

European Heart Journal (2011) **32**, 680–685 doi:10.1093/eurheartj/ehq484 REVIEW

Basic science for the clinician

When are pro-inflammatory cytokines SAFE in heart failure?

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The cytokine hypothesis presently suggests that an excessive production of pro-inflammatory cytokines, such as tumour necrosis factor alpha (TNF) and interleukin 6 (IL6), contributes to the pathogenesis of heart failure. The concept, successfully proved in genetically modified animal models, failed to translate to humans. Recently, accumulation of apparently paradoxical experimental data demonstrates that, under certain conditions, production of pro-inflammatory cytokines can initiate the activation of a pro-survival cardioprotective signalling pathway. This novel path that involves the activation of a transcription factor, signal transducer and activator of transcription 3 (STAT3), has been termed the survival activating factor enhancement (SAFE) pathway. In this review, we will discuss whether targeting the SAFE pathway may be considered as a preventive and/or therapeutic measure for the treatment of heart failure.

Keywords

Heart failure • Pro-inflammatory cytokines • Tumour necrosis factor alpha • Signal transducer and activator of transcription • Prosurvival pathways

Introduction

In the 1990s, the introduction of the cytokine hypothesis proposed that an excessive production of pro-inflammatory cytokines contributes to heart failure (HF).^{1,2} Despite encouraging animal studies and small clinical trials, larger clinical trials targeting the cytokines in HF have failed. The guestion as to whether the increased levels of inflammatory cytokines observed with HF are a cause or consequence of the disease still remains unresolved. In the new millennium, data from numerous animal studies have lead to a better understanding of the beneficial vs. deleterious effect of pro-inflammatory cytokines in pathophysiological conditions. Recent experimental work currently suggests that activation of these cytokines, including tumour necrosis factor alpha (TNF) and the interleukin 6 family, can promote a pro-survival signalling pathway termed the SAFE (survivor activating factor enhancement) pathway to protect against myocardial infarction (MI).^{3,4} In this review, we will summarize evidence for and against the cytokine hypothesis in both experimental models and clinical conditions and we will discuss whether targeting the

SAFE pathway may be considered as a preventive and/or therapeutic measure for the treatment of HF.

Pro- and anti-inflammatory cytokines imbalance in heart failure

Pro-inflammatory cytokines in heart failure

The interest of TNF (also known as cachectin) in HF evolved from the observation that cachexia is a common phenomenon associated with severe HF.⁵ In the 1990s, numerous studies evidenced an increase in circulating TNF, function to the severity and the outcome of HF.^{6,7} The production of TNF in HF may originate from the periphery (liver) or the myocardium (see review⁸). In HF patients, cachexia was associated with a further increase in circulating TNF receptors type I and type II (TNFR1 and TNR2), through which TNF is believed to exert its function.⁶ Tumour necrosis factor receptor type I is

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considered as a major predictor of mortality and new-onset HF in patients with acute MI (C-ALPHA study). 9

The main components of the interleukin 6 family cytokines are interleukin 6 (IL6), leucaemia inhibitory factor (LIF), and cardiotrophin 1. In animal models and in man, these three cytokines are increased in HF. Similar to TNF, the circulating levels of IL6 increase with the severity of HF and it may be considered as a prognostic marker in patients with congestive HF.^{10,11} The production of IL6 in the disease may originate from either the periphery or the myocardium depending on the disease stage.^{11,12} In addition to TNF and IL6, myocardial gene expression of both cardiotrophin 1 and LIF also increase in the failing heart.^{13,14} Most of the effects associated with these cytokines occur after binding to the receptor gp130 which is also increased in HF, to a greater extent in patients with dilated cardiomyopathy than valvular or ischaemic cardiomyopathy. This suggests that the role of gp130 in the pathogenesis of chronic HF may vary as a function of the aetiology of HF.^{10,14}

These studies demonstrate that pro-inflammatory cytokines can be considered as a biomarker in HF. Unfortunately, levels of cytokines measured from one study to another may vary considerably, which is related essentially to the commercial kit/technique used (*Table 1*).

Table ITumour necrosis factor and interleukin 6levels in various pathophysiological conditions

In humans	TNF (pg/mL) mean	IL6 (pg/mL)
Heart failure (HF)		
Ferrari et al. ¹⁰⁴ (congestive heart failure)	Control: 14.3 ± 7; HF: 33.5 ± 13.1	
Milani <i>et al.</i> ⁴³ (acute decompensated chronic heart failure)	Control: 0.9 \pm 0.4; HF: 3.0 \pm 0.4	
Testa et al. ¹⁰² (congestive heart failure)	Range: 0.5–32	3–300
Petretta et al. ¹⁰³ (mild to severe CHF)	Class II: 8.3 ± 1.0	Class II: 0.7 \pm 0.7
	Class III: 6.0 \pm 1.6	Class III: 5.2 ± 1.3
	Class IV: 15.0 ± 1.1	Class IV: 5.6 ± 1.2
Stumpf et al. ¹⁶ (chronic heart failure)	Control: 2.5 ± 1.8; HF: 6.5 ± 2.9	
Hirota et <i>al.</i> ¹⁰ (congestive heart failure)		Control: 1.7 ± 1.6
		Class II: 3.8 \pm 2.0
		Class III, IV: 18.5 ± 11.8
Rheumatoid arthritis (RA)	
Paramalingam et <i>al</i> . (2007)	Control: 9; RA : 40	Control: 2; RA: 14
Septic shock (S)		
Damas et al. ¹⁰⁵	471	10049
Charalambos et al. ¹⁰⁶	With shock: 80 \pm 6; No shock: 61 \pm 6	With shock: 266 \pm 28; No shock: 142 \pm 28

Anti-inflammatory cytokines in heart failure

Overall, the production of anti-inflammatory cytokines has been much less studied than the pro-inflammatory cytokines in the human heart. The main anti-inflammatory cytokine studied in HF is interleukin 10 (IL10). Produced in mononuclear cells (essentially macrophages and T cells), IL10 inhibits the production of pro-inflammatory cytokines as well as the matrix metalloproteinases by activated monocytes.¹⁵ Compared with healthy control subjects, plasma levels of IL10 are decreased in patients with HF, in particular, in patients suffering from advanced HF NYHA class III and IV.¹⁶

Circulating thrombospondin-1, another anti-inflammatory cytokine released upon activation of platelets, is also reduced in HF, together with tumour growth factor $\beta 1.^{17,18}$

The cytokine hypothesis: evidence for and against

Evidence for a contribution of cytokines in the development and pathogenesis of the disease

The adverse effect of pro-inflammatory cytokines in the heart has been widely demonstrated in animal models, using mainly cytokine depleted or overexpressing mouse models. In experimental models, inflammatory cytokines promote left ventricular remodelling, acute reversible contractile dysfunction, and uncouple myocardial β -adrenergic receptors (see review¹⁹). Cardiac specific overexpression of TNF causes cardiac failure²⁰ while pathophysiologically relevant concentrations of TNF promote progressive left ventricular dysfunction and remodelling in rats.²¹ Interleukin 10 knockout animals exhibit increased mortality and larger infarct size compared with their littermate controls.²² All these data suggest that limiting the cytokine imbalance observed in pathophysiological conditions, such as HF may limit the development of the disease and improve its outcome. Such modulation of the cytokine balance in HF can be approached in two ways: (i) by enhancing the level of anti-inflammatory cytokines and (ii) by targeting a decrease in pro-inflammatory cytokines. These two approaches have been considered, to varying degrees, in both experimental models and clinical conditions.

Increase of anti-inflammatory cytokines

The concept of raising the anti-inflammatory cytokines in HF has been poorly studied. In animal studies, treatment with recombinant IL10 limits myocardial lesions in viral myocarditis in rats²³ and improves left ventricular function in rats with HF after experimental MI.²⁴ This effect was associated with a decrease in circulating inflammatory cytokines TNF and IL6 and a reduction in myocardial macrophage infiltration.²⁴ Similarly, chronic treatment with atorvastatin increases plasma levels of IL10, decreases the TNF/IL10 ratio, ameliorates left ventricular remodelling, and improves left ventricular function in rats with HF subsequent to MI.²⁵ In fact, the simplest and safest way to increase IL10 is with physical exercise. In a rat model of chronic HF, regular swimming improves

plasma levels of IL10 and cardiac function and reduces muscle cellular damage. $^{26}\,$

In human studies, specifically targeting an increase of IL10 in HF has been essentially ignored. Intense treatment with atorvastatin in patients with stable coronary disease reduces subsequent hospitalization for HF.²⁷ Immunomodulation therapy with intravenous immunoglobulin in patients with chronic HF improved the left ventricular ejection fraction and increased plasma IL10 levels.²⁸ A recent Cochrane systematic review and meta-analysis shows that exercise training, known to increase IL10 levels, reduces HF-related hospitalization and results in clinical improvement in health-related quality-of-life.²⁹ Similarly, the HF-ACTION study demonstrates that participation in an exercise training programme provides improvement in HF patients-reported health status compared with usual care.³⁰ Targeting the anti-inflammatory cytokines to limit the development of the pathology in HF certainly merits further investigation.

Decrease of pro-inflammatory cytokines

The concept of decreasing the pro-inflammatory cytokines has been more readily explored in both clinical and experimental settings but this has led to conflicting and confusing data.

In a viral myocarditis mouse model, anti-TNF improved survival and myocardial lesions.³¹ Tumour necrosis factor neutralization attenuated the impairment of left ventricular function 10 weeks after MI in rats.³² Similarly, contractile dysfunction was attenuated in TNF R1 knockout mice with MI³³ or with TNF antibodies after microembolization.³⁴ Conversely, adenoviral transfection of TNFR1 increased contractile dysfunction following MI.³⁵

In a small randomized preclinical trial with 18 patients of NYHA class III, a single intravenous infusion of the TNF antagonist Etanercept resulted in a decrease in TNF bioactivity and a significant overall increase in quality-of-life scores.³⁶ Similarly, pentoxifilline, a putative TNF inhibitor, was beneficial in small clinical trials with HF patients. Pentoxyfilline therapy of ischaemic cardiomypathy for 6 months improved the ejection fraction.³⁷ The benefit with regard to symptoms of HF and cardiac function was seen in all grades of severity of HF (class I–IV) and in patients with ischaemic and idio-pathic dilated cardiomyopathy.^{37,38} However, the beneficial effect has not consistently been associated with a reduction in pro-inflammatory cytokines, therefore suggesting that pentoxyfilline, which is a phosphodiesterase inhibitor, may protect independently of an immunomodulatory effect (see review³⁹).

Diet enriched with omega-3 polyunsaturated fatty acids (ω -3 PUFA) may decrease pro-inflammatory cytokines in patients with HF but further studies are needed to determine the optimal source and dosing of ω -3 PUFA.⁴⁰

Evidence against a contribution of cytokines in the disease

In contrast to the preclinical trials, large randomized placebo-controlled clinical trials with anti-TNF therapies were disappointing. No beneficial effect was observed with Etanercept in patients with chronic HF (NYHA class II–IV) as reported in RENEWAL, RENAISSANCE, and RECOVER (see review⁴¹). The ACCLAIM study tested the effect of immunomodulation therapy in patients with chronic HF. The rationale of the study was to have a non-specific but broad immunomodulatory effect by reducing pro-inflammatory cytokines and increasing anti-inflammatory cytokines.⁴² The trial did not show any significant reduction in mortality or cardiovascular-related hospitalization. Unfortunately, the levels of cytokine activation were not assessed in the study. Of note, two specific sub-group of patients, those without a previous history of MI and those with NYHA II had significant reduction in their primary endpoint therefore suggesting that this therapy may benefit certain subgroups with HF.

The beneficial effect of several drugs in HF also brings into question the contribution of pro-inflammatory cytokines in the disease. In patients hospitalized with advanced acutely decompensated congestive HF, traditional therapy leads to clinical improvement but no reduction in pro-inflammatory cytokine levels.⁴³ Similarly, treatment with amiodarone in patients with ischaemic cardiomyopathy is associated with an increase in circulating TNF levels but this increase is not associated with an adverse effect on survival.⁴⁴

Although the severity of HF correlates with an increase in plasma inflammatory cytokines levels in patients, it is important to mention that these levels are much lower compared with inflammatory diseases, such as septic conditions (*Table 1*). Does this mean that their contribution to the disease, if any, may be relatively modest? Or are there other explanations?

Pro-inflammatory cytokines initiate a cardioprotective signalling cascade in the heart: the survival activating factor enhancement pathway

With the new millennium, a large body of experimental work conducted in various animal models has brought more insights with respect to the disappointment of multiple clinical trials targeting the inflammatory cytokines in HF. In fact, activation of the immune system (with TNF and/or IL6) can promote the activation of an intrinsic cell survival signalling pathway that involves activation of a transcription factor, the signal transducer and activator of transcription 3 (STAT3). This path, recently discovered in MI, has been termed the SAFE (survivor activating factor enhancement) pathway.^{4,45}

Tumour necrosis factor and interleukin 6 cytokines can protect against ischaemia-reperfusion injury

As observed in HF, an increase in pro-inflammatory cytokines occurs in patients with MI and circulating IL6, TNF, and their respective receptors are increased further after reperfusion (see review⁴⁶). Both cytokines are thought to contribute to contractile dysfunction,⁴⁷⁻⁵⁰ most likely as a result of perturbation in calcium homoeostasis and formation of free radicals.^{51,52}

However, as early as 1998, there was experimental data disputing the contribution of TNF or IL6 to the damage associated with MI. Hence, exogenous addition of TNF protects against hypoxic injury in cardiomyocytes⁵³ and mice lacking both TNFR1 and TNFR2 are more susceptible to MI than their littermate controls.⁵⁴ Similarly, expression of IL6 occurs in the viable border zone of a myocardial infarct. $^{\rm 55}$

In fact, the cardioprotective effect of TNF is influenced by several factors including dose, sex, time, and type of receptors activated. Hence, TNF protects against ischaemia-reperfusion in a dose-dependent manner with small amounts of exogenous TNF (0.5 ng/mL, *in vitro*) given prior to ischaemia-reperfusion enhancing cell survival while higher concentrations (10–20 ng/mL, *in vitro*) are cytotoxic.^{3,56,57} Deficiency of TNFR1 protects the myocardium through IL6 following TNF infusion,⁵⁸ and the activation of the TNFR2 seems to convey the protective effect of TNF.^{59–61}

Both cytokines are also important components in the powerful protective effect of preconditioning, which fails in mice in the absence of either TNF or IL6.^{56,62-65} Tumour necrosis factor can mimic pre- or post-conditioning *in vitro* and *in vivo*.^{47,56,66-68}

Signal transducer and activator of transcription 3 is a downstream target of tumour necrosis factor and interleukin 6

Once TNF and IL6 bind to their specific receptors (TNFR and gp130), a common signalling path, called the Janus kinase (JAK)/STAT3 pathway can be activated (*Figure 1*). Janus kinases are a family of tyrosine kinases that are associated with the cytoplasmic domain of cytokine and growth factor receptors (including TNFR and gp130) and play a major role in transducing signals from the cytosol to the nucleus (see reviews^{69–71}). Upon activation of

the receptors, JAK proteins phosphorylate and create a docking site for STAT proteins, which in turn are activated by phosphorylation (Figure 1). While the mechanisms involved in IAK activation by the gp130 and TNFR1 have been clearly characterized, the activation of JAK 2 by TNFR2 still remains to be elucidated. Tumour necrosis factor receptor II does not contain an intrinsic protein tyrosine kinase but the phosphorylation of IAK2 by this receptor may occur via TNF receptor-associated factor 2 (TRAF2) or nuclear factor-kappaB activation, both components previously implicated in TNF-mediated cardioprotection.72,73 The STAT family of transcription factor proteins consists of seven identified members: only STAT3 will be considered in this review. Tyrosine phosphorylation of STAT3 enables it to homodimerize and translocate to the nucleus. Both tyrosine and serine phosphorylated STAT3 are also present within the mitochondria.^{74,75} Under physiological conditions, this pathway can be regulated by the suppressor of cytokine signalling (SOCS) proteins that serve as a negative regulator of JAK/STAT signalling. Activation of STAT3 induces the expression of SOCS which in turn binds to the tyrosine 757 residue of membrane receptor which is necessary for the docking of JAK and STAT3 at the receptors to activate the JAK/ STAT3 pathway (see reviews^{71,76}).

Activation of the SAFE pathway with TNF/JAK/STAT3 or IL6/ JAK/STAT3 signalling is required for the cardioprotective effect of ischaemic pre- and post-conditioning as neither TNF knockout, IL6 knockout or cardiomyocyte STAT3 knockout mice could be protected with a conditioning stimulus.^{62,63,77-79} Activation of

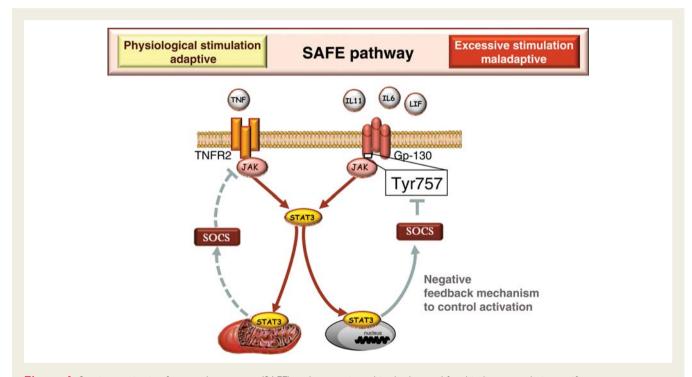


Figure I Survivor activating factor enhancement (SAFE) pathway activated in the heart. After binding onto their specific receptors, tumour necrosis factor and interleukin 6 family cytokines activate the janus kinase/signal transducer and activator of transcription 3 (JAK/STAT3) signalling path. After phosphorylation, signal transducer and activator of transcription 3 translocates either in the nucleus or in the mitochondria. The activation of the survivor activating factor enhancement pathway is regulated by the activation of the suppressor of cytokine signalling (SOCS). Moderate stimulation of the survivor activating factor enhancement pathway leads to cell survival but intense stimulation is detrimental. the JAK/STAT3 pathway also occurs with many other cardioprotective agents, such as erythropoietin, cannabinoid agonists, insulin, prostaglandins, and high density lipoproteins (HDL).^{4,80,81}

Downstream effectors of the SAFE pathway following cytokine and STAT3 activation still need to be defined. Several targets of STAT3 have been identified including proteins that are involved in cell survival and proliferation, such as Bcl-2, Bcl-xL, Mcl-1, Fas, cyclin D1, E1, p21, some growth factors (VEGF), and other transcription factors (see review⁴). In the preconditioning setting, STAT3 plays a critical role by increasing the anti-apoptotic gene Bcl-2 and reducing the pro-apoptotic gene bax.⁸² Also, STAT3 mediates cardioprotection via the phosphorylation and inactivation of the pro-apoptotic factor Bad.⁶⁴ In late preconditioning, STAT3 activates superoxide dismutase,⁸³ inducible nitric oxide synthase,⁶² and cyclooxygenase 2⁶² expression. Similarly, the JAK/STAT pathway protects in opioid-induced cardioprotection via phosphorylation and inactivation of glycogen synthase kinase 3B (GSK3 β).⁸⁴ Mitochondrial connexion 43, a key element of the signal transduction cascade of the protection by ischaemic preconditioning,⁸⁵ may also be a downstream target of STAT3.⁸⁶

By definition, STAT3 is a transcription factor but its effects observed in a setting of ischaemia/reperfusion or classic preconditioning do not seem to occur at a transcription level, as the time-frame between the activation of STAT3 and its action is too short to allow an effect at the gene level. This observation suggests that STAT3 has additional effects, such as direct phosphorylation of target molecules. With the recent discovery that STAT3 in preand post-conditioning is likely to be mediated by modulating the opening of the mitochondrial permeability transition pore.⁷⁵

Tumour necrosis factor can also promote cardioprotection via TRAF2, nuclear factor kappaB, reactive oxygen species, and protein kinase C.^{4,72,73} With regards to sphingolipids, TNF-induced cardioprotection is mediated via ceramide but sphingosine-1 phosphate protects via TNF and STAT3.^{3,87} Whether these intermediates form part of the SAFE pathway or constitute an alternative protective signalling path activated by TNF merits further investigation.

How to activate the survival activating factor enhancement pathway?

The SAFE pathway being recently discovered, few upstream targets have been delineated so far. Initially discovered as a downstream target of ischaemic preconditioning and ischaemic postconditioning^{64,67,88} (see *Figure 2*), it is now recognized that various cardio-protective agents can activate both TNF and STAT3, such as bradykinin, adrenoreceptors, leptin, opioids, and cannabinoids.⁸⁹ Similarly, HDL (whose low levels are associated with poor prognosis in patients with HF⁹⁰) or one of its components sphingosine-1 phosphate, confers protection via the activation of TNF and/or STAT3.^{80,87} Moderate consumption of red wine may also confer cardioprotection via the activation of the SAFE pathway. Hence, resveratrol and two biogenic amines present in red wine (ethanolamine and melatonin) protect the ischaemic isolated mouse heart against MI but this effect was lost in TNF knockout mice or STAT3 knockout mice.^{91,92} Surprisingly, adenosine, a well-known

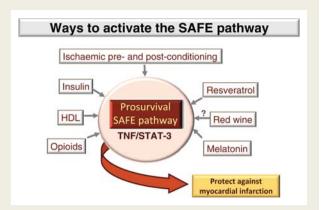


Figure 2 Multiple ways of activating the SAFE pathway to protect against myocardial infarction. SAFE, survivor activating factor enhancement; HDL, high density lipoprotein cholesterol.

preconditioning mimetic does not seem to activate the SAFE pathway, therefore confirming the existence of alternative cardioprotective signalling paths, such as the well-described reperfusion injury salvage kinase pathway or protein kinase G-dependent pathway.^{63,77,93}

Activation of the survival activating factor enhancement pathway in heart failure: a safe therapy to consider?

Evidence for a survival activating factor enhancement therapy in heart failure

As demonstrated in the previous section, experimental data provide evidence that activation of the SAFE pathway is protective in MI, but what about HF?

In humans, the concept that activation of the SAFE pathway may protect against HF is supported by the alteration in left ventricular STAT3 and gp130 proteins and phosphorylation of JAK observed in patients with dilated cardiomyopathy.^{14,94} The expression of SOCS is also attenuated in terminally failing human hearts.⁹⁴ Similarly, women with postpartum cardiomyopathies have a decrease in STAT3 levels compared with healthy women.⁹⁵

In experimental studies, although cardiomyocyte STAT3 deficient mice do not develop any cardiac abnormalities of function or morphology at a young age,⁹⁶ abnormalities appear with aging and severe cardiac fibrosis is observed at 6 months (see review⁷⁰). Female cardiomyocyte STAT3 deficient mice develop postpartum cardiomyopathy suggesting that activation of STAT3 is essential to protect the maternal heart from postpartum oxidative stress.⁹⁵ Mice with cardiomyocyte overexpression of STAT3 activate a hypertrophic signal but also a protective signal against doxorubicin-induced cardiomyopathy by inhibiting reduction in cardiac contractile gene expression and inducing cardiac protective factors.⁹⁷ Similarly, mice with cardiac restriction of gp130 display rapid onset of dilated cardiomyopathy and massive

induction of myocyte apoptosis during aortic pressure overload vs. the control mice that exhibit compensatory hypertrophy.⁹⁸ Also, activation of STAT3 by granulocyte colony stimulating factor in dilated cardiomyopathy improves survival and cardiac function.⁹⁹ Furthermore, activation of gp130/STAT3 with IL11 attenuates cardiac fibrosis following MI.¹⁰⁰

Evidence against a survival activating factor enhancement therapy in heart failure

These promising data in favour of a protective effect of the SAFE activation in HF should however be considered with precaution for several reasons. First, as mentioned earlier, TNF can activate two types of receptors, TNFR1 and TNFR2. Activation of the SAFE pathway involves the activation of TNFR2 only.⁶⁷ In experimental ischaemic HF, TNFR1 and TNFR2 have disparate and opposing effects on remodelling, hypertrophy, inflammation and apoptosis with TNFR1 exacerbating, and TNFR2 ameliorating these effects.^{59,61} Potential drugs targeting the activation of the SAFE pathway should therefore be specific to TNFR2. Second, continuous gp130-mediated IAK/STAT3 activation obtained by blockage of the tyrosine 757 on the gp130 site (and therefore inefficacy of SOCS to regulate the pathway) promotes cardiac inflammation, adverse remodelling and HF in mice with MI.¹⁰¹ These data suggest that an overstimulation of the SAFE pathway would lead to an adverse outcome. Third, the putative beneficial role of cytokines/STAT3 in ischaemic HF may not translate in non-ischaemic HF. The kinetics of production of cytokines varies as a function of the aetiology of HF with, for instance, gp130 increased to a greater extent in patients with dilated cardiomyopathy than ischaemic cardiomyopathy.¹⁴ Thus defining beneficial/deleterious roles of the SAFE pathway in HF will require intensive investigation taking into account the aetiology of the disease.

Conclusion

In HF, there is an undoubted imbalance between pro-inflammatory and anti-inflammatory cytokines. This imbalance correlates with the severity of the disease and the aetiology of HF. Animal studies conducted essentially on ischaemic HF models with overstimulation or repression of the pro-inflammatory cytokines are in favour of a causal role of these cytokines in the pathogenesis of HF. In contrast, while human studies suggest that measuring cytokines can be used as a biomarker for HF, they presently do not support a contribution of pro-inflammatory cytokines in the development of the disease. Over the last decade, new experimental data and human studies have demonstrated the Janus nature of cytokines, being either cytoprotective or detrimental. It underlines the dynamic aspect of cytokine action, where questions such as concentration, the time when produced and the type of receptors that they activate need to be addressed. The protective effect of the cytokines has been clearly demonstrated when there is a moderate activation of the SAFE pathway. Conversely, an uncontrolled activation of the SAFE pathway may be detrimental. Targeting a controlled activation of the SAFE pathway in HF is undoubtedly a valid therapeutic objective but the precise kinetic and bioactivity

characteristics of pro-inflammatory cytokines in either chronic or acute HF should be carefully delineated before this can be considered.

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References

- Seta Y, Shan K, Bozkurt B, Oral H, Mann DL. Basic mechanisms in heart failure: the cytokine hypothesis. J Card Fail 1996;2:243–249.
- Oral H, Kapadia S, Nakano M, Torre-Amione G, Lee J, Lee-Jackson D, Young JB, Mann DL. Tumor necrosis factor-alpha and the failing human heart. *Clin Cardiol* 1995;**18**:IV20–IV27.
- Lecour S, Smith RM, Woodward B, Opie LH, Rochette L, Sack MN. Identification of a novel role for sphingolipid signaling in TNF alpha and ischemic preconditioning mediated cardioprotection. J Mol Cell Cardiol 2002;34:509–518.
- Lecour S. Activation of the protective survivor activating factor enhancement (SAFE) pathway against reperfusion injury: does it go beyond the RISK pathway? J Mol Cell Cardiol 2009;47:32–40.
- Levine B, Kalman J, Mayer L, Fillit H, Packer M. Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. N Engl J Med 1990;323: 236-241.
- McMurray J, Abdullah I, Dargie HJ, Shapiro D. Increased concentrations of tumour necrosis factor in 'cachectic' patients with severe chronic heart failure. Br Heart J 1991;66:356–358.
- Deswal A, Petersen NJ, Feldman AM, Young JB, White BG, Mann DL. Cytokines and cytokine receptors in advanced heart failure : an analysis of the cytokine database from the Vesnarinone trial (VEST). *Circulation* 2001;**103**:2055–2059.
- Kleinbongard P, Heusch G, Schulz R. TNFalpha in atherosclerosis, myocardial ischemia/reperfusion and heart failure. *Pharmacol Ther* 2010;**127**:295–314.
- Valgimigli M, Ceconi C, Malagutti P, Merli E, Soukhomovskaia O, Francolini G, Cicchitelli G, Olivares A, Parrinello G, Percoco G, Guardigli G, Mele D, Pirani R, Ferrari R. Tumor necrosis factor-alpha receptor 1 is a major predictor of mortality and new-onset heart failure in patients with acute myocardial infarction: the Cytokine-Activation and Long-Term Prognosis in Myocardial Infarction (C-ALPHA) study. *Circulation* 2005;**111**:863–870.
- Hirota H, Izumi M, Hamaguchi T, Sugiyama S, Murakami E, Kunisada K, Fujio Y, Oshima Y, Nakaoka Y, Yamauchi-Takihara K. Circulating interleukin-6 family cytokines and their receptors in patients with congestive heart failure. *Heart* Vessels 2004;**19**:237–241.
- 11. Tsutamoto T, Hisanaga T, Wada A, Maeda K, Ohnishi M, Fukai D, Mabuchi N, Sawaki M, Kinoshita M. Interleukin-6 spillover in the peripheral circulation increases with the severity of heart failure, and the high plasma level of interleukin-6 is an important prognostic predictor in patients with congestive heart failure. J Am Coll Cardiol 1998;**31**:391–398.
- Kubota T, Miyagishima M, Alvarez RJ, Kormos R, Rosenblum WD, Demetris AJ, Semigran MJ, Dec GW, Holubkov R, McTiernan CF, Mann DL, Feldman AM, McNamara DM. Expression of proinflammatory cytokines in the failing human heart: comparison of recent-onset and end-stage congestive heart failure. *J Heart Lung Transplant* 2000;**19**:819–824.
- Eiken HG, Oie E, Damas JK, Yndestad A, Bjerkeli V, Aass H, Simonsen S, Geiran OR, Tonnessen T, Christensen G, Froland SS, Gullestad L, Attramadal H, Aukrust P. Myocardial gene expression of leukaemia inhibitory factor, interleukin-6 and glycoprotein 130 in end-stage human heart failure. *Eur J Clin Invest* 2001;**31**:389–397.
- Kurdi M, Booz GW. Can the protective actions of JAK-STAT in the heart be exploited therapeutically? Parsing the regulation of interleukin-6-type cytokine signaling. J Cardiovasc Pharmacol 2007;50:126–141.
- Silvestre JS, Mallat Z, Tamarat R, Duriez M, Tedgui A, Levy BI. Regulation of matrix metalloproteinase activity in ischemic tissue by interleukin-10: role in ischemia-induced angiogenesis. *Circ Res* 2001;89:259–264.

- Stumpf C, Lehner C, Yilmaz A, Daniel WG, Garlichs CD. Decrease of serum levels of the anti-inflammatory cytokine interleukin-10 in patients with advanced chronic heart failure. *Clin Sci (Lond)* 2003;**105**:45–50.
- Batlle M, Perez-Villa F, Lazaro A, Garcia-Pras E, Vallejos I, Sionis A, Castel MA, Roig E. Decreased expression of thrombospondin-1 in failing hearts may favor ventricular remodeling. *Transplant Proc* 2009;41:2231–2233.
- Sakata Y, Chancey AL, Divakaran VG, Sekiguchi K, Sivasubramanian N, Mann DL. Transforming growth factor-beta receptor antagonism attenuates myocardial fibrosis in mice with cardiac-restricted overexpression of tumor necrosis factor. *Basic Res Cardiol* 2008;**103**:60–68.
- Chen D, Assad-Kottner C, Orrego C, Torre-Amione G. Cytokines and acute heart failure. *Crit Care Med* 2008;36:S9–16.
- Kubota T, McTiernan CF, Frye CS, Slawson SE, Lemster BH, Koretsky AP, Demetris AJ, Feldman AM. Dilated cardiomyopathy in transgenic mice with cardiac- specific overexpression of tumor necrosis factor-alpha. *Circ Res* 1997; 81:627–635.
- Bozkurt B, Kribbs SB, Clubb FJ Jr, Michael LH, Didenko VV, Hornsby PJ, Seta Y, Oral H, Spinale FG, Mann DL. Pathophysiologically relevant concentrations of tumor necrosis factor- alpha promote progressive left ventricular dysfunction and remodeling in rats. *Circulation* 1998;97:1382–1391.
- Yang Z, Zingarelli B, Szabo C. Crucial role of endogenous interleukin-10 production in myocardial ischemia/reperfusion injury. *Circulation* 2000;**101**: 1019–1026.
- Nishio R, Matsumori A, Shioi T, Ishida H, Sasayama S. Treatment of experimental viral myocarditis with interleukin-10. *Circulation* 1999;100:1102–1108.
- Stumpf C, Seybold K, Petzi S, Wasmeier G, Raaz D, Yilmaz A, Anger T, Daniel WG, Garlichs CD. Interleukin-10 improves left ventricular function in rats with heart failure subsequent to myocardial infarction. *Eur J Heart Fail* 2008;**10**:733–739.
- Stumpf C, Petzi S, Seybold K, Wasmeier G, Arnold M, Raaz D, Yilmaz A, Daniel WG, Garlichs CD. Atorvastatin enhances interleukin-10 levels and improves cardiac function in rats after acute myocardial infarction. *Clin Sci* (*Lond*) 2009;**116**:45–52.
- Nunes RB, Tonetto M, Machado N, Chazan M, Heck TG, Veiga AB, Dall'Ago P. Physical exercise improves plasmatic levels of IL-10, left ventricular end-diastolic pressure, and muscle lipid peroxidation in chronic heart failure rats. J Appl Physiol 2008;104:1641–1647.
- Khush KK, Waters DD, Bittner V, Deedwania PC, Kastelein JJ, Lewis SJ, Wenger NK. Effect of high-dose atorvastatin on hospitalizations for heart failure: subgroup analysis of the Treating to New Targets (TNT) study. *Circulation* 2007;**115**:576–583.
- Gullestad L, Aass H, Fjeld JG, Wikeby L, Andreassen AK, Ihlen H, Simonsen S, Kjekshus J, Nitter-Hauge S, Ueland T, Lien E, Froland SS, Aukrust P. Immunomodulating therapy with intravenous immunoglobulin in patients with chronic heart failure. *Circulation* 2001;**103**:220–225.
- Davies EJ, Moxham T, Rees K, Singh S, Coats AJ, Ebrahim S, Lough F, Taylor RS. Exercise training for systolic heart failure: Cochrane systematic review and meta-analysis. *Eur J Heart Fail* **12**:706–715.
- Flynn KE, Pina IL, Whellan DJ, Lin L, Blumenthal JA, Ellis SJ, Fine LJ, Howlett JG, Keteyian SJ, Kitzman DW, Kraus WE, Miller NH, Schulman KA, Spertus JA, O'Connor CM, Weinfurt KP. Effects of exercise training on health status in patients with chronic heart failure: HF-ACTION randomized controlled trial. *Jama* 2009;**301**:1451–1459.
- Matsumori A, Sasayama S. Immunomodulating agents for the management of heart failure with myocarditis and cardiomyopathy—lessons from animal experiments. *Eur Heart J* 1995;16(Suppl. O):140–143.
- 32. Berthonneche C, Sulpice T, Boucher F, Gouraud L, de Leiris J, O'Connor SE, Herbert JM, Janiak P. New insights into the pathological role of TNF-alpha in early cardiac dysfunction and subsequent heart failure after infarction in rats. *Am J Physiol Heart Circ Physiol* 2004;**287**:H340–350.
- Ramani R, Mathier M, Wang P, Gibson G, Togel S, Dawson J, Bauer A, Alber S, Watkins SC, McTiernan CF, Feldman AM. Inhibition of tumor necrosis factor receptor-1-mediated pathways has beneficial effects in a murine model of postischemic remodeling. *Am J Physiol Heart Circ Physiol* 2004;287:H1369–1377.
- 34. Thielmann M, Dorge H, Martin C, Belosjorow S, Schwanke U, van De Sand A, Konietzka I, Buchert A, Kruger A, Schulz R, Heusch G. Myocardial dysfunction with coronary microembolization: signal transduction through a sequence of nitric oxide, tumor necrosis factor-alpha, and sphingosine. *Circ Res* 2002;**90**: 807–813.
- Monden Y, Kubota T, Inoue T, Tsutsumi T, Kawano S, Ide T, Tsutsui H, Sunagawa K. Tumor necrosis factor-alpha is toxic via receptor 1 and protective via receptor 2 in a murine model of myocardial infarction. *Am J Physiol Heart Circ Physiol* 2007;**293**:H743–753.

- Deswal A, Bozkurt B, Seta Y, Parilti-Eiswirth S, Hayes FA, Blosch C, Mann DL. Safety and efficacy of a soluble P75 tumor necrosis factor receptor (Enbrel, etanercept) in patients with advanced heart failure. *Circulation* 1999;99:3224–3226.
- Sliwa K, Skudicky D, Candy G, Wisenbaugh T, Sareli P. Randomised investigation of effects of pentoxifylline on left-ventricular performance idiopathic dilated cardiomyopathy. *Lancet* 1998;**351**:1091–1093.
- Sliwa K, Woodiwiss A, Kone VN, Candy G, Badenhorst D, Norton G, Zambakides C, Peters F, Essop R. Therapy of ischemic cardiomyopathy with the immunomodulating agent pentoxifylline: results of a randomized study. *Circulation* 2004;**109**:750–755.
- Shaw SM, Shah MK, Williams SG, Fildes JE. Immunological mechanisms of pentoxifylline in chronic heart failure. *Eur J Heart Fail* 2009;**11**:113–118.
- Lavie CJ, Milani RV, Mehra MR, Ventura HO. Omega-3 polyunsaturated fatty acids and cardiovascular diseases. J Am Coll Cardiol 2009;54:585–594.
- Anker SD, Coats AJ. How to RECOVER from RENAISSANCE? The significance of the results of RECOVER, RENAISSANCE, RENEWAL and ATTACH. Int J Cardiol 2002;86:123–130.
- 42. Torre-Amione G, Anker SD, Bourge RC, Colucci WS, Greenberg BH, Hildebrandt P, Keren A, Motro M, Moye LA, Otterstad JE, Pratt CM, Ponikowski P, Rouleau JL, Sestier F, Winkelmann BR, Young JB. Results of a nonspecific immunomodulation therapy in chronic heart failure (ACCLAIM trial): a placebo-controlled randomised trial. *Lancet* 2008;**371**:228–236.
- Milani RV, Mehra MR, Endres S, Eigler A, Cooper ES, Lavie CJ Jr, Ventura HO. The clinical relevance of circulating tumor necrosis factor-alpha in acute decompensated chronic heart failure without cachexia. *Chest* 1996;**110**:992–995.
- Oral H, Fisher SG, Fay WP, Singh SN, Fletcher RD, Morady F. Effects of amiodarone on tumor necrosis factor-alpha levels in congestive heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. *Am J Cardiol* 1999;83: 388–391.
- Lecour S. Multiple protective pathways against reperfusion injury: a SAFE path without Aktion? J Mol Cell Cardiol 2009;46:607–609.
- Kleinbongard P, Schulz R, Heusch G. TNFalpha in myocardial ischemia/reperfusion, remodeling and heart failure. *Heart Fail Rev* 2010.
- Skyschally A, Gres P, Hoffmann S, Haude M, Erbel R, Schulz R, Heusch G. Bidirectional role of tumor necrosis factor-alpha in coronary microembolization: progressive contractile dysfunction versus delayed protection against infarction. *Circ Res* 2007;**100**:140–146.
- 48. Yang S, Zheng R, Hu S, Ma Y, Choudhry MA, Messina JL, Rue LW III, Bland KI, Chaudry IH. Mechanism of cardiac depression after trauma-hemorrhage: increased cardiomyocyte IL-6 and effect of sex steroids on IL-6 regulation and cardiac function. Am J Physiol Heart Circ Physiol 2004;287:H2183–2191.
- Finkel MS, Oddis CV, Jacob TD, Watkins SC, Hattler BG, Simmons RL. Negative inotropic effects of cytokines on the heart mediated by nitric oxide. *Science* 1992;**257**:387–389.
- Pagani FD, Baker LS, Hsi C, Knox M, Fink MP, Visner MS. Left ventricular systolic and diastolic dysfunction after infusion of tumor necrosis factor-alpha in conscious dogs. J Clin Invest 1992;90:389–398.
- Yokoyama T, Vaca L, Rossen RD, Durante W, Hazarika P, Mann DL. Cellular basis for the negative inotropic effects of tumor necrosis factor-α in the adult mammalian heart. J Clin Invest 1993;92:2303-2312.
- Canton M, Skyschally A, Menabo R, Boengler K, Gres P, Schulz R, Haude M, Erbel R, Di Lisa F, Heusch G. Oxidative modification of tropomyosin and myocardial dysfunction following coronary microembolization. *Eur Heart J* 2006;27: 875–881.
- Nakano M, Knowlton AA, Dibbs Z, Mann DL. Tumor necrosis factor-alpha confers resistance to hypoxic injury in the adult mammalian cardiac myocyte. *Circulation* 1998;97:1392–1400.
- Kurrelmeyer KM, Michael LH, Baumgarten G, Taffet GE, Peschon JJ, Sivasubramanian N, Entman ML, Mann DL. Endogenous tumor necrosis factor protects the adult cardiac myocyte against ischemic-induced apoptosis in a murine model of acute myocardial infarction. *Proc Natl Acad Sci U S A* 2000; 97:5456–5461.
- Gwechenberger M, Mendoza LH, Youker KA, Frangogiannis NG, Smith CW, Michael LH, Entman ML. Cardiac myocytes produce interleukin-6 in culture and in viable border zone of reperfused infarctions. *Circulation* 1999;99:546–551.
- Deuchar GA, Opie LH, Lecour S. TNFalpha is required to confer protection in an in vivo model of classical ischaemic preconditioning. *Life Sci* 2007;80: 1686–1691.
- Lacerda L, McCarthy J, Mungly SF, Lynn EG, Sack MN, Opie LH, Lecour S. TNFalpha protects cardiac mitochondria independently of its cell surface receptors. *Basic Res Cardio* 2010;**105**:751–762.
- Wang M, Markel T, Crisostomo P, Herring C, Meldrum KK, Lillemoe KD, Meldrum DR. Deficiency of TNFR1 protects myocardium through SOCS3 and IL-6 but not p38 MAPK or IL-1beta. *Am J Physiol Heart Circ Physiol* 2007;292: H1694–1699.

- Hamid T, Gu Y, Ortines RV, Bhattacharya C, Wang G, Xuan YT, Prabhu SD. Divergent tumor necrosis factor receptor-related remodeling responses in heart failure: role of nuclear factor-kappaB and inflammatory activation. *Circulation* 2009;**119**:1386–1397.
- Crisostomo PR, Wang M, Herring CM, Markel TA, Meldrum KK, Lillemoe KD, Meldrum DR. Gender differences in injury induced mesenchymal stem cell apoptosis and VEGF, TNF, IL-6 expression: role of the 55 kDa TNF receptor (TNFR1). J Mol Cell Cardiol 2007;42:142-149.
- Schulz R, Heusch G. Tumor necrosis factor-alpha and its receptors 1 and 2: Yin and Yang in myocardial infarction? *Circulation* 2009;119:1355–1357.
- 62. Dawn B, Xuan YT, Guo Y, Rezazadeh A, Stein AB, Hunt G, Wu WJ, Tan W, Bolli R. IL-6 plays an obligatory role in late preconditioning via JAK-STAT signaling and upregulation of iNOS and COX-2. *Cardiovasc Res* 2004;**64**:61–71.
- Smith RM, Suleman N, McCarthy J, Sack MN. Classic ischemic but not pharmacologic preconditioning is abrogated following genetic ablation of the TNFα gene. *Cardiovasc Res* 2002;55:553–560.
- 64. Lecour S, Suleman N, Deuchar GA, Somers S, Lacerda L, Huisamen B, Opie LH. Pharmacological preconditioning with tumor necrosis factor-alpha activates signal transducer and activator of transcription-3 at reperfusion without involving classic prosurvival kinases (Akt and extracellular signal-regulated kinase). *Circulation* 2005;**112**:3911–3918.
- Smith RM, Lecour S, Sack MN. Innate immunity and cardiac preconditioning: a putative intrinsic cardioprotective program. *Cardiovasc Res* 2002;55:474–482.
- Lecour S, Rochette L, Opie L. Free radicals trigger TNFalpha-induced cardioprotection. *Cardiovasc Res* 2005;65:239–243.
- Lacerda L, Somers S, Opie LH, Lecour S. Ischaemic postconditioning protects against reperfusion injury via the SAFE pathway. *Cardiovasc Res* 2009;84: 201–208.
- Lacerda L, Smith RM, Opie L, Lecour S. TNFalpha-induced cytoprotection requires the production of free radicals within mitochondria in C(2)C(12) myotubes. Life Sci 2006.
- 69. Imada K, Leonard WJ. The Jak-STAT pathway. Mol Immunol 2000;37:1-11.
- Boengler K, Hilfiker-Kleiner D, Drexler H, Heusch G, Schulz R. The myocardial JAK/STAT pathway: from protection to failure. *Pharmacol Ther* 2008;**120**: 172–185.
- Kurdi M, Booz GW. JAK redux: a second look at the regulation and role of JAKs in the heart. Am J Physiol Heart Circ Physiol 2009;297:H1545–1556.
- Somers S, Lacerda L, Opie L, Lecour S. NFkB triggers TNF induced cardioprotection in C2C12. J Mol Cell Cardiol 2005;38:1040–1041.
- 73. Burchfield JS, Dong JW, Sakata Y, Gao F, Tzeng HP, Topkara VK, Entman ML, Sivasubramanian N, Mann DL. The cytoprotective effects of tumor necrosis factor are conveyed through tumor necrosis factor receptor-associated factor 2 in the heart. *Circ Heart Fail* 2010;**3**:157–164.
- 74. Wegrzyn J, Potla R, Chwae YJ, Sepuri NB, Zhang Q, Koeck T, Derecka M, Szczepanek K, Szelag M, Gornicka A, Moh A, Moghaddas S, Chen Q, Bobbili S, Cichy J, Dulak J, Baker DP, Wolfman A, Stuehr D, Hassan MO, Fu XY, Avadhani N, Drake JI, Fawcett P, Lesnefsky EJ, Larner AC. Function of mitochondrial Stat3 in cellular respiration. *Science* 2009;**323**:793–797.
- Boengler K, Hilfiker-Kleiner D, Heusch G, Schulz R. Inhibition of permeability transition pore opening by mitochondrial STAT3 and its role in myocardial ischemia/reperfusion. *Basic Res Cardiol* 2010;**105**:771–785.
- Fischer P, Hilfiker-Kleiner D. Role of gp130-mediated signalling pathways in the heart and its impact on potential therapeutic aspects. *Br J Pharmacol* 2008; 153(Suppl. 1):S414–S427.
- Smith RM, Suleman N, Lacerda L, Opie LH, Akira S, Chien KR, Sack MN. Genetic depletion of cardiac myocyte STAT-3 abolishes classical preconditioning. *Cardiovasc Res* 2004;63:611–616.
- 78. Dawn B, Guo Y, Rezazadeh A, Wang OL, Stein AB, Hunt G, Varma J, Xuan YT, Wu WJ, Tan W, Zhu X, Bolli R. Tumor necrosis factor-alpha does not modulate ischemia/reperfusion injury in naive myocardium but is essential for the development of late preconditioning. J Mol Cell Cardiol 2004;37:51–61.
- Xuan YT, Guo Y, Han H, Zhu Y, Bolli R. An essential role of the JAK-STAT pathway in ischemic preconditioning. *Proc Natl Acad Sci U S A* 2001;98: 9050–9055.
- Frias MA, Lang U, Gerber-Wicht C, James RW. Native and reconstituted HDL protect cardiomyocytes from doxorubicin-induced apoptosis. *Cardiovasc Res* 2010;85:118–126.
- Frias MA, James RW, Gerber-Wicht C, Lang U. Native and reconstituted HDL activate Stat3 in ventricular cardiomyocytes via ERK1/2: role of sphingosine-1phosphate. *Cardiovasc Res* 2009;82:313–323.
- Hattori R, Maulik N, Otani H, Zhu L, Cordis G, Engelman RM, Siddiqui MAQ, Das DK. Role of STAT3 in ischemic preconditioning. J Mol Cell Cardiol 2001; 33:1929–1936.

- Jung JE, Kim GS, Narasimhan P, Song YS, Chan PH. Regulation of Mn-superoxide dismutase activity and neuroprotection by STAT3 in mice after cerebral ischemia. J Neurosci 2009;29:7003–7014.
- 84. Gross ER, Hsu AK, Gross GJ. The JAK/STAT pathway is essential for opioid-induced cardioprotection: JAK2 as a mediator of STAT3, Akt, and GSK-3 beta. Am J Physiol Heart Circ Physiol 2006;291:H827-834.
- Schulz R, Boengler K, Totzeck A, Luo Y, Garcia-Dorado D, Heusch G. Connexin 43 in ischemic pre- and postconditioning. *Heart Fail Rev* 2007;12:261–266.
- 86. Ozog MA, Bernier SM, Bates DC, Chatterjee B, Lo CW, Naus CC. The complex of ciliary neurotrophic factor-ciliary neurotrophic factor receptor alpha up-regulates connexin43 and intercellular coupling in astrocytes via the Janus tyrosine kinase/signal transducer and activator of transcription pathway. *Mol Biol Cell* 2004;**15**:4761–4774.
- Somers S, Opie L, Lecour S. Sphingosine-1-phosphate (S1P) can mimic ischaemic postconditioning via activation of the STAT-3 pathway. *European Heart Journal* 2009;**30**:729.
- Suleman N, Somers S, Smith R, Opie LH, Lecour SC. Dual activation of STAT-3 and Akt is required during the trigger phase of ischaemic preconditioning. *Cardiovasc Res* 2008;**79**:127–133.
- Hausenloy DJ, Lecour S, Yellon DM. RISK and SAFE pro-survival signalling pathways in ischaemic postconditioning: two sides of the same coin. *Antioxid Redox Signal* 2010; doi:10.1089/ars.2010.3360. Published online ahead of print 26 October 2010.
- Mehra MR, Uber PA, Lavie CJ, Milani RV, Park MH, Ventura HO. High-density lipoprotein cholesterol levels and prognosis in advanced heart failure. J Heart Lung Transplant 2009;28:876–880.
- Kelly RF, Lamont KT, Somers S, Hacking D, Lacerda L, Thomas P, Opie LH, Lecour S. Ethanolamine is a novel STAT-3 dependent cardioprotective agent. *Basic Res Cardiol* 2010;**105**:763–770.
- Lamont K, Opie LH, Lecour S. Sleep your way to cardioprotection:red wine induced cardioprotection is mediated via melatonin and STAT3. J Mol Cell Cardiol 2010;48:S158–S159.
- Heusch G, Boengler K, Schulz R. Cardioprotection: nitric oxide, protein kinases, and mitochondria. *Circulation* 2008;**118**:1915–1919.
- Podewski EK, Hilfiker-Kleiner D, Hilfiker A, Morawietz H, Lichtenberg A, Wollert KC, Drexler H. Alterations in Janus kinase (JAK)-signal transducers and activators of transcription (STAT) signaling in patients with end-stage dilated cardiomyopathy. *Circulation* 2003;**107**:798–802.
- Hilfiker-Kleiner D, Kaminski K, Podewski E, Bonda T, Schaefer A, Sliwa K, Forster O, Quint A, Landmesser U, Doerries C, Luchtefeld M, Poli V, Schneider MD, Balligand JL, Desjardins F, Ansari A, Struman I, Nguyen NQ, Zschemisch NH, Klein G, Heusch G, Schulz R, Hilfiker A, Drexler H. A cathepsin D-cleaved 16 kDa form of prolactin mediates postpartum cardiomyopathy. *Cell* 2007;**128**:589–600.
- 96. Jacoby JJ, Kalinowski A, Liu MG, Zhang SS, Gao Q, Chai GX, Ji L, Iwamoto Y, Li E, Schneider M, Russell KS, Fu XY. Cardiomyocyte-restricted knockout of STAT3 results in higher sensitivity to inflammation, cardiac fibrosis, and heart failure with advanced age. *Proc Natl Acad Sci U S A* 2003;**100**:12929–12934.
- 97. Kunisada K, Negoro S, Tone E, Funamoto M, Osugi T, Yamada S, Okabe M, Kishimoto T, Yamauchi-Takihara K. Signal transducer and activator of transcription 3 in the heart transduces not only a hypertrophic signal but a protective signal against doxorubicin-induced cardiomyopathy. *Proc Natl Acad Sci U S A* 2000;**97**:315–319.
- Hirota H, Chen J, Betz UA, Rajewsky K, Gu Y, Ross J Jr, Muller W, Chien KR. Loss of a gp130 cardiac muscle cell survival pathway is a critical event in the onset of heart failure during biomechanical stress. *Cell* 1999; 97:189–198.
- Miyata S, Takemura G, Kawase Y, Li Y, Okada H, Maruyama R, Ushikoshi H, Esaki M, Kanamori H, Li L, Misao Y, Tezuka A, Toyo-Oka T, Minatoguchi S, Fujiwara T, Fujiwara H. Autophagic cardiomyocyte death in cardiomyopathic hamsters and its prevention by granulocyte colony-stimulating factor. *Am J Pathol* 2006;**168**:386–397.
- 100. Obana M, Maeda M, Takeda K, Hayama A, Mohri T, Yamashita T, Nakaoka Y, Komuro I, Matsumiya G, Azuma J, Fujio Y. Therapeutic activation of signal transducer and activator of transcription 3 by interleukin-11 ameliorates cardiac fibrosis after myocardial infarction. *Circulation* 2010;**121**:684–691.
- 101. Hilfiker-Kleiner D, Shukla P, Klein G, Schaefer A, Stapel B, Hoch M, Muller W, Scherr M, Theilmeier G, Ernst M, Hilfiker A, Drexler H. Continuous glycoprotein-130-mediated signal transducer and activator of transcription-3 activation promotes inflammation, left ventricular rupture, and adverse outcome in subacute myocardial infarction. *Circulation* 2010;**122**:145–155.
- 102. Testa M, Yeh M, Lee P, Fanelli R, Loperfido F, Berman JW, LeJemtel TH. Circulating levels of cytokines and their endogenous modulators in patients with mild to severe congestive heart failure due to coronary artery disease or hypertension. J Am Coll Cardiol 1996;28:964–971.

- 103. Petretta M, Condorelli GL, Spinelli L, Scopacasa F, de Caterina M, Leosco D, Vicario ML, Bonaduce D. Circulating levels of cytokines and their site of production in patients with mild to severe chronic heart failure. *Am Heart J* 2000; 140:E28.
- 104. Ferrari R, Bachetti T, Confortini R, Opasich C, Febo O, Corti A, Cassani G, Visioli O. Tumor necrosis factor soluble receptors in patients with various degrees of congestive heart failure. *Circulation* 1995;**92**:1479–1486.
- Damas P, Ledoux D, Nys M, Vrindts Y, De Groote D, Franchimont P, Lamy M. Cytokine serum level during severe sepsis in human IL-6 as a marker of severity. Ann Surg 1992;215:356–362.
- 106. Gogos CA, Drosou E, Bassaris HP, Skoutelis A. Pro- versus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. J Infect Dis 2000;181:176-180.