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Immobilization of Chondroitin Sulfate A into Monolithic Epoxy Column for Chiral Separation

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Abstract

Chondroitin sulfate A was successfully immobilized into epoxy monolithic column at a concentration of 3% (w/v) in the presence of ethylene diamine. The epoxy group of monolithic column was first converted to aldehyde group by successive hydrolization and oxidation. A Schiff base reaction at pH 8.0 was used to attach the diamine-spacer to aldehyde group. The chondroitin sulfate A was introduced into the monolithic column by circulating the solution at a flow rate of 0.1 mL/min for 24 hours. The chondroitin sulfate A-immobilized epoxy column was evaluated for chiral separation of verapamil enantiomers under optimized HPLC conditions at a wavelength of 230 nm. As a mobile phase, 20 mM Na₂HPO₄ (pH 2.9) was used. A resolution (R_s) of about 1.5 was achieved for the separation of verapamil enantiomers. A good repeatability of the retention time at two concentration levels (n=8) with RSD < 1% was obtained. The linear responses of verapamil enantiomers were in the range of 1.0-3.0 ppm with R² of about 0.994.

Overview

The high recognition capacity and enantioselectivity of polysaccharides make them ideal chiral selectors [1-3]. Several polysaccharide-based chiral HPLC columns are currently commercially available [4]. Chondroitin sulfate A belongs to mucopolysaccharides. It has shown an initial promising chiral recognition ability on capillary electrophoresis [5], but has not been tried on HPLC yet. Macroporous epoxy-based columns establish low backpressure on HPLC system and have good separation performance [6]. Having porous backbone makes the monolithic stationary phase a good candidate for the immobilization process [6, 7]. In this study, the immobilization of chondroitin sulfate A into the monolithic epoxy column and the evaluation of its chiral recognition ability were carried out.

Experimental



Figure 1. Monolithic silica column.





Figure 2. Structure of chondroitin sulfate A sodium salt.



Figure 3. Hydrolysis of epoxy groups (monolithic epoxy silica column) and addition of diamine-spacer by Schiff base reaction.



Figure 5. Immobilization of polysaccharide into monolithic column.

Results



Minutes

Figure 6. Chromatographic separation of racemic verapamil; mobile phase: Na₂HPO₄ 20 mM pH 2.9; UV detection at 230 nm; *R*_s 1.52.

Figure 7. Calibration curve for verapamil enantiomers.

Conclusion

- Immobilization of chondroitin sulfate A 3% (w/v) in monolithic epoxy column (Chromolith® widepore epoxy 100-4.6 mm) was successful in the presence of ethylene diamine through a Schiff base reaction.
- The immobilized chondroitin sulfate A has shown initial chiral separation ability for verapamil enantiomers.

References

- [1] Ali I., Aboul-Enein Y., Chiral separation techniques: Role of polysaccharides in chiral separations by liquid chromatography and capillary electrophoresis, Subramanian G. Editor, *Wiley-VCH Verlag GmbH & Co.KGaA* (2007), 3rd Edition, p 29-97.
- [2] Aboul-Enein H.Y., Ali I., Chiral separations by liquid chromatography and related technologies: An introduction, *Marcel Dekker Inc.* (2003), Volume 20, p 1-20.
- [3] Ikai T., Okamoto Y., Chiral recognition mechanism in enantiomers separations methods (mechanism and applications): Preparation and chiral recognition of polysaccharide-based selector, Berthod A. Editor, Springer-Verlag (2010), p 33 52.
- [4] Berthod A., Chiral recognition mechanism in enantiomers separations methods (mechanism and applications): A general view, Berthod A. Editor, Springer-Verlag (2010), p 1-32.
- [5] Nishi H., Enantiomer separation on basic drugs by capillary electrophoresis using ionic and neutral polysaccharides as chiral selector, *Journal of Chromatography A* (2006) 735:345-351.
- [6] Ott S., Niessner R., Seidel M., Preparation of epoxy-based macroporous monolithic column and for the fast and efficient immunofiltration of Staphylococcus aureus, Journal of Separation Science (2011), 34(16-17):2181-92.
- [7] Declerck S., Vander Heyden Y., Mangelings D., Enantioseparations of pharmaceuticals with capillary electrochromatography: A review, Journal of Pharmaceutical and Biomedical Analysis (2016), 130:81-99.

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