

## Dinucleotide repeat polymorphism within ERCC5 gene

S.Samec, S.G.Clarkson\*, J.Blaschak<sup>1</sup>, A.Chakravarti<sup>1</sup>, M.A.Morris, D.Scherly and S.E.Antonarakis

Department of Genetics and Microbiology, Centre Médical Universitaire (C.M.U.), 9 avenue de Champel, 1211 Geneva 4, Switzerland and <sup>1</sup>Department of Human Genetics, University of Pittsburgh, Pittsburgh, PA 15261, USA

**Source/Description:** A human genomic DNA library in EMBL3 was screened with a cDNA probe encoding the conserved I-region of ERCC5 (1), corrects the DNA repair defects in both xeroderma pigmentosum complementation group G and rodent ERCC group 5 (1–3). A 2.6 kb *EcoRI* fragment of one positive phage, phage #10, was found to contain a (CA)<sub>19</sub> repeat. The repeat and surrounding sequences (EMBL accession no. X75341) lie within an intron that disrupts the codon for Gly652.

### PCR Primers:

CA strand: 5'-GCAATGACTCGGTATTGG-3'  
GT strand: 5'-TGGATGCTAACAAAGTGGG-3'

### Polymorphism:

Allele	bp	Frequency	Allele	bp	Frequency
A1	277	.01	A7	265	.06
A2	275	.05	A8	263	.14
A3	273	.13	A9	261	.34
A4	271	.04	A10	257	.04
A5	269	.03	A11	245	.01
A6	267	.15			

Heterozygosity = 0.84 observed in 121 unrelated individuals in the CEPH reference pedigrees.

### Allele Identification in Selected CEPH Individuals:

13291-01: A3/A9 1333-01: A9/A11 1331-02: A7/A8  
1333-01: A9/A11 13291-02: A4/A10 1333-02: A2/A7

**Mendelian Inheritance:** Observed in all 40 CEPH pedigrees.

**Chromosomal Localization:** 13q33 by FISH.

**Mapping by Linkage Analyses:** Two-point linkage analysis in CEPH families showed no recombination with D13S158 ( $\theta = 0.00$ ; lod = 25.59). Multipoint analysis placed XPG between D13S122 and COL4A1 with odds > 1000:1.

**PCR Conditions:** The 22  $\mu$ l reactions contained 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl<sub>2</sub>, 50  $\mu$ M of each dNTP, 400 ng of genomic DNA, 0.75 pmoles of CA strand primer 5'-end labelled with [ $\gamma$ -<sup>32</sup>P]ATP by T4 polynucleotide kinase, 10 pmoles of CA strand primer, 20 pmoles of GT strand primer, and 1 unit of Taq polymerase (Perkin-Elmer/Cetus). Samples were heated for 7 min at 94°C then subjected to 35 cycles of 30 sec at 94°C, 30 sec at 51°C, and 45 sec at 72°C. Amplified products were electrophoresed through 6% denaturing polyacrylamide gels.

**Acknowledgements:** Supported by Swiss National Science Foundation grants 31-36481.92 to S.G.C. and 31-33965.92 to S.E.A., and NIH grant HG00344 to A.C.

**References:** 1) Scherly, D. *et al.* (1993) *Nature* 363, 182–185.  
2) O'Donovan, A. and Wood, R.D. (1993) *Nature* 363, 185–188.  
3) MacInnes, M.A. *et al.* (1993) *Mol. Cell. Biol.* 13, 6393–6402.

## Tetranucleotide repeat polymorphism at the D8S346 locus

Robyn Riley<sup>1</sup>, Lesa Nelson<sup>1</sup>, Jun Lu<sup>1</sup>, Margaret Robertson<sup>2</sup> and Kenneth Ward<sup>1,3,\*</sup>

<sup>1</sup>Department of Obstetrics and Gynecology, <sup>2</sup>Howard Hughes Medical Institute and <sup>3</sup>Department of Human Genetics, University of Utah School of Medicine, 50 North Medical Drive, Room 2B200, Salt Lake City, UT 84132, USA

**Source/Description:** The polymorphic (A<sub>3</sub>T) repeat KW400 was isolated from a *Sau* 3A subclone of a flow-sorted chromosome 8 specific cosmid library (LA08NC01) (1) by hybridization to an (A<sub>3</sub>T)<sub>20</sub> oligomer. The predicted length of the amplified sequence is approximately 240 bp (GenBank accession number L12264).

### PCR Primers:

KW400A 5'-TGC ATG ACA GCC TGG GTG ACA GAA  
KW400B 5'-CCA CAG GTG CCC ACG CTA AGA CA

**Frequency:** Estimated from 200 chromosomes of unrelated Caucasian individuals.

Observed Heterozygosity Index = 63%.

Allele	bp	Frequency	Allele	bp	Frequency
A1	252	0.005	A4	240	0.515
A2	248	0.025	A5	236	0.380
A3	244	0.065	A6	232	0.010
CEPH 884.1–5,5 884.2–3,4					

**Mendelian Inheritance:** Co-dominant segregation was observed in 5 informative Caucasian families.

**Chromosomal Localization:** KW400 has been assigned to chromosome 8q (Region I) by PCR of a somatic cell hybrid mapping panel (2).

**PCR Conditions:** PCR is performed in 25  $\mu$ l containing 200 ng DNA, 0.5  $\mu$ M of each primer, 1.5 mM MgCl<sub>2</sub>, 10 mM Tris-HCl (pH 8.4), 40 mM NaCl, 200  $\mu$ M of each dNTP, 0.25 mM spermidine HCl, 0.625 units of Taq polymerase (Ampli-Taq, Perkin-Elmer/Cetus), and 0.01  $\mu$ M of radiolabelled KW400B (5' end-labelled with  $\gamma$ -<sup>32</sup>P ATP (3,000 Ci/mmol) using 30  $\mu$ Ci/10 pmol primer). Initial denaturation was for 5 minutes at 94°C. Amplification was for 30 cycles with denaturation at 94°C for 30 seconds, annealing at 60°C for 30 seconds, and extension at 72°C for 60 seconds. The products were separated by electrophoresis on a 32% formamide, 5.7 M urea, 7% polyacrylamide denaturing gel and analysed by autoradiography.

**Acknowledgements:** This work was supported in part by a grant from the Shriner's Research Foundation (#15962).

**References:** 1) Evans, G.A., Lewis, K. and Rothenberg, B.E. (1989) *Gene* 79, 9–20. 2) Wagner, M.J., Ge, Y., Siciliano, M. and Wells, D.E. (1991) *Genomics* 10, 114–125.

\* To whom correspondence should be addressed