

## Brief note

# Differences in intracellular localization of corn stunt spiroplasmas in magnesium treated maize.

CLAUDIA NOME<sup>1\*</sup>, PAULO CÉSAR MAGALHÃES<sup>2</sup>, ELIZABETH OLIVEIRA<sup>2</sup>, SERGIO NOME AND IRMA GRACIELA LAGUNA<sup>1</sup>

1. Instituto Nacional de Investigaciones Agropecuarias – Instituto de Fitopatología y Fisiología Vegetal (INTA-IFFIVE). Córdoba. Argentina
2. Embrapa Milho e Sorgo. Caixa Postal 151 – 35701-970, Sete Lagoas, MG, Brasil.

**Key words:** mollicutes, cytopathology, plant pathogen, magnesium deficiency

**ABSTRACT:** Maize plants infected with *Spiroplasma kunkelii* show symptoms similar to that of plants in a magnesium-deficient soil, and it has been shown that the spiroplasma alters the plants' magnesium absorption. In the current study we compared changes associated to either spiroplasma infection, two soil magnesium levels and their combinations. Plant symptoms were recorded and correlated with transmission electron microscopy observations. Plants grown on a high magnesium treatment showed no macroscopical alterations nor organelle ultrastructural alterations, while plants on a low magnesium treatment showed macroscopical vein yellowing and, ultrastructurally, they had most chloroplasts and mitochondrial membranes altered. Infected plants on a low magnesium treatment had an ageing aspect, ultrastructurally showed chloroplasts and mitochondrial alterations similar to those non-infected and grown on a low magnesium treatment, and spiroplasma cells were found in phloem cells, but outside their cytoplasm. Infected plants on a high magnesium treatment showed similar symptoms and ultrastructural alterations as either non-infected plants on the low magnesium treatment or in infected plants on the low magnesium treatment, but differ from them in that the spiroplasma cells were located inside the cytoplasm. Results suggest that magnesium is involved in the plant-pathogen interaction.

Plant pathogens include viruses, viroids, fungi, bacteria, and within the latter, the Mollicutes, a bacterial class including the families Mycoplasmataceae, Spiroplasmataceae and Acheloplasmataceae. They are the smallest known prokaryotes and they lack a cell wall. Their genome oscillates from 577 to 2220 kpb, and they

are extremely demanding from the nutritional point of view, therefore very difficult to grow in culture medium (Bové, 1988).

The Spiroplasmataceae include the genus *Spiroplasma*, whose name derives from the helical morphology and the motility of cells in liquid media. They have been recognized in diseased plants less than thirty years ago, and they have been recognized also as pathogens of warm-blooded animals and insects (Maramorosch, 1981; Bastian *et al.*, 2007). They are diagnosed by serological and molecular methods, and by electron microscopy. The genus name *Phytoplasma* has not yet been formally recognized, but it is currently at "candidatus

\*Address correspondence to: Claudia Nome. Instituto Nacional de Investigaciones Agropecuarias - Instituto de Fitopatología y Fisiología Vegetal (INTA-IFFIVE). Camino 60 cuadras Km 5 1/ 2. X5020ICA Córdoba, ARGENTINA.  
E-mail: [cfnome@yahoo.com.ar](mailto:cfnome@yahoo.com.ar)  
Received: May 23, 2008. Revised version received: June 1, 2009. Accepted: June 2, 2009.

status” and is used for [Spiroplasmataceae] bacteria that cannot be cultured, and they belong to the monophyletic order Acholeplasmatales (Lee *et al.*, 2000). They are only detected by molecular methods, electron microscopy, and by the response of the host plant to antibiotic treatment. Phytoplasmas exist both in plants and in insects feeding on phloem of infected plants. As for spiroplasmas, such insects have been described as the pathogen vectors. Inside the plants, both phytoplasmas and spiroplasmas are confined out of the cytoplasm of the sieve cells, while in insects they are distributed in several tissues (Conci *et al.*, 2001; Ammar and Hogenhout, 2005).

*Spiroplasma kunkelii* is a maize pathogen transmitted by the leafhopper *Dalbulus maydis* (A+Y), producing a disease named corn stunt disease. It has been reported that the infected plants show a lower magnesium absorption, Oliveira *et al.* (2002, 2005).

The symptoms of maize plants grown on magnesium deficient soil (concentrations below 0,5 Cmol c dm<sup>-3</sup>) are leaf veins yellowing/whitening all along their length, and purple coloring on the tips of older leaves, Coelho and França (1995). Almost identical symptoms are the ones produced by the spiroplasma infection (Tsai and Miller, 1995).

The aim of this work was to compare plant alterations associated with low soil magnesium levels and by the infection with *S. kunkelii*, and to assess the possibility of spiroplasma-induced alterations were reverted by a high magnesium availability.

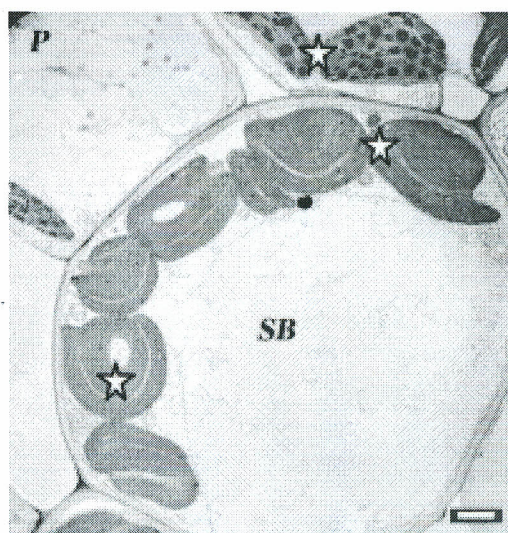
*Zea mays* plants, cv. Pop Zélia, were bred on soil from the Brazilian Cerrado (savanna) with naturally shows low magnesium levels, to which magnesium was added to obtain either 0.20 or 1.0 Cmol c dm<sup>-3</sup> (they will be referred to as the low and the high magnesium treatments). They were kept in greenhouse with stable conditions of 27°C, 16 hours of lighting and 60% of humidity. Some plants were kept uninfected, while others were exposed to the leafhopper *Dalbulus maydis* for inoculation with *S. kunkelii* after seven days of plant emergence.

Sheath tissue samples of maize leaves (2 x 3 mm) sixty seven days old (sixty days after inoculation) were collected and fixed in Karnovsky fluid, post fixed in osmium tetroxide 1%, contrasted in uranyl acetate 0,5%, dehydrated in acetone solution series, pre-embedded in acetone-epoxy resin, and embedded in low viscosity Spurr resin (polymerized at 70°C for 2 days). Ultrathin sections were made with a diamond blade and sections were contrasted with lead citrate pH 12 and with uranyl acetate 2%. Sections were observed with a transmission electron microscope Jeol 1220 JEM EXII.

Uninfected plants on the high magnesium treatment showed no abnormal symptoms, while maize plants on the low magnesium treatment showed vein yellowing



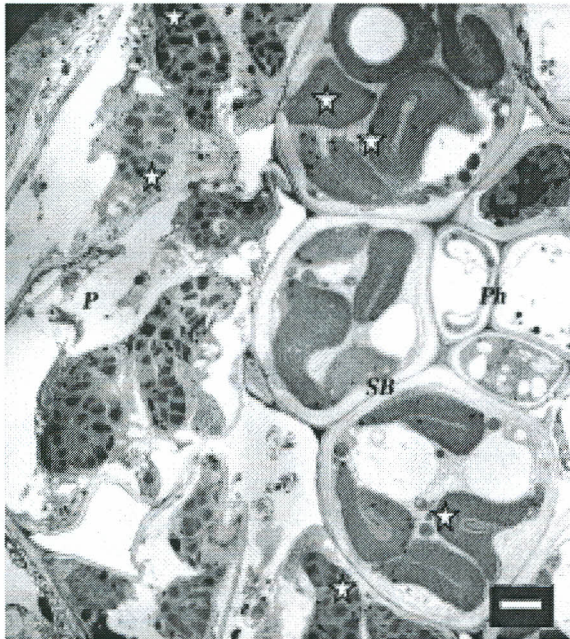
**FIGURE 1.** Uninfected maize cells on the high magnesium treatment. Normal chloroplasts are seen in the sheath bundle and the parenchyma. Vascular bundle shows no spiroplasmas. In this and in all subsequent figures, stars indicate chloroplasts; arrows indicate spiroplasmas; Cy is cytoplasm; CW, cell wall; SB, sieve bundle; X, xylem; Ph, phloem; P, phloem parenchyma. Scale bar represents 7  $\mu$ m.



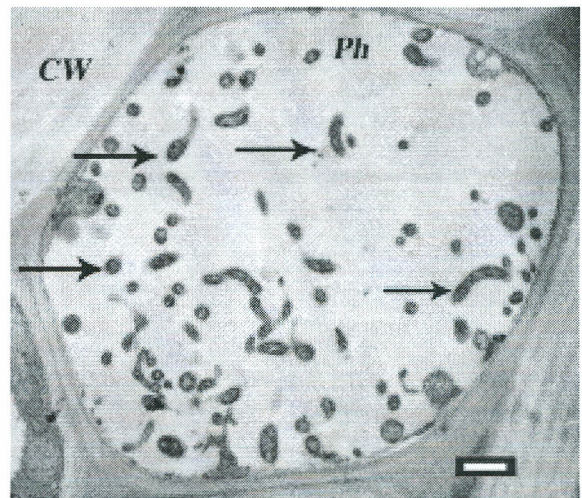
**FIGURE 2.** Uninfected maize cells on the low magnesium treatment. Picture shows distorted chloroplasts of sheath bundle and normal parenchymal chloroplasts. Scale bar represents 2  $\mu$ m.

all along their leaves and some whitening down on the sheath. Also, infected plants cultivated on low magnesium treated soil appeared old, but showed a more intense purple coloring and sometimes even leaf apex necrosis. Infected plants on the high magnesium treatment also showed vein yellowing all along the sheath and purple coloring on older leaves; such plants appeared old, though they were less damaged than spiroplasma infected plants on the low magnesium treatment.

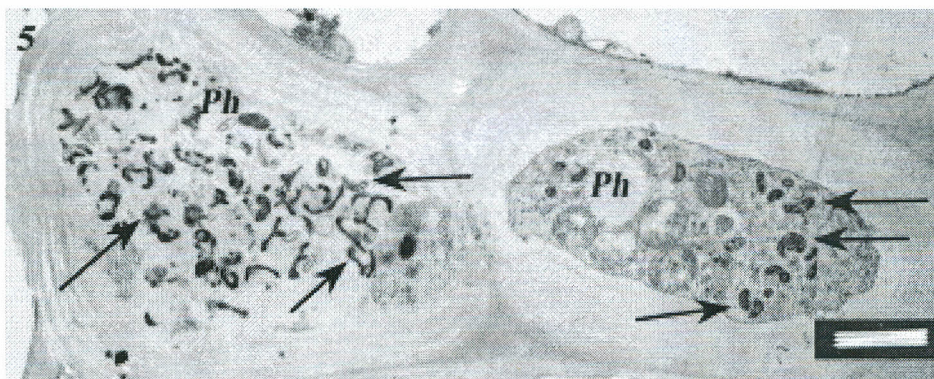
As expected, uninfected plants on high magnesium treatment showed no ultrastructural alterations (Fig. 1). However, uninfected plants on the low magnesium treatment exhibited chloroplast alterations, swollen chloroplasts with disordered grana, in all of the studied tissues (Fig. 2). Spiroplasma infected plants on the low magnesium treatment also showed disorganized chloroplasts, mainly in their veins, and the pathogen was seen inside phloem sieve tubes, but not in the cytoplasm (Figs. 3, 5). In contrast, infected plants on the high magnesium treatment, showed the spiroplasma both outside and inside the cytoplasm of phloem cells (Figs. 4, 6).



**FIGURE 3.** Spiroplasma infected maize cells on the low magnesium treatment. Micrograph shows devastated chloroplasts both in sheath bundle and the parenchyma. Scale bar represents 2  $\mu$ m.



**FIGURE 4.** Sieve tube of infected maize plant on the low magnesium treatment with spiroplasmas. Ordinary image of spiroplasmas spread loose out of the cytoplasm. Scale bar represents 500 nm.



**FIGURE 5.** Sieve tube of an infected maize plant on the high magnesium treatment showing spiroplasmas within their cells. Notably, the pathogen is not only within sieve tubes but also inside cytoplasm of young phloem cells and phloem parenchyma. Scale bar represents 1  $\mu$ m.

It is known that low magnesium concentrations induce maize plants to show leaf vein yellowing (INFOPOS, 2001; Bull, 1993). Earlier studies have shown that *S. kunkelii* infection reduces magnesium concentration in maize plants (Oliveira *et al.*, 2002, 2005; Ammar *et al.*, 2005). We may suggest, therefore, that the symptom expression may be related to a competition for the cation between the plant and the pathogen.

Infected plants on the low magnesium treatment were more damaged than infected plants on the high magnesium treatment. It is notable, however, that the spiroplasma locates *inside* of the young phloem cells of

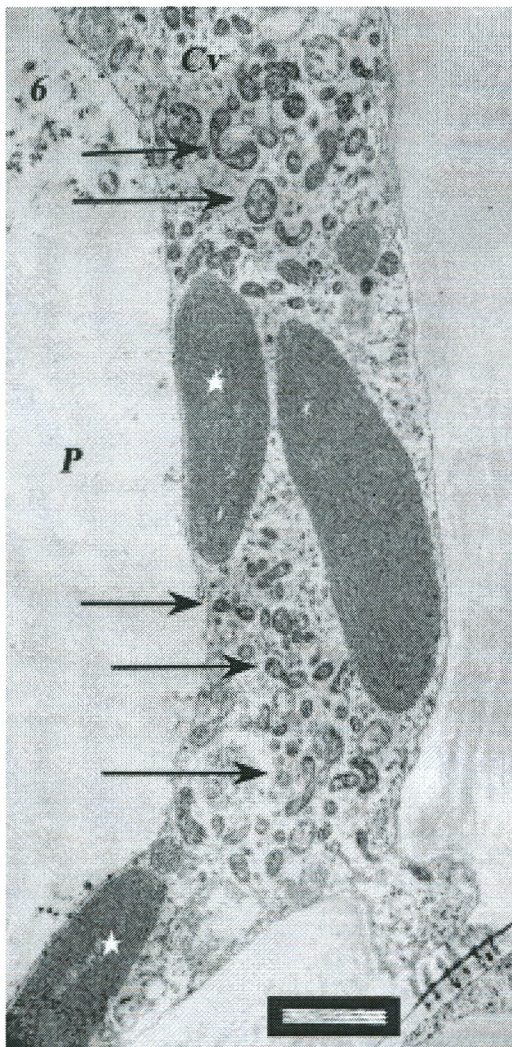


FIGURE 6. Same as in Fig. 5. Scale bar represents 1  $\mu$ m.

infected plants on the high magnesium treatment. This may suggest that magnesium influences the way that the pathogen invades the plant, and that it allows the spiroplasma to introduce inside the young cells, and to remain alive, and to proliferate within their cytoplasm.

## References

- Aminar E, Hogenhout S (2005). Use of Immunofluorescence Confocal Laser Scanning Microscopy to Study Distribution of the Bacterium Corn Stunt Spiroplasma in Vector Leafhoppers (Hemiptera: Cicadellidae) and in Host Plants. *Annals of the Entomological Society of America* **98**: 820-826.
- Bastian F, Sanders D, Forbes W, Hagius S, Walker J, Henk W, Enright FM, Elzer PH (2007). *Spiroplasma* spp. from transmissible spongiform encephalopathy brains or ticks induce spongiform encephalopathy in ruminants. *Journal of Medical Microbiology* **56**: 1235-1242.
- Bull L (1993). Nutrição mineral do milho. In: Simposio sobre fatores que afetam a produtividade do milho e do sorgo. In: Bull, L.T.; Cantarella, H. (Eds.). *Cultura do milho; fatores que afetam a produtividade*. Piracicaba: POTAFOS, p. 63-145.
- Bové J (1988). Plant mollicutes: phloem-restricted agents and surface contaminants. *Acta Horticulturae* **225**: 215-222.
- Coe'ho A, França G (1995). Nutrição e Adução. 2 ed. Aum. In: Arquivo Agrônomo, n.º 2, POTAFÓS. (Piracicaba, SP). *Seja o doutor de seu milho*. Piracicaba: p. 1-9.
- Conci L, Torres L, Galdeano E, Meneguzzi N, Nome C, Docampo D, Nome S, Paradell S, Conci VC, Ramallo JC (2001). Fitoplasmas. Caracterización molecular, diagnóstico y epidemiología de fitoplasmas que afectan cultivos de importancia económica. In: *Memoria anual INTA-IFFIVE*. INTA. Buenos Aires, Argentina, pp. 103.
- INFOPOS (2001). *Conozca y Resuelva los Problemas del Maíz*. International Plant Nutrition Institute. Trípico SP-0801.
- Lee I, Davis R, Gundersen-Rindal D (2000). Phytoplasma: Phytopathogenic Mollicutes. *Annual Review of Microbiology* **54**: 221-55.
- Malavota E, Haang H, de Mello F, Brasil Sobr M (1964). Los elementos nutritivos esenciales. In: *La nutrición mineral de algunas cosechas tropicales*. Malavota E, Haang HP, de Mello FAF and Brasil Sobr MOC. Eds. Instituto Nacional de la potasa Bernal, Suiza, pp. 24-25.
- Maramorosch K (1981). Spiroplasmas: Agents of Animal and Plant Diseases. *BioScience* **31**: 374-380.
- Oliveira C, Cruz I, Schaffert R (2002). Growth and nutrition of mollicute infected maize. *Plant Disease* **86**: 945-949.
- Oliveira E, Oliveira C, Souza I, Magalhães P, Andrade C, Hogenhout S (2005). Spiroplasma and phytoplasma reduce kernel production and nutrient and water contents of several but not all maize cultivars. *Maydica* **50**: 171-178.
- Tsai J, Miller J (1995). Corn Stunt Spiroplasma. *Plant Pathology Circular* 373.