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**Homocysteine and folate plasma concentrations in mother and baby at delivery after pre-eclamptic or normotensive pregnancy; influence of parity.**

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**Short title:** Homocysteine, folate, pre-eclampsia & parity

**Keywords:** Homocysteine, folate, maternal nutrition, parity; pre-eclampsia.

1 **Abstract**

2 Pre-eclampsia affects between 2-7% of pregnant women. There are conflicting data on plasma  
3 homocysteine and folate in pre-eclampsia, and little about fetal concentrations.

4 **Objectives:** To compare the concentrations of homocysteine and folate in maternal and paired fetal  
5 (umbilical venous) plasma samples from normotensive or pre-eclamptic pregnancies at delivery and to  
6 identify any effect of parity on these concentrations.

7 **Study design:** Hospital based cross-sectional study of 24 normotensive and 16 pre-eclamptic pregnant  
8 women from whom maternal and fetal plasma samples were collected at delivery.

9 **Main outcome measures:** Maternal and fetal plasma homocysteine and folate concentrations between  
10 normotensive and pre-eclamptic pregnancies with varying parity.

11 **Results:** There were no significant differences in either maternal or fetal plasma homocysteine or folate  
12 concentrations between normotensive and pre-eclamptic pregnancies. In both the normotensive and  
13 pre-eclamptic women, plasma folate concentration was higher in paired fetal compared to maternal  
14 plasma ( $P < 0.001$  and  $P = 0.047$  respectively). Both maternal and fetal plasma folate concentrations  
15 were lower in parous women ( $P = 0.001$ ;  $P = 0.017$  respectively), the lowest concentrations being in  
16 pre-eclamptic parous women ( $P = 0.004$ ), but homocysteine concentrations were similar.

17 **Conclusions:** The low plasma folate in parous women is an interesting finding and, when intake is also  
18 low, may contribute to adverse pregnancy outcomes, particularly in relation to pre-eclampsia.

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## 26 **Introduction**

27 Pre-eclampsia is estimated to occur in 2-7% of all pregnancies and is a leading cause of maternal and  
28 perinatal mortality and morbidity in the Western world [1]; together with other hypertensive disorders  
29 of pregnancy it is responsible for approximately 60,000 deaths each year [2]. Pre-eclampsia is now  
30 commonly regarded as being a state of oxidative stress [3]. It is thought that primarily inadequate  
31 placental perfusion results in excessive production of reactive oxygen species giving rise to endothelial  
32 cell dysfunction and thus clinical manifestations of pre-eclampsia [4].

33 Homocysteine is a metabolic product of methyl-group donation by the amino acid methionine;  
34 remethylation is catalysed by folate. Small increases in plasma homocysteine concentrations are  
35 associated with increased risk of vascular disease [5], Alzheimer's disease [6] and neural tube defects  
36 [7] in the general population. Elevated homocysteine concentrations contribute to oxidative stress and  
37 endothelial dysfunction [8] and are thus also potentially implicated in the pathogenesis of pre-  
38 eclampsia. Some studies report raised homocysteine concentrations in pre-eclampsia (e.g. [9, 10])  
39 while others have shown no significant differences [11, 12]. Maternal and fetal plasma homocysteine  
40 concentrations have been reported to be directly correlated in healthy nulliparae [13] and do not appear  
41 to change significantly during the course of pregnancy [14]. If plasma homocysteine concentrations are  
42 indeed raised in pre-eclampsia, and there is similar parallelism, then the fetus will be exposed to  
43 potentially damaging levels of homocysteine even before birth, which might have long-term  
44 consequences. Important factors influencing homocysteine concentrations are folate and vitamin B<sub>12</sub>  
45 status and the methylenetetrahydrofolate reductase (MTHFR) polymorphisms [15]; once again the data  
46 on these in pre-eclampsia is conflicting [16, 17].

47 There is considerably less, but similarly conflicting, evidence linking folate concentrations and pre-  
48 eclampsia [18, 19]. The fetus must receive an adequate supply of folate for growth and development;  
49 inadequate concentrations will also, *inter alia*, impede the remethylation of homocysteine. Both  
50 metabolites have active transport systems in the placenta [20] and impaired placentation is believed to

51 be central to the pathogenesis of pre-eclampsia. Folate has recently been shown to possibly play a  
52 direct role in extravillous trophoblast invasion [21], thus highlighting the need for adequate folate  
53 concentrations pre-pregnancy and during the early stages of pregnancy. Studies linking homocysteine  
54 and folate in the mother and fetus are very limited [17]. We therefore felt it to be important to measure  
55 fetal, as well as maternal, concentrations of homocysteine and folate at delivery.

56 Folate intake among women of reproductive age in the UK is reported to be low [22]. There is an  
57 increased demand for folate during pregnancy, which can result in suboptimal folate status [23],  
58 although there is some evidence to suggest that folate turnover during pregnancy does not appear to  
59 change if folate intake is adequate [24]. Increasing parity has been associated with decreasing plasma  
60 folate concentrations [25, 26], presumably because lactation is a further drain on folate reserves [27]. If  
61 dietary intake is suboptimal, these stores may not be replenished before a subsequent pregnancy,  
62 especially in cases of short inter-pregnancy intervals [25, 28]. We are unaware of any studies relating  
63 these low folate levels to increased plasma homocysteine concentrations in either parous women as a  
64 group, or parous women who develop pre-eclampsia. However, there is indirect evidence for an  
65 interaction between parity and raised plasma homocysteine concentrations which has been related to  
66 increased risk of pre-eclampsia [29, 30]. We hypothesised that plasma folate concentrations would be  
67 lower, and homocysteine higher, in pre-eclamptic women and their babies than in normotensive  
68 pregnant controls. We also opportunistically examined the effect of parity on maternal plasma folate  
69 concentrations in normal and pre-eclamptic pregnancy.

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76 **Methods**

77 *Subjects:* These investigations formed part of a detailed study of selenium and glutathione peroxidases  
78 in pregnancy, requiring recruitment of 25 pregnant women to each arm (see: [31]). The current study  
79 had a power of 80% to detect a difference of 1SD in maternal plasma folate concentration between the  
80 24 normotensive and 16 pre-eclamptic women at the 5% level from whom paired maternal and fetal  
81 samples were available. Approval for these investigations was given by the Nottingham Hospital Ethics  
82 Committee and written informed consent was obtained from each participant. The study population  
83 consisted of two groups of White European women: normotensive and pre-eclamptics (Table 1). Paired  
84 fetal samples were also obtained via umbilical venous plasma. Cases were defined on admission with a  
85 clinical diagnosis of pre-eclampsia, using the International Society for the Study of Hypertension in  
86 Pregnancy definition of blood pressure  $\geq 140/90$  mm Hg (Korokoff V) on 2 occasions after 20 weeks  
87 gestation and proteinuria  $> 300$  mg/L, 500 mg/day or 2+ on dipstick [32]. Controls were healthy with  
88 no pregnancy or medical complications.

89 *Sample collection:* Non-fasting venous blood samples were taken from mothers before delivery; where  
90 possible, umbilical venous samples were also taken immediately after placental delivery, in EDTA.  
91 Samples were transported on ice to the laboratory and centrifuged at 1,400 x g for 10 minutes at 4°C  
92 and plasma was immediately stored at -80°C until analysis.

93 *Homocysteine measurements:* Measurements of plasma homocysteine concentrations were obtained via  
94 fluorescence polarization immunoassay on the Abbott AxSym analyser. This assay has an analytical  
95 range of 1-50  $\mu\text{mol/L}$  and reproducibility (CV) of 5.5% at 7.4  $\mu\text{mol/L}$ , 6.2% at 13.5  $\mu\text{mol/L}$ , and 5.4%  
96 at 25.9  $\mu\text{mol/L}$ .

97 *Folate assay:* A microbiological technique with a chloramphenicol-resistant strain of *Lactobacillus*  
98 *rhamnosus* (*L. rhamnosus*) was used to assess the plasma folate concentrations as described previously  
99 [33]; 15 mg/ml manganese sulphate was added to the buffer to avoid artificially low plasma folate  
100 values [33, 34]. The inter- and intra-assay CVs were 5% and 4% respectively.

101 *Statistical analysis:* All tests were performed using SPSS for Windows version 14.0. Summary data  
102 are presented as medians [interquartile range (IQR)] or means  $\pm$  SD depending on the distribution. The  
103 Kolmogorov-Smirnov test was used to assess the distribution of the data. Between group comparisons  
104 were made using Kruskal-Wallis tests or 2-tailed Student's *t* tests/Mann-Whitney *U*-tests depending on  
105 the distribution. Wilcoxon Signed Ranks tests were used to carry out paired comparison. Multiple  
106 regression analysis for maternal folate and parity correcting for inter-pregnancy interval and number of  
107 previous pregnancies were conducted. Correlations between the parameters were tested using  
108 Spearman's Ranks correlation test. The null hypothesis was rejected where  $P < 0.05$ .

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126 **Results**

127 *Subjects:* Both groups conceived spontaneously, carried singleton pregnancies and all neonates  
128 survived. Normotensive pregnant women delivered without developing hypertension or proteinuria,  
129 having infants weighing > 2500 g, delivered at 37 weeks' gestation or later. The systolic and diastolic  
130 blood pressure levels were, by definition, significantly raised in pre-eclampsia compared to normal  
131 pregnancy ( $P < 0.0001$  for both; Table 1). Overall, the pre-eclamptic women all had moderate-to-  
132 severe disease (see[31]).

133 *Biochemical and molecular measurements:* Maternal and fetal plasma homocysteine and folate  
134 concentrations are given in Table 1. Plasma homocysteine concentrations were very similar in  
135 normotensive and pre-eclamptic women; fetal concentrations were also very similar in the two groups.  
136 Maternal and fetal homocysteine concentrations were significantly positively correlated ( $r = 0.68$ ,  $R^2 =$   
137  $0.68$ ,  $P < 0.0001$ ). Fetal homocysteine concentrations were higher than maternal in normotensive ( $P =$   
138  $0.002$ ), but not pre-eclamptic ( $P > 0.1$ ) pregnancy.

139 Maternal folate appeared higher in normal than pre-eclamptic pregnancy, but there this was not  
140 statistically significant due to the large range of data. Fetal plasma folate concentration did not differ  
141 across groups (Table 1) and were positively correlated with maternal concentrations ( $r = 0.44$ ,  $R^2 =$   
142  $0.20$ ,  $P = 0.005$ ). Fetal folate concentrations were higher than maternal in normotensive ( $P < 0.001$ ) and  
143 pre-eclamptic ( $P = 0.047$ ) pregnancies. Correlation analysis showed no evidence for an association  
144 between plasma homocysteine and folate concentrations in either maternal or umbilical venous plasma  
145 ( $r = 0.07$ ,  $R^2 = 0.03$ ,  $P > 0.6$ ;  $r = 0.09$ ,  $R^2 = 0.023$ ,  $P > 0.4$  respectively).

146 *Folate concentrations and parity:* Overall, parous women and their babies had significantly lower  
147 plasma folate concentrations (Maternal: 6.0 [4.9, 6.8]; Fetal 18.2 [10.8, 22.2]) than nulliparous women  
148 (13.5 [7.5, 19.3];  $P = 0.001$ ) and babies (23.8 [17.2, 33.4];  $P = 0.017$ ; Figure 1a and b). Moreover, in  
149 both normotensive and pre-eclamptic women, parous women had significantly lower folate  
150 concentrations (normotensive: 6.2 [4.8, 10.4] cf pre-eclamptic: 5.4 [5.0, 6.2];  $P = 0.006$ ) compared to



151 nulliparous women (normotensive: 14.3 [10.2, 20.8] cf pre-eclamptic: 11.4 [6.2, 17.6];  $P = 0.015$ ;  
152 Figure 1b). Parous pre-eclamptic women had the lowest plasma folate concentrations (ANOVA;  $P =$   
153 0.004). This difference was maintained when considered with respect to fetal plasma folate but only in  
154 the normotensive group (parous: 13.8 [10.4, 21.8] cf nulliparous 17.3 [11.5, 25.5];  $P = 0.015$ ). A  
155 Kruskal-Wallis test showed that maternal folate concentrations were significantly different ( $P = 0.006$ )  
156 between nulliparous women and women who had had one or two previous pregnancies. Subsequent  
157 Mann-Whitney  $U$  tests indicated that the maternal folate concentrations were significantly higher in  
158 nulliparous women compared to parous women with 1 ( $P = 0.005$ ) and 2 previous ( $P = 0.037$ )  
159 pregnancies (Table 2). Multiple regression analysis correcting for confounding factors thought to  
160 influence maternal plasma folate concentrations (inter-pregnancy intervals, number of previous  
161 pregnancies) indicated parity as an independent factor. No similar effect was seen with respect to  
162 homocysteine.

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165 **Discussion**

166 Although small, this cross-sectional study reports both maternal and fetal homocysteine and folate  
167 concentrations in normotensive and pre-eclamptic pregnancies; there appears to be little comparable  
168 maternal:fetal data from European populations [17]. Our maternal data are consistent with the results of  
169 other European studies in which no association was found with respect to homocysteine or folate  
170 concentrations and normotensive and pre-eclamptic pregnancies [11, 12]. Homocysteine has been  
171 potentially linked to processes such as inflammation [35] and hypercoagulation [8], which are present  
172 in severe pre-eclampsia. However, this does not mean that hyperhomocysteinaemia is a requirement for  
173 the development of pre-eclampsia and the elevated homocysteine concentrations seen in some pre-  
174 eclamptic patients may be an associated, or pre-disposing, rather than a causal factor.

175 Folate plays an essential role in the growth of the placenta from early pregnancy; folate deficiency has  
176 been linked to placental abruption and thus restricted fetal growth [36]. We observed no significant  
177 differences between maternal or fetal samples from normotensive pregnancy or pre-eclampsia. Most  
178 folate concentrations were within the reference range (7 – 46 nmol/L [37]) for non-pregnant UK adults,  
179 using this methodology, but 9 normotensive pregnant (7 parous) and 12 pre-eclamptic (6 parous)  
180 women had concentrations below this level and the pre-eclamptic patients had the lowest median  
181 values, which is consistent with previous studies examining folate concentrations [10, 17, 38]. Having  
182 folate concentrations at the lowest end of the range may contribute to the pathological consequences of  
183 oxidative stress as folate has been reported to have antioxidant properties [39]. The placenta takes up  
184 folate from the maternal circulation against a concentration gradient through a high-affinity binding  
185 site, the folate receptor- $\alpha$  [40], and releases it into the fetal circulation through the reduced folate  
186 carrier. Further functional studies are required to ascertain the changes folate transport in adequate and  
187 deficient folate concentrations.

188 Women in the UK have moderately low folate intakes for the developed world [41]; in our laboratory, a  
189 group of normotensive, non-pregnant women of comparable age had a median folate concentration of  
190 13.9 nmol/L, relatively low in the reference range [37]. Periconceptual dietary supplementation with  
191 folic acid is widely advocated for the prevention of neural tube defects. However, only four women in  
192 normotensive and two women in the pre-eclamptic group (Table 1) took folic acid supplementation in  
193 this study. Unless there is dietary supplementation with folic acid, maternal folate concentrations in  
194 both plasma and red blood cells fall from about the fifth month of pregnancy [42].

195 Although sample sizes were small, we observed lower plasma folate concentrations in parous mothers  
196 and their babies in our study (Fig 1a), a trend exacerbated when pre-eclampsia supervened (Fig 1b). We  
197 do not know whether the parous women began pregnancy with lower stores or whether the uptake or  
198 renal reabsorption of folate differs in parous women. Larger studies are required to investigate this  
199 further especially by the findings from our group to indicate that folate may be essential early in  
200 pregnancy possibly directly promoting extravillous trophoblast invasion [21]. As only two women had  
201 3 previous pregnancies, statistical tests could not be performed. These women did however have  
202 maternal plasma concentrations of 6.2 and 6.6 nmol/L respectively, which are all approximately half of  
203 the nulliparous folate concentrations (Table 2). This suggests that regardless of how many previous  
204 pregnancies a woman has had, her folate concentrations appear to be significantly lower than  
205 nulliparous women. Qvist *et al* also showed that by 2 -3 months after delivery, a third of all mothers  
206 can have subnormal concentrations of folate in serum and red blood cells, both those who are and those  
207 who are not breast-feeding [42]. Furthermore, it has been reported that by 6 months postpartum, 20% of  
208 mothers were still folate deficient [43]. The low plasma folate concentration in parous women  
209 independent of inter-pregnancy interval suggest possible beneficial effects of pre-pregnancy folic acid  
210 supplementation particularly in parous women who have previously suffered from pre-eclampsia.

211 Despite public health campaigns recommending the use of pre-conceptual folic acid supplementation,

212 the dietary intake of folic acid is still inadequate in about 13 million people within the UK [44]. Further  
213 investigations using larger cohorts are underway to further examine these fascinating initial data.

214

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222

## 223 **Contributors**

224 HDM completed this study as part of a PhD funded by BBSRC and wrote the majority of this article.  
225 JM completed the main laboratory experiments supervised by HDM and LOK as part of her BMedSci  
226 research project. FBP and MES were the principal investigators; MMR provided the clinical input for  
227 this study.

228

## 229 **Conflict of Interest**

230 None of the authors had a personal or financial conflict of interest.

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237 **References**

- 238 [1] B. Sibai, G. Dekker, and M. Kupferminc, Pre-eclampsia. *Lancet* 365 (2005) 785-99.  
 239 [2] F. Broughton Pipkin, Risk factors for preeclampsia. *N Engl J Med* 344 (2001) 925-6.  
 240 [3] L. Poston, The Role of Oxidative Stress. in: H. Critchley, A. MacLean, L. Poston, and J. Walker,  
 241 (Eds.), *Pre-eclampsia*, RCOG Press, London, 2004.  
 242 [4] C.A. Hubel, Oxidative stress in the pathogenesis of preeclampsia. *Proc Soc Exp Biol Med* 222  
 243 (1999) 222-35.  
 244 [5] E.S. Ford, S.J. Smith, D.F. Stroup, K.K. Steinberg, P.W. Mueller, and S.B. Thacker,  
 245 Homocyst(e)ine and cardiovascular disease: a systematic review of the evidence with special  
 246 emphasis on case-control studies and nested case-control studies. *Int J Epidemiol* 31 (2002) 59-  
 247 70.  
 248 [6] S. Seshadri, A. Beiser, J. Selhub, P.F. Jacques, I.H. Rosenberg, R.B. D'Agostino, P.W. Wilson, and  
 249 P.A. Wolf, Plasma homocysteine as a risk factor for dementia and Alzheimer's disease. *N Engl*  
 250 *J Med* 346 (2002) 476-83.  
 251 [7] J.L. Mills, J.M. Scott, P.N. Kirke, J.M. McPartlin, M.R. Conley, D.G. Weir, A.M. Molloy, and Y.J.  
 252 Lee, Homocysteine and neural tube defects. *J Nutr* 126 (1996) 756S-760S.  
 253 [8] J.M. Roberts, R.N. Taylor, T.J. Musci, G.M. Rodgers, C.A. Hubel, and M.K. McLaughlin,  
 254 Preeclampsia: an endothelial cell disorder. *Am J Obstet Gynecol* 161 (1989) 1200-4.  
 255 [9] A. Gurbuz, A. Karateke, and M. Mengulluoglu, Elevated plasma homocysteine levels in  
 256 preeclampsia and eclampsia. *Int J Gynaecol Obstet* 87 (2004) 165-6.  
 257 [10] L.E. Mignini, P.M. Latthe, J. Villar, M.D. Kilby, G. Carroli, and K.S. Khan, Mapping the theories  
 258 of preeclampsia: the role of homocysteine. *Obstet Gynecol* 105 (2005) 411-25.  
 259 [11] K. Mayerhofer, L. Hefler, H. Zeisler, C. Tempfer, K. Bodner, S. Stockler-Ipsiroglu, A. Muhl, A.  
 260 Kaider, C. Schatten, S. Leodolter, P. Husslein, and C. Kainz, Serum homocyst(e)ine levels in  
 261 women with preeclampsia. *Wien Klin Wochenschr* 112 (2000) 271-5.  
 262 [12] W. Herrmann, U. Hubner, I. Koch, R. Obeid, U. Retzke, and J. Geisel, Alteration of homocysteine  
 263 catabolism in pre-eclampsia, HELLP syndrome and placental insufficiency. *Clin Chem Lab*  
 264 *Med* 42 (2004) 1109-16.  
 265 [13] M.R. Malinow, A. Rajkovic, P.B. Duell, D.L. Hess, and B.M. Upson, The relationship between  
 266 maternal and neonatal umbilical cord plasma homocyst(e)ine suggests a potential role for  
 267 maternal homocyst(e)ine in fetal metabolism. *Am J Obstet Gynecol* 178 (1998) 228-33.  
 268 [14] H. Watanabe, H. Fukuoka, T. Sugiyama, Y. Nagai, K. Ogasawara, and N. Yoshiike, Dietary folate  
 269 intake during pregnancy and birth weight in Japan. *Eur J Nutr* 47 (2008) 341-7.  
 270 [15] A.M. Molloy, J.L. Mills, J. McPartlin, P.N. Kirke, J.M. Scott, and S. Daly, Maternal and fetal  
 271 plasma homocysteine concentrations at birth: the influence of folate, vitamin B12, and the 5,10-  
 272 methylenetetrahydrofolate reductase 677C-->T variant. *Am J Obstet Gynecol* 186 (2002) 499-  
 273 503.  
 274 [16] G. Makedos, A. Papanicolaou, A. Hitoglou, I. Kalogiannidis, A. Makedos, V. Vrazioti, and M.  
 275 Goutzioulis, Homocysteine, folic acid and B12 serum levels in pregnancy complicated with  
 276 preeclampsia. *Arch Gynecol Obstet* 275 (2007) 121-4.  
 277 [17] K. Braekke, P.M. Ueland, N.K. Harsem, A. Karlsen, R. Blomhoff, and A.C. Staff, Homocysteine,  
 278 Cysteine, and Related Metabolites in Maternal and Fetal Plasma in Preeclampsia. *Pediatr Res*  
 279 (2007).  
 280 [18] A. Rajkovic, P.M. Catalano, and M.R. Malinow, Elevated homocyst(e)ine levels with  
 281 preeclampsia. *Obstet Gynecol* 90 (1997) 168-71.  
 282 [19] S.E. Sanchez, C. Zhang, M. Rene Malinow, S. Ware-Jauregui, G. Larrabure, and M.A. Williams,  
 283 Plasma folate, vitamin B(12), and homocyst(e)ine concentrations in preeclamptic and  
 284 normotensive Peruvian women. *Am J Epidemiol* 153 (2001) 474-80.

- 285 [20] T.R. Regnault, Y. Kudo, J. Glazier, S. Roos, R.M. Lewis, and T. Jansson, Heterodimeric amino  
286 acid transporters in the placenta--a workshop report. *Placenta* 28 Suppl A (2007) S103-6.
- 287 [21] P.J. Williams, J.N. Bulmer, B.A. Innes, and F. Broughton Pipkin, Folic acid: novel roles in  
288 placentation?, Society for Gynecologic Investigations, Reproductive Sciences, Orlando, Florida,  
289 2010, pp. 217.
- 290 [22] T. Mouratidou, F. Ford, F. Prountzou, and R. Fraser, Dietary assessment of a population of  
291 pregnant women in Sheffield, UK. *Br J Nutr* 96 (2006) 929-35.
- 292 [23] M.A. Caudill, J.F. Gregory Iii, A.D. Hutson, and L.B. Bailey, Folate Catabolism in Pregnant and  
293 Nonpregnant Women with Controlled Folate Intakes. *J. Nutr.* 128 (1998) 204-208.
- 294 [24] J.F. Gregory, 3rd, M.A. Caudill, F.J. Opalko, and L.B. Bailey, Kinetics of folate turnover in  
295 pregnant women (second trimester) and nonpregnant controls during folic acid  
296 supplementation: stable-isotopic labeling of plasma folate, urinary folate and folate catabolites  
297 shows subtle effects of pregnancy on turnover of folate pools. *J Nutr* 131 (2001) 1928-37.
- 298 [25] L.J. Smits, and G.G. Essed, Short interpregnancy intervals and unfavourable pregnancy outcome:  
299 role of folate depletion. *Lancet* 358 (2001) 2074-7.
- 300 [26] M.A. Caudill, J.F. Gregory, A.D. Hutson, and L.B. Bailey, Folate catabolism in pregnant and  
301 nonpregnant women with controlled folate intakes. *J Nutr* 128 (1998) 204-8.
- 302 [27] E.A. Letsky, The Haematological System. in: G.V. Chamberlain, and F. Broughton Pipkin, (Eds.),  
303 *Clinical Physiology in Obstetrics*, Blackwell Science Ltd, Oxford, 1998, pp. 71-110.
- 304 [28] M. van Eijsden, L.J. Smits, M.F. van der Wal, and G.J. Bonsel, Association between short  
305 interpregnancy intervals and term birth weight: the role of folate depletion. *Am J Clin Nutr* 88  
306 (2008) 147-53.
- 307 [29] A. Rajkovic, K. Mahomed, M.R. Malinow, T.K. Sorenson, G.B. Woelk, and M.A. Williams,  
308 Plasma Homocyst(e)ine Concentrations in Eclamptic and Preeclamptic African Women  
309 Postpartum. *Obstet Gynecol* 94 (1999) 355-360.
- 310 [30] S.E. Sanchez, C. Zhang, M. Rene Malinow, S. Ware-Jauregui, G. Larrabure, and M.A. Williams,  
311 Plasma Folate, Vitamin B12, and Homocyst(e)ine Concentrations in Preeclamptic and  
312 Normotensive Peruvian Women. *Am. J. Epidemiol.* 153 (2001) 474-480.
- 313 [31] H.D. Mistry, V. Wilson, M.M. Ramsay, M.E. Symonds, and F. Broughton Pipkin, Reduced  
314 selenium concentrations and glutathione peroxidase activity in pre-eclamptic pregnancies.  
315 *Hypertension* 52 (2008) 881-888.
- 316 [32] M.A. Brown, M.D. Lindheimer, M. de Swiet, A. Van Assche, and J.M. Moutquin, The  
317 classification and diagnosis of the hypertensive disorders of pregnancy: statement from the  
318 International Society for the Study of Hypertension in Pregnancy (ISSHP). *Hypertens*  
319 *Pregnancy* 20 (2001) IX-XIV.
- 320 [33] A.M. Molloy, and J.M. Scott, Microbiological Assay for Serum, Plasma, and Red Cell Folate  
321 Using, Cryopreserved, Microtiter Plate Method. *Methods in Enzymology* 281 (1997) 43-53.
- 322 [34] T. Tamura, L.E. Freeberg, and P.E. Cornwell, Inhibition of EDTA of growth of *Lactobacillus casei*  
323 in the folate microbiological assay and its reversal by added manganese or iron. *Clin Chem* 36  
324 (1990) 1993.
- 325 [35] I.L. Sargent, S.J. Germain, G.P. Sacks, S. Kumar, and C.W. Redman, Trophoblast deportation and  
326 the maternal inflammatory response in pre-eclampsia. *J Reprod Immunol* 59 (2003) 153-60.
- 327 [36] J. Rolschau, K. Kristoffersen, M. Ulrich, P. Grinsted, E. Schaumburg, and N. Foged, The influence  
328 of folic acid supplement on the outcome of pregnancies in the county of Funen in Denmark.  
329 Part I. *Eur J Obstet Gynecol Reprod Biol* 87 (1999) 105-10; discussion 103-4.
- 330 [37] Scientific Advisory Committee on Nutrition, Folate and Disease Prevention, HMSO for the Food  
331 Standards Agency and The Department of Health, Norwich, UK, 2006.

- 332 [38] R.W. Powers, R.W. Evans, A.K. Majors, J.I. Ojimba, R.B. Ness, W.R. Crombleholme, and J.M.  
333 Roberts, Plasma homocysteine concentration is increased in preeclampsia and is associated with  
334 evidence of endothelial activation. *Am J Obstet Gynecol* 179 (1998) 1605-11.
- 335 [39] R. Joshi, S. Adhikari, B.S. Patro, S. Chattopadhyay, and T. Mukherjee, Free radical scavenging  
336 behavior of folic acid: evidence for possible antioxidant activity. *Free Radic Biol Med* 30  
337 (2001) 1390-9.
- 338 [40] T. Green, and H.C. Ford, Human placental microvilli contain high-affinity binding sites for folate.  
339 *Biochem J* 218 (1984) 75-80.
- 340 [41] T. Mouratidou, F. Ford, F. Prountzou, and R. Fraser, Dietary assessment of a population of  
341 pregnant women in Sheffield, UK. *British Journal of Nutrition* 96 (2006) 929-935.
- 342 [42] I. Qvist, M. Abdulla, M. Jagerstad, and S. Svensson, Iron, zinc and folate status during pregnancy  
343 and two months after delivery. *Acta Obstet Gynecol Scand* 65 (1986) 15-22.
- 344 [43] H.W. Bruinse, and H. van den Berg, Changes of some vitamin levels during and after normal  
345 pregnancy. *Eur J Obstet Gynecol Reprod Biol* 61 (1995) 31-7.
- 346 [44] P.S. Shah, and A. Ohlsson, Effects of prenatal multimicronutrient supplementation on pregnancy  
347 outcomes: a meta-analysis. *CMAJ* 180 (2009) E99-108.
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351 **Table 1.** Maternal and fetal venous plasma homocysteine and folate and participant details  
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Parameter	Normotensive pregnant	Pre-eclampsia
	n = 24	n = 16
Maternal age (years) <sup>†</sup>	29 ± 6.6	31 ± 5.8
Booking body mass index (kg/m <sup>2</sup> ) <sup>†</sup>	25.9 ± 5.6	25.7 ± 3.8
Primipara n (%)	15 (62.5)	11 (68.8)
Max. systolic blood pressure (mm Hg) <sup>†</sup>	116 ± 4.3	157 ± 7.4 <sup>#</sup>
Max. diastolic blood pressure (mm Hg) <sup>†</sup>	76 ± 2.5	98 ± 4.9 <sup>#</sup>
Proteinuria (g/L) <sup>‡</sup>	-	1.0 [0.5, 1.8]
Gestation age at delivery, wks	39.8 ± 1.0	38.1 ± 2.0 <sup>#</sup>
Folate supplementation n (%)	4 (16.7)	2 (12.5)
Maternal homocysteine (μmol/L) <sup>‡</sup>	8.7 [ 6.8, 11.0]	8.2 [7.0, 10.6]
Fetal homocysteine (μmol/L) <sup>‡</sup>	10.5 [9.0, 12.4]*	9.5 [7.4, 11.9]
Maternal folate (nmol/L) <sup>‡</sup>	10.7 [6.2, 19.4]	6.8 [5.3, 14.1]
Fetal folate (nmol/L) <sup>‡</sup>	22.2 [17.8, 32.7]*	16.4 [11.3, 23.4]*

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354 <sup>#</sup> *P* < 0.05 between normotensive and pre-eclamptic pregnancies; \**P* < 0.05 between maternal and fetal  
 355 samples. <sup>†</sup>Values represented as means ± SD; <sup>‡</sup>values represented as median [IQR]. Further  
 356 demographic and pregnancy outcome details have been previously published[31].

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362 **Table 2.** Maternal plasma folate concentrations in women with different parities.

<b>Parity</b>	<b>Maternal Folate, nmol/L</b>
	<b>(median [IQR])</b>
0 (n = 24)	13.5 [6.9, 19.4]
1 (n = 11)	5.6 [ 4.5, 8.0]**
2 (n = 5)	6.0 [6.0, 6.8]*

363

364 \**P* = 0.037 between nulliparous and two previous pregnancies; \*\**P* = 0.005 between nulliparous and

365 one previous pregnancy.

366

367 **Figure Legends**

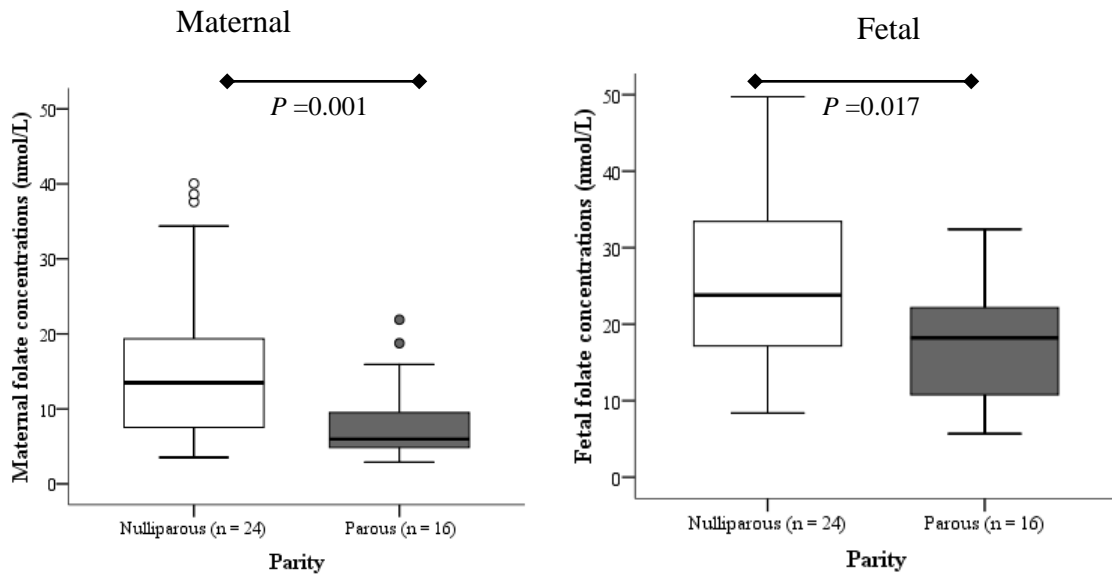
368 **Figure 1.** a) Maternal and fetal plasma folate concentrations by parity (nulliparous: white or parous:  
369 grey) in all pregnancies.

370 b) Maternal plasma folate concentrations by both parity (nulliparous: white or parous: grey) and the  
371 presence or absence of pre-eclampsia. There was a significant effect across parity:group for both  
372 maternal ( $P = 0.006$ ) and fetal ( $P = 0.015$ ) samples. Maternal and plasma folate concentrations were  
373 lowest in pre-eclamptic, parous women.

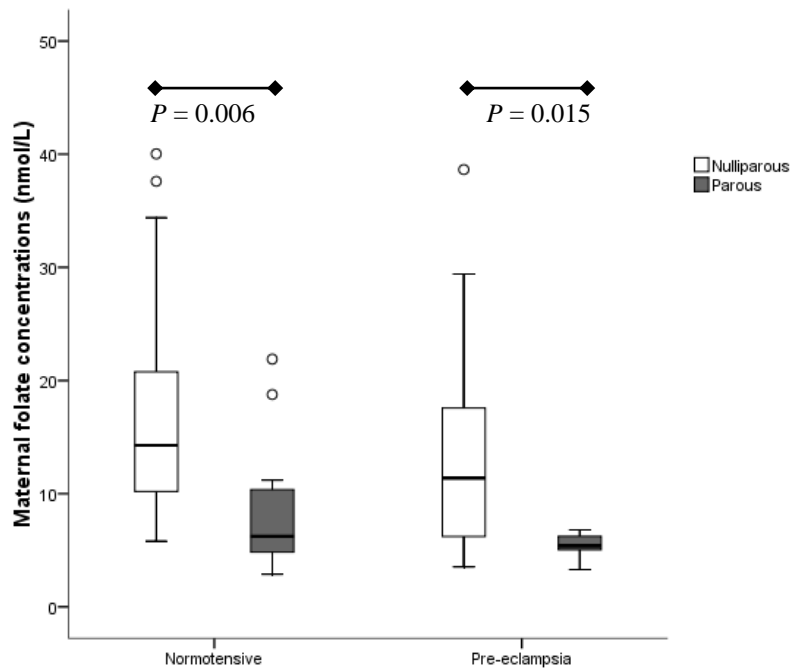
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375 **Figure 1**

376 **a)**



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379 **b)**  
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