

## Increase of phenotypic variance in stressful environments

Hans Burla and Charles E. Taylor

**ABSTRACT:** The hypothesis that genetic homeostasis breaks down to yield greater genetic variance in more stressful environments was examined. Environmental stress was measured by mean development time and by wing length, it being expected that more stress generally gives rise to longer development times and shorter wings. The correlations between mean values and genetic variance were predicted to be positive for development time and negative for wing length. The correlations were not always statistically significant, but were in the predicted direction in 7 out of 8 tests. Quite possibly this phenomenon contributes to observed increases of genetic variation in marginal environments and more rapid evolution during periods of special stress.

IT WOULD SEEM plausible that populations of plants and animals show more genetic variation when in new or stressful environments than in familiar and favorable ones. This is because, as Lewontin<sup>3</sup> has stated: "The zone of canalization corresponds to the range of environments that have been historically the most common in the species, but in the new environments much greater variance appears." There is some evidence that this is the case. Developmental geneticists have long known it is easier to identify genetic variants with heat stress or ether<sup>2</sup>, and plant breeders generally recognize that differences among crop strains are more noticeable in stressed than in favorable environments<sup>1</sup>. But there is surprisingly little known about the extent that stress accentuates genetic differences. In evolutionary processes this might be important, for stress would lead to increased variation in ecologically marginal populations and increased rates of evolution during periods of special stress. We have attempted to explore further this phenomenon by measuring the association between genetic variation and environmental harshness in experimental populations of *Drosophila*.

For our analysis we used data that had been collected for another purpose and have been reported by Taylor and Condra<sup>4</sup>. Briefly, 12 strains of *Drosophila pseudoobscura* were raised on each of 10 different media and at two temperatures. The strains were only partly inbred, but each carried one or more visible mutants unique to it. Eggs were collected from spoons and then 100 were counted out onto a small piece of paper towel and placed on a shell vial containing 10 cc of medium. Larval crowding was minimal in these circumstances. The media was composed of our regular cornmeal-molasses medium, an

agar base without sugar, or the agar base supplemented with one sugar or another (maltose, mannitol, dextran, lactose, galactose, glucose, sucrose, or xylose). They were then stored at 15 or 25°C. The number of flies to emerge each day was recorded and the wing length of three males and three females from each vial was measured. There was a total 12 strains × 10 media × 2 temperatures × 5 replicates = 1200 vials.

Environmental stress was measured operationally by taking the mean value of the trait in question across all strains. If, on average, the flies took a longer time to develop or had smaller wings on one medium than that medium was regarded as more stressful than one in which the flies developed rapidly or achieved a larger size. Within either temperature the two measurements of stress used here were similar (see Table I). Nearly all correlations among measures were 0.80 or better.

Genetic variance was measured by first calculating the variance of mean values across all strains on the medium in question. From this value was subtracted the mean value of the variance within strains for that medium divided by the number of replicates. This is not quite the same as genetic variance per se, because the strains were not entirely inbred, but it should be approximately correct.

The relations between genetic variance and environmental stress are shown by correlation coefficients in Table II. Those media giving longer wings also produced a smaller genetic variance of wing length. Likewise, the genetic variance of development time and mean development time produced correlations from 0.21 to 0.93.

Using two-tailed tests not all correlations are statistically significant, but nearly all (7 out of 8) are in the direction expected. In an effort to eliminate scaling problems that might have been present we also divided the genetic variances by mean values of the traits and repeated the correlations. There were no substantial differences. Thus, it is evident that genetic variances were generally greater in environments that are more stressed.

The only exception to the general pattern of correlations was for male wing length at 25°C.

**Table I. Correlations among operational measures of environmental stress for laboratory populations of *Drosophila pseudoobscura*. Greater stress should be reflected by longer development times (for males, Daysm, and females, Daysf) and smaller wing sizes (for males, Wingsm, and females, Wingsf) at each temperature**

	25°C				15°C			
	Daysm	Daysf	Wingsm	Wingsf	Daysm	Daysf	Wingsm	Wingsf
25°C								
Daysm	1.00							
Daysf		1.00						
Wingsm	-0.80	-0.80	1.00					
Wingsf	-0.81	-0.82	0.92	1.00				
15°C								
Daysm	0.97	0.96	-0.78	-0.81	1.00			
Daysf	0.98	0.97	-0.76	-0.79	0.99	1.00		
Wingsm	-0.91	-0.91	0.92	0.96	-0.92	-0.91	1.00	
Wingsf	-0.89	-0.88	0.82	0.89	-0.94	-0.93	0.96	1.00

The authors are affiliated, respectively, with the Zoological Museum, University of Zürich, 8057, Zürich, Switzerland; and Department of Biology, University of California, Los Angeles, CA 90024. © 1982, American Genetic Association.

**Table II. Correlations between operational measures of environmental stress and genetic variation for laboratory populations of *Drosophila pseudoobscura***

	Development time		Wing length	
	males	females	males	females
15°C	0.91*	0.93*	-0.24	-0.82*
25°C	0.21	0.57	0.35	-0.24

\* P < 0.01

It appears that this was due principally to one medium, cornmeal-molasses, on which male flies have larger wings but also are more variable. When this medium is excluded the correlation drops from +0.35 to +0.03. Subsequent exclusions, however, had little effect.

In general, the correlations tended to be lower at 25°C than at 15°C. In view of the smaller body size, emergence, and reduced vigor of the flies at 25°C it would seem that *D. pseudoobscura* is more stressed at this temperature than at 15°C. Additional information would be needed to explain the role of temperature in this context. We speculate that 15°C is somewhat lower than the optimal temperature for *D. pseudoobscura* and that 25°C is higher. However, coldness and warmth differ in their physiological effect on the developing flies and cannot be expected to influence the functional dependence of genotypic expression on media in the same way.

### References

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