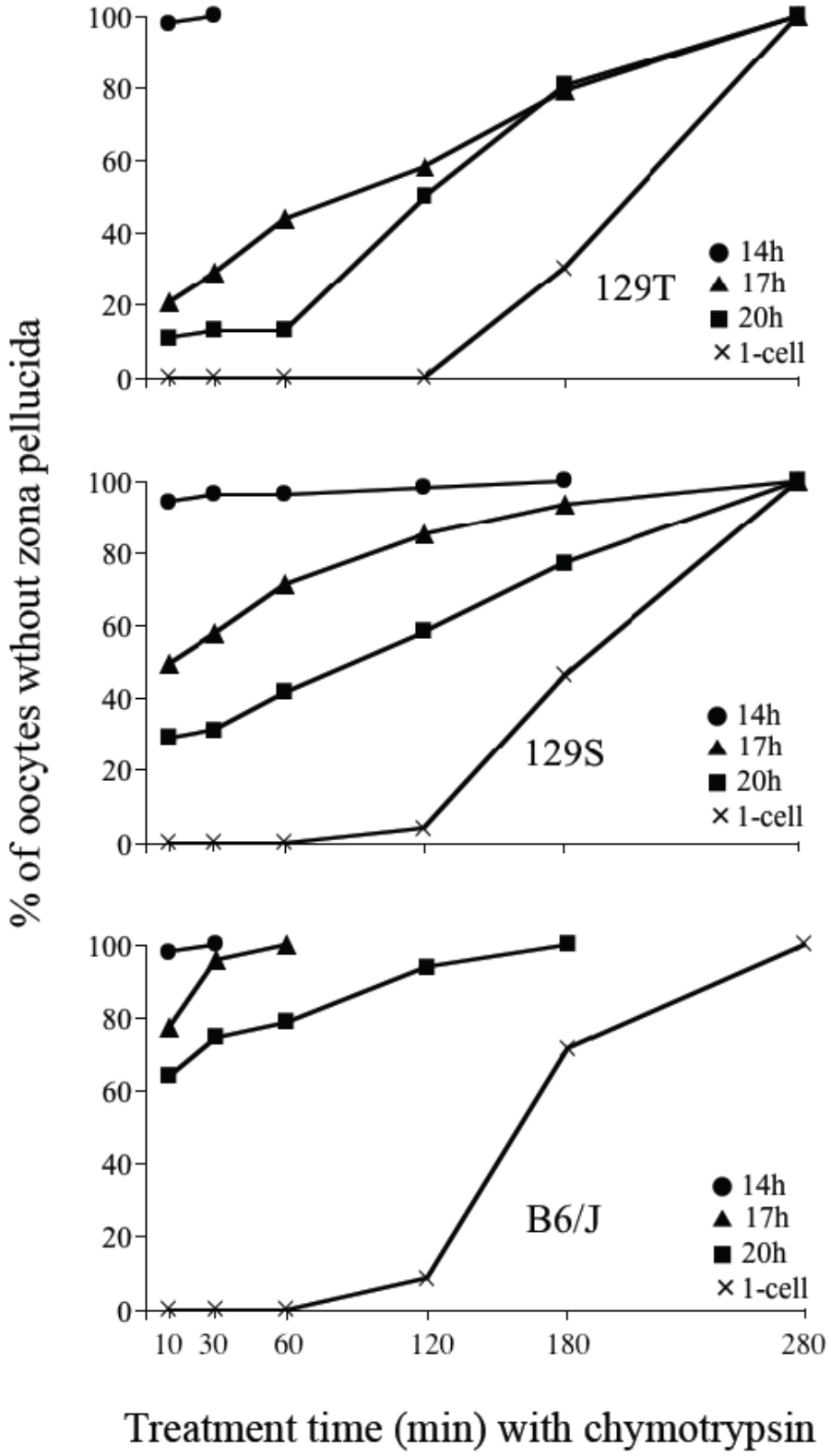
% of oocytes wthout zona pellucida
100
80
60
40
20
0
14h
17h
20h
1-cell
129T
129S
B6/J
10
30
60
120
180
280
Treatment time (min) with chymotrypsin

Figure 2