

TIIA MÖLLER

Mapping and modelling of
the spatial distribution of benthic
macrovegetation in the NE Baltic Sea
with a special focus on the eelgrass
Zostera marina Linnaeus, 1753



TIIA MÖLLER

Mapping and modelling of
the spatial distribution of benthic
macrovegetation in the NE Baltic Sea
with a special focus on the eelgrass
Zostera marina Linnaeus, 1753



Estonian Marine Institute and Department of Zoology, Institute of Ecology and Earth Sciences, Faculty of Science and Technology, University of Tartu, Estonia

The thesis was accepted for the commencement of the degree of *Doctor philosophiae* in hydrobiology at the University of Tartu on 8 May 2017 by the Scientific Council of the Institute of Ecology and Earth Sciences, University of Tartu.

Supervisor: Ph.D. Georg Martin, University of Tartu, Estonia
Ph.D. Jonne Kotta, University of Tartu, Estonia

Opponent: Ph.D. Marianne Holmer, University of Southern Denmark,
Denmark

Commencement: Room 301, 46 Vanemuise Street, Tartu, on 8 September 2017
at 10.15 a.m.

Publication of this thesis is granted by the Institute of Ecology and Earth Sciences, University of Tartu and by the Estonian Marine Institute, University of Tartu.

ISSN 1024-6479
ISBN 978-9949-77-471-5 (print)
ISBN 978-9949-77-472-2 (pdf)

Copyright: Tiia Möller, 2017

University of Tartu Press
www.tyk.ee

*“... I signed onto a sailing ship
My very first day at sea
I seen the Mermaid in the waves,
Reaching out to me
Come live with me in the sea said she,
Down on the ocean floor
And I’ll show you a million wonderous things
You’ve never seen before...”*

*Excerpt from the poem “The Mermaid”
by Shel Silverstein*

CONTENTS

LIST OF ORIGINAL PUBLICATIONS	8
1. INTRODUCTION.....	10
1.1. Direct observation, optical remote sensing and spatial modelling methods for mapping marine benthic habitats.....	11
1.2. Seagrasses in the changing environment.....	13
Aims of the thesis.....	15
2. MATERIAL AND METHODS	16
2.1. Study area	16
2.2. Field sampling	17
2.3. Environmental data used	17
2.4. Statistics and modelling.....	19
3. RESULTS AND DISCUSSION	21
3.1. In situ mapping of benthic shallow water communities in the NE Baltic Sea and the spatial spread of the eelgrass habitats.....	21
3.2. Linking spatial scale, observation method and community perception	23
3.3. Eelgrass community characteristics.....	24
3.4. Mapping eelgrass communities using remote sensing techniques.....	29
3.5. Modelling current patterns and possible futures of eelgrass	31
4. CONCLUSIONS.....	36
SUMMARY IN ESTONIAN	38
ACKNOWLEDGEMENTS	41
REFERENCES.....	43
PUBLICATIONS	53
CURRICULUM VITAE	143
ELULOOKIRJELDUS.....	145

LIST OF ORIGINAL PUBLICATIONS

This thesis is based on the following papers, which are referred to in the text by Roman numerals. The papers are reproduced by kind permission of the publishers: Estonian Academy Publishers (I, II), John Wiley and Sons (III), Springer (IV) and Elsevier (V).

- I. Martin, G., Kotta, J., Möller, T., Herkül, K. 2013. Spatial distribution of marine benthic habitats in the Estonian coastal sea, northeastern Baltic Sea. *Estonian Journal of Ecology* 62, 165–191.
- II. Möller, T., Kotta, J., Martin, G. 2014. Spatiotemporal variability in the eelgrass *Zostera marina* L. in the north-eastern Baltic Sea: Canopy structure and associated macrophyte and invertebrate communities. *Estonian Journal of Ecology* 63, 90–108.
- III. Möller, T., Kotta, J., Martin, G. 2009. Effect of observation method on the perception of community structure and water quality in a brackish water ecosystem. *Marine Ecology* 30, 105–112.
- IV. Vahtmäe, E., Kutser, T., Kotta, J., Pärnoja, M., Möller, T., Lennuk, L. 2012. Mapping Baltic Sea shallow water environments with airborne remote sensing. *Oceanology* 52, 803–809.
- V. Kotta, J., Möller, T., Orav-Kotta, H., Pärnoja, M. 2014. Realized niche width of a brackish water submerged aquatic vegetation under current environmental conditions and projected influences of climate change. *Marine Environmental Research* 102, 88–101.

Authors' contribution to the papers was as follows:

	I	II	III	IV	V
Original idea	GM, JK	TM, JK	TM, JK	EV, TK, JK	JK
Study design	JK, GM, TM, KH	TM	TM, JK	TK, JK	JK, TM
Data collection	MG, JK, TM, KH,*	TM, *	TM, *	EV, TK, MP, TM, *	JK, TM, *
Data analyses	JK, KH, TM	TM	TM, JK	EV, MP	JK, TM, HOK, MP
Manuscript preparation	GM, JK, TM, KH	TM, JK, GM	TM, JK, GM	EV, TK, JK, MP, TM, LL	JK, TM

TM Tiia Möller
GM Georg Martin
JK Jonne Kotta
KH Kristjan Herkül
EV Ele Vahtmäe
TK Tiit Kutser
MP Merli Pärnoja
LL Lennart Lennuk
HOK Helen Orav-Kotta
***** Co-workers

The author's contribution to papers II and III was substantial including the generation of the original idea, data collection and analyses and manuscript preparation. As for publications I and V the author was most involved in working out the study design, data collection and analysis processes and writing the manuscript. In publication IV the author's main activities were participation in the data collection and manuscript preparation.

1. INTRODUCTION

Our planet is changing fast, which is primarily caused by human activity. Since the 18th century and particularly after the industrial breakthrough in the 1850s the variety and speed of changes in the environment have grown exponentially in parallel with the growth of economy and human population. Oceans and seas cover over 70% of the Earth's surface and host an extraordinarily rich biodiversity (Gjerde, 2006). Nearshore areas in particular are among the most valuable biomes and responsible for 90% of the world's marine primary production (Kaiser et al., 2011). At the same time it is estimated that 90% of the oceans are unexplored (Gjerde, 2006). Vastness, opacity and stratification are the main reasons why scientific knowledge on the marine environment is sparse compared to the terrestrial environment, given also that the marine environment is harsh to access, observe and collect samples from (Norse and Crowder, 2005; Robinson et al., 2011). The gradual increase and diversification in the use of coastal natural resources jeopardize the stability of the marine environment and have resulted in a necessity for stock-taking and protective measures for a variety of marine species (Halpern et al., 2008). Obtaining information on the distribution of coastal marine species has become an important goal.

Coastal habitats (e.g. seagrass beds, kelp beds, saltmarshes, coral reefs, mussel beds, macroalgal beds, rocky bottom, and mariculture beds) have high ecological value (Moberg and Folke, 1999; Seitz et al., 2014). Among these seagrasses form a diverse ecological group of phanerogams that inhabit about 10% of the intertidal and shallow sublittoral areas along temperate and tropical coastlines (den Hartog, 1970; Larkum et al., 2006). They form extensive meadows in sheltered and semiexposed near-coastal zones (Reusch et al., 2005; Larkum et al., 2006), which are among the most productive habitats worldwide (Duarte, 2002). Furthermore, these meadows provide a range of ecological functions such as coastline protection, sediment stabilization, wave attenuation, land-derived nutrient filtration and carbon fixation, just to name a few; thereby providing some of the most valuable ecosystem services on the Earth (Costanza et al., 1997; Short et al., 2011; Nordlund et al., 2016). Seagrasses are also regarded as ecosystem engineering species because they are important as food, shelter and space for a large number of invertebrates and fishes, many of which are socio-economically important (Hemminga and Duarte, 2000; Seitz et al., 2014).

This thesis gives a basic overview of main methods applied in marine benthic communities research and investigates the distribution of a soft-bottom habitat-forming seagrass species in relation to abiotic and biotic forcings in the NE Baltic Sea. The Baltic Sea is among the largest semi-enclosed brackish inland seas in the world, it is a vulnerable ecosystem and the predicted increase in the diversity and intensity of anthropogenic pressures together with climate change will challenge all the aquatic species (Elmgren, 1989; Elmgren, 2001; Koch et al., 2013). Prior to this thesis, the published background information on the seagrass communities in the Baltic Sea was very scattered (e.g. Boström et al.,

2003; Möller and Martin, 2007) and owing to differences in methodologies there was no way to systematize such knowledge.

1.1. Direct observation, optical remote sensing and spatial modelling methods for mapping marine benthic habitats

To date a number of quantitative methods have been developed to collect information on marine benthic species and map benthic macrophyte communities (for more detailed overview, see e.g. Eleftheriou, 2014). The current methods span from local small-scale mapping to regional remote sensing and modelling exercises. In situ methods allow investigating marine benthic communities with very high precision. These methods are currently considered the only techniques to provide true data on the actual taxonomic composition of the seafloor. The main methods include diving to survey and sample the underwater environment and/or sampling the benthic habitat remotely using benthic grabs. These basic sampling methods have remained fairly unchanged since they came into use. When compared with other indirect methods, the cost of diving and grab sampling is the highest as it is time consuming, requires special equipment and skills and the explored area per time unit is small. Moreover, the laboratory work associated to sample analyses is also time consuming and expensive. Due to this, the studies usually result in a low number of observations within an investigated region and may fail to give accurate estimates at the seascape/regional level. With the introduction of digital video cameras with greatly improved durability and video quality, filming and assessing benthic communities along transects has become an increasingly practical and popular method (Murdoch and Aronson, 1999; Riegl et al., 2001; Houk and Van Woessik, 2006; Bucas et al., 2007; Mallet and Pelletier, 2014). Videos are most useful when the absolute accuracy is not needed, e.g. for mapping the distribution of key species or habitats.

However, as direct benthic habitat mapping is very expensive and time consuming, it is still impossible to get maps covering large seascapes by relying solely on ground-based data. Here remote sensing from aircraft and space-based platforms offers unique large-scale mapping possibilities. Remote sensing is widely used in the terrestrial environment (Townshend and Justice, 2002). Its use in aquatic ecosystems remains challenging due to a strong absorbance of the water medium, but the availability of environmental data gathered via remote sensing has significantly increased also in the marine environment (Brown et al., 2011). High concentrations of optically active substances in the water column complicate the mapping of benthic substrates and communities in eutrophied coastal areas. Nevertheless, the main macroalgal groups (green, brown and red algae) are distinguishable from one another in shallow areas (Vahtmäe et al., 2006). As multispectral instruments have limited usability (e.g. Kutser et al. 2006), airborne hyperspectral instruments have been widely tested and offer

new possibilities of mapping large seascapes with reasonable taxonomic resolution also in the Baltic Sea coastal area.

Due to difficulties faced in studying the marine environment, the knowledge on the distribution of different habitats in the sea is still comparatively fragmented. In order to promote sustainable coastal zone management the knowledge on the spatio-temporal distribution of marine habitats, communities and species needs to be increased. This in turn demands harmonized large-scale benthic habitat mapping. Large-scale analysis of spatial patterns of coastal marine habitats makes it possible to adequately estimate the status of coastal marine habitats, provide better evidence for environmental changes and describe processes that are behind the changes. To identify the most important governing factors one needs to determine also the scales at which communities have the largest variability and where the links between environmental and biotic patterns are the strongest (Platt and Denman, 1975; Steele and Henderson, 1994). Benthic communities have high structural variability at a multitude of scales and this variability is closely linked with physical setting. Therefore the mapping studies should incorporate the relevant scales of variability to understand factors and processes generating patterns in biotic and abiotic components of ecosystems (Menge and Olson, 1990; Levin, 1992; Karlson and Cornell, 1998).

Most of our knowledge on marine species and habitats is based on small-scale studies, which is not bad, as for the effective management and conservation of coastal ecosystems as well as for assessing the impacts of human activities the understanding of the spatial distribution of biota at local scales (10s or 100s of metres) is especially required (Kotta et al., 2008c). Yet, even if extensive distribution mapping has been conducted, such campaigns are often constrained to country case studies. Up-to-date regional overviews on the distribution and ecology of marine species are still rare, but their compilation is an emerging trend. Marine ecosystems have been deteriorating in status over the last decades, which points to the need for regional management efforts that carefully consider local and regional stressors when trying to offset human-caused impacts. Much of effective management, however, involves regional databases on the environment and human uses.

The urgent need for large-scale spatial data on benthic species and communities has intensified the evolving of different distribution modelling techniques that are able to describe ecological systems and predict their future behaviour (e.g. Müller et al., 2009; Reiss et al., 2014). In the spatial modelling field, point data are first collected by e.g. a diver or a video device. These data are used to build ecologically meaningful functional form relationships between the environment and biota and then these relationships are used to predict species patterns at large seascapes. However, many of these models perform poorly because very little is known about how organisms might interactively respond to multiple pressures, e.g. of natural and anthropogenic origin (Hoffman et al., 2003; Reynaud et al., 2003) and it is difficult to deal with complex and non-linear systems, such as those seen in the marine environment (see Byrne and Przeslaswki (2013) for an overview). Novel machine learning methods

mainly use an algorithm to discover the relationship between the response and its predictors (Hastie et al., 2009) and automatically handle interaction effects between predictors. Due to their strong predictive performance, such machine learning methods are increasingly used in ecological studies (Elith et al., 2008; Reiss et al., 2014).

1.2. Seagrasses in the changing environment

The key environmental variables affecting the distribution of seagrasses are light climate (Peralta et al., 2002; Krause-Jensen et al., 2008), temperature (Perez-Llorens and Niell, 1993; Marba et al., 1996; Glemarec et al., 1997) and nutrient concentrations in the water column (Orth, 1977) and/or in the sediment (Viarioli et al., 1997). Also hydrodynamic conditions (Schanz and Asmus, 2003), nature of the substrate (Viarioli et al., 1997; De Boer, 2007) and salinity (Wortmann et al., 1997) are important factors. In addition to these, ice conditions are also significant in high-latitude regions (Robertson and Mann, 1984).

In recent decades seagrasses have gone through an unusually fast transition in terms of areal decline in habitat and loss of species richness (Orth et al., 2006; Waycott et al., 2009; Short et al., 2011). The composition of communities was formerly seen as the outcome of local-scale processes, in recent decades this view has been challenged by emphasizing the importance of large-scale processes, including climate change, which may result in dramatic shifts in species distribution patterns and thereby affect community species composition, diversity, structure and productivity (Hawkins et al., 2013). Overviewed also in Paper V, according to most climate change scenarios, mean global surface temperatures will rise by 1.4–4.0 °C in the next 100 years and northern high-latitude regions are expected to experience more severe warming compared to low-latitude regions (IPCC, 2013). On average the prognoses for the Baltic Sea area predict a 5 °C rise in temperature in winter and a 4 °C rise in summer by the end of the century. The increase in temperature would significantly reduce the ice extent (>50%) and therefore indirectly amplifying the effects of wind stress. Furthermore, it is also expected that the mean daily wind speed over sea areas would increase by up to 18% in winter. Also the average salinity of the Baltic Sea is projected to decrease by 25% of the recent level (BACC, 2008). Such shifts plausibly result in the doubling of phytoplankton biomass (Hense et al., 2013). In addition, water transparency is expected to be reduced as a function of water salinity (Stramska and Swirgon, 2014).

Future water temperature likely dictates the distribution change of seagrasses as increasing temperature causes steady decrease of the photosynthesis to respiration ratio in most seagrass species (Marsh et al., 1986; Zimmerman et al., 1989; Glemarec et al., 1997). In addition, physical disturbance by intensifying heavy storms reduces seagrass cover and increases fragmentation within seagrass beds (Fonseca and Bell, 1998; Fonseca et al., 2000). At northern latitudes, also ice-scouring destroys submerged aquatic vegetation (Robertson and Mann, 1984;

Schneider and Mann, 1991), but contemporary climate change may release vegetation from such a disturbance. Also discussed in Paper V, due to non-linear responses of biota to the environment, even gradual changes in future anthropogenic pressures and/or climate may provoke sudden and perhaps unpredictable shifts in seagrass communities that grow at their physiological tolerance limit. Present worldwide estimates suggest already a 30% reduction of areal cover of seagrasses over the past 30 years (Waycott et al., 2009).

Out of the 72 known seagrasses (Short et al., 2011), only four species are found in the Baltic Sea. Due to diluted salinity conditions only three, the eelgrass *Zostera marina* Linnaeus, 1753, the widgeongrass *Ruppia maritima* Linnaeus, 1753 and the ditchgrass *Ruppia cirrhosa* (Petagna) Grande, prevail in the northeastern part of the Baltic Sea. Eelgrass is the only higher plant species of marine origin within the region as species of *Ruppia* are best described as salt-tolerant freshwater species. Eelgrass is the most wide-ranging marine flowering plant in the Northern Hemisphere (den Hartog, 1970) and it is restricted to the cooler waters in the North Atlantic, North Pacific and is also present in the Arctic region. In the Baltic Sea the species grows mainly in sandy sediments in moderately exposed bays and is totally submerged. The plant is rhizomatous with long, green, ribbon-like leaves and a rhizome growing horizontally through the substrate. Eelgrass is a perennial plant, but some populations may be annual. In the inner Baltic Sea eelgrass reproduces vegetatively, flowering is rare (den Hartog, 1970).

Presently the distribution area of eelgrass in the Baltic Sea is estimated at 1200 km² as a minimum (Boström et al., 2014). In Scandinavia, large-scale losses of eelgrass have been documented in Denmark since the 1900s and at smaller scales in the Swedish West coast and in Poland since the mid-1980s (Baden et al., 2003; Boström et al., 2003, overview in Boström et al., 2014). The shoot density of eelgrass has significantly declined in time with more severe effects recorded at deeper areas. Such losses were generally recorded in nutrient-rich areas and indicates the importance of the interactive effect of light climate and eutrophication on seagrass (Boström et al., 2014 and references therein). The disappearance of seagrass is very dramatic for the Baltic Sea ecosystem as here eelgrass is largely the only phanerogam on moderately exposed sandy habitats. Eelgrass meadows are one of the most diverse coastal habitats in the Baltic Sea – on otherwise species-poor sandy substrates, eelgrass promotes floral and faunal richness within its canopy (see also overview in Boström et al., 2014; Baden and Boström, 2001; Fredriksen et al., 2005; Jephson et al., 2008, II), which, in turn, supports diverse fish communities (Pihl et al., 2006). In addition, the complex root systems facilitate the existence of diverse infaunal communities (Boström et al., 2002; Fredriksen et al., 2010). Moreover, when the detached eelgrass is washed on the coast, numerous insects and other invertebrates inhabit the wrack (Jedrzejczak, 2002). Thus, any significant loss transfers to the reduced functioning of the coastal ecosystems of the Baltic Sea. The failure of eelgrass to re-establish despite reduction in background nutrient levels signals complex recovery trajectories and calls for much greater

conservation effort to protect existing eelgrass meadows. To prevent further loss of eelgrass, region-specific management and monitoring actions are also needed to identify and control the local loss drivers (Boström et al., 2014). Although eelgrass is listed as ‘Least concerned’ in the IUCN Red List (Short et al., 2010), considering the alarming current and predicted trends, it is not unlikely that the marginal habitats (i.e. habitats supporting only a few species or individuals because of the limiting environmental conditions) of the Baltic Sea will ultimately lose important functions performed by eelgrass (Waycott et al., 2009; Short et al., 2011).

Aims of the thesis

The current thesis is based on a detailed inventory of the eelgrass habitats performed within the whole Estonian coastal range covering the most important environmental gradients in its marginal habitats. This inventory allowed us for the first time to characterize the eelgrass habitats in terms of benthic biodiversity, species composition and dominance structure; to provide a knowledge base for follow-up biological and ecological studies; and thus, to provide important information for managing our marine resources.

The aims of the thesis are:

- (1) to provide a basic overview of the main methods applied in marine benthos mapping in Estonian coastal waters (**I–V**); to more specifically test and determine the most suitable video observation method in assessing the cover of benthic macroalgal species (**III**);
- (2) to compile a large-scale up-to-date distribution map of eelgrass (**I, V**);
- (3) to give an overview of eelgrass community characteristics in the coastal waters of Estonia (**II**);
- (4) to evaluate the use of optical remote sensing in mapping eelgrass habitats in the optically complex waters of the NE Baltic Sea (**IV**);
- (5) to identify the most important environmental variables defining the cover of eelgrass, to specify the spatial scales where such relationships are the strongest and to predict changes in the distributional pattern of eelgrass from the current to plausible future climate (**V**).

2. MATERIAL AND METHODS

2.1. Study area

The study area is located in North Europe, in the north-eastern part of the Baltic Sea. Papers I and V cover the whole Estonian coastal sea area, whereas the other studies were carried out in different Baltic Sea sub-basins: in the West Estonian Archipelago Sea (II), the Baltic Proper (III, IV) and the Gulf of Finland (II, III) (Figure 1).

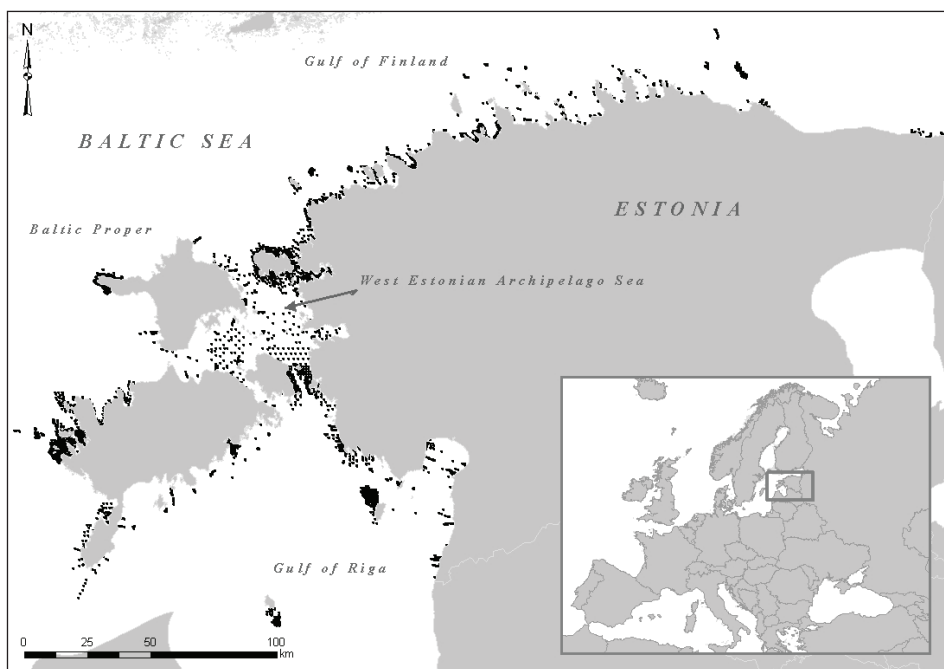


Figure 1. Map of sampling stations in the study area. The sampling effort was carried out in the depth range 0–10 m in 2005–2015. (Redrawn from Paper V.)

The Baltic Sea is a geologically young semi-enclosed sea and one of the largest brackish water basins in the world. Due to the short evolutionary history, low salinity and strong seasonality in temperature and light conditions of this sea, the species richness is small, characterized by a mixture of marine, brackish or freshwater origin species (Hällfors et al., 1981). In the study area there is a strong permanent salinity gradient from west (higher values) to east. In coastal areas the dynamics of seawater temperatures is directly coupled with air temperatures. The average sea surface temperature is around 2 °C in winter and may rise up to 20 °C in August. The study area has a wide coastal zone with diverse bottom topography and underwater habitats. Soft sediments such as sand, silt and clay prevail with hard substrata usually found in shallow and exposed

coastal areas. Due to the prevalence of clay substrate and strong wave action, the water transparency is often very low, especially in the West Estonian Archipelago Sea and the Gulf of Riga (Kotta et al., 2008a; 2008b) (see also Table 1 in Section 2.3.).

2.2. Field sampling

Prior to fieldwork activities a regularly spaced grid of sampling points was generated using ArcGIS software (ESRI, 2011); the distance between the sampling points varied from 10 m to 1 km (I, IV, V). Denser sampling grids were applied in the shallow (mainly <5 m) marine areas due to their higher heterogeneity. The sampling stations and areas were selected so as to cover the full diversity of the benthic habitats and the range of environmental gradients (I, IV, V). Information from previous underwater investigations and the available geological maps and nautical charts were also considered (I–V).

In this thesis traditional on-site methods such as diving and collecting biomass samples with grab or frame samplers were the most commonly used techniques (samples were analysed according to HELCOM (2015) guidelines; for details see Papers I, II, IV, V). In addition, qualitative sampling was mostly performed with hand-held drop cameras (I, III–V). Paper III focuses solely on remote underwater video observations and tests different approaches to analysing video material. A hyperspectral imager was used to obtain airborne imagery for larger seascape areas (IV) and a spatial modelling method incorporating machine learning techniques was applied to predict current and future patterns along the whole Estonian coastal sea (V).

2.3. Environmental data used

The following environmental information was used when assessing species patterns under current environmental conditions (see more detailed information in the referred Papers): sediment character (I, V), depth (I–V), slope (I, II, V), exposure to waves (I, II, V), water temperature (I, II, V), salinity (I, II, V), current velocity (I, II, V), ice cover (I, II, V), water transparency (V) and chlorophyll *a* content (I, V). The ESRI Spatial Analyst tool was used to calculate the average of all abiotic and biotic variables (those obtained from field sampling as well as from modelling) for local (i.e. sampling), 1 km and 10 km spatial scales (V). These values were used to link environmental and biotic patterns at larger spatial scales. The abiotic environmental variables with means, minima and maxima used in Paper V are presented in Table 1. In Paper V predictions of the same variables were used to analyse how global climate change will potentially affect species distribution patterns (Table 1).

Table 1. List of environmental variables with their means, minima and maxima in different water bodies around the Estonian coastal sea under current and projected climate conditions. Water basins are denoted as follows: GOF – Gulf of Finland, WEAS – West Estonian Archipelago Sea, GOR – Gulf of Riga, BP – Baltic Proper. Environmental variables are as follows: Temp – average water temperature, Salinity – average water salinity, Velocity – average current velocity, K_d – average water attenuation coefficient, Chl a – average chlorophyll a content in water, Slope – inclination of coastal slope, Soft sediment – percentage cover of soft sediment fractions, Ice cover – average ice cover over the study site. (Table modified from Paper V in which more information on the studied variables can be found.)

Variable	De- scriptive statistics	Current				Projected			
		GOF	WEAS	GOR	BP	GOF	WEAS	GOR	BP
Temp, ° C	Mean	12.9	14.2	13.4	12.8	16.9	18.2	17.4	16.8
	Min	10.3	11.4	10.2	11.0	14.3	15.4	14.2	14.9
	Max	17.3	19.1	18.5	18.1	21.3	23.1	22.5	22.1
Salinity	Mean	5.3	7.0	5.5	7.3	3.9	5.2	4.1	5.5
	Min	3.3	6.0	3.4	6.4	2.5	4.5	2.6	4.8
	Max	7.5	7.3	6.8	7.8	5.6	5.5	5.1	5.9
Velocity, cm s ⁻¹	Mean	1.9	5.5	2.8	4.0	1.9	5.5	2.8	4.0
	Min	0.1	0.1	0.2	0.1	0.1	0.1	0.2	0.1
	Max	5.2	11.7	7.4	9.7	5.2	11.7	7.4	9.7
K_d	Mean	1.4	1.3	1.2	1.1	1.7	1.5	1.5	1.3
	Min	0.8	0.6	0.7	0.4	1.0	0.7	0.9	0.5
	Max	2.7	2.7	2.9	2.9	3.3	3.2	3.5	3.5
Chl a , mg m ⁻³	Mean	25.4	17.9	20.3	12.1	38.2	26.9	30.5	18.2
	Min	6.8	7.6	8.5	3.8	10.2	11.4	12.8	5.7
	Max	45.0	47.7	47.2	45.8	67.5	71.6	70.8	68.7
Slope, °	Mean	0.5	0.1	0.1	0.2	0.5	0.1	0.1	0.2
	Min	0.0	0.0	0.0	0.0	0	0	0	0
	Max	22.9	5.1	5.9	11.1	22.9	5.1	5.9	11.1
Soft sediment, %	Mean	66.9	86.6	68.7	48.7	66.9	86.6	68.7	48.7
	Min	3.1	12.2	12.0	1.1	3.1	12.2	12.0	1.1
	Max	98.8	99.2	96.0	95.4	98.8	99.2	96.0	95.4
Ice cover, %	Mean	30.1	32.6	33.5	15.9	15.1	16.3	16.7	8.0
	Min	19.4	23.4	19.4	4.7	9.7	11.7	9.7	2.3
	Max	38.1	36.3	41.9	32.7	19.0	18.1	20.9	16.4
Depth, m	Mean	38.0	4.9	26.0	55.0	38.0	4.9	26.0	55.0
	Min	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	Max	115.0	24.0	67.0	459.0	115.0	24.0	67.0	459.0

2.4. Statistics and modelling

Spearman's rank correlations between different environmental variables, eelgrass parameters and associated flora and fauna were found (II). When analysing effects of study methods on the perception of benthic community structure, factorial analysis of variance (ANOVA, Statistica version 7.1 and 8.0) was used (III). The repeated measures analysis of variance (RM ANOVA) was applied to test differences in sediment organic matter and eelgrass community parameters between depths, sites and months (II). ANOSIM (Primer version 6.1; Clarke et al., 2014) analyses were used to examine differences in the patterns of variation in species composition (I, III) and between time, depth and site (II). Taxa responsible for the observed differences were identified by SIMPER analysis (Primer version 6.1; Clarke, 1993), where the cut-off percentage was set at 90% (I, II). Canonical correspondence analysis (CCA) was used to visualize variability of different habitats along a multitude of abiotic environmental gradients (I). The analyses in Paper I were done in the statistical software R version 2.15.1 (RDC Team, 2013) using the vegan package (Oksanen et al., 2017). The relationship between different eelgrass community parameters and environmental variables was examined using the BIOENV procedure (Primer version 6.1; Clarke and Ainsworth, 1993) (II).

In Paper IV a supervised classification procedure was performed, which allows automatic categorization of all image pixels into previously defined classes. Field data from groundtruth stations, as well as spectrally similar areas close to the field control points and experts knowledge on the area, were used for image classification. Two different supervised classification algorithms were tested using ENVI software: Maximum Likelihood (ML) and Spectral Angle Mapper (SAM). The quality of benthic habitat information derived from the airborne CASI data was determined by the quantitative process of accuracy assessment.

The niche breadth of eelgrass and other higher plants was assessed using analysis of outlying mean index (OMI) (V). This index measures the distance between the mean habitat conditions used by the species (niche centre) and the mean habitat conditions of the sampling area (Doledec et al., 2000). The higher the value of OMI of a species, the higher is its habitat specialization. The package 'ade4' (Dray and Dufour, 2007) was used for running OMI analysis in the statistical software R (RDC Team, 2013). The environmental niche space of submerged aquatic vegetation was visualized by drawing a convex hull over the points of OMI ordination where the species was present. When drawing the border of the niche space, 5% of the most distant observations of species occurrences were considered as outliers and excluded from the analysis.

In Paper V the predictive modelling technique called Boosted Regression Trees (BRT) was used. The method combines the strength of machine learning and statistical modelling and due to its strong predictive performance is increasingly used in ecology (Elith et al., 2008). The BRT models quantified the contribution of different environmental variables to the coverage of eelgrass and

these models were also used to predict the species coverage for the whole study area given ambient and projected climate conditions (V). Both present and future predictions were modelled over a 200×200 m grid covering water depths of 0–10 m. The BRT modelling was done in the statistical software R using the gbm package (RDC Team, 2013).

3. RESULTS AND DISCUSSION

3.1. In situ mapping of benthic shallow water communities in the NE Baltic Sea and the spatial spread of the eelgrass habitats

Submersible video systems that consist of a hand-held submersible camera and recorder were first introduced in Estonia in 2005 (I). That year also marks the beginning of large-scale mapping of marine benthic habitats in the Estonian coastal sea as this technique allowed researchers to significantly increase the number of samples to be collected. Systematic marine inventories in Estonia started in 1959 and were based mostly on grab sampling for several decades. By the end of 2016 the marine benthos database of the Estonian Marine Institute stored in total over 16 000 observations and over 22 000 samples (Figure 2). As of 13.12.2016 the total numbers were as follows: 16 451 visual observations (11 927 video observations and 4524 estimations by diver) and 22 857 biomass samples (14 835 grab samples and 8022 frame samples).

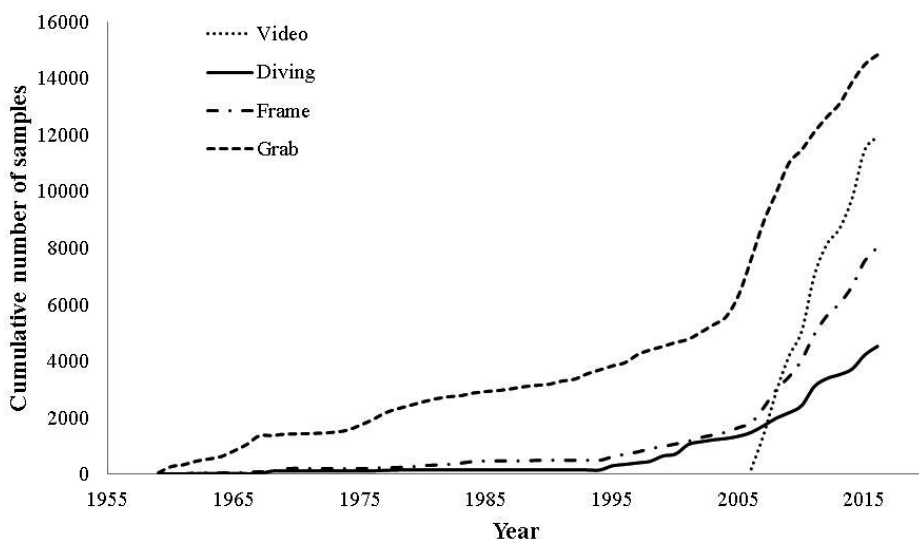


Figure 2. The use of different in situ methods in the marine benthos surveys in the Estonian coastal sea in 1959–2016 (the database of Estonian Marine Institute).

The first detailed marine benthos inventory project for the Estonian sea area was carried out in 2005–2009; prior to this inventory no detailed benthic habitat maps had been produced for this region (I). Although large sea areas were mapped, we are still very data-limited as only a minor fraction of the studied seascapes were actually sampled and vast areas between sampling stations remained unstudied. Nevertheless, this huge data set enabled establishing a marine benthic habitat classification system of 18 habitat classes to meet the

local conservation purposes (EBHAB – Eastern Baltic marine benthic HABitats, **I**). As the EBHAB classification scheme had substrate, wave exposure, light and key species as classifiers (see Paper **I** for details) it was also easy to link this initiative to the EU level EUNIS (the European Nature Information System) habitat system (<http://eunis.eea.europa.eu/>). Overall, eelgrass was found within eight different habitats with one being recognized as a true eelgrass habitat (namely ‘Moderately exposed soft bottoms with *Z. marina*’) (**I**).



Figure 3. The distribution of eelgrass in the Estonian coastal waters in 2005–2015 in relation to marine protected areas. The spatial distribution of the eelgrass habitat follows the EBHAB classification scheme.

According to Paper **I**, the eelgrass habitat (eelgrass coverage $\geq 10\%$) was found on substrates ranging from fine to coarse sands. The habitat was found in a depth range of 1–9 m (see also Section 3.3 for specific comments) at salinities down to 4. The habitat was largely dominated by the higher plant eelgrass and filamentous brown algae with other higher plants occurring occasionally. This habitat type hosted high numbers of plant and invertebrate species (for more details on the eelgrass community composition see Section 3.3). The total area of eelgrass habitat was estimated at minimum at 155 km² (Figure 3). The distribution of the eelgrass habitat was linked to sediment characteristics and salinity, whereas the distribution of higher plants communities excluding eelgrass was mostly related to ice cover, water velocity and temperature (**I**). This resembles the findings in **V** and is discussed in more detail in Section 3.5

(future conditions). In terms of benthic species dominances the habitat class ‘Moderately exposed soft bottoms with higher plants excluding *Z. marina*’ highly resembled the eelgrass habitat. Both habitats were found in the same depth and salinity range and hosted similar numbers of macrophyte and invertebrate species (I, see also Papers II and V and Section 3.3 for more details), but the habitats differed in species composition (I).

In order to fulfil the needs of an efficient protection strategy, the management focus should be broader than just the valuable habitats where eelgrass is currently found. Thus, in order to have positive effects on biodiversity on a larger spatial scale, the action of designating protected areas has to involve analyses of species distribution on different levels (e.g. species, habitat, ecosystem). Specifically, some areas may not host the habitat-forming species due to natural succession and/or anthropogenic disturbances (Kendrick et al., 2000; Kotta et al., 2012). Therefore, it is rewarding to perform a supplementary habitat suitability modelling (see Paper V for the current ranges of the eelgrass habitat) in order to predict the potential of the environment for the species distribution of high conservation value (Araújo and New, 2007) and use these modelled layers together with the actual mapping results when creating the boundaries of a protected area (Rioja-Nieto and Sheppard, 2008). It has to be also taken into consideration that in low salinity areas such as the NE Baltic Sea, eelgrass reproduces asexually (e.g. Reusch et al., 1999) and the pattern seen in its distribution is rather a function of a colonization history that reflects the conditions from decades to centuries (e.g. Kendrick et al., 2000). As such the eelgrass habitat is very fragile and valuable and all efforts should be made to ensure that its further deterioration is avoided.

3.2. Linking spatial scale, observation method and community perception

The study on video methodology reported in Paper III showed that (1) there were no clear local spatial scales in which the variability of benthic communities was maximized; (2) the eelgrass community was only poorly predicted by the spatial arrangement of sediment characteristics; (3) the selection of method had no effect on the estimates of macrophyte cover, but the method had independently of habitat type an impact on error estimates of macrophyte cover.

As discussed in Paper III, it is plausible that the weak effects of the choice of the methods (continuous video, different sets (5, 10, 20, 35) of uniform or random picture samples from video) can be attributed to the homogeneous distribution of benthic macrophyte communities in the study area and to the small range of scales investigated. The spatial pattern of the eelgrass community varied much along transects but it was weakly related to sediment patterns (III). Some transects had the largest variability at 20 or 50 m scales (i.e. the patch size of macrophyte communities was on average 20 or 50 m) (III), which suggests that exposure to waves rather than sediment characteristics determines the

spatial patterns of the eelgrass community (see also Paper V). As also discussed in Paper III, large waves may cause considerable resuspension of sediments and result in prolonged periods of poor light conditions (Madsen et al., 2001) but, similarly, they may cause mechanical disturbance of the benthic communities (Dernie et al., 2003; Kotta et al., 2007; Schiel and Lilley, 2007). Nevertheless, in the Baltic Sea the bottom substrate is often very heterogeneous. Substrate types either form fine-grained mosaics (e.g. patches of sand or stones that do not exceed 50–200 cm in diameter) or, alternatively, different sediments are mixed at the same location (e.g. mixture of clay, sand and pebbles) (Kotta et al., 2008a; 2008b). In otherwise favourable conditions (exposure, depth etc.) but mixed substrates, the occurrence of suitable sandy patches is crucial for eelgrass presence (e.g. Küdema Bay, where eelgrass and *Fucus vesiculosus* Linnaeus, 1753 (key species on hard substrate) grow side by side; personal observations).

The video-observations caught in total eight species of macrophytes and one species of invertebrates in eelgrass stands (III). The small number of species is related to both the small number of samples (5 transects) and the ability of the method to detect mainly macrophytes, species that are either large enough (e.g. *Chorda filum* (Linnaeus) Stackhouse, 1797) or with unique appearance/traits (e.g. *Cladosiphon zosterae* (J. Agardh) Kylin, 1940). Taken that the maximum number of benthic species (both macrophytes and invertebrates) found in one eelgrass meadow in the NE Baltic Sea is 36 (II, Sõru), the video method captures at least 20% of the total species richness within an eelgrass habitat. Both video and still picture mode captured precisely the coverage of key species and cover patterns of annual and perennial algae (III). Thus, considering the generally low diversity of macrophyte communities in the study area (Kotta et al., 2008a, 2008b), already a few observations (or pictures) may capture the pattern of macrophyte communities and provide accurate estimates of their parameters. In a long-term monitoring programme high precision (ability to detect differences) is more important than high accuracy (ability to detect true value) (Andrew and Mapstone, 1987). Altogether, our estimates in a continuous video mode and in a still picture mode based on 35 and 20 photographs produced less variable results compared to other methods (III). The continuous video method is precise enough for monitoring the changes in key species coverage and now it has been included into e.g. benthic habitat mapping (e.g. I), national monitoring programmes of the marine benthic environment and water quality assessment in Estonia.

3.3. Eelgrass community characteristics

In 2005–2015 eelgrass was found in more than 300 locations (Figure 3) on soft substrates with standalone specimens growing even on mixed sediments of pebbles, gravel and coarse sand (westernmost area of Hiiumaa) (I, V) with the total estimated area about 400 km² (Herkül et al., unpublished). Plants were recorded up to 8 km from the shore (V). Recent findings suggest that eelgrass is

more common than it was previously expected and that the species can inhabit areas with salinity below 5 (V). The observed values indicate that eelgrass beds grow down to 7 m depth with the main distribution depth at 2–5 m in the Estonian coastal waters (II, V). A few specimens of eelgrass have been found down to 9 m (I), but according to the present knowledge, they do not form a distinguishable stand (Möller and Martin, 2007; II, V). The main distribution range is similar to Danish and Swedish waters where eelgrass grows mainly at 2–5 m depth and the total depth range of eelgrass is 1–10 m (Boström et al., 2003). In our study sites (II) the depth distribution of eelgrass follows the same pattern as observed in Øresund, Denmark (Krause-Jensen et al., 2003), i.e. eelgrass creates many small shoots in dense patches in exposed shallow waters with high light intensity and in deeper areas larger but fewer shoots are found in sparse coverage. This growth pattern can be regarded as a photoadaptive response along the water depth gradient (Dennison and Alberte, 1986), and also the upper depth limit is mainly controlled by wave action and ice-scouring (Middelboe et al., 2003).

In the West-Estonian Archipelago Sea eelgrass was found only in shallow areas (II), the species grows at depths > 4 m rarely and sparsely (Estonian Marine Institute database). In this area clay sediments prevail at depths > 5 m and fluctuations in water transparency (and in light climate) due to wind-induced water movement are very common (Suursaar et al., 1998). In areas where sand prevails and light conditions are more stable (e.g. Gulf of Finland), dense eelgrass stands were also observed deeper down (I, II, V). As discussed in Paper II, substrate availability and light conditions with the light climate along the depth gradient controlling photosynthetic activity determine the eelgrass depth distribution (Moore and Short, 2006). Both light intensity and the duration of the daily light period at which light equals or exceeds the photosynthetic light saturation point are important in seagrass growth and survival, especially for plants at or near their maximum depth distribution (Touchette and Burkholder, 2000 and references therein).

The presence of eelgrass significantly contributes to sediment trapping and erosion (II). On average the content of organic matter in the sediment varied between 0.38% and 1.47% for the eelgrass bed and between 0.29% and 1.1% for the bare sand (II). Our study showed that the organic content in the sediment was higher in the eelgrass stands compared to unvegetated areas only in areas where the movement of soft sediments is higher due to the combination of slope, exposure and area-specific water regime. In moderately exposed habitats the content of sediment organic matter between the eelgrass stands and the adjacent unvegetated areas did not differ (II). The measured sediment organic contents within eelgrass stands are in good accordance with the values determined for the Finnish Archipelago Sea (0.5–1.5%) (Boström et al., 2003).

In the eelgrass habitats the formation of new leaves was observed throughout the study period (May–September), but the growth in shoot biomass varied among depth strata and months (II). Different eelgrass parameters such as shoot biomass, shoot density, number of leaves per shoot and shoot length were best explained by depth and temperature (mean temperature of 2 preceding months)

with an increasing depth reducing and an elevating temperature raising the shoot density and biomass of the eelgrass community (II). As discussed in Paper II, here temperature should be regarded as a proxy of seasonality in e.g. temperature, light climate and their interaction. The highest densities often coincided with a low biomass (II). Also shown by Sand-Jensen and Borum (1983), such decrease in the biomass of eelgrass shoots may be a result of self-shading at high densities. For the Baltic Sea area the maximum biomass of eelgrass has been observed in July and August (Duarte, 1989; Olesen and Sand-Jensen, 1994; Boström et al., 2004), but our findings suggest an increase of shoot biomass and length also in September (II). Such later termination of the growing season in colder areas was also suggested by Clausen et al. (2014).

The average biomass of eelgrass was 7–70 g dw per m² (II). As discussed in Paper II, apart from the seasonal maximum of shoot biomass, comparison of our results with the existing data on eelgrass populations in the Baltic Sea in terms of density, aboveground biomass and length (Boström et al., 2003, 2004, 2014) does not show any major differences. The biomass values are similar to those estimated also by Trei (1973) for some eelgrass communities in the West-Estonian Archipelago Sea in earlier decades (128–300 g ww m⁻²). The values below 100 g dw m⁻² are typical for the Baltic Sea (Boström et al., 2014; Röhr et al., 2016). Thus, in spite of different environmental conditions compared to northern shores of the Baltic Sea, the response of eelgrass to the abiotic environment is similar.

The average density of eelgrass in all our study sites varied between (50)133 and 1300 shoots per m² with the maximum of 1725 shoots per m² (II). As discussed in Paper II, these densities are somewhat higher compared to the values reported in Finland where the shoot density ranges from 50 to 800 per m² (Boström et al., 2003, 2004, 2006). However, the Estonian values were lower compared to the Swedish and Danish observations (the maxima 3600 and 3500 shoots per m², respectively) (Sand-Jensen, 1975; Wium-Andersen and Borum, 1984; Krause-Jensen et al., 2000; Boström et al., 2003). As compared to the values reported for the whole Baltic range, the density of eelgrass in the Estonian coastal sea is surprisingly high. This agrees with results by Boström et al. (2014) according to which salinity defines the distribution range of eelgrass but does not largely affect eelgrass growth and density at the salinity minima. Nevertheless, low salinity areas compared to e.g. the Danish straits seem to have greater variability in shoot numbers (II; Röhr et al., 2016).

The growth of associated algae also follows the general seasonal pattern (II; Wallentinus, 1984; Lotze et al., 1999). Owing to the spring-time bloom of ephemeral algae, the coverage of drifting macroalgae was greater in May compared to the following months (II). Macroalgal mats refer to eutrophication and are common in all coastal regions of the Baltic Sea (e.g. Kiirikki and Blomster, 1996; Bäck et al., 2000; Vahteri et al., 2000; Paalme et al., 2004). Also discussed in Paper II, macroalgal blooms can reduce eelgrass shoot density (Nelson and Lee, 2001), shoot size and biomass and also the distribution area (Bintz et al., 2003; overview in McGlathery, 2001). Also the presence of

loose algae can be one of the factors triggering a high abundance of herbivores (Philippart, 1995; Drury, 2004; Gil et al., 2006) and other epibenthic fauna (Pihl et al., 1995). Although our eelgrass stands were often covered by loose-lying algae we did not observe any sign of stress related to these ephemeral algae (**I**, **II**). The dense macroalgal mats that cause heavy light limitation have lethal effects on northern Baltic eelgrass populations after 4 weeks of suffocation (Salo et al., 2015). In natural conditions the suffocative effect for a month is not often met and as suggested by Rasmussen et al. (2013), the highly dynamic nature of loose algae in small scale may reduce suffocation stress in eelgrass communities. Besides, the Estonian coastal sea is characterized by a good water exchange (coastal–offshore and surface–bottom) and the sediment–water interface is usually well oxygenated (Kotta et al., 2008a).

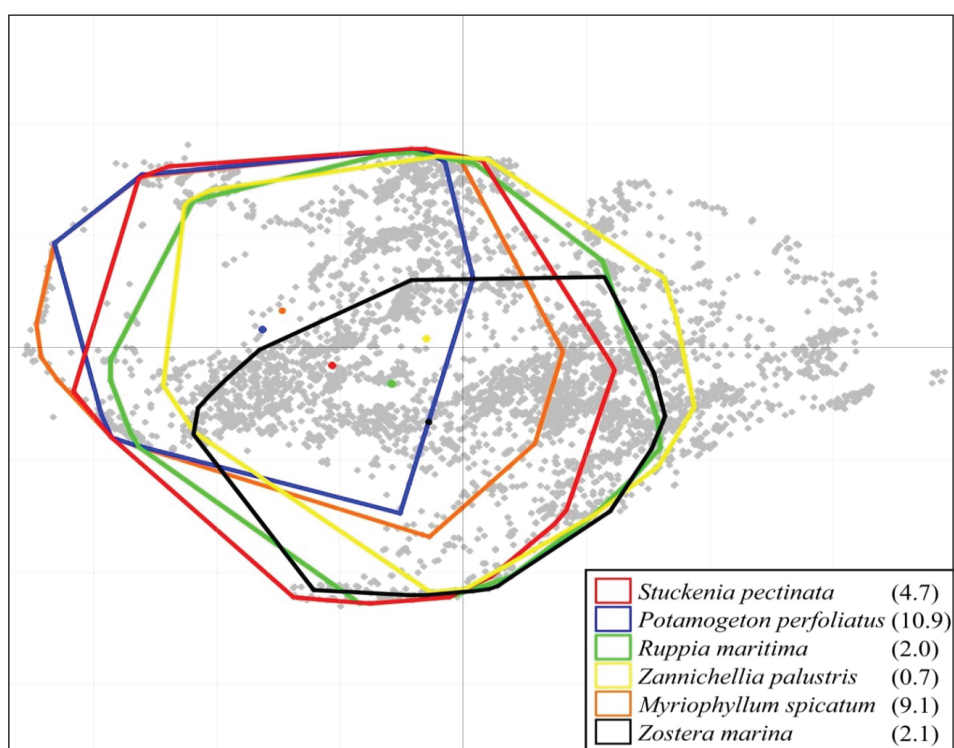


Figure 4. Niche breadth analysis of the studied species. The borders of niche space and separation of habitat niche between the studied submerged aquatic species are shown by coloured lines. The OMI identifies ordination axes that optimize separation between species and the observations are positioned in the multidimensional space as a function of environmental variables. Coloured dots on the plot indicate the centres of niche space of the species. Grey dots represent sampling sites. The value of outlying mean index (OMI) is shown in brackets. OMI measures the distance between the mean habitat conditions used by species (niche centre) and the mean habitat conditions of the sampling area. The higher the value of the OMI index of a species, the higher is its habitat specialization. (Redrawn from Paper V.)

A total of 33 macrophytobenthic taxa were found in samples from eelgrass stands (I), in the four studied communities in Paper II the species number was 19. Taken separately, the eelgrass meadows inhabited 9–15 species of macrophytes (II). All these macrophytes are common in the coastal waters of Estonia, only the the distribution of *Cladosiphon zosterae* (J.Agardh) Kylin, 1940 (epiphyte on eelgrass) is restricted to eelgrass habitats. Filamentous brown algae *Pylaiella littoralis* (Linnaeus) Kjellman, 1872, which is one of the most common species found in loose algal mats, was recorded in all four studied sites (II) and contributed most to the overall similarity within the eelgrass habitat (I). The vascular plants occurring together with eelgrass are all common on soft substrates in the Estonian coastal waters and can be also found as single-species stands in the depth range of 1–7 m (Trei, 1991). The most common species co-occurring with eelgrass are sago pondweed *Stuckenia pectinata* (L.) Börner, 1912 and clasping-leaf pondweed *Potamogeton perfoliatus* Linnaeus, 1753 (I, II, V). In addition, horned pondweed *Zannichellia palustris* Linnaeus, 1753, bird's-nest stonewort *Tolypella nidifica* (O.F.Müller) Leonhardi, 1864 and in some occasions *R. maritima* and charophytes *Chara* spp. are common (I, II, V). Similarly, the niche modelling indicated that there was a large overlap in niche space among eelgrass and other submerged plant species (V). Although eelgrass and *P. perfoliatus* inhabited somewhat opposing niche space (V, Figure 4), otherwise there was a large overlap in niche space of studied submerged plants. However, eelgrass is the only species forming continuous meadows outside sheltered bays and inhabiting depths > 3 m in the moderately exposed coastal sea of Estonia (I, V; Herkül et al., unpublished). Both interspecific competition and genetics has an important role in niche differentiation among brackish and marine seagrass populations.

The eelgrass stands in Estonian coastal sea areas support high biodiversity of invertebrates: overall 35 species have been found in the eelgrass stands (I) with about 23 faunal species being more common (II). This corresponds to about a quarter of the invertebrate richness found in shallow waters of the Estonian coastal sea (about 130 species). The study also showed that a higher biomass of eelgrass supported an elevated density of invertebrates (II). Taken separately, the eelgrass meadows inhabited 8–21 species of invertebrates (II). Our findings are similar to those reported for north-eastern Baltic Sea (Homziak et al., 1982; Boström and Bonsdorff, 2000; Moore and Short, 2006). Also discussed in Paper II, the abundance of benthic invertebrates was about 100 times lower than recorded for nearby eelgrass communities in Finland but was within the range of values estimated from e.g. the coastal sea of Great Britain. Variant sampling methodology with likely over- and underestimations of abundance may explain these large differences (Boström et al., 2006 and references therein). However, some disparity can be attributed to true habitat differences. Often low faunal diversity and density are related to neighbourhood habitats. If the belt of eelgrass is narrow and is surrounded by defaunated coarse unvegetated sand, a poor representation of benthic invertebrates in adjacent habitats may be one of the most plausible reasons for the observed low faunal diversity in such habitat (II).

Also discussed in Paper II, the species composition of plants has a strong effect on the abundance and biomass of benthic invertebrates. For example, mixed eelgrass stands, especially with the presence of *P. perfoliatus*, are expected to favour gammarid amphipods (Gustafsson and Boström, 2009). Similarly in our study area the sites that lacked *P. perfoliatus* were devoid of gammarids. However, it has been also suggested that the three-dimensional structure of the macrophyte habitat is more important for the richness of associated fauna than the macrophyte identity, i.e. species belonging to the community (Heck et al., 2003). Anyhow, the species that contributed most to the overall similarity of eelgrass stands were burrowing filter-feeding clams *Limecola balthica* (Linnaeus, 1758) (formerly *Macoma balthica*) and *Cerastoderma glaucum* (Bruguère, 1789) (I). These species are most common in shallow sandy sediments and eelgrass is not vital for their presence; moreover, lower densities of clams have been reported in continuous vegetation compared to patches of plants and bare sand (Boström et al., 2010). Thus the common (and numerous) presence of burrowing clams in eelgrass stands also suggests the patchy distribution of eelgrass within its habitat in the Estonian coastal sea (II).

3.4. Mapping eelgrass communities using remote sensing techniques

In remote sensing habitat mapping it is important to define an appropriate habitat mapping scheme that is also meaningful from the ecological point of view. In Paper IV two different classification schemes were developed and used in the classification of the CASI hyperspectral image. With this combined method we were able to classify nine benthic habitat classes with three of them referring to the possible presence of eelgrass. These habitats were as follows: (1) dense higher order vegetation at depths < 2 m with total macrophyte coverage more than 50%, prevailing species higher plants; (2) higher order vegetation on bright bottom at depths < 2 m with sand or silt coverage, total macrophyte coverage less than 50%, prevailing species higher plants and finally (3) vegetated communities at 2–6 m depth. A coarse classification scheme of six habitat classes returned two classes that possibly host eelgrass. These habitats are (1) green algae and higher vegetation at depths < 2 m (green macroalgae and higher plants vegetation with coverage more than 10%) and (2) vegetated area at 2–6 m depth containing either vegetation or bare substrate.

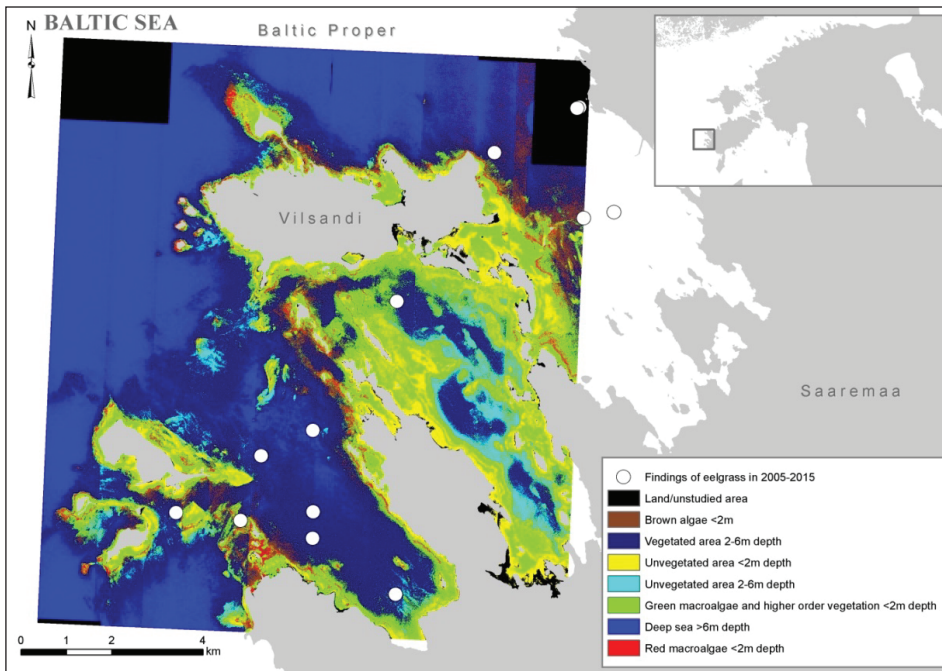


Figure 5. Benthic habitat map of the Vilsandi National Park area produced from the CASI airborne imagery describing six benthic communities. Within search for eelgrass it is rewarding to focus the search effort on areas classified as ‘vegetated area 2–6m depth’. White dots indicate real findings of eelgrass during earlier in situ surveys. (Modified from Paper IV.)

In Paper IV it was not possible to distinguish different species but instead we assessed broader habitat classes. However, even such broad habitat classes may be useful when planning future habitat mapping of eelgrass. Specifically, habitat features that potentially contain eelgrass can be validated by a targeted mapping of these sea areas (Figure 5). Consequently, the method allows saving time and money as well as covering much broader areas of interest and thereby offers unique large-scale synoptic data to address the complex nature of coastal waters. More recently, the used hyperspectral remote sensing was combined with spatial modelling techniques involving machine learning algorithms (Kotta et al., 2013). Such ensemble models succeeded in identifying and assessing the coverage of eelgrass in the optically complex seawater of the Baltic Sea. Thus, it would be rewarding to seek a generic standardized procedure for mapping multiple species in multiple areas. Such maps would greatly expand our capacity to understand biotic patterns, their changes and causes and thereby improve ecological theory and potentially preserve endangered seascapes for future generations.

3.5. Modelling current patterns and possible futures of eelgrass

In Paper V Boosted Regression Trees (BRT) modelling was used to relate the cover of eelgrass to the abiotic environment in the brackish Baltic Sea. The established functional relationships were then used to predict current and future patterns of eelgrass in the whole Estonian coastal sea. Paper V covered a large gradient of water temperature, salinity, wave exposure etc. and the modelled ranges of environmental variability did not exceed those observed in the field. The analyses in Paper V showed eelgrass to be the most sensitive to changes in water temperature, current velocity and bottom topography. Water salinity and eutrophication have little impact on the distributional pattern of the species. Eelgrass spreads all over the Estonian coastal sea, except the easternmost parts of the Gulf of Finland and the turbid and diluted Pärnu Bay area (V, Figure 6). All submerged plant species (of both marine and freshwater origin) in the coastal waters of Estonia seem to benefit from climate change, and eelgrass coverage is expected to increase under the projected climate change (V, Figure 6).

The local-scale environmental variability explained 45% of the eelgrass distribution; for 10 km scale (seascape-scale) the variance explained was 29% (BRT, V). The results of Paper V suggest that both local- and seascape-scale environmental variability affect the cover patterns of eelgrass with local variability exceeding seascape-scale variability and that species distributional patterns seem to have scale invariance in the Baltic Sea (*sensu* Halley, 1996; Gisiger, 2001). Our results in Paper V support the current understanding that the response of macrophyte species to environmental forcing is highly variable through a wide range of spatial scales (e.g. Kendrick et al., 2008).

As discussed in Paper V, it is likely that the high variability in submerged plant species at small scales is related to the mosaic of sediment and bottom topography at this scale in the study area (Kotta et al., 2008a, 2008b). Firstly, the availability of soft substrate is a pre-requisite for the establishment of the submerged plant species. Secondly, sediment modulates the flow above the seabed (e.g. Prasad et al., 2000; Hokinson and Eckhell, 2005) and the intensity of flows is directly related to the cover pattern of macrophytes (van Katwijk and Hermus, 2000; Madsen et al., 2001). In soft sediments, water flow also determines the light climate; i.e. large waves may cause considerable re-suspension of sediments and prolonged periods of poor light conditions (Madsen et al., 2001). Thirdly, small-scale topographic heterogeneity may provide the species refuge from physical disturbances including ice-scouring and mechanical stress due to waves (Kautsky, 1988; Heine, 1989). High variability in macrophyte communities at seascape scales is related to broad patterns of seawater warming, exposure to waves and winter ice scour, and an interaction of all these variables defines the suitability of a seascape for the growth of submerged aquatic vegetation (Kautsky and van der Maarel, 1990).

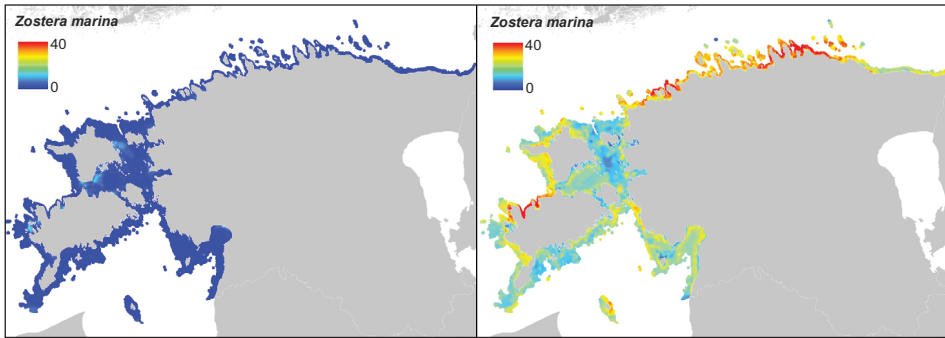


Figure 6. Modelled distribution of eelgrass for current climate conditions (left) and for projected future climate (end of this century) (right). Diagram shows the species coverage in percentage. The Booster Regression Tree technique was used for modelling. (Modified from Paper V.)

According to Short et al. (2010) there has been a global decline of area covered by eelgrass by 1.4% per year (based on researches conducted in 1990-2000). There are regions with significant large-scale decline of eelgrass cover (e.g. due to wasting disease or major pollution), areas where there has been no decline or the species distribution area has widened and areas where species has completely disappeared. Declines are mostly reported for developed and populated areas in Europe and North America (Short et al., 2010). The modelling exercise in Paper V suggests that elevated coverages of eelgrass associated to the future climate in the NE Baltic Sea are largely triggered by elevated temperatures. Water temperature affects plant physiological processes, growth rates and reproduction patterns and determines the geographic distribution of species based on their temperature tolerances (Short and Neckles, 1999). Higher temperatures generally alter the growth rates of the submerged plants (Short and Neckles, 1999) up to an optimum value and further temperature increase results in a dramatic plant net photosynthesis decrease and mortality (Díaz-Almela et al., 2009; Oviatt, 2004; Reusch et al., 2005). Temperature stresses are most obvious at the edges of species ranges, e.g. low salinity areas. Optimum salinity and temperature values for eelgrass growth are 10–30 and 10–20 °C, respectively (Nejrup and Pedersen, 2008; Salo and Pedersen, 2014). In a 5-week experiment the combination of low salinity (5) and high temperatures (25 °C) was shown to significantly increase the mortality of eelgrass adult shoots (Salo and Pedersen, 2014). At salinity 5 the plant is stressed but if other environmental conditions are suitable, eelgrass can survive at salinity as low as 2 (den Hartog, 1970; Salo et al., 2014). Eelgrass is also sensitive to a short-term rapid temperature increase in summer and the heat stress can lead to serious diebacks, declines in net primary production have been reported already above 23 °C (Moore et al., 2014). As discussed in Paper V, presently the Baltic Sea is a rather cold environment with a short vegetation season and in general here the mean temperature increase of 25% (i.e. summer maximum temperatures from the present 19 °C to 23 °C) will

likely not exceed the optimum growth values for submerged species. Elevated temperatures are expected to prolong the vegetation season, i.e. increase the growth of the submerged aquatic vegetation and shoot to root ratio (Zimmerman et al., 1989). However, more frequent heat waves affect seagrass communities all over the world, both positive and negative impacts have been reported (e.g. Díaz-Almela et al., 2007; Lefcheck et al., 2017; see overview in Short et al., 2016). Not considered in our modelling exercise in Paper V, but in shallow coastal areas of the Baltic Sea unforeseen heat waves combined with other stressors may cause a total collapse of eelgrass populations as the recovery of meadows is complicated both due to altered environmental conditions and slow clonal growth of the plant.

As discussed in Paper V, water motion is known to affect the plant structure of submerged aquatic vegetation (Fonseca et al., 1983; Worcester, 1995) and elevating current velocity will increase leaf biomass and width as well as canopy height (Fonseca and Kenworthy, 1987; Short, 1987). However, our study shows opposite results: eelgrass was disfavoured by elevated current velocities (V). A probable reason behind this pattern is specific light conditions in our study area – already wind speeds higher than 5 m s^{-1} cause considerable resuspension of prevailing clayey sediments and reduction of water transparency in the coastal water (discussed also in Section 3.3). A combination of high temperatures and low water transparency is considered lethal for eelgrass populations (Lefcheck et al., 2017).

Salinity plays a great role in the species distribution in the brackish Baltic Sea; however, the impact of salinity in local models of benthic vegetation is marginal (V; Rosqvist et al., 2010). In the Baltic Sea range low salinity values do not significantly decrease the growth rates of eelgrass (II; Boström et al. 2014; Salo et al., 2014). Nevertheless, the lowering salinity favours submerged plant species of freshwater origin, and higher interspecific competition between these and eelgrass is expected (Riddin and Adams, 2010). In addition, changed ice conditions also modify the competition between submerged plants. Models in Paper V suggest that reduction of ice cover duration favours slow-growing species such as eelgrass while fast-growing species such as *M. spicatum*, *P. perfoliatus* and *S. pectinata* lose their advantage from periodically destructive ice.

Although our model in Paper V predicted that all submerged plant species in the NE Baltic Sea would gain from climate change we have to keep in mind the combined influence of several stressors and non-linearities between the environment and biota, which can provoke unpredictable changes in biological communities (e.g. Hoffman et al., 2003). There are several environmental forcings that our study did not consider. For example, in addition to heat waves, the increased temperature under the projected climate is expected to enhance the growth of ephemeral epiphytic macroalgae (Lotze et al., 1999; Taylor et al., 2001) and macroalgal blooms will decrease eelgrass photosynthetic performance mainly due to changes in light climate (see Paper V and also Section 3.3 for a short overview). Light climate is expected to change also due to changes in the sea level: the IPCC (2013) prognoses the minimum global mean level rise of

0.63 m by 2100. In case of global sea level rise of 0.70 ± 0.30 m, the prognoses of relative sea level change for the Baltic Sea vary between sea level fall of 0.35 m (in northern areas) and sea level rise of 0.60 m (in southern areas). In the coastal areas where the slope is relatively slight, the increase in the water column height will automatically induce a lower depth limit of seagrasses whenever the limit is beyond the compensation depth. For a slope of 2%, an increase of 2 cm in the depth of the water column (vertical) corresponds to a linear regression of 1 m of the position of the lower limit (horizontal) of the seagrass meadow (Pergent et al., 2015). This again might be compensated by the predicted CO₂ enrichment, which will enhance the maximum photosynthetic rate of eelgrass and reduce the daily light requirement of plants (Zimmerman et al., 1997).

Not considered in our study, but genotypic diversity (aka clonal diversity) is another important factor affecting the stability and longevity of eelgrass meadows under fluctuating environmental conditions (see e.g. Boström et al., 2014 for more details). The clonal diversity of seagrass is relatively high in the North Sea, Scandinavian west coast and the Kattegat area. Less is known about eelgrass population genetics in the southern and eastern areas of the Baltic Sea, but low clonal diversity has been described for eelgrass growing in isolated lagoons and fjords in the Kattegat area (e.g. Limfjorden, Denmark; Ferber et al., 2008). In inner parts of the Baltic Sea eelgrass reproduces asexually due to low salinity and the probability of the occurrence of large, 800–1600 years old clones also increases (Reusch et al., 1999). These populations are expected to be highly sensitive to extrinsic stressors and potentially have a very low genetic adaptation potential (Lasker and Coffroth, 1999; Santamaría, 2002). At the same time, these genotypes are characterized by wide reaction norms enabling the persistence of species under highly fluctuating environmental conditions in terms of ice cover and eutrophication. However, according to the present knowledge, the clonal variability of eelgrass in the Baltic Sea is highly variable and neither related to present salinity conditions nor different sub-basins. Gonciarz et al. (2014) revealed that eelgrass populations located across the Baltic Sea (specifically in the coastal waters of Germany, Poland and Estonia) are genetically distinct, differing both in the clonal structure and in the level of genetic polymorphism. Surprisingly, no clones were discovered in the easternmost low-salinity study area (24 individuals studied) and the allelic richness was similar to that observed in the North Sea populations. Taken separately, eelgrass populations at the southern Baltic Sea have low clonal diversity; however, when the total (southern) distribution range is considered, the diversity is as high as in the North Sea (Diekmann and Serrao, 2012; Gonciarz et al., 2014). Recently, the genome of eelgrass was fully sequenced (Olsen et al., 2016) and this will contribute highly to the future studies e.g. on eelgrass adaptation to different salinity regimes and climate change effects on the species distribution.

Several marine foundation species in the Baltic Sea are expected to reduce their distribution range due to lower salinity, e.g. *F. vesiculosus*, *Furcellaria*

lumbricalis (Hudson) J.V.Lamouroux, 1813 (Vuorinen et al., 2015). Losing these species would impoverish species complexity and structure on hard substrates. The loss of diversity would make marine ecosystems more vulnerable and less resilient to climate change and other environmental shifts caused by disease, alien invasive species and the cascading effects of overexploitation (Gjerde, 2006). Our modelling in **V** showed that the key species on soft substrates, eelgrass, will benefit from climate change. In the NE Baltic Sea the species already grows on its physical tolerance limit, the eelgrass meadows are often narrow, patchy and sparse (**II**) and due to predominantly vegetative reproduction the recovery of disturbed meadows is limited. The estimated area of eelgrass habitats makes up less than 0.5% of the total Estonian sea area (36 500 km²), the distribution area of macrophytobenthic key-species habitats on hard substrate is even smaller (**I**). As discussed in Paper **I**, a holistic approach is needed to managing the marine environment and modelled distribution maps and predictions of the coverage of key species under future climate conditions are essential for effective conservational planning and would contribute to minimizing the risk of biodiversity loss.

4. CONCLUSIONS

One of the aims of the thesis was to provide a methodological overview of studies of coastal benthic species and habitats with eelgrass as the case species. Traditional survey methods such as diving and physical sampling of seabed have been used in studies of Estonian coastal sea benthic communities since 1959, and as these offer the most detailed information, they are most commonly used also today. Anyhow, as these methods require a lot of labour power, are very expensive and time consuming, mapping of large sea areas needs a different approach. Video observations came into use in 2005 and this method has been widely used as it provides a quicker and cheaper way to estimate the status of most key species in benthic habitats (I). In our studies a continuous video mode and a still picture mode based on 35 and 20 photographs produced the best results (III). Therefore these methods should be preferred in the mapping of benthic macrophyte communities as well as in the assessment of water quality in the relatively homogeneous environments of the northern Baltic Sea.

Eelgrass is an ecosystem engineering species in moderately exposed sandy substrates of the NE Baltic Sea. In 2005–2015 eelgrass was found in more than 300 locations (I–V). The species spreads all over the Estonian coastal sea, except the easternmost low salinity parts of the Gulf of Finland and the turbid and diluted Pärnu Bay area (I, V). Its main depth range is 2–5 m (I, II, V). The area of eelgrass coverage of higher than 10% is estimated at about 150 km² (I). For comparison – in the whole Baltic Sea the distribution area of eelgrass is estimated at 1200 km².

The seasonal variability of the eelgrass habitat was low whereas spatially the biomass of eelgrass varied largely, often as a function of depth (i.e. light, temperature, sediment characteristics) with some areas characterized by extensive but low-cover eelgrass meadows whereas other sites had only narrow but dense eelgrass belts (II). The average biomass of eelgrass was from 7 to 70 g dw per m² and the shoot density was on average 133–1300 shoots per m² (II). According to literature, these findings are similar across the species distribution range in the Baltic Sea. Contrary to other areas of the Baltic Sea, a significant growth of eelgrass occurred in the Estonian coastal sea also in September (II).

A total of 33 macrophytobenthic and 35 invertebrate taxa were found in samples from eelgrass stands in the coastal waters of Estonia (I). Taken separately, the eelgrass meadows inhabited 9–15 species of macrophytes and 8–21 species of invertebrates (II). All species found in eelgrass meadows are also common in the coastal waters of Estonia (except the eelgrass epiphyte *Cladosiphon zosterae*) and findings correspond to about a quarter of total benthic species richness in the area. Eelgrass habitats hosted one of the richest mesograzers community in the Baltic Sea (I, II). Our niche modelling indicated that there was a large overlap in niche space among eelgrass and other submerged plant species (V), and although the total number of species found

either in eelgrass or other submerged plant species habitats is similarly high, the habitats differ significantly in species composition (I). A generic relationship was found between the biomass of the eelgrass community and the abundance of associated benthic invertebrates (II), suggesting that the local abiotic environment does not modulate how benthic invertebrates respond to eelgrass canopy parameters.

Many novel tools such as hyperspectral remote sensing of benthic habitats, spatial predictive modelling and machine learning are becoming more and more common. Combining the traditional and novel tools is at present the best approach in order to understand biotic patterns and their change in the marine realm. The used remote sensing classification schemes produced accurate high-resolution maps at 0–6 m depth with a potential to distinguish larger groups of macrophytes (IV). Although not able to distinguish species, the results provide an important cost-effective input when planning targeted large-scale mapping of e.g. eelgrass. In this thesis, as an example of modelling benefits, we used the predictive modelling technique called Boosted Regression Trees to quantify non-linear relationships between the cover of eelgrass and the environment as well as to predict species cover under current and future environmental conditions (V).

Our results confirmed that small- and large-scale environmental variability contribute both separately and interactively to the variability in the cover of eelgrass (V). Under future climate change, physical disturbances such as seawater warming, elevated wave-induced current velocity and reduced ice scour are predicted to override the effects of salinity reduction, elevated turbidity and pelagic production (V). Our modelling study showed that eelgrass is very resilient to a broad range of environmental perturbation and biomass gains are expected when seawater temperature increases (V). This is mainly because vegetation will develop faster in spring and will have a longer growing season under the projected climate change scenario. Nevertheless, opposite changes in the abiotic environment might also occur under other climate change scenarios and these may have catastrophic effects on local eelgrass populations (e.g. due to unforeseen heat waves). As the species is growing at its stress limits, a modelling of the cover of eelgrass under the future climate is essential in order to help managers to establish marine protected areas that can resist the projected influences of climate change and thereby minimize the loss of biodiversity.

SUMMARY IN ESTONIAN

Läänemere kirdeosa põhjataimestiku leviku kaardistamine ja modelleerimine pika meriheina *Zostera marina* Linnaeus, 1753 näitel

Tänapäeval on merekeskkond inimtegevusest tugevalt mõjutatud, kaasnevad peamised ohud on looduslike varude ületarbimine, elupaikade hävimine, reostus, võõrliikide invasioon ning keskkonnatingimuste muutumine kliimamuutuse tõttu. Mereressursside laialdane kasutamine on viinud liikide ning elupaikade säästva majandamise ning kaitsmise vajaduseni, mille edukaks läbiviimiseks on vajalik liikide levikumustrite kaardistamine ning muutuste prognoosimine. Võrreldes maismaaga on merekeskkonnas uuringute läbiviimine ning proovide kogumine keerukam ning seetõttu on ka meie teadmised puudulikumad.

Mere põhjaelustik täidab ökosüsteemi toimimisel mitmeid olulisi funktsioone. Merepõhja elupaikadest on meriheinakooslused ühed produktiivsemad alad. Maal, neil on oluline roll rannajoone kaitsel, sette stabiliseerimisel, lainete summutamisel, maismaalt pärinevate toitainete filtreerimisel ja süsiniku sidumisel. Samuti on nad olulised paljudele selgrootutele, kaladele ning imetajatele toitumis-, varje- või elupaigana. „Meriheinad“ on kokkuleppeline termin, mis tähistab ökoloogilist rühma, kuhu kuuluvad merevees elavad kõrgemad taimed, mis asustavad nii tõusu-mõõna kui sublitoraalseid piirkondi troopilistel ja parasvöötmelistel rannikutel. Meriheinakoosluste levila on viimastel aastakümnetel oluliselt vähenenud ning lisaks lokaalsetele teguritele (sh eutrofeerumisele) on oluline roll ülemaailmsel protsessidel (sh kliimamuutustel). Kliimamuutuste tagajärjel tekkivad muutused veetemperatuuris ning tugevate tormide sagedam esinemine võivad põhjustada meriheinakoosluste leviku muutumist ja mõjutada nii nende liigilist koosseisu, mitmekesisust, struktuuri kui ka produktsiooni. Eutrofeerumine soodustab efemeersete makrovetikate vohamist ja madalas rannikumeres meriheinakoosluste asendumist vetikatega. Riimveelises Läänemeres kasvab 72st meriheina liigist vaid neli. Vaid üks merelise päritoluga liik – pikk merihein (*Zostera marina*) levib Läänemere lääne- ning põhjaosas. Läänemere piirkonna kliimaprognoos näitab kasvuperioodi pikenemist, jääkatte vähenemist ning muutusi tuule- ja sademete mustris, mis omakorda mõjutavad meres valgustingimusi ning soolsust. Mittelineaarsete elustiku ja keskkonna vaheliste seoste tõttu võivad isegi väikesed muutused inimtegevuses ja/või kliimas põhjustada muutuseid meriheina kooslustes, mis kasvavad juba niigi oma füsioloogilisel taluvuspiiril, ning viia oluliste, pika meriheina poolt täidetavate funktsioonide kadumiseni Läänemeres.

Käesolevas töös on kasutatud Eesti rannikumeres 2005–2015. aastatel laialatuslike merepõhjakoosluste kaardistamistöde käigus kogutud andmestikke. Töö põhieesmärkideks oli: (1) anda lühiülevaade Eesti rannikumere põhjakoosluste peamistest uuringumeetoditest (I–V), sh määrata kindlaks merepõhja koosluste võtmeliikide kaardistamiseks sobivaim videovaatlus meetod (III);

(2) kaasajastada pika meriheina levikukaart Eesti rannikumeres (**I, V**); (3) anda ülevaade pika meriheina koosluse põhiparameetritest Eesti rannikumeres (**II**); (4) uurida kaugseire võimalusi meriheina koosluste kaardistamisel Eesti rannikumeres (**IV**); (5) määrata kindlaks kõige olulisemad pika meriheina esinemist mõjutavad keskkonnamuutujad ning ennustada muutusi meriheina levikumustris tänapäeva ja tuleviku kliima tingimustes (**V**).

Traditsioonilised merekeskkonna uurimismeetodid – merepõhjust proovide kogumine kas sukelduja poolt või pinnalt põhjaammute abil on Eesti rannikumere põhjakoosluste uurimisel kasutusel alates 1959. aastast. Mõlemad meetodid võimaldavad koguda detailset infot koosluste kohta ning on laialt kasutusel ka tänapäeval. Siiski, kuna eelnimetatud meetodid on tööjõumahukad, kallid ning aeganõudvad, on suurte merealade kaardistamisel vajalik rakendada kui muid uuringumeetodeid. Veepinnalt teostatavad videovaatlused on Eestis kasutusel alates 2005. aastast ning kuna meetod võimaldab koguda infot võtmelekkide katvuse kohta kiiremini ning odavamalt kui sukelduja või põhjaammuti, on see tänapäeval merepõhjaelustiku kaardistamisel laialt kasutusel (**I, V**). Väljatõtatud videopõhised või videost pärineva 20 või 35 foto põhised hinnangud on piisava täpsusega ning sobivad kasutamiseks Läänemere kirdeosa põhjaelustiku koosluste kaardistamisel (**III**).

Pikk merihein on Eesti rannikumeres avatud liivastel põhjadel üks olulisemaid võtmeliike (**I, II**). Aastatel 2005–2015 tuvastati üle 300 pika meriheina kasvukoha. Liik on tavapärane kogu Eesti rannikumeres, välja arvatud madala soolsusega piirkonnad (Soome lahe idaosa ning Pärnu laht) (**I–V**). Liik kasvab peamiselt sügavusvahemikus 2–5 m (**I, II, V**). Pika meriheina levikuala, kus liigi katvus ületab 10%, on hinnanguliselt 150 km² (<0,5% Eesti mereala pindalast) (**I**). Läänemeres on pika meriheina praegune areaal hinnanguliselt 1200 km². Liigi katvuse sesoonne varieeruvus oli väike, ruumiline varieeruvus oli enam mõjutatud sügavusest (sh valgus, temperatuur, sette iseloom) ning varieerus kitsastest tihedatest meriheinavöönditest ulatuslike hõredate kooslusteni (**II**). Meriheina biomass varieerus vahemikus 7–70 g/m⁻² (kuivkaal) ning võsude tihedus koosluses varieerus peamiselt vahemikus (50)133–1300 tk/m⁻² (**II**) – antud väärtused on kirjanduse andmetel sarnased kogu Läänemere ulatuses. Erinevalt teistest Läänemere piirkondadest jätkus pika meriheina kasv Eesti rannikumeres ka septembris (**II**). Pika meriheina kooslustest leiti kokku 35 põhjataimestiku ning 33 põhjaloomastiku liiki, tavapäraselt esines ühes meriheinakoosluses 9–15 põhjataimestiku ning 8–21 põhjaloomastiku liiki (**I, II**). Pika meriheina kooslustes esinevad liigid on tavapärased kogu Eesti rannikumeres (ainult *Cladosiphon zosterae* esinemine on seotud pika meriheina levikuga) (**I, II**), leitud liikide arv on umbes neljandik piirkonna põhjaelustiku liikide arvust. Pika meriheina ning teiste Eesti rannikumeres levinud kõrgemate taimede elupaigad kattuvad suures osas (**V**) ning liikide arv on kõrge kõikides kõrgemate taimedega elupaikades, samas on nende koosluste liigiline koosseis erinev (**I, II**). Pika meriheina koosluste biomassi ning kooslusega seotud põhjaloomastiku liikide arvukuse vahel on tugev lineaarne seos (**II**), mis viitab

sellele, et meriheina poolt pakutav struktuur ja elukeskkond on loomastiku jaoks olulisemad kui muud lokaalsed keskkonnatingimused.

Mitmed uued meetodid, nagu merepõhja elupaikade hüperspektraalne kaugseire, ennustav ruumiline modelleerimine ja masinõpe on järjest enam kasutatavad. Nende kasutamine on osutunud eriti väärtuslikuks madalas rannikumeres, kus esineb suur ruumiline varieeruvus, mida traditsiooniliste uurimismeetodite abil pole võimalik kaardistada. Traditsiooniliste ja uute meetodite kombineerimine on hetkel parim lähenemine, mõistmaks liikide levikumustreid ning nende muutusi merekeskkonnas. Käesolevas töös välja töötatud kaugseire klassifikatsiooniskeem võimaldas luua täpsed kõrge lahutusvõimega põhjakoosluste levikukaardid 0–6 m sügavusel rannikuvees, sh eristades enimlevinud põhjataimestiku rühmad – rohe-, pruun- ja punavetikad, mändvetikad ja kõrgemad taimed (IV). Kuigi kõikide koosluses esinevate liikide eristamine ei ole alati detailselt võimalik, võimaldavad väljatöötatud levikukaardid ka liigipõhiste uuringute optimaalsemat planeerimist. Demonstreerimaks ruumilise modelleerimise võimalusi merepõhjakoosluste leviku hindamisel, kasutati doktoritöös näitena võimendatud regressioonipuu meetodit (boosted regression tree – BRT), mis võimaldas hinnata mittelineaarseid seoseid pika meriheina katvuse ning keskkonna vahel ning ennustada muutusi liigi katvuses nii praeguse kui tulevase kliima tingimustes (V).

Modelleerimise tulemused kinnitasid, et väikese- ja suureskaalaline keskkonnamuutlikkus mõjutavad nii eraldi kui koos vaadatuna pika meriheina katvust ning levikut (V). Tuleviku kliima stsenaariumi kohaselt on füüsikalistel muutujatel, sh merevee soojenemisel, lainetuse poolt põhjustatud vee liikumiskiiruse tõusul ning laguneva jää poolt põhjustatud mehaanilisel häiringul madalas rannikumeres suurem mõju pika meriheina levikule kui soolsuse vähenemisel, vee hägususe ning pelaagilise produktsiooni suurenemisel (V). Meie modelleerimine näitas, et pikk merihein on vastupidav väga suurtele keskkonnatingimuste muutustele ning merevee temperatuuri tõusuga kaasneb eeldatavalt liigi biomassi suurenemine (V). Vastavalt levinud tuleviku kliima stsenaariumile on pika meriheina biomassi suurenemine peamiselt seostatav kiirema taimestiku arenguga kevadel ning pikema kasvuperioodiga. Antud tulevikuennustused pika meriheina leviku muutustest võimaldavad merekaitsealade paremat planeerimist ning aitavad tagada liigilise mitmekesisuse säilumist Läänemere idaosas. Pika meriheina koosluste püsimise tagamiseks on Läänemeres vajalik ka laiapõhjalisem koostöö ning üldine teadlikkuse tõstmine nende koosluste unikaalsusest ning väärtuslikkusest.

ACKNOWLEDGEMENTS

Completing this thesis took longer than expected, but it surely has been a great opportunity and challenge. I am truly grateful to my supervisor Georg Martin, who introduced me to the marine biology research field and guided me to the underwater world and seagrasses. Thank you for your great support and patient teaching for more than a decade.

My deepest gratitude belongs also to my second supervisor Jonne Kotta, whose enthusiasm, knowledge, wisdom and friendliness have been a great example and motivation at all times. Your help with the research and compiling the thesis has been enormous.

My co-author and former room-mate at work Kristjan Herkül – I still don't know how you put up with all my endless questions whether these were about life, love, weather, GIS, statistics, articles, teaching or whatever from day to day from morning to late at night – what does it mean, what does this do, how do I get it fixed, where did it go, how does it work etc. etc. etc. Most of the time your 'magic presence' was just about enough to solve a problem. Thank you and Priit Kersen and Teemar Püss, who shared the same room – thank you for tolerating me!

I want to thank Kaire Kaljurand for all the help with diving and for long-long, sometimes too long (for others, that is) hours of fieldwork. It was a pleasure to be on the same boat with you.

I also want to thank Helen Orav-Kotta for all the help in the office and in personal matters and last but not least for introducing me to the world of ceramics.

I am glad to know Maria Pöldma, who has been a great friend and support during the process of writing the thesis. Talking for hours in the middle of the night at an empty car park – I really thought I had lost that skill, but no – you just brought it back to life.

I am grateful to all the colleagues and co-authors in the Estonian Marine Institute situated in different cities. The sea is a place where one should never be alone. Thanks to you I never felt alone whether on sea or in the office.

Thank you – Kristiina Jürgens, Maili Peterson, Kaile Peet for keeping my two feet on the ground since the beginning of university studies. Somehow the phrase 'how does the lizard get to the island' keeps popping up when I think about our studies. And yes, I can picture your harried faces when reading it. Nevertheless, your friendship means a lot to me.

I am grateful to Volunteer Rescue Service and for the people that came into my life through that institution – just to name a few – Eve Salumaa, Tiit Umbaar, Krista Jaamul, Tiina Sutt, Liis Truubon, Ülle Sirkas – you were all a great help and having a cup of tea or a hike in the seaside or forest with you was fun and most needed in some difficult times. Also thank you for help in the fieldwork – you were there when it was needed.

My thanks go to Tiia Kaare and Tiina Paalme for revising the thesis and Margret Sisask for arranging the necessary paperwork.

My family has always supported my studies and work. My mother and deceased father – thank you for believing in me and letting me do things my own way (with some debates surely). Kaie – thank you for always being close, even if being on the other side of the world. I will try to distribute all the kindness you have shown to me. Thank you Mati for being someone I could always look up to and be proud of. Kristel, Maria, Kaspar, Marta-Marie, Olaf, Oliver, Martin – it is my honour to know you.

Alo, Joosep, Jesper and Leeni – you are the main reason why my years of being in the university list just piled and piled. Yet without having you and all the endless hullabaloo through days and nights, my life would be meaningless. Thank you for being such a big part of my life and heart!

Most of the fieldwork was done in the framework of the European Union LIFE project ‘Marine Protected Areas in the Eastern Baltic Sea (Baltic MPAs)’ (LIFE05NAT/LV/000100) and the Central Baltic Interreg Iva Programme Hispares. Funding for the research was also provided by Estonian Targeted Financing Grants SF0180013s08 and SF0180009sll of the Estonian Ministry of Education and Research, Institutional research funding grant IUT2-20 of the Estonian Research Council and Estonian Science Foundation grants 5927, 6015, 7813, 8254, 8807 and 9439. The studies also received funding from the BONUS project BIO-C3, the joint Baltic Sea research and development programme (Art 185), funded jointly from the European Union’s Seventh Programme for research, technological development and demonstration and from the Estonian Research Council. The study has been also supported by the projects ‘The status of marine biodiversity and its potential futures in the Estonian coastal sea’ grant no 3.2.0801.11-0029 and ‘Estkliima’ grant no 3.2.0802.11-0043 of the Environmental Protection and Technology Programme of the European Regional Development Fund.

Scholarships from the Archimedes Foundation, Nordic Seagrass Network and Doctoral School of Earth Sciences and Ecology were a great financial support that helped me to discover the aspects of seagrass ecosystems in different parts of the world.

REFERENCES

- Andrew, N.L., Mapstone, B.D. 1987. Sampling and the description of spatial patterns in marine ecology. *Oceanogr. Mar. Biol. Annu. Rev.* 25, 39–90.
- Anonymous. 2003a. Common Implementation Strategy for the Water Framework Directive (2000/60/EC). Monitoring under the Water Framework Directive. Guidance Document No 7. Produced by Working Group 2.7 – Monitoring: 160 pp.
- Anonymous. 2003b. Common Implementation Strategy for the Water Framework Directive (2000/60/EC). Guidance on typology, reference conditions and classification systems for transitional and coastal waters. Produced by Working Group 2.4 – Coast: 121 pp.
- Araújo, M.B., New, M. 2007. Ensemble forecasting of species distributions. *Trends Ecol. Evol.* 22, 42–47.
- BACC Author Team, ed. 2008. Assessment of Climate Change for the Baltic Sea Basin. Springer Verlag: 474 pp.
- Bäck, S., Lehvo, A., Blomster, J. 2000. Mass occurrence of unattached *Enteromorpha intestinalis* on the Finnish Baltic Sea coast. *Ann. Bot. Fenn.* 37, 155–161.
- Baden, S., Boström, C. 2001. The leaf canopy of seagrass beds: faunal community structure and function in a salinity gradient along the Swedish coast. In: *Ecological Comparisons of Sedimentary Shores*, (Reise, K. ed.), pp. 213–236. *Ecological Studies* 151, Springer Verlag, Berlin/Heidelberg.
- Baden, S., Gullström, M., Lundén, B., Pihl, L., Rosenberg, R. 2003. Vanishing seagrass *Zostera marina*, L. in Swedish coastal waters. *Ambio* 32, 374–377.
- Baden, S., Emanuelsson, A., Pihl, L., Svensson, C.J., Åberg, P. 2012. Shift in seagrass food web structure over decades is linked to overfishing. *Mar. Ecol. Prog. Ser.* 451, 61–73.
- Bintz, J.C., Nixon, S.W., Buckley, B.A., Granger, S.L. 2003. Impacts of temperature and nutrients on coastal lagoon plant communities. *Estuaries* 26, 765–776.
- Boström, C., Bonsdorff, E. 2000. Zoobenthic community establishment and habitat complexity the importance of seagrass shoot-density, morphology and physical disturbance for faunal recruitment. *Mar. Ecol. Prog. Ser.* 205, 123–138.
- Boström, C., Bonsdorff, E., Kangas, P., Norkko, A. 2002. Longterm changes in a brackish water *Zostera marina* community indicate effects of eutrophication. *Estuar. Coast. Shelf Sci.* 55, 795–804.
- Boström, C., Baden, S.P., Krause-Jensen, D. 2003. The seagrasses of Scandinavia and the Baltic Sea. In: *The World Atlas of Seagrasses* (Green, P., Short, F.T., eds.), pp. 27–37. University of California Press, Berkeley, USA.
- Boström, C., Roos, C., Rönnberg, O. 2004. Shoot morphometry and production dynamics of eelgrass in the northern Baltic Sea. *Aquat. Bot.* 79, 145–161.
- Boström, C., O'Brien, K., Roos, C., Ekeboom, J. 2006. Environmental variables explaining structural and functional diversity of seagrass macrofauna in an archipelago landscape. *J. Exp. Mar. Biol. Ecol.* 335, 52–73.
- Boström, C., Törnroos, A., Bonsdorff, E. 2010. Invertebrate dispersal and habitat heterogeneity: expression of biological traits in a seagrass landscape. *J. Exp. Mar. Biol. Ecol.* 390, 106–117.
- Boström, C., Baden, S., Bockelmann, A., Dromph, K., Frederiksen, S., Gustafsson, C., Krause-Jensen, D., Möller, T., Nielsen, S.L., Olesen, B., Olsen, J., Pihl, L., Rinde, E. 2014. Distribution, structure and function of Nordic seagrass ecosystems: impli-

- cations for coastal management and conservation. *Aquat. Conservat. Mar. Freshwat. Ecosyst.* 24, 410–434.
- Brown, C. J., Smith, S. J., Lawton, P., Anderson, J. T. 2011. Benthic habitat mapping: a review of progress towards improved understanding of the spatial ecology of the seafloor using acoustic techniques. *Estuar. Coast. Shelf Sci.* 92: 502–520.
- Bucas, M., Daunys, D., Olenin, S. 2007. Overgrowth patterns of the red algae *Furcellaria lumbricalis* at an exposed Baltic Sea coast: the result of a remote underwater video data analysis. *Estuar. Coast. Shelf Sci.* 75, 308–316.
- Byrne, M., Przeslawski, R. 2013. Multistressor impacts of warming and acidification of the ocean on marine invertebrates' life histories. *Integr. Comp. Biol.* 53, 582–596.
- Clarke, K.R. 1993. Non-parametric multivariate analysis of changes in community structure. *Aust. J. Ecol.* 18, 117–143.
- Clarke, K.R., Ainsworth, M. 1993. A method of linking multivariate community structure to environmental variables. *Mar. Ecol. Prog. Ser.* 92, 205–219.
- Clarke, K.R., Gorley, R.N., Somerfield, P.J., Warwick, R.M. 2014. Change in marine communities: an approach to statistical analysis and interpretation, 3rd edition. PRIMER-E, Plymouth: 260 pp.
- Clausen, K.K., Krause-Jensen, D., Olesen, B., Marba, N. 2014. Seasonality of eelgrass biomass across gradients in temperature and latitude. *Mar. Ecol. Prog. Ser.* 506, 71–85.
- Costanza, R., d'Arge, R., Groot, R., Farber, S., Grasso, M., Hannon, B., Limburg, K., Naeem, R.V., Paruelo, J., Raskin, R.G., Sutton, P., van den Belt, M. 1997. The value of the world's ecosystem services and natural capital. *Nature* 387, 253–260.
- De Boer, W. 2007. Seagrass-sediment interactions, positive feedbacks and critical thresholds for occurrence: A review. *Hydrobiologia* 591, 5–24.
- den Hartog, C. 1970. The Seagrasses of the World. North Holland Publ. Co, Amsterdam: 275 pp.
- Dennison, W.C., Alberte, R.S. 1986. Photoadaptation and growth of *Zostera marina* L. (eelgrass) transplants along a depth gradient. *J. Exp. Mar. Biol. Ecol.* 98, 265–282.
- Dernie, K.M., Kaiser, M.J., Warwick, R.M. 2003. Recovery rates of benthic communities following physical disturbance. *J. Anim. Ecol.* 72, 1043–1056.
- Diaz-Almela, E., Marba, N., Martinez, R., Santiago, R., Duarte, C.M. 2009. Seasonal dynamics of *Posidonia oceanica* in Magalluf Bay (Mallorca, Spain): temperature effects on seagrass mortality. *Limnol. Oceanogr.* 54, 2170–2182.
- Diekmann, O.E., Serrao, E.A., 2012. Range-edge genetic diversity: locally poor extant southern patches maintain a regionally diverse hotspot in the seagrass *Zostera marina*. *Mol. Ecol.* 21, 1647–1657.
- Doledec, S., Chessel, D., Gimaret-Carpentier, C. 2000. Niche separation in community analysis: a new method. *Ecology* 81, 2914–2927.
- Dray, S., Dufour, A.B. 2007. The ade4 package: implementing the duality diagram for ecologists. *J. Stat. Softw.* 22, 1–20.
- Drury, D. M. 2004. Effects of grass shrimp (*Palaemonetes* spp.) density manipulations and nutrient enrichment on widgeongrass (*Ruppia maritima*) condition, epiphyte load, and epiphyte functional groups. Dissertations. 1971. http://aquila.usm.edu/theses_dissertations/1971 (accessed 03.04.2014).
- Duarte, C.M. 1989. Temporal biomass variability and production/biomass relationships of seagrass communities. *Mar. Ecol. Prog. Ser.* 51, 269–276.
- Duarte, C.M. 2002. The future of seagrass meadows. *Environ. Conserv.* 29, 192–206.

- Eleftheriou, A. (ed.) 2014. *Methods for the Study of Marine Benthos*, 4th Edition. Wiley-Blackwell: 496 pp.
- Elith, J., Leathwick, J.R., Hastie, T. 2008. A working guide to boosted regression trees. *J. Anim. Ecol.* 77, 802–813.
- Elmgren, R. 1989. Man's impact on the ecosystem of the Baltic Sea: energy flows today and at the turn of the century. *Ambio* 18, 326–332.
- Elmgren, R. 2001. Understanding human impact on the Baltic ecosystem: changing views in recent decades. *Ambio* 30, 222–231.
- ESRI. 2011. *ArcGIS Desktop: Release 10*. Environmental Systems Research Institute, Redlands, CA.
- Ferber, S., Stam, W.T., Olsen, J.L. 2008. Genetic diversity and connectivity remain high in eelgrass *Zostera marina* populations in the Wadden Sea, despite major impacts. *Mar. Ecol. Prog. Ser.* 372, 87–96.
- Fonseca, M.S., Bell, S.S. 1998. Influence of physical setting on seagrass landscapes near Beaufort, North Carolina. *Mar. Ecol. Prog. Ser.* 171, 109–121.
- Fonseca, M.S., Kenworthy, W.J. 1987. Effects of current on photosynthesis and distribution of seagrass. *Aquat. Bot.* 27, 59–78.
- Fonseca, M.S., Zieman, J.C., Thayer, G.W., Fisher, J.S. 1983. The role of current velocity in structuring eelgrass *Zostera marina* meadows. *Estuar. Coast. Shelf Sci.* 17, 367–380.
- Fonseca, M.S., Kenworthy, W.J., Whitfield, P.E. 2000. Temporal dynamics of seagrass landscapes: a preliminary comparison of chronic and extreme disturbance events. In: *Proceedings of the Fourth International Seagrass Biology Workshop*. September 25–October 2, 2000, Corsica, France (Pergent, G., Pergent-Martini, C., Buia, M.C., Gambi, M.C. eds.), *Biologia Marina Mediterranea*, Instituto di Zoologia, Genova, Italy: pp. 373–376.
- Fredriksen, S., Christie, H., Sæthre, B.A. 2005. Species richness in macroalgae and macrofauna assemblages on *Fucus serratus* L. (Phaeophyceae) and *Zostera marina* L. (Angiospermae) in Skagerrak, Norway. *Mar. Biol. Res.* 1, 2–19.
- Fredriksen, S., De Backer, A., Boström, C., Christie, H. 2010. Infauna from *Zostera marina* (L.) meadows in Norway. Differences in vegetated and unvegetated areas. *Mar. Biol. Res.* 6, 189–200.
- Gil, M., Armitage, A.R., Fourqurean, J.W. 2006. Nutrient impacts on epifaunal density and species composition in a subtropical seagrass bed. *Hydrobiologia* 569, 437–447.
- Gisiger, T. 2001. Scale invariance in biology: coincidence or footprint of a universal mechanism? *Biol. Rev.* 76, 161–209.
- Gjerde, K.M. 2006. *Ecosystems and biodiversity in deep waters and high seas*. UNEP Regional Seas Reports and Studies 178, UNEP/IUCN, Gland: 58 pp.
- Glemarec, M., LeFaou, Y., Cuq, F. 1997. Long-term changes of seagrass beds in the Glenan Archipelago (South Brittany). *Oceanol. Acta* 20, 217–227.
- Gonciarz, M., Wiktor, J., Tatarek, A., Węgleński, P., Stanković, A. 2014. Genetic characteristics of three Baltic *Zostera marina* populations. *Oceanologia* 56, 2014, 549–564.
- Gustafsson, C., Boström, C. 2009. Effects of plant species richness and composition on epifaunal colonization in brackish water angiosperm communities. *J. Exp. Mar. Biol. Ecol.* 382, 8–17.
- Håkanson, L., Eckhell, J. 2005. Suspended particulate matter (SPM) in the Baltic Sea – new empirical data and models. *Ecol. Model.* 189, 130–150.
- Halley, J.M. 1996. Ecology, evolution and 1/f-noise. *Trends Ecol. Evol.* 11, 33–37.

- Hällfors, G., Niemi, Å., Ackefors, H., Lassig, J., Leppäkoski, E. 1981. Biological oceanography (chapter 5). In: The Baltic Sea (Voipio, A. ed.), Elsevier Oceanography Series 30, pp. 219–274. Amsterdam, The Netherlands.
- Halpern, B.S., Walbridge, S., Selkoe, K.A., Kappel, C.V., Micheli, F., D'Agrosa, C., Bruno, J.F., Casey, K.S., Ebert, C., Fox, H.E., Fujita, R., Heinemann, D., Lenihan, H.S., Madin, E.M.P., Perry, M.T., Selig, E.R., Spalding, M., Steneck, R., Watson, R. 2008. A global map of human impact on marine ecosystems. *Science* 319, 948–952.
- Hastie, T., Tibshirani, R., Friedman, J.H. 2009. *The Elements of Statistical Learning: Data Mining, Inference, and Prediction*. Springer-Verlag, New York: 744 pp.
- Hawkins, S.J., Firth, L.B., McHugh, M., Poloczanska, E.S., Herbert, R.J.H., Burrows, M.T., Kendall, M.A., Moore, P.J., Thompson, R.C., Jenkins, S.R., Sims, D.W., Genner, M.J., Mieszkowska, N. 2013. Data rescue and re-use: recycling old information to address new policy concerns. *Mar. Policy* 42, 91–98.
- Heck, K.L. Jr., Hays, G., Orth, R.J. 2003. Critical evaluation of the nursery role hypothesis for seagrass meadows. *Mar. Ecol. Prog. Ser.* 253, 123–136.
- Heine, J.N. 1989. Effects of ice scour on the structure of sublittoral marine algal assemblages of St. Lawrence and St. Matthew Islands, Alaska. *Mar. Ecol. Prog. Ser.* 52, 253–260.
- HELCOM 2015. *Manual for Marine Monitoring in the COMBINE Programme of HELCOM*. Helsinki. <http://helcom.fi/action-areas/monitoring-and-assessment/manuals-and-guidelines/combine-manual> (accessed 09.12.2016).
- Hemminga, M.A., Duarte, C.M. 2000. *Seagrass Ecology*. Cambridge University Press, Cambridge, UK: 298 pp.
- Hense, I., Meier, H.E.M., Sonntag, S. 2013. Projected climate change impact on Baltic Sea cyanobacteria. *Clim. Change* 119, 391–406.
- Hoffman, J.R., Hansen, L.J., Klinger, T. 2003. Interactions between UV radiation and temperature limit inferences from single-factor experiments. *J. Phycol.* 39, 268–272.
- Homziak, J., Fonseca, M.S., Kenworthy, W.J. 1982. Macrobenthic community structure in a transplanted eelgrass (*Zostera marina*) meadow. *Mar. Ecol. Prog. Ser.* 9, 211–221.
- Houk, P., Van Woesik, R. 2006. Coral reef benthic video survey facilitates long-term monitoring in the Commonwealth of the Northern Mariana Islands: towards an optimal sampling strategy. *Pac. Sci.* 60, 177–189.
- International Panel on Climate Change (IPCC), 2013. *Climate Change 2013: The Physical Science Basis*. Cambridge University Press, Cambridge, UK: 222 pp. https://www.ipcc.ch/pdf/assessment-report/ar5/wg1/WG1AR5_SummaryVolume_FINAL.pdf (accessed 01.12.2016).
- Jedrzejcak, M. F. 2002. Stranded *Zostera marina* L. vs wrack fauna community interactions on a Baltic sandy beach (Hel, Poland): A short term pilot study, Part II. *Oceanologia* 44, 367–87.
- Jephson, T., Nyström, P., Moksnes, P., Baden, S. 2008. Trophic interactions in *Zostera marina* beds along the Swedish coast. *Mar. Ecol. Prog. Ser.* 369, 63–76.
- Kaiser, M.J., Attrill, M.J., Jennings, S., Thomas, D.N., Barnes, D.K.A., Brierley, A.S., Hiddink, J.G., Kaartokallio, H., Polunin, N.V.C., Raffaelli, D.G. 2011. *Marine Ecology. Processes, Systems and Impacts*. Second edition. Oxford University Press, New York: 528 pp.
- Karlson, R.H., Cornell, H.V. 1998. Scale-dependent variation in local vs. regional effects on coral species richness. *Ecol. Monogr.* 68, 259–274.

- Kautsky, L. 1988. Life strategies of aquatic soft bottom macrophytes. *Oikos* 53, 126–135.
- Kautsky, H., van der Maarel, E. 1990. Multivariate approaches to the variation in phytobenthic communities and environmental vectors in the Baltic Sea. *Mar. Ecol. Prog. Ser.* 60, 169–184.
- Kendrick, G.A., Hegge, B.J., Wyllie, A., Davidson, A., Lord, D.A. 2000. Changes in seagrass cover on Success and Parmelia Banks, Western Australia between 1965 and 1995. *Estuar. Coast. Shelf Sci.* 50, 341–353.
- Kendrick, G.A., Holmes, K.W., van Niel, K.P. 2008. Multi-scale patterns of three seagrass species with different growth dynamics. *Ecography* 31, 191–200.
- Kiirikki, M., Blomster, J. 1996. Wind induced upwelling as a possible explanation for mass occurrences of epiphytic *Ectocarpus siliculosus* (Phaeophyta) in the northern Baltic Proper. *Mar. Biol.* 127, 353–358.
- Koch, M., Bowes, G., Ross, C., Zhang, X.-H. 2013. Climate change and ocean acidification effects on seagrasses and marine macroalgae. *Glob. Change Biol.* 19, 103–132.
- Kotta, J., Lauringson, V., Kotta, I. 2007. Response of zoobenthic communities to changing eutrophication in the northern Baltic Sea. *Hydrobiologia* 580, 97–108.
- Kotta, J., Jaanus, A., Kotta, I. 2008a. Haapsalu and Matsalu Bays. In: *Ecology of Baltic Coastal Waters* (Schiewer, U. Ed.), *Ecol. Stud.* 197, pp. 245–258, Springer.
- Kotta, J., Lauringson, V., Martin, G., Simm, M., Kotta, I., Herkül, K., Ojaveer, H. 2008b. Gulf of Riga and Pärnu Bay. In: *Ecology of Baltic Coastal Waters* (Schiewer, U. Ed.), *Ecol. Stud.* 197, pp. 217–243, Springer.
- Kotta, J., Paalme, T., Püss, T., Herkül, K., Kotta, I. 2008c. Contribution of scale-dependent environmental variability on the biomass patterns of drift algae and associated invertebrates in the Gulf of Riga, northern Baltic Sea. *J. Marine Syst.* 74, S116–S123.
- Kotta, J., Lauringson, V., Kaasik, A., Kotta, I. 2012. Defining the coastal water quality in Estonia based on benthic invertebrate communities. *Estonian J. Ecol.* 61, 86–105.
- Kotta, J., Kutser, T., Teeveer, K., Vahtmäe, E., Pärnoja, M. 2013. Predicting species cover of marine macrophyte and invertebrate species combining hyperspectral remote sensing, machine learning and regression techniques. *PlosOne* 8, e63946.
- Krause-Jensen, D., Middelboe, A L., Sand-Jensen, K., Christensen, P.B. 2000. Eelgrass, *Zostera marina*, growth along depth gradients: upper boundaries of the variation as a powerful predictive tool. *Oikos* 91, 233–244.
- Krause-Jensen, D., Pedersen, M.F., Jensen, C., 2003. Regulation of eelgrass (*Zostera marina*) cover along depth gradients in Danish coastal waters. *Estuaries* 26, 866–877.
- Krause-Jensen, D., Sagert, S., Schubert, H., Boström, C. 2008. Empirical relationships linking distribution and abundance of marine vegetation to eutrophication. *Ecol. Indic.* 8, 515–529.
- Kutser, T., Vahtmäe, E., Martin, G. 2006. Assessing suitability of multispectral satellites for mapping benthic macroalgal cover in turbid coastal waters by means of model simulations. *Estuar. Coast. Shelf Sci.* 67, 521–529.
- Larkum, A.W.D., Orth, R.J., Duarte, C. (eds). 2006. *Seagrasses: Biology, Ecology and Conservation*. Springer, Dordrecht, Netherlands: 691 pp.
- Lasker, H.R., Coffroth, M.A. 1999. Responses of clonal reef taxa to environmental change. *Am. Zool.* 39, 92–103.

- Lefcheck, J., Wilcox, D.J., Murphy, R.B., Marion, S.R., Orth, R.J. 2017. Multiple stressors threaten the imperiled coastal foundation species eelgrass (*Zostera marina*) in Chesapeake Bay, USA. *Global Change Biol.*, doi: 10.1111/gcb.13623.
- Levin, S.A. 1992. The problem of pattern and scale in ecology. *Ecology* 73, 1943–1967.
- Lotze, H.K., Schramm, W., Schories, D., Worm, B. 1999. Control of macroalgal blooms at early developmental stages: *Pilayella littoralis* versus *Enteromorpha* spp. *Oecologia* 119, 46–54.
- Lundberg, C. 2005. Eutrophication in the Baltic Sea – from area-specific biological effects to interdisciplinary consequences. PhD thesis. Faculty of Mathematics and Natural Sciences, Department of Biology, Abo Akademi University, Abo: 166 pp.
- Madsen, J.D., Chambers, P.A., James, W.F., Koch, E.W., Westlake, D.F. 2001. The interaction between water movement, sediment dynamics and submersed macrophytes. *Hydrobiologia* 444, 71–84.
- Mallet, D., Pelletier, D. 2014. Underwater video techniques for observing coastal marine biodiversity: a review of sixty years of publications (1952–2012). *Fish. Res.* 154, 44–62.
- Marba, N., Cebrian, J., Enriquez, S., Duarte, C.M. 1996. Growth patterns of Western Mediterranean seagrasses: species-specific responses to seasonal forcing. *Mar. Ecol. Prog. Ser.* 133, 203–215.
- Marsh, J.A. Jr., Dennison, W.C., Alberte, R.S. 1986. Effects of temperature on photosynthesis and respiration in eelgrass (*Zostera marina* L.). *J. Exp. Mar. Biol. Ecol.* 101, 257–267.
- McGlathery, K.J. 2001. Macroalgal blooms contribute to the decline of seagrass in nutrient enriched coastal waters. *J. Phycol.* 37, 453–456.
- Menge, B.A., Olson, A.M. 1990. Role of scale and environmental factors in regulation of community structure. *Trends Ecol. Evol.* 5, 52–57.
- Middelboe, A.L., Sand-Jensen, K., Krause-Jensen, D. 2003. Spatial and interannual variations with depth in eelgrass populations. *J. Exp. Mar. Biol. Ecol.* 291, 1–15.
- Moberg, F., Folce, C. 1999. Ecological goods and services of coral reef ecosystems. *Ecol. Econ.* 29, 215–233.
- Möller, T., Martin, G. 2007. Distribution of the eelgrass *Zostera marina* L. in the coastal waters of Estonia, NE Baltic Sea. *Proc. Est. Acad. Sci. Biol. Ecol.* 56, 270–277.
- Moore, K.A., Short, F.T. 2006. *Zostera*: biology, ecology, and management. In: *Seagrasses: Biology, Ecology and Conservation* (Larkum, A. W. D., Orth, R. J., Duarte, C., eds.), pp. 361–386. Springer, Dordrecht, Netherlands.
- Moore, K.A., Shields, E.C., Parrish, D.B. 2014. Impacts of varying estuarine temperature and light conditions on *Zostera marina* (eelgrass) and its interactions with *Ruppia maritima* (widgeon grass). *Estuar. Coast.* 37, 20–30.
- Morin, A. 1985. Variability of density estimates and the optimization of sampling programs for stream benthos. *Can. J. Fish. Aquat. Sci.* 42, 1530–1534.
- Müller, R., Laepple, T., Bartsch, I., Wiencke, C. 2009. Impact of oceanic warming on the distribution of seaweeds in polar and cold-temperate waters. *Bot. Mar.* 52, 617–638.
- Murdoch, T.J.T., Aronson, R.B. 1999. Scale-dependent spatial variability of coral assemblages along the Florida reef tract. *Coral Reefs* 18, 341–351.
- Nejrup, L.B., Pedersen, M.F. 2008. Effects of salinity and water temperature on the ecological performance of *Zostera marina*. *Aquat. Bot.* 88, 239–246.

- Nelson, T.A., Lee, A. 2001. A manipulative experiment demonstrates that blooms of the macroalga *Ulvaria obscura* can reduce eelgrass shoot density. *Aquat. Bot.* 71, 149–154.
- Nordlund, L.M., Koch, E.W., Barbier, E.B., Creed, J.C. 2017. Seagrass ecosystem services and their variability across genera and geographical regions. *PLOS ONE* 12(1): e0169942.
- Norse, E., Crowder, L.B. 2005. Why marine conservation biology? In: *Marine Conservation Biology* (Norse, E., Crowder, L.B. eds.), pp. 1–18. Island Press, Washington, DC.
- Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R., O'Hara, B., Simpson, G.L., Solymos, P., Stevens, H.M.H., Szoecs, E., Wagner, H. 2017. *vegan: Community Ecology Package*. R package version 2.0–10. <http://CRAN.R-project.org/package=vegan>, (accessed 20.07.2013).
- Olesen, B., Sand-Jensen, K. 1994. Demography of shallow eelgrass (*Zostera marina*) populations – shoot dynamics and biomass development. *J. Ecol.* 82, 379–390.
- Olsen, J., Rouzé, P., Verhelst, B., Lin, Y.-C., Bayer, T., Collen, J., Dattolo, E., De Paoli, E., Dittami, S., Maumus, F., Michel, G., Kersting, A., Lauritano, C., Lohaus, R., Töpel, M., Tonon, T., Vanneste, K., Amirebrahimi, M., Brakel, J., Boström, C., Chovatia, M., Grimwood, J., Jenkins, J.W., Jueterbock, A., Mraz, A., Stam, W.T., Tice, H., Bornberg-Bauer, E., Green, P.J., Pearson, G.A., Procaccini, G., Duarte, C.M., Schmutz, J., Reusch, T.B.H., Van de Peer, Y. 2016. The genome of the seagrass *Zostera marina* reveals angiosperm adaptation to the sea. *Nature* 530, 331–335.
- Orth, R.J. 1977. Effect of nutrient enrichment on growth of the seagrass *Zostera marina* in the Chesapeake Bay, Virginia, USA. *Mar. Biol.* 44, 187–194.
- Orth, R., Carruthers, T.J.B., Dennison, W.C., Duarte, C.M., Fourqurean, J.W., Heck, J.K.L., Hughes, A.R., Kendrick, G., Kenworthy, W.J., Olyarnik, S., Short, F.T., Waycott, M., Williams, S.L. 2006. A global crisis for seagrass ecosystems. *BioScience* 56, 987–996.
- Oviatt, C.A. 2004. The changing ecology of temperate coastal waters during a warming trend. *Estuaries* 27, 895–904.
- Paalme, T., Martin, G., Kotta, J., Kukk, H., Kaljurand, K. 2004. Distribution and dynamics of drifting macroalgal mats in Estonian coastal waters during 1995–2003. *Proc. Est. Acad. Sci. Biol. Ecol.* 53, 260–268.
- Peralta, G., Perez-Llorens, J.L., Hernandez, I., Vergara, J.J. 2002. Effects of light availability on growth, architecture and nutrient content of the seagrass *Zostera noltii* Hornem. *J. Exp. Mar. Biol. Ecol.* 269, 9–26.
- Perez-Llorens, J.L., Niell, F.X. 1993. Temperature and emergence effects on the net photosynthesis of 2 *Zostera noltii* Hornem. morphotypes. *Hydrobiologia* 254, 53–64.
- Pergent, G., Pergent-Martini, C., Bein, A., Dedeken, M., Oberti, P., Orsini, A., Santucci, J.F., Short, F. 2015. Dynamic of *Posidonia oceanica* seagrass meadows in the northwestern Mediterranean: Could climate change be to blame? *C R Biol.* 338, 484–493.
- Philippart, C.J.M. 1995. Effect of periphyton grazing by *Hydrobia ulvae* on the growth of *Zostera noltii* on a tidal flat in the Dutch Wadden Sea. *Mar. Biol.* 122, 431–437.
- Pihl, I., Isaksson, I., Wennhage, H., Moksnes, P.-O. 1995. Recent increase of filamentous algae in shallow Swedish bays, effects on the community structure of epibenthic fauna and fish. *Neth. J. Aquat. Ecol.* 29, 349–358.

- Pihl, L., Baden, S., Kautsky, N., Rönnbäck, P., Söderqvist, T., Troell, M., Wennhage, H. 2006. Shift in fish assemblage structure due to loss of seagrass *Zostera marina* habitats in Sweden. *Estuar. Coast. Shelf Sci.* 67, 123–132.
- Platt T., Denman K.L. 1975. Spectral analysis in ecology. *Annu. Rev. Ecol. Syst.* 6, 189–210.
- Prasad, S.N., Pal, D., Römkens, M.J.M. 2000. Wave formation on a shallow layer of flowing grains. *J. Fluid Mech.* 413, 89–110.
- Rasmussen, J.R., Pedersen, M.F., Olesen, B., Nielsen, S.L., Pedersen, T.M. 2013. Temporal and spatial dynamics of ephemeral drift-algae in eelgrass, *Zostera marina*, beds. *Estuar. Coast. Shelf Sci.* 119, 167–175.
- RDC Team. 2013. R: a Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.Rproject.org/> (accessed 20.07.2013).
- Reusch, T.B.H., Boström, C., Stam, W.T., Olsen, J.L. 1999. An ancient eelgrass clone in the Baltic Sea. *Mar. Ecol. Prog. Ser.* 183, 301–304.
- Reusch, T.B.H., Ehlers, A., Hämmerli, A., Worm, B. 2005. Ecosystem recovery after climatic extremes enhanced by genotypic diversity. *Proc. Nat. Acad. Sci. USA*, 102, 2826–2831.
- Reynaud, S., Leclercq, N., Romaine-Lioud, S., Ferrier-Pages, C., Jaubert, J., Gattuso, J.-P. 2003. Interacting effects of CO₂ partial pressure and temperature on photosynthesis and calcification in a scleractinian coral. *Glob. Change Biol.* 9, 1660–1668.
- Riddin, T., Adams, J.B. 2010. The effect of a storm surge event on the macrophytes of a temporarily open/closed estuary, South Africa. *Estuar. Coast. Shelf Sci.* 89, 119–123.
- Riegl, B., Korrubel, J.L., Martin, C. 2001. Mapping and monitoring of coral communities and their spatial patterns using a surface-based video method from a vessel. *B. Mar. Sci.* 69, 869–880.
- Rioja-Nieto, R., Sheppard, C. 2008. Effects of management strategies on the landscape ecology of a Marine Protected Area. *Ocean Coast. Manage.* 51, 397–404.
- Robertson, A.I., Mann, K.H. 1984. Disturbance of ice and life-history adaptations of the seagrass *Zostera marina*. *Mar. Biol.* 80, 131–141.
- Robinson, L.M., Elith, J., Hobday, A.J., Pearson, R.G., Kendall, B.E., Possingham, H.P., Richardson, A.J. 2011. Pushing the limits in marine species distribution modelling: lessons from the land present challenges and opportunities. *Global Ecol. Biogeogr.* 20, 789–802.
- Röhr, M.E., Boström, C., Canal-Vergés, P., Holmer, M. 2016. Blue carbon stocks in Baltic Sea eelgrass (*Zostera marina*) meadows. *Biogeosciences* 13, 6139–6153.
- Rosqvist, K., Mattila, J., Sandstrom, A., Snickars, M., Westerborn, M. 2010. Regime-shifts in vegetation composition of Baltic Sea coastal lagoons. *Aquat. Bot.* 93, 39–46.
- Salo, T., Pedersen, M.F. 2014. Synergistic effects of altered salinity and temperature on estuarine eelgrass (*Zostera marina*) seedlings and clonal shoots. *J. Exp. Mar. Biol. Ecol.* 457, 143–150.
- Salo, T., Pedersen, M.F., Boström, C. 2014. Population specific salinity tolerance in eelgrass (*Zostera marina*). *J. Exp. Mar. Biol. Ecol.* 461, 425–429.
- Salo, T., Reusch, T.B.H., Boström, C. 2015. Genotype-specific responses to light stress in eelgrass *Zostera marina*, a marine foundation plant. *Mar. Ecol. Prog. Ser.* 519, 129–140.

- Sand-Jensen, K. 1975. Biomass, net production and growth dynamics in an eelgrass (*Zostera marina* L.) population in Vellerup Vig, Denmark. *Ophelia* 14, 185–201.
- Sand-Jensen, K., Borum, J. 1983. Regulation of growth of eelgrass (*Zostera marina* L.) in Danish coastal waters. *Mar. Technol. Soc. J.* 17, 15–21.
- Santamaría, L. 2002. Why are most aquatic plants widely distributed? Dispersal, clonal growth and small-scale heterogeneity in a stressful environment. *Acta Oecol.* 23, 137–154.
- Schanz, A., Asmus, H. 2003. Impact of hydrodynamics on development and morphology of intertidal seagrasses in the Wadden Sea. *Mar. Ecol. Prog. Ser.* 261, 123–134.
- Schiel, D.R., Lilley, S.A. 2007. Gradients of disturbance to an algal canopy and the modification of an intertidal community. *Mar. Ecol. Prog. Ser.* 339, 1–11.
- Schneider, F.I., Mann, K.H. 1991. Rapid recovery of fauna following simulated ice rafting in a Nova Scotian seagrass bed. *Mar. Ecol. Prog. Ser.* 78, 57–70.
- Seitz, R.D., Wennhage, H., Bergström, U., Lipcius, R.N., Ysebaert, T. 2014. Ecological value of coastal habitats for commercially and ecologically important species. *ICES J. Mar. Sci.* 71, 648–665.
- Short, F.T. 1987. Effects of sediment nutrients on seagrasses: literature review and mesocosm experiment. *Aquat. Bot.* 27, 41–57.
- Short, F.T., Neckles, A.H. 1999. The effects of global climate change on seagrasses. *Aquat. Bot.* 63, 169–196.
- Short, F.T., Carruthers, T.J.R., Waycott, M., Kendrick, G.A., Fourqurean, J.W., Callabine, A., Kenworthy, W.J., Dennison, W.C. 2010. *Zostera marina*. The IUCN Red List of Threatened Species 2010: e.T153538A4516675. <http://dx.doi.org/10.2305/IUCN.UK.2010-3.RLTS.T153538A4516675.en>. Downloaded on 06 May 2017.
- Short, F.T., Polidoro, B., Livingstone, S.R., Carpenter, K.E., Bandeira, S., Bujang, J.S., Calumpong, H.P., Carruthers, T.J.B., Coles, R.G., Dennison, W.C., Erftemeijer, P.L.A., Fortes, M.D., Freeman, A.S., Jagtap, T.G., Kamal, A.H.M., Kendrick, G.A., Kenworthy, W.J., Nafie, Y.A.L., Nasution, I.M., Orth, R.J., Prathep, A., Sanciangco, J.C., van Tussenbroek, B., Vegara, S.G., Waycott, M., Zieman, J.C. 2011. Extinction risk assessment of the world's seagrass species. *Biol. Conserv.* 144, 1961–1971.
- Short, F.T., Kosten, S., Morgan, P.A., Malone, S., Moore E.A. 2016. Impacts of climate change on submerged and emergent wetland plants. *Aquat. Bot.* 135, 3–17.
- Spalding, M., Taylor, M., Ravilious, C., Short, F.T., Green, F. 2003. Global overview: the distribution and status of seagrasses. In: *World Atlas of Seagrasses* (Green, E.P., Short, F.T., eds.), pp. 5–26. UNEP World Conservation Monitoring Centre, University of California Press, Berkeley.
- StatSoft, Inc. 2006. STATISTICA (data analysis software system), version 7.1 and 8. www.statsoft.com
- Steele, J.H., Henderson, E.W. 1994. Coupling between physical and biological scales. *Philos. T. R. Soc. B.* 343, 5–9.
- Stramska, M., Swirgon, M. 2014. Influence of atmospheric forcing and freshwater discharge on interannual variability of the vertical diffuse attenuation coefficient at 490 nm in the Baltic Sea. *Remote Sens. Environ.* 140, 155–164.
- Suursaar, Ü., Astok, V., Otsmann, M. 1998. The front of Väinameri. *EMI Report Series* 9, 23–33.
- Svensson, C.J., Baden, S., Moksnes, P.-O., Åberg, P. 2012. Timely herbivores can prevent algal blooms in nutrient enriched environments. *Mar. Ecol. Prog. Ser.* 471, 61–71.

- Taylor, R., Fletcher, R.L., Raven, J.A. 2001. Preliminary studies on the growth of selected 'Green Tide' algae in laboratory culture: effects of irradiance, temperature, salinity and nutrients on growth rate. *Bot. Mar.* 44, 327–336.
- Touchette, B.W., Burkholder, J.M. 2000. Overview of the physiological ecology of carbon metabolism in seagrasses. *J. Exp. Mar. Biol. Ecol.* 250, 169–205.
- Townshend, J.R.G., Justice, C.O. 2002. Towards operational monitoring of terrestrial systems by moderate-resolution remote sensing. *Remote Sens. Environ.* 83, 351–359.
- Trei, T. 1973. Lääne-Eesti rannikuvete fütobentos [The phytobenthos of West-Estonian coastal waters]. Dissertation. Eesti Mereihüloogia laboratoorium, Tallinn (in Estonian). 34 pp.
- Trei, T. 1991. Taimed Läänemere põhjal [Plants on the bottom of the Baltic Sea]. Valgus, Tallinn (in Estonian). 144 pp.
- Vahteri, P., Mackinen, A., Salovius, S., Vuorinen, I. 2000. Are drifting algal mats conquering the bottom of the Archipelago Sea, SW Finland? *Ambio* 29, 338–343.
- Vahtmäe, E., Kutser, T., Martin, G., Kotta, J. 2006. Feasibility of hyperspectral remote sensing for mapping benthic macroalgal cover in turbid coastal waters – a Baltic Sea case study. *Remote Sens. Environ.* 101, 342–351.
- Van Katwijk, M.M., Hermus, D.C.R. 2000. Effects of water dynamics on *Zostera marina*: transplantation experiments in the intertidal Dutch Wadden Sea. *Mar. Ecol. Prog. Ser.* 208, 107–118.
- Viaroli, P., Bartoli, M., Fumagalli, I., Giordani, G. 1997. Relationship between benthic fluxes and macrophyte cover in a shallow brackish lagoon. *Water Air Soil Pollut.* 99, 533–540.
- Vuorinen, I., Hänninen, J., Rajasilta, M., Laine, P., Eklund, J., Montesino-Pouzols, F., Corona, F., Junker, K., Meier, H.E.M., Dippner, J.W. 2015. Scenario simulations of future salinity and ecological consequences in the Baltic Sea and adjacent North Sea areas – implications for environmental monitoring. *Ecol. Indic.* 50, 196–205.
- Wallentinus, I. 1984. Partitioning of nutrient uptake between annual and perennial seaweeds in a Baltic archipelago area. *Hydrobiologia* 116/117, 363–370.
- Waycott, M., Duarte, C.M., Carruthers, T.J.B., Orth, R.J., Dennison, W.C., Olyarnik, S., Calladine, A., Fourqurean, J.W., Heck, J.K.L., Hughes, A.R., Kendrick, G.A., Kenworthy, W.J., Short, F.T., Williams, S.L. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proc. Natl. Acad. Sci. USA* 106, 12377–12381.
- Wium-Andersen, S., Borum, J. 1984. Biomass variations and autotrophic production of an epiphyte-macrophyte community in a coastal Danish area: I. Eelgrass (*Zostera marina* L.) biomass and net production. *Ophelia* 23, 33–46.
- Worcester, S.E. 1995. Effects of eelgrass beds on advection and turbulent mixing in low current and low shoot density environments. *Mar. Ecol. Prog. Ser.* 126, 223–232.
- Wortmann, J., Hearne, J.W., Adams, J.B. 1997. A mathematical model of an estuarine seagrass. *Ecol. Model.* 98, 137–149.
- Zimmerman, R.C., Smith, R.D., Alberte, R.S. 1989. Thermal acclimation and whole-plant carbon balance in *Zostera marina* L. (eelgrass). *J. Exp. Mar. Biol. Ecol.* 130, 93–109.
- Zimmerman, R.C., Kohrs, D.G., Steller, D.L., Alberte, R.S. 1997. Impacts of CO₂ enrichment on productivity and light requirements of eelgrass. *Plant. Physiol.* 115, 599–607.

PUBLICATIONS

CURRICULUM VITAE

Name: Tiia Möller
Date of Birth: 11.06.1982
Citizen: Estonian
Address: Mäealuse 14, 12618 Tallinn, Estonia
Phone: +372 5027672
E-mail: tiia.moller@ut.ee

Education:

2007 MSc, University of Tartu, hydrobiology
2004 BSc, University of Tartu, hydrobiology
2000 Hugo Treffner Gymnasium

Professional career:

2010–... University of Tartu, Faculty of Science and Technology, Estonian Marine Institute, University of Tartu, Researcher
2004–2010 University of Tartu, Faculty of Science and Technology, Estonian Marine Institute, University of Tartu, laboratory technician
1999–2000 Mäksa Municipality Government, editor of county's newspaper

Language skills: estonian, english

Scientific and research activity

Research topics:

Mapping of marine benthic habitats in coastal waters of the Baltic Sea.
Distribution and variability of eelgrass communities in NE Baltic Sea in relation to interactive effect of small and large scale environmental variability.

Scientific publications:

Herkül, K.; Torn, K.; Möller, T. Environmental niche separation between charophytes and angiosperms in the northern Baltic Sea. Submitted to Botany Letters.
Möller, T.; Kotta, J. (2017). *Rangia cuneata* (G.B.Sowerby I, 1831) continues its invasion in the Baltic Sea: the first record in Pärnu Bay, Estonia. Bioinvasions Records, 6, 167–172.
Kotta, J.; Möller, T.; Orav-Kotta, H.; Pärnoja, M. (2014). Realized niche width of a brackish water submerged aquatic vegetation under current environmental conditions and projected influences of climate change. Marine Environmental Research, 102, 88–101.
Boström, C.; Baden, S.; Bockelmann, A.; Dromph, K.; Frederiksen, S.; Gustafsson, C.; Krause-Jensen, D.; Möller, T.; Nielsen, S.L.; Olesen, B.;

- Olsen, J.; Pihl, L.; Rinde, E. (2014).** Distribution, structure and function of Nordic seagrass ecosystems: implications for coastal management and conservation. *Aquatic Conservation-Marine and Freshwater Ecosystems*, 24 (3), 410–434.
- Möller, T.; Kotta, J.; Martin, G. (2014).** Spatiotemporal variability in eelgrass *Zostera marina* L. in the northeastern Baltic Sea: Canopy structure and associated macrophyte and invertebrate communities. *Estonian Journal of Ecology*, 63 (2), 90–108.
- Martin, G.; Kotta, J.; Möller, T.; Herkül, K. (2013).** Spatial distribution of marine benthic habitats in the Estonian coastal sea, northeastern Baltic Sea. *Estonian Journal of Ecology*, 62, 165–191.
- Vahtmäe, E.; Kutser, T.; Kotta, J.; Pärnoja, M.; Möller, T.; Lennuk, L. (2012).** Mapping Baltic Sea shallow water environments with airborne remote sensing. *Oceanology*, 52 (6), 803–809.
- Jüssi, I.; Kalamees, A.; Kuris, M.; Kuus, A.; Martin, G.; Möller, T.; Vetemaa, M. (2011).** Väärtuslikud avameremadalikud Eesti vetes (Valuable offshore shoals in Estonian waters). Tallinn: Balti Keskkonnafoorum.
- Möller, T.; Kotta, J.; Martin, G. (2010).** Mapping of structural and functional characteristics of reef habitats. *IEEE Conference Proceedings: The First International Conference on Biodiversity and Invasion Control*; Cancun, Mexico; March 7–13, 2010. Ed. Dini, Petre; Popescu, Manuela. IEEE, 150–154.
- Möller, T.; Kotta, J.; Martin, G. (2009).** Effect of observation method on the perception of community structure and water quality in a brackish water ecosystem. *Marine Ecology*, 30, 105–112.
- Kotta, J.; Möller, T. (2009).** Important scales of distribution patterns of the key benthic species in Gretagrund area, the central Gulf of Riga. *Estonian Journal of Ecology*, 58, 259–269.
- Kotta, J.; Paalme, T.; Kersen, P.; Martin, G.; Herkül, K.; Möller, T. (2008).** Density dependent growth of the red algae *Furcellaria lumbricalis* and *Coccotylus truncatus* in the West-Estonian Archipelago Sea, northern Baltic Sea. *Oceanologia*, 50 (4), 577–585.
- Möller, T.; Martin, G. (2007).** The distribution of the eelgrass *Zostera marina* in the coastal waters of Estonia, NE Baltic Sea. *Proceedings of the Estonian Academy of Sciences. Biology. Ecology*, 56 (4), 270–277.

ELULOOKIRJELDUS

Nimi: Tiia Möller
Sünniaeg: 11.06.1982
Kodakondsus: Eesti
Aadress: Haapsalu mnt 72, Pärnu
Telefon: +372 5027672
E-mail: tiia.moller@ut.ee

Hariduskäik

2007 MSc, Tartu Ülikool, hüdrobioloogia eriala
2004 BSc, Tartu Ülikool, hüdrobioloogia eriala
2000 Hugo Treffneri Gümnaasium

Teenistuskäik

2010–... Tartu Ülikooli Eesti Mereinstituut, merebioloogia teadur
2003–2010 Tartu Ülikooli Eesti Mereinstituut, laborant
1999–2000 Mäksa Vallavalitsus, valla infolehe toimetaja

Keelteoskus: eesti, inglise

Teadustöö ja arendustegevus

Teadustöö põhisuunad:

Merepõhjakoosluste kaardistamine Läänemeres.
Meriheina koosluste levik ja varieeruvus suure- ning väikeskaalalise keskkonnategurite koosmõjus.

Publikatsioonid:

Herkül, K.; Torn, K.; Möller, T. Environmental niche separation between charophytes and angiosperms in the northern Baltic Sea. Esitatud ajakirja Botany Letters.

Möller, T.; Kotta, J. (2017). *Rangia cuneata* (G.B.Sowerby I, 1831) continues its invasion in the Baltic Sea: the first record in Pärnu Bay, Estonia. Bioinvasions Records 6, 167–172.

Kotta, J.; Möller, T.; Orav-Kotta, H.; Pärnoja, M. (2014). Realized niche width of a brackish water submerged aquatic vegetation under current environmental conditions and projected influences of climate change. Marine Environmental Research, 102, 88–101.

Boström, C.; Baden, S.; Bockelmann, A.; Dromph, K.; Frederiksen, S.; Gustafsson, C.; Krause-Jensen, D.; Möller, T.; Nielsen, S.L.; Olesen, B.; Olsen, J.; Pihl, L.; Rinde, E. (2014). Distribution, structure and function of

- Nordic seagrass ecosystems: implications for coastal management and conservation. *Aquatic Conservation-Marine and Freshwater Ecosystems*, 24 (3), 410–434.
- Möller, T.; Kotta, J.; Martin, G. (2014).** Spatiotemporal variability in eelgrass *Zostera marina* L. in the northeastern Baltic Sea: Canopy structure and associated macrophyte and invertebrate communities. *Estonian Journal of Ecology*, 63 (2), 90–108.
- Martin, G.; Kotta, J.; Möller, T.; Herkül, K. (2013).** Spatial distribution of marine benthic habitats in the Estonian coastal sea, northeastern Baltic Sea. *Estonian Journal of Ecology*, 62, 165–191.
- Vahtmäe, E.; Kutser, T.; Kotta, J.; Pärnoja, M.; Möller, T.; Lennuk, L. (2012).** Mapping Baltic Sea shallow water environments with airborne remote sensing. *Oceanology*, 52 (6), 803–809.
- Jüssi, I.; Kalamees, A.; Kuris, M.; Kuus, A.; Martin, G.; Möller, T.; Vetemaa, M. (2011).** Väärtuslikud avameremadalikud Eesti vetes (Valuable offshore shoals in Estonian waters). Tallinn: Balti Keskkonnafoorum.
- Möller, T.; Kotta, J.; Martin, G. (2010).** Mapping of structural and functional characteristics of reef habitats. *IEEE Conference Proceedings: The First International Conference on Biodiversity and Invasion Control*; Cancun, Mexico; March 7–13, 2010. Ed. Dini, Petre; Popescu, Manuela. IEEE, 150–154.
- Möller, T.; Kotta, J.; Martin, G. (2009).** Effect of observation method on the perception of community structure and water quality in a brackish water ecosystem. *Marine Ecology*, 30, 105–112.
- Kotta, J.; Möller, T. (2009).** Important scales of distribution patterns of the key benthic species in Gretagrund area, the central Gulf of Riga. *Estonian Journal of Ecology*, 58, 259–269.
- Kotta, J.; Paalme, T.; Kersen, P.; Martin, G.; Herkül, K.; Möller, T. (2008).** Density dependent growth of the red algae *Furcellaria lumbricalis* and *Coccolytus truncatus* in the West-Estonian Archipelago Sea, northern Baltic Sea. *Oceanologia*, 50 (4), 577–585.
- Möller, T.; Martin, G. (2007).** The distribution of the eelgrass *Zostera marina* in the coastal waters of Estonia, NE Baltic Sea. *Proceedings of the Estonian Academy of Sciences. Biology. Ecology*, 56 (4), 270–277.

DISSERTATIONES BIOLOGICAE UNIVERSITATIS TARTUENSIS

1. **Toivo Maimets.** Studies of human oncoprotein p53. Tartu, 1991, 96 p.
2. **Enn K. Seppet.** Thyroid state control over energy metabolism, ion transport and contractile functions in rat heart. Tartu, 1991, 135 p.
3. **Kristjan Zobel.** Epifüütsete makrosamblike väärtus õhu saastuse indikaatoritena Hamar-Dobani boreaalsetes mägimetsades. Tartu, 1992, 131 lk.
4. **Andres Mäe.** Conjugal mobilization of catabolic plasmids by transposable elements in helper plasmids. Tartu, 1992, 91 p.
5. **Maia Kivisaar.** Studies on phenol degradation genes of *Pseudomonas* sp. strain EST 1001. Tartu, 1992, 61 p.
6. **Allan Nurk.** Nucleotide sequences of phenol degradative genes from *Pseudomonas* sp. strain EST 1001 and their transcriptional activation in *Pseudomonas putida*. Tartu, 1992, 72 p.
7. **Ülo Tamm.** The genus *Populus* L. in Estonia: variation of the species biology and introduction. Tartu, 1993, 91 p.
8. **Jaanus Remme.** Studies on the peptidyltransferase centre of the *E.coli* ribosome. Tartu, 1993, 68 p.
9. **Ülo Langel.** Galanin and galanin antagonists. Tartu, 1993, 97 p.
10. **Arvo Käär.** The development of an automatic online dynamic fluorescence-based pH-dependent fiber optic penicillin flowthrough biosensor for the control of the benzylpenicillin hydrolysis. Tartu, 1993, 117 p.
11. **Lilian Järvekülg.** Antigenic analysis and development of sensitive immunoassay for potato viruses. Tartu, 1993, 147 p.
12. **Jaak Palumets.** Analysis of phytomass partition in Norway spruce. Tartu, 1993, 47 p.
13. **Arne Sellin.** Variation in hydraulic architecture of *Picea abies* (L.) Karst. trees grown under different environmental conditions. Tartu, 1994, 119 p.
13. **Mati Reeben.** Regulation of light neurofilament gene expression. Tartu, 1994, 108 p.
14. **Urmas Tartes.** Respiration rhythms in insects. Tartu, 1995, 109 p.
15. **Ülo Puurand.** The complete nucleotide sequence and infections *in vitro* transcripts from cloned cDNA of a potato A potyvirus. Tartu, 1995, 96 p.
16. **Peeter Hõrak.** Pathways of selection in avian reproduction: a functional framework and its application in the population study of the great tit (*Parus major*). Tartu, 1995, 118 p.
17. **Erkki Truve.** Studies on specific and broad spectrum virus resistance in transgenic plants. Tartu, 1996, 158 p.
18. **Illar Pata.** Cloning and characterization of human and mouse ribosomal protein S6-encoding genes. Tartu, 1996, 60 p.
19. **Ülo Niinemets.** Importance of structural features of leaves and canopy in determining species shade-tolerance in temperature deciduous woody taxa. Tartu, 1996, 150 p.

20. **Ants Kurg.** Bovine leukemia virus: molecular studies on the packaging region and DNA diagnostics in cattle. Tartu, 1996, 104 p.
21. **Ene Ustav.** E2 as the modulator of the BPV1 DNA replication. Tartu, 1996, 100 p.
22. **Aksel Soosaar.** Role of helix-loop-helix and nuclear hormone receptor transcription factors in neurogenesis. Tartu, 1996, 109 p.
23. **Maido Remm.** Human papillomavirus type 18: replication, transformation and gene expression. Tartu, 1997, 117 p.
24. **Tiiu Kull.** Population dynamics in *Cypripedium calceolus* L. Tartu, 1997, 124 p.
25. **Kalle Olli.** Evolutionary life-strategies of autotrophic planktonic microorganisms in the Baltic Sea. Tartu, 1997, 180 p.
26. **Meelis Pärtel.** Species diversity and community dynamics in calcareous grassland communities in Western Estonia. Tartu, 1997, 124 p.
27. **Malle Leht.** The Genus *Potentilla* L. in Estonia, Latvia and Lithuania: distribution, morphology and taxonomy. Tartu, 1997, 186 p.
28. **Tanel Tenson.** Ribosomes, peptides and antibiotic resistance. Tartu, 1997, 80 p.
29. **Arvo Tuvikene.** Assessment of inland water pollution using biomarker responses in fish *in vivo* and *in vitro*. Tartu, 1997, 160 p.
30. **Urmas Saarma.** Tuning ribosomal elongation cycle by mutagenesis of 23S rRNA. Tartu, 1997, 134 p.
31. **Henn Ojaveer.** Composition and dynamics of fish stocks in the gulf of Riga ecosystem. Tartu, 1997, 138 p.
32. **Lembi Lõugas.** Post-glacial development of vertebrate fauna in Estonian water bodies. Tartu, 1997, 138 p.
33. **Margus Pooga.** Cell penetrating peptide, transportan, and its predecessors, galanin-based chimeric peptides. Tartu, 1998, 110 p.
34. **Andres Saag.** Evolutionary relationships in some cetrarioid genera (Lichenized Ascomycota). Tartu, 1998, 196 p.
35. **Aivar Liiv.** Ribosomal large subunit assembly *in vivo*. Tartu, 1998, 158 p.
36. **Tatjana Oja.** Isoenzyme diversity and phylogenetic affinities among the eurasian annual bromes (*Bromus* L., Poaceae). Tartu, 1998, 92 p.
37. **Mari Moora.** The influence of arbuscular mycorrhizal (AM) symbiosis on the competition and coexistence of calcareous grassland plant species. Tartu, 1998, 78 p.
38. **Olavi Kurina.** Fungus gnats in Estonia (*Diptera: Bolitophilidae, Keroplattidae, Macroceridae, Ditomyiidae, Diadocidiidae, Mycetophilidae*). Tartu, 1998, 200 p.
39. **Andrus Tasa.** Biological leaching of shales: black shale and oil shale. Tartu, 1998, 98 p.
40. **Arnold Kristjuhan.** Studies on transcriptional activator properties of tumor suppressor protein p53. Tartu, 1998, 86 p.
41. **Sulev Ingerpuu.** Characterization of some human myeloid cell surface and nuclear differentiation antigens. Tartu, 1998, 163 p.

42. **Veljo Kisand**. Responses of planktonic bacteria to the abiotic and biotic factors in the shallow lake Võrtsjärv. Tartu, 1998, 118 p.
43. **Kadri Põldmaa**. Studies in the systematics of hypomyces and allied genera (Hypocreales, Ascomycota). Tartu, 1998, 178 p.
44. **Markus Vetemaa**. Reproduction parameters of fish as indicators in environmental monitoring. Tartu, 1998, 117 p.
45. **Heli Talvik**. Prepatent periods and species composition of different *Oesophagostomum* spp. populations in Estonia and Denmark. Tartu, 1998, 104 p.
46. **Katrin Heinsoo**. Cuticular and stomatal antechamber conductance to water vapour diffusion in *Picea abies* (L.) karst. Tartu, 1999, 133 p.
47. **Tarmo Annilo**. Studies on mammalian ribosomal protein S7. Tartu, 1998, 77 p.
48. **Indrek Ots**. Health state indices of reproducing great tits (*Parus major*): sources of variation and connections with life-history traits. Tartu, 1999, 117 p.
49. **Juan Jose Cantero**. Plant community diversity and habitat relationships in central Argentina grasslands. Tartu, 1999, 161 p.
50. **Rein Kalamees**. Seed bank, seed rain and community regeneration in Estonian calcareous grasslands. Tartu, 1999, 107 p.
51. **Sulev Kõks**. Cholecystokinin (CCK) – induced anxiety in rats: influence of environmental stimuli and involvement of endopioid mechanisms and serotonin. Tartu, 1999, 123 p.
52. **Ebe Sild**. Impact of increasing concentrations of O₃ and CO₂ on wheat, clover and pasture. Tartu, 1999, 123 p.
53. **Ljudmilla Timofejeva**. Electron microscopical analysis of the synaptosomal complex formation in cereals. Tartu, 1999, 99 p.
54. **Andres Valkna**. Interactions of galanin receptor with ligands and G-proteins: studies with synthetic peptides. Tartu, 1999, 103 p.
55. **Taavi Virro**. Life cycles of planktonic rotifers in lake Peipsi. Tartu, 1999, 101 p.
56. **Ana Rebane**. Mammalian ribosomal protein S3a genes and intron-encoded small nucleolar RNAs U73 and U82. Tartu, 1999, 85 p.
57. **Tiina Tamm**. Cocksfoot mottle virus: the genome organisation and translational strategies. Tartu, 2000, 101 p.
58. **Reet Kurg**. Structure-function relationship of the bovine papilloma virus E2 protein. Tartu, 2000, 89 p.
59. **Toomas Kivisild**. The origins of Southern and Western Eurasian populations: an mtDNA study. Tartu, 2000, 121 p.
60. **Niilo Kaldalu**. Studies of the TOL plasmid transcription factor XylS. Tartu, 2000, 88 p.
61. **Dina Lepik**. Modulation of viral DNA replication by tumor suppressor protein p53. Tartu, 2000, 106 p.

62. **Kai Vellak**. Influence of different factors on the diversity of the bryophyte vegetation in forest and wooded meadow communities. Tartu, 2000, 122 p.
63. **Jonne Kotta**. Impact of eutrophication and biological invasions on the structure and functions of benthic macrofauna. Tartu, 2000, 160 p.
64. **Georg Martin**. Phytobenthic communities of the Gulf of Riga and the inner sea the West-Estonian archipelago. Tartu, 2000, 139 p.
65. **Silvia Sepp**. Morphological and genetical variation of *Alchemilla L.* in Estonia. Tartu, 2000. 124 p.
66. **Jaani Liira**. On the determinants of structure and diversity in herbaceous plant communities. Tartu, 2000, 96 p.
67. **Priit Zingel**. The role of planktonic ciliates in lake ecosystems. Tartu, 2001, 111 p.
68. **Tiit Teder**. Direct and indirect effects in Host-parasitoid interactions: ecological and evolutionary consequences. Tartu, 2001, 122 p.
69. **Hannes Kollist**. Leaf apoplastic ascorbate as ozone scavenger and its transport across the plasma membrane. Tartu, 2001, 80 p.
70. **Reet Marits**. Role of two-component regulator system PehR-PehS and extracellular protease PrtW in virulence of *Erwinia Carotovora* subsp. *Carotovora*. Tartu, 2001, 112 p.
71. **Vallo Tilgar**. Effect of calcium supplementation on reproductive performance of the pied flycatcher *Ficedula hypoleuca* and the great tit *Parus major*, breeding in Northern temperate forests. Tartu, 2002, 126 p.
72. **Rita Hõrak**. Regulation of transposition of transposon Tn4652 in *Pseudomonas putida*. Tartu, 2002, 108 p.
73. **Liina Eek-Piirsoo**. The effect of fertilization, mowing and additional illumination on the structure of a species-rich grassland community. Tartu, 2002, 74 p.
74. **Krõõt Aasamaa**. Shoot hydraulic conductance and stomatal conductance of six temperate deciduous tree species. Tartu, 2002, 110 p.
75. **Nele Ingerpuu**. Bryophyte diversity and vascular plants. Tartu, 2002, 112 p.
76. **Neeme Tõnisson**. Mutation detection by primer extension on oligonucleotide microarrays. Tartu, 2002, 124 p.
77. **Margus Pensa**. Variation in needle retention of Scots pine in relation to leaf morphology, nitrogen conservation and tree age. Tartu, 2003, 110 p.
78. **Asko Lõhmus**. Habitat preferences and quality for birds of prey: from principles to applications. Tartu, 2003, 168 p.
79. **Viljar Jaks**. p53 – a switch in cellular circuit. Tartu, 2003, 160 p.
80. **Jaana Männik**. Characterization and genetic studies of four ATP-binding cassette (ABC) transporters. Tartu, 2003, 140 p.
81. **Marek Sammul**. Competition and coexistence of clonal plants in relation to productivity. Tartu, 2003, 159 p.
82. **Ivar Ilves**. Virus-cell interactions in the replication cycle of bovine papillomavirus type 1. Tartu, 2003, 89 p.

83. **Andres Männik**. Design and characterization of a novel vector system based on the stable replicator of bovine papillomavirus type 1. Tartu, 2003, 109 p.
84. **Ivika Ostonen**. Fine root structure, dynamics and proportion in net primary production of Norway spruce forest ecosystem in relation to site conditions. Tartu, 2003, 158 p.
85. **Gudrun Veldre**. Somatic status of 12–15-year-old Tartu schoolchildren. Tartu, 2003, 199 p.
86. **Ülo Väli**. The greater spotted eagle *Aquila clanga* and the lesser spotted eagle *A. pomarina*: taxonomy, phylogeography and ecology. Tartu, 2004, 159 p.
87. **Aare Abroi**. The determinants for the native activities of the bovine papillomavirus type 1 E2 protein are separable. Tartu, 2004, 135 p.
88. **Tiina Kahre**. Cystic fibrosis in Estonia. Tartu, 2004, 116 p.
89. **Helen Orav-Kotta**. Habitat choice and feeding activity of benthic suspension feeders and mesograzers in the northern Baltic Sea. Tartu, 2004, 117 p.
90. **Maarja Öpik**. Diversity of arbuscular mycorrhizal fungi in the roots of perennial plants and their effect on plant performance. Tartu, 2004, 175 p.
91. **Kadri Tali**. Species structure of *Neotinea ustulata*. Tartu, 2004, 109 p.
92. **Kristiina Tambets**. Towards the understanding of post-glacial spread of human mitochondrial DNA haplogroups in Europe and beyond: a phylogeographic approach. Tartu, 2004, 163 p.
93. **Arvi Jõers**. Regulation of p53-dependent transcription. Tartu, 2004, 103 p.
94. **Lilian Kadaja**. Studies on modulation of the activity of tumor suppressor protein p53. Tartu, 2004, 103 p.
95. **Jaak Truu**. Oil shale industry wastewater: impact on river microbial community and possibilities for bioremediation. Tartu, 2004, 128 p.
96. **Maire Peters**. Natural horizontal transfer of the *pheBA* operon. Tartu, 2004, 105 p.
97. **Ülo Maiväli**. Studies on the structure-function relationship of the bacterial ribosome. Tartu, 2004, 130 p.
98. **Merit Otsus**. Plant community regeneration and species diversity in dry calcareous grasslands. Tartu, 2004, 103 p.
99. **Mikk Heidemaa**. Systematic studies on sawflies of the genera *Dolerus*, *Empria*, and *Caliroa* (Hymenoptera: Tenthredinidae). Tartu, 2004, 167 p.
100. **Ilmar Tõnno**. The impact of nitrogen and phosphorus concentration and N/P ratio on cyanobacterial dominance and N₂ fixation in some Estonian lakes. Tartu, 2004, 111 p.
101. **Lauri Saks**. Immune function, parasites, and carotenoid-based ornaments in greenfinches. Tartu, 2004, 144 p.
102. **Siiri Rootsi**. Human Y-chromosomal variation in European populations. Tartu, 2004, 142 p.
103. **Eve Vedler**. Structure of the 2,4-dichloro-phenoxyacetic acid-degradative plasmid pEST4011. Tartu, 2005. 106 p.

104. **Andres Tover.** Regulation of transcription of the phenol degradation *pheBA* operon in *Pseudomonas putida*. Tartu, 2005, 126 p.
105. **Helen Udras.** Hexose kinases and glucose transport in the yeast *Hansenula polymorpha*. Tartu, 2005, 100 p.
106. **Ave Suija.** Lichens and lichenicolous fungi in Estonia: diversity, distribution patterns, taxonomy. Tartu, 2005, 162 p.
107. **Piret Lõhmus.** Forest lichens and their substrata in Estonia. Tartu, 2005, 162 p.
108. **Inga Lips.** Abiotic factors controlling the cyanobacterial bloom occurrence in the Gulf of Finland. Tartu, 2005, 156 p.
109. **Kaasik, Krista.** Circadian clock genes in mammalian clockwork, metabolism and behaviour. Tartu, 2005, 121 p.
110. **Juhan Javoš.** The effects of experience on host acceptance in ovipositing moths. Tartu, 2005, 112 p.
111. **Tiina Sedman.** Characterization of the yeast *Saccharomyces cerevisiae* mitochondrial DNA helicase Hmi1. Tartu, 2005, 103 p.
112. **Ruth Agurauja.** Hawaiian endemic fern lineage *Diellia* (Aspleniaceae): distribution, population structure and ecology. Tartu, 2005, 112 p.
113. **Riho Teras.** Regulation of transcription from the fusion promoters generated by transposition of Tn4652 into the upstream region of *pheBA* operon in *Pseudomonas putida*. Tartu, 2005, 106 p.
114. **Mait Metspalu.** Through the course of prehistory in india: tracing the mtDNA trail. Tartu, 2005, 138 p.
115. **Elin Lõhmussaar.** The comparative patterns of linkage disequilibrium in European populations and its implication for genetic association studies. Tartu, 2006, 124 p.
116. **Priit Kopper.** Hydraulic and environmental limitations to leaf water relations in trees with respect to canopy position. Tartu, 2006, 126 p.
117. **Heili Iives.** Stress-induced transposition of Tn4652 in *Pseudomonas Putida*. Tartu, 2006, 120 p.
118. **Silja Kuusk.** Biochemical properties of Hmi1p, a DNA helicase from *Saccharomyces cerevisiae* mitochondria. Tartu, 2006, 126 p.
119. **Kersti Püssa.** Forest edges on medium resolution landsat thematic mapper satellite images. Tartu, 2006, 90 p.
120. **Lea Tummeleht.** Physiological condition and immune function in great tits (*Parus major* L.): Sources of variation and trade-offs in relation to growth. Tartu, 2006, 94 p.
121. **Toomas Esperk.** Larval instar as a key element of insect growth schedules. Tartu, 2006, 186 p.
122. **Harri Valdmann.** Lynx (*Lynx lynx*) and wolf (*Canis lupus*) in the Baltic region: Diets, helminth parasites and genetic variation. Tartu, 2006. 102 p.
123. **Priit Jõers.** Studies of the mitochondrial helicase Hmi1p in *Candida albicans* and *Saccharomyces cerevisia*. Tartu, 2006. 113 p.
124. **Kersti Lilleväli.** Gata3 and Gata2 in inner ear development. Tartu, 2007, 123 p.

125. **Kai Rünk.** Comparative ecology of three fern species: *Dryopteris carthusiana* (Vill.) H.P. Fuchs, *D. expansa* (C. Presl) Fraser-Jenkins & Jermy and *D. dilatata* (Hoffm.) A. Gray (Dryopteridaceae). Tartu, 2007, 143 p.
126. **Aveliina Helm.** Formation and persistence of dry grassland diversity: role of human history and landscape structure. Tartu, 2007, 89 p.
127. **Leho Tedersoo.** Ectomycorrhizal fungi: diversity and community structure in Estonia, Seychelles and Australia. Tartu, 2007, 233 p.
128. **Marko Mägi.** The habitat-related variation of reproductive performance of great tits in a deciduous-coniferous forest mosaic: looking for causes and consequences. Tartu, 2007, 135 p.
129. **Valeria Lulla.** Replication strategies and applications of Semliki Forest virus. Tartu, 2007, 109 p.
130. **Ülle Reier.** Estonian threatened vascular plant species: causes of rarity and conservation. Tartu, 2007, 79 p.
131. **Inga Jüriado.** Diversity of lichen species in Estonia: influence of regional and local factors. Tartu, 2007, 171 p.
132. **Tatjana Krama.** Mobbing behaviour in birds: costs and reciprocity based cooperation. Tartu, 2007, 112 p.
133. **Signe Saumaa.** The role of DNA mismatch repair and oxidative DNA damage defense systems in avoidance of stationary phase mutations in *Pseudomonas putida*. Tartu, 2007, 172 p.
134. **Reedik Mägi.** The linkage disequilibrium and the selection of genetic markers for association studies in european populations. Tartu, 2007, 96 p.
135. **Priit Kilgas.** Blood parameters as indicators of physiological condition and skeletal development in great tits (*Parus major*): natural variation and application in the reproductive ecology of birds. Tartu, 2007, 129 p.
136. **Anu Albert.** The role of water salinity in structuring eastern Baltic coastal fish communities. Tartu, 2007, 95 p.
137. **Kärt Padari.** Protein transduction mechanisms of transportans. Tartu, 2008, 128 p.
138. **Siiri-Liis Sandre.** Selective forces on larval colouration in a moth. Tartu, 2008, 125 p.
139. **Ülle Jõgar.** Conservation and restoration of semi-natural floodplain meadows and their rare plant species. Tartu, 2008, 99 p.
140. **Lauri Laanisto.** Macroecological approach in vegetation science: generality of ecological relationships at the global scale. Tartu, 2008, 133 p.
141. **Reidar Andreson.** Methods and software for predicting PCR failure rate in large genomes. Tartu, 2008, 105 p.
142. **Birgot Paavel.** Bio-optical properties of turbid lakes. Tartu, 2008, 175 p.
143. **Kaire Torn.** Distribution and ecology of charophytes in the Baltic Sea. Tartu, 2008, 98 p.
144. **Vladimir Vimberg.** Peptide mediated macrolide resistance. Tartu, 2008, 190 p.
145. **Daima Örd.** Studies on the stress-inducible pseudokinase TRB3, a novel inhibitor of transcription factor ATF4. Tartu, 2008, 108 p.

146. **Lauri Saag.** Taxonomic and ecologic problems in the genus *Lepraria* (*Stereocaulaceae*, lichenised *Ascomycota*). Tartu, 2008, 175 p.
147. **Ulvi Karu.** Antioxidant protection, carotenoids and coccidians in greenfinches – assessment of the costs of immune activation and mechanisms of parasite resistance in a passerine with carotenoid-based ornaments. Tartu, 2008, 124 p.
148. **Jaanus Remm.** Tree-cavities in forests: density, characteristics and occupancy by animals. Tartu, 2008, 128 p.
149. **Epp Moks.** Tapeworm parasites *Echinococcus multilocularis* and *E. granulosus* in Estonia: phylogenetic relationships and occurrence in wild carnivores and ungulates. Tartu, 2008, 82 p.
150. **Eve Eensalu.** Acclimation of stomatal structure and function in tree canopy: effect of light and CO₂ concentration. Tartu, 2008, 108 p.
151. **Janne Pullat.** Design, functionlization and application of an *in situ* synthesized oligonucleotide microarray. Tartu, 2008, 108 p.
152. **Marta Putrinš.** Responses of *Pseudomonas putida* to phenol-induced metabolic and stress signals. Tartu, 2008, 142 p.
153. **Marina Semtšenko.** Plant root behaviour: responses to neighbours and physical obstructions. Tartu, 2008, 106 p.
154. **Marge Starast.** Influence of cultivation techniques on productivity and fruit quality of some *Vaccinium* and *Rubus* taxa. Tartu, 2008, 154 p.
155. **Age Tats.** Sequence motifs influencing the efficiency of translation. Tartu, 2009, 104 p.
156. **Radi Tegova.** The role of specialized DNA polymerases in mutagenesis in *Pseudomonas putida*. Tartu, 2009, 124 p.
157. **Tsipe Aavik.** Plant species richness, composition and functional trait pattern in agricultural landscapes – the role of land use intensity and landscape structure. Tartu, 2009, 112 p.
158. **Kaja Kiiver.** Semliki forest virus based vectors and cell lines for studying the replication and interactions of alphaviruses and hepaciviruses. Tartu, 2009, 104 p.
159. **Meelis Kadaja.** Papillomavirus Replication Machinery Induces Genomic Instability in its Host Cell. Tartu, 2009, 126 p.
160. **Pille Hallast.** Human and chimpanzee Luteinizing hormone/Chorionic Gonadotropin beta (*LHB/CGB*) gene clusters: diversity and divergence of young duplicated genes. Tartu, 2009, 168 p.
161. **Ain Vellak.** Spatial and temporal aspects of plant species conservation. Tartu, 2009, 86 p.
162. **Triinu Remmel.** Body size evolution in insects with different colouration strategies: the role of predation risk. Tartu, 2009, 168 p.
163. **Jaana Salujõe.** Zooplankton as the indicator of ecological quality and fish predation in lake ecosystems. Tartu, 2009, 129 p.
164. **Ele Vahtmäe.** Mapping benthic habitat with remote sensing in optically complex coastal environments. Tartu, 2009, 109 p.

165. **Liisa Metsamaa**. Model-based assessment to improve the use of remote sensing in recognition and quantitative mapping of cyanobacteria. Tartu, 2009, 114 p.
166. **Pille Säälük**. The role of endocytosis in the protein transduction by cell-penetrating peptides. Tartu, 2009, 155 p.
167. **Lauri Peil**. Ribosome assembly factors in *Escherichia coli*. Tartu, 2009, 147 p.
168. **Lea Hallik**. Generality and specificity in light harvesting, carbon gain capacity and shade tolerance among plant functional groups. Tartu, 2009, 99 p.
169. **Mariliis Tark**. Mutagenic potential of DNA damage repair and tolerance mechanisms under starvation stress. Tartu, 2009, 191 p.
170. **Riinu Rannap**. Impacts of habitat loss and restoration on amphibian populations. Tartu, 2009, 117 p.
171. **Maarja Adojaan**. Molecular variation of HIV-1 and the use of this knowledge in vaccine development. Tartu, 2009, 95 p.
172. **Signe Altmäe**. Genomics and transcriptomics of human induced ovarian folliculogenesis. Tartu, 2010, 179 p.
173. **Triin Suvi**. Mycorrhizal fungi of native and introduced trees in the Seychelles Islands. Tartu, 2010, 107 p.
174. **Velda Lauringson**. Role of suspension feeding in a brackish-water coastal sea. Tartu, 2010, 123 p.
175. **Eero Talts**. Photosynthetic cyclic electron transport – measurement and variably proton-coupled mechanism. Tartu, 2010, 121 p.
176. **Mari Nelis**. Genetic structure of the Estonian population and genetic distance from other populations of European descent. Tartu, 2010, 97 p.
177. **Kaarel Krjutškov**. Arrayed Primer Extension-2 as a multiplex PCR-based method for nucleic acid variation analysis: method and applications. Tartu, 2010, 129 p.
178. **Egle Köster**. Morphological and genetical variation within species complexes: *Anthyllis vulneraria* s. l. and *Alchemilla vulgaris* (coll.). Tartu, 2010, 101 p.
179. **Erki Õunap**. Systematic studies on the subfamily Sterrhinae (Lepidoptera: Geometridae). Tartu, 2010, 111 p.
180. **Merike Jõesaar**. Diversity of key catabolic genes at degradation of phenol and *p*-cresol in pseudomonads. Tartu, 2010, 125 p.
181. **Kristjan Herkül**. Effects of physical disturbance and habitat-modifying species on sediment properties and benthic communities in the northern Baltic Sea. Tartu, 2010, 123 p.
182. **Arto Pulk**. Studies on bacterial ribosomes by chemical modification approaches. Tartu, 2010, 161 p.
183. **Maria Põllupüü**. Ecological relations of cladocerans in a brackish-water ecosystem. Tartu, 2010, 126 p.
184. **Toomas Silla**. Study of the segregation mechanism of the Bovine Papillomavirus Type 1. Tartu, 2010, 188 p.

185. **Gyaneshwer Chaubey**. The demographic history of India: A perspective based on genetic evidence. Tartu, 2010, 184 p.
186. **Katrin Kepp**. Genes involved in cardiovascular traits: detection of genetic variation in Estonian and Czech populations. Tartu, 2010, 164 p.
187. **Virve Sõber**. The role of biotic interactions in plant reproductive performance. Tartu, 2010, 92 p.
188. **Kersti Kangro**. The response of phytoplankton community to the changes in nutrient loading. Tartu, 2010, 144 p.
189. **Joachim M. Gerhold**. Replication and Recombination of mitochondrial DNA in Yeast. Tartu, 2010, 120 p.
190. **Helen Tammert**. Ecological role of physiological and phylogenetic diversity in aquatic bacterial communities. Tartu, 2010, 140 p.
191. **Elle Rajandu**. Factors determining plant and lichen species diversity and composition in Estonian *Calamagrostis* and *Hepatica* site type forests. Tartu, 2010, 123 p.
192. **Paula Ann Kivistik**. ColR-ColS signalling system and transposition of Tn4652 in the adaptation of *Pseudomonas putida*. Tartu, 2010, 118 p.
193. **Siim Sõber**. Blood pressure genetics: from candidate genes to genome-wide association studies. Tartu, 2011, 120 p.
194. **Kalle Kipper**. Studies on the role of helix 69 of 23S rRNA in the factor-dependent stages of translation initiation, elongation, and termination. Tartu, 2011, 178 p.
195. **Triinu Siibak**. Effect of antibiotics on ribosome assembly is indirect. Tartu, 2011, 134 p.
196. **Tambet Tõnissoo**. Identification and molecular analysis of the role of guanine nucleotide exchange factor RIC-8 in mouse development and neural function. Tartu, 2011, 110 p.
197. **Helin Räägel**. Multiple faces of cell-penetrating peptides – their intracellular trafficking, stability and endosomal escape during protein transduction. Tartu, 2011, 161 p.
198. **Andres Jaanus**. Phytoplankton in Estonian coastal waters – variability, trends and response to environmental pressures. Tartu, 2011, 157 p.
199. **Tiit Nikopensius**. Genetic predisposition to nonsyndromic orofacial clefts. Tartu, 2011, 152 p.
200. **Signe Värv**. Studies on the mechanisms of RNA polymerase II-dependent transcription elongation. Tartu, 2011, 108 p.
201. **Kristjan Välk**. Gene expression profiling and genome-wide association studies of non-small cell lung cancer. Tartu, 2011, 98 p.
202. **Arno Põllumäe**. Spatio-temporal patterns of native and invasive zooplankton species under changing climate and eutrophication conditions. Tartu, 2011, 153 p.
203. **Egle Tammeleht**. Brown bear (*Ursus arctos*) population structure, demographic processes and variations in diet in northern Eurasia. Tartu, 2011, 143 p.

205. **Teele Jairus**. Species composition and host preference among ectomycorrhizal fungi in Australian and African ecosystems. Tartu, 2011, 106 p.
206. **Kessy Abarenkov**. PlutoF – cloud database and computing services supporting biological research. Tartu, 2011, 125 p.
207. **Marina Grigorova**. Fine-scale genetic variation of follicle-stimulating hormone beta-subunit coding gene (*FSHB*) and its association with reproductive health. Tartu, 2011, 184 p.
208. **Anu Tiitsaar**. The effects of predation risk and habitat history on butterfly communities. Tartu, 2011, 97 p.
209. **Elin Sild**. Oxidative defences in immunoeological context: validation and application of assays for nitric oxide production and oxidative burst in a wild passerine. Tartu, 2011, 105 p.
210. **Irja Saar**. The taxonomy and phylogeny of the genera *Cystoderma* and *Cystodermella* (Agaricales, Fungi). Tartu, 2012, 167 p.
211. **Pauli Saag**. Natural variation in plumage bacterial assemblages in two wild breeding passerines. Tartu, 2012, 113 p.
212. **Aleksei Lulla**. Alphaviral nonstructural protease and its polyprotein substrate: arrangements for the perfect marriage. Tartu, 2012, 143 p.
213. **Mari Järve**. Different genetic perspectives on human history in Europe and the Caucasus: the stories told by uniparental and autosomal markers. Tartu, 2012, 119 p.
214. **Ott Scheler**. The application of tmRNA as a marker molecule in bacterial diagnostics using microarray and biosensor technology. Tartu, 2012, 93 p.
215. **Anna Balikova**. Studies on the functions of tumor-associated mucin-like leukosialin (CD43) in human cancer cells. Tartu, 2012, 129 p.
216. **Triinu Kõressaar**. Improvement of PCR primer design for detection of prokaryotic species. Tartu, 2012, 83 p.
217. **Tuul Sepp**. Hematological health state indices of greenfinches: sources of individual variation and responses to immune system manipulation. Tartu, 2012, 117 p.
218. **Rya Ero**. Modifier view of the bacterial ribosome. Tartu, 2012, 146 p.
219. **Mohammad Bahram**. Biogeography of ectomycorrhizal fungi across different spatial scales. Tartu, 2012, 165 p.
220. **Annely Lorents**. Overcoming the plasma membrane barrier: uptake of amphipathic cell-penetrating peptides induces influx of calcium ions and downstream responses. Tartu, 2012, 113 p.
221. **Katrin Männik**. Exploring the genomics of cognitive impairment: whole-genome SNP genotyping experience in Estonian patients and general population. Tartu, 2012, 171 p.
222. **Marko Prouš**. Taxonomy and phylogeny of the sawfly genus *Empria* (Hymenoptera, Tenthredinidae). Tartu, 2012, 192 p.
223. **Triinu Visnapuu**. Levansucrases encoded in the genome of *Pseudomonas syringae* pv. tomato DC3000: heterologous expression, biochemical characterization, mutational analysis and spectrum of polymerization products. Tartu, 2012, 160 p.

224. **Nele Tamberg**. Studies on Semliki Forest virus replication and pathogenesis. Tartu, 2012, 109 p.
225. **Tõnu Esko**. Novel applications of SNP array data in the analysis of the genetic structure of Europeans and in genetic association studies. Tartu, 2012, 149 p.
226. **Timo Arula**. Ecology of early life-history stages of herring *Clupea harengus membras* in the northeastern Baltic Sea. Tartu, 2012, 143 p.
227. **Inga Hiiesalu**. Belowground plant diversity and coexistence patterns in grassland ecosystems. Tartu, 2012, 130 p.
228. **Kadri Koorem**. The influence of abiotic and biotic factors on small-scale plant community patterns and regeneration in boreonemoral forest. Tartu, 2012, 114 p.
229. **Liis Andresen**. Regulation of virulence in plant-pathogenic pectobacteria. Tartu, 2012, 122 p.
230. **Kaupo Kohv**. The direct and indirect effects of management on boreal forest structure and field layer vegetation. Tartu, 2012, 124 p.
231. **Mart Jüssi**. Living on an edge: landlocked seals in changing climate. Tartu, 2012, 114 p.
232. **Riina Klais**. Phytoplankton trends in the Baltic Sea. Tartu, 2012, 136 p.
233. **Rauno Veeroja**. Effects of winter weather, population density and timing of reproduction on life-history traits and population dynamics of moose (*Alces alces*) in Estonia. Tartu, 2012, 92 p.
234. **Marju Keis**. Brown bear (*Ursus arctos*) phylogeography in northern Eurasia. Tartu, 2013, 142 p.
235. **Sergei Põlme**. Biogeography and ecology of *alnus*- associated ectomycorrhizal fungi – from regional to global scale. Tartu, 2013, 90 p.
236. **Liis Uusküla**. Placental gene expression in normal and complicated pregnancy. Tartu, 2013, 173 p.
237. **Marko Lõoke**. Studies on DNA replication initiation in *Saccharomyces cerevisiae*. Tartu, 2013, 112 p.
238. **Anne Aan**. Light- and nitrogen-use and biomass allocation along productivity gradients in multilayer plant communities. Tartu, 2013, 127 p.
239. **Heidi Tamm**. Comprehending phylogenetic diversity – case studies in three groups of ascomycetes. Tartu, 2013, 136 p.
240. **Liina Kangur**. High-Pressure Spectroscopy Study of Chromophore-Binding Hydrogen Bonds in Light-Harvesting Complexes of Photosynthetic Bacteria. Tartu, 2013, 150 p.
241. **Margus Leppik**. Substrate specificity of the multisite specific pseudouridine synthase RluD. Tartu, 2013, 111 p.
242. **Lauris Kaplinski**. The application of oligonucleotide hybridization model for PCR and microarray optimization. Tartu, 2013, 103 p.
243. **Merli Pärnoja**. Patterns of macrophyte distribution and productivity in coastal ecosystems: effect of abiotic and biotic forcing. Tartu, 2013, 155 p.
244. **Tõnu Margus**. Distribution and phylogeny of the bacterial translational GTPases and the Mqsr/YgiT regulatory system. Tartu, 2013, 126 p.

245. **Pille Mänd.** Light use capacity and carbon and nitrogen budget of plants: remote assessment and physiological determinants. Tartu, 2013, 128 p.
246. **Mario Plaas.** Animal model of Wolfram Syndrome in mice: behavioural, biochemical and psychopharmacological characterization. Tartu, 2013, 144 p.
247. **Georgi Hudjašov.** Maps of mitochondrial DNA, Y-chromosome and tyrosinase variation in Eurasian and Oceanian populations. Tartu, 2013, 115 p.
248. **Mari Lepik.** Plasticity to light in herbaceous plants and its importance for community structure and diversity. Tartu, 2013, 102 p.
249. **Ede Leppik.** Diversity of lichens in semi-natural habitats of Estonia. Tartu, 2013, 151 p.
250. **Ülle Saks.** Arbuscular mycorrhizal fungal diversity patterns in boreo-nemoral forest ecosystems. Tartu, 2013, 151 p.
251. **Eneli Oitmaa.** Development of arrayed primer extension microarray assays for molecular diagnostic applications. Tartu, 2013, 147 p.
252. **Jekaterina Jutkina.** The horizontal gene pool for aromatics degradation: bacterial catabolic plasmids of the Baltic Sea aquatic system. Tartu, 2013, 121 p.
253. **Helen Vellau.** Reaction norms for size and age at maturity in insects: rules and exceptions. Tartu, 2014, 132 p.
254. **Randel Kreitsberg.** Using biomarkers in assessment of environmental contamination in fish – new perspectives. Tartu, 2014, 107 p.
255. **Krista Takkis.** Changes in plant species richness and population performance in response to habitat loss and fragmentation. Tartu, 2014, 141 p.
256. **Liina Nagirnaja.** Global and fine-scale genetic determinants of recurrent pregnancy loss. Tartu, 2014, 211 p.
257. **Triin Triisberg.** Factors influencing the re-vegetation of abandoned extracted peatlands in Estonia. Tartu, 2014, 133 p.
258. **Villu Soon.** A phylogenetic revision of the *Chrysis ignita* species group (Hymenoptera: Chrysididae) with emphasis on the northern European fauna. Tartu, 2014, 211 p.
259. **Andrei Nikonov.** RNA-Dependent RNA Polymerase Activity as a Basis for the Detection of Positive-Strand RNA Viruses by Vertebrate Host Cells. Tartu, 2014, 207 p.
260. **Eele Õunapuu-Pikas.** Spatio-temporal variability of leaf hydraulic conductance in woody plants: ecophysiological consequences. Tartu, 2014, 135 p.
261. **Marju Männiste.** Physiological ecology of greenfinches: information content of feathers in relation to immune function and behavior. Tartu, 2014, 121 p.
262. **Katre Kets.** Effects of elevated concentrations of CO₂ and O₃ on leaf photosynthetic parameters in *Populus tremuloides*: diurnal, seasonal and interannual patterns. Tartu, 2014, 115 p.

263. **Külli Lokko**. Seasonal and spatial variability of zoopsammon communities in relation to environmental parameters. Tartu, 2014, 129 p.
264. **Olga Žilina**. Chromosomal microarray analysis as diagnostic tool: Estonian experience. Tartu, 2014, 152 p.
265. **Kertu Lõhmus**. Colonisation ecology of forest-dwelling vascular plants and the conservation value of rural manor parks. Tartu, 2014, 111 p.
266. **Anu Aun**. Mitochondria as integral modulators of cellular signaling. Tartu, 2014, 167 p.
267. **Chandana Basu Mallick**. Genetics of adaptive traits and gender-specific demographic processes in South Asian populations. Tartu, 2014, 160 p.
268. **Riin Tamme**. The relationship between small-scale environmental heterogeneity and plant species diversity. Tartu, 2014, 130 p.
269. **Liina Remm**. Impacts of forest drainage on biodiversity and habitat quality: implications for sustainable management and conservation. Tartu, 2015, 126 p.
270. **Tiina Talve**. Genetic diversity and taxonomy within the genus *Rhinanthus*. Tartu, 2015, 106 p.
271. **Mehis Rohtla**. Otolith sclerochronological studies on migrations, spawning habitat preferences and age of freshwater fishes inhabiting the Baltic Sea. Tartu, 2015, 137 p.
272. **Alexey Reshchikov**. The world fauna of the genus *Lathrolestes* (Hymenoptera, Ichneumonidae). Tartu, 2015, 247 p.
273. **Martin Pook**. Studies on artificial and extracellular matrix protein-rich surfaces as regulators of cell growth and differentiation. Tartu, 2015, 142 p.
274. **Mai Kukumägi**. Factors affecting soil respiration and its components in silver birch and Norway spruce stands. Tartu, 2015, 155 p.
275. **Helen Karu**. Development of ecosystems under human activity in the North-East Estonian industrial region: forests on post-mining sites and bogs. Tartu, 2015, 152 p.
276. **Hedi Peterson**. Exploiting high-throughput data for establishing relationships between genes. Tartu, 2015, 186 p.
277. **Priit Adler**. Analysis and visualisation of large scale microarray data, Tartu, 2015, 126 p.
278. **Aigar Niglas**. Effects of environmental factors on gas exchange in deciduous trees: focus on photosynthetic water-use efficiency. Tartu, 2015, 152 p.
279. **Silja Laht**. Classification and identification of conopeptides using profile hidden Markov models and position-specific scoring matrices. Tartu, 2015, 100 p.
280. **Martin Kesler**. Biological characteristics and restoration of Atlantic salmon *Salmo salar* populations in the Rivers of Northern Estonia. Tartu, 2015, 97 p.
281. **Pratyush Kumar Das**. Biochemical perspective on alphaviral nonstructural protein 2: a tale from multiple domains to enzymatic profiling. Tartu, 2015, 205 p.

282. **Priit Palta**. Computational methods for DNA copy number detection. Tartu, 2015, 130 p.
283. **Julia Sidorenko**. Combating DNA damage and maintenance of genome integrity in pseudomonads. Tartu, 2015, 174 p.
284. **Anastasiia Kovtun-Kante**. Charophytes of Estonian inland and coastal waters: distribution and environmental preferences. Tartu, 2015, 97 p.
285. **Ly Lindman**. The ecology of protected butterfly species in Estonia. Tartu, 2015, 171 p.
286. **Jaanis Lodjak**. Association of Insulin-like Growth Factor I and Corticosterone with Nestling Growth and Fledging Success in Wild Passerines. Tartu, 2016, 113 p.
287. **Ann Kraut**. Conservation of Wood-Inhabiting Biodiversity – Semi-Natural Forests as an Opportunity. Tartu, 2016, 141 p.
288. **Tiit Örd**. Functions and regulation of the mammalian pseudokinase TRIB3. Tartu, 2016, 182. p.
289. **Kairi Käiro**. Biological Quality According to Macroinvertebrates in Streams of Estonia (Baltic Ecoregion of Europe): Effects of Human-induced Hydromorphological Changes. Tartu, 2016, 126 p.
290. **Leidi Laurimaa**. *Echinococcus multilocularis* and other zoonotic parasites in Estonian canids. Tartu, 2016, 144 p.
291. **Helerin Margus**. Characterization of cell-penetrating peptide/nucleic acid nanocomplexes and their cell-entry mechanisms. Tartu, 2016, 173 p.
292. **Kadri Runnel**. Fungal targets and tools for forest conservation. Tartu, 2016, 157 p.
293. **Urmo Võsa**. MicroRNAs in disease and health: aberrant regulation in lung cancer and association with genomic variation. Tartu, 2016, 163 p.
294. **Kristina Mäemets-Allas**. Studies on cell growth promoting AKT signaling pathway – a promising anti-cancer drug target. Tartu, 2016, 146 p.
295. **Janeli Viil**. Studies on cellular and molecular mechanisms that drive normal and regenerative processes in the liver and pathological processes in Dupuytren's contracture. Tartu, 2016, 175 p.
296. **Ene Kook**. Genetic diversity and evolution of *Pulmonaria angustifolia* L. and *Myosotis laxa sensu lato* (Boraginaceae). Tartu, 2016, 106 p.
297. **Kadri Peil**. RNA polymerase II-dependent transcription elongation in *Saccharomyces cerevisiae*. Tartu, 2016, 113 p.
298. **Katrin Ruisu**. The role of RIC8A in mouse development and its function in cell-matrix adhesion and actin cytoskeletal organisation. Tartu, 2016, 129 p.
299. **Janely Pae**. Translocation of cell-penetrating peptides across biological membranes and interactions with plasma membrane constituents. Tartu, 2016, 126 p.
300. **Argo Ronk**. Plant diversity patterns across Europe: observed and dark diversity. Tartu, 2016, 153 p.

301. **Kristiina Mark.** Diversification and species delimitation of lichenized fungi in selected groups of the family Parmeliaceae (Ascomycota). Tartu, 2016, 181 p.
302. **Jaak-Albert Metsoja.** Vegetation dynamics in floodplain meadows: influence of mowing and sediment application. Tartu, 2016, 140 p.
303. **Hedvig Tamman.** The GraTA toxin-antitoxin system of *Pseudomonas putida*: regulation and role in stress tolerance. Tartu, 2016, 154 p.
304. **Kadri Pärtel.** Application of ultrastructural and molecular data in the taxonomy of helotialean fungi. Tartu, 2016, 183 p.
305. **Maris Hindrikson.** Grey wolf (*Canis lupus*) populations in Estonia and Europe: genetic diversity, population structure and -processes, and hybridization between wolves and dogs. Tartu, 2016, 121 p.
306. **Polina Degtjarenko.** Impacts of alkaline dust pollution on biodiversity of plants and lichens: from communities to genetic diversity. Tartu, 2016, 126 p.
307. **Liina Pajusalu.** The effect of CO₂ enrichment on net photosynthesis of macrophytes in a brackish water environment. Tartu, 2016, 126 p.
308. **Stoyan Tankov.** Random walks in the stringent response. Tartu, 2016, 94 p.
309. **Liis Leitsalu.** Communicating genomic research results to population-based biobank participants. Tartu, 2016, 158 p.
310. **Richard Meitern.** Redox physiology of wild birds: validation and application of techniques for detecting oxidative stress. Tartu, 2016, 134 p.
311. **Kaie Lokk.** Comparative genome-wide DNA methylation studies of healthy human tissues and non-small cell lung cancer tissue. Tartu, 2016, 127 p.
312. **Mihhail Kurašin.** Processivity of cellulases and chitinases. Tartu, 2017, 132 p.
313. **Carmen Tali.** Scavenger receptors as a target for nucleic acid delivery with peptide vectors. Tartu, 2017, 155 p.
314. **Katarina Oganjan.** Distribution, feeding and habitat of benthic suspension feeders in a shallow coastal sea. Tartu, 2017, 132 p.
315. **Taavi Paal.** Immigration limitation of forest plants into wooded landscape corridors. Tartu, 2017, 145 p.
316. **Kadri Õunap.** The Williams-Beuren syndrome chromosome region protein WBSCR22 is a ribosome biogenesis factor. Tartu, 2017, 135 p.
317. **Riin Tamm.** In-depth analysis of factors affecting variability in thiopurine methyltransferase activity. Tartu, 2017, 170 p.
318. **Keiu Kask.** The role of RIC8A in the development and regulation of mouse nervous system. Tartu, 2017, 184 p.