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Methamphetamine: effects on the brain, gut and immune system

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ABSTRACT

Methamphetamine (METH) is a powerful central nervous system stimulant which elevates mood, alertness, energy levels and concentration in the short-term. However, chronic use and/or at higher doses METH use often results in psychosis, depression, delusions and violent behavior. METH was formerly used to treat conditions such as obesity and attention deficit hyperactivity disorder, but now is primarily used recreationally. Its addictive nature has led to METH abuse becoming a global problem. At a cellular level, METH exerts a myriad of effects on the central and peripheral nervous systems, immune system and the gastrointestinal system. Here we present how these effects might be linked and their potential contribution to the pathogenesis of neuropsychiatric disorders. In the long term, this pathway could be targeted therapeutically to protect people from the ill effects of METH use. This model of METH use may also provide insight into how gut, nervous and immune systems might break down in other conditions that may also benefit from therapeutic intervention.

Keywords: Anxiety Depression Ice Immune system Methamphetamine Nervous system Gastrointestinal system

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1. Introduction

Methamphetamine (METH; also called crystal, chalk or ice) is an addictive stimulant that can be administered orally, smoked, snorted or injected. Smoking or intravenous injection delivers METH to the brain rapidly, resulting in immediate and intense euphoria (1). METH use is associated with severe neurological and physical consequences (e.g. paranoia, violent behaviour, psychosis, anxiety and depression) and has become a serious public health problem worldwide (2, 3).

METH was discovered in Japan in 1919 and was commercially used in 1938 under the brand name Pervitin. It was especially popular for tired night-shift workers and was used during WWII by Germany to treat fatigue in tired army troops (4). METH became widely available from 1943 to treat a range of disorders including narcolepsy, depression, obesity, alcoholism and attention deficit hyperactivity disorder (ADHD). As METH decreased appetite it was also well marketed to women for weight loss. Although prolonged METH use can cause severe neurological damage, prescribed METH is still legally available under the brand name Desoxyn to treat severe obesity, narcolepsy and ADHD (5-7).

In recent years METH use has increased dramatically. In the USA, approximately 1.3 million people over the age of 12 have reported using METH. According to the 2011 United Nations survey, about 2.5 % of Australians have tried METH, which is 3-5 times higher than USA, Canada and UK (United Nations, 2011). In 2013, 7 % of Australians over the age of 14 years reported having used METH, with 50 % having used ice, the purest form of METH (8).

Immediate effects of acute or short-term METH use include increased alertness, heart rate, blood pressure, body temperature and a loss of appetite. Long-term, regular METH use can lead to severe tooth decay, infection, weight loss, malnutrition, kidney damage, liver damage, respiratory issues, paranoia, violent behaviour, psychosis, severe anxiety and depression. Even when individuals stop taking METH, the symptoms may persist for many years (9-14).

METH has more potent effects in women than men. In fact, 6-fold greater vulnerability to relapse of METH-seeking behavior is evident in experimental female rats as compared to male rats (15). Changes in brain morphology, such as hippocampus volume reduction, were seen in METH-abstinent females but not in males (16). In addition, females that are undergoing treatment for METH abuse have higher instances of psychological and physical trauma compared to males (17).

Herein, we review the findings on METH-related neurological and immunological effects, particularly neuro-immune cell stability, alteration of cytokine production, inflammation, immunosuppression, signal transduction and gene regulation.

2. METH and the Blood-Brain Barrier

METH increases blood brain barrier (BBB) permeability, inducing damage by altering the structure of proteins that are involved in BBB stability in mice (18). BBB permeability is also affected by body temperature, oxidative stress and inflammation, all of which are impacted by METH use (Fig. 1). Both hyperthermia and hypothermia alter BBB permeability, although hypothermia has less effect (19). Oxidative stress and excess inflammation is also associated with BBB damage in a number of neurodegenerative disorders (20-24). Recently, liquid chromatography-mass spectrometry (LC-MS/MS) analysis of extracts from rat brains following METH exposure identified changes in 18 proteins (11 from the hippocampus and 7 in the olfactory bulb); 13 of which were upregulated and 5 were downregulated. The modified proteins were predominantly involved in cell death, inflammation, oxidation and apoptotic pathways (25). In addition, alterations of endothelial cell structure and function, with increased levels of ROS, are observed in METH-related BBB disruptions (26, 27).

METH induces peripheral kidney and liver damage that leads to toxic ammonia levels in the blood and subsequently, the brain. Ammonia that is not cleared by the liver as normal accumulates and causes oxidative damage of endothelial cells, activation of matrix metalloproteases (MMPs) and neuro-inflammation via microglia and astrocyte activation, leading to BBB disruption (28-30) (Fig. 1). Furthermore, METH alters BBB permeability via dysregulation of tight junction proteins including occludin, claudin-5, and ZO family proteins (18, 26, 27, 31). Cytoskeletal rearrangement is also perturbed, with increased actin polymerization and expression of actin-binding protein Arp2/3 complex observed following METH administration (18). Interestingly, galectin-1, which is highly expressed in endothelial cells involved in BBB remodeling, alleviates the METH-induced increase in BBB permeability, thus acting as a neuroprotective molecule (32).

3. Neurological effects of METH

The euphoric effects of METH occur due to release of the neurotransmitter dopamine, which is involved in the experience of pleasure, motivation and motor function. However, long-term use of METH causes molecular changes in the dopamine system, contributing to nerve terminal damage in the brain and leading to impaired motor skills, rapid cognitive decline, increased anxiety, psychotic disorders, violent behaviour, hallucination, delusions and depression (Fig. 2) (33). These brain changes persist for many years after METH use has ceased (34).

Acute METH use causes an increase in neurotransmitter release, leading to potential damage to the terminal ends of neurons and ultimately alters brain function. A single high dose of METH causes neurotoxicity to dopamine and serotonin producing neurons in rodents (35). Positron

emission tomography (PET) and magnetic resonance spectroscopy (MRS) studies in abstinent METH users indicate a reduction of dopamine transporters (DAT) (36, 37) and serotonin transporters (SERT) (38, 39) that lasts up to 3 years after cessation of METH use. Brain tissues from rodents exposed to METH and post-mortem brain tissues isolated from chronic METH users demonstrate decreased levels of dopamine, serotonin, DAT and SERT in areas highly innervated by dopaminergic and serotoninergic axon terminals (40).

Gamma-aminobutyric acid (GABA) is a major inhibitory neurotransmitter in the brain. Disruption of inhibition via GABA receptors can lead to dopamine and serotonin dysfunction and promote depression, anxiety, stress and cognition. Similar reductions in neurotransmitters are also observed in a number of chronic neurological disorders such as Parkinson's and Alzheimer's disease (41-43).

Trace amine-associated receptor 1 (TAAR1) is a G-protein coupled receptor expressed on astrocytes, lymphocytes and neurons and negatively regulates neurotransmission via dopamine, norepinephrine and serotonin in the central nervous system (CNS) (44-46). It is an intracellular receptor predominantly found in the cytoplasm of presynaptic terminals and is poorly expressed on the cell membrane (45). Activated TAAR1 reduces dopamine receptor activity and increases cyclic adenosine monophosphate (cAMP), protein kinase A and protein kinase C activation. Subsequently, DAT is phosphorylated, leading to inhibition of dopamine transport (47, 48). TAAR1 signaling also activates transcription factor cAMP response element-binding protein (CREB) and nuclear factor of activated T-cell (NFAT), which are associated with immune cell activation and proliferation (49, 50). There are numerous studies that examine the effect of METH on TAAR1. METH directly activates TAAR1 in vitro and increases the intracellular cAMP levels in human HEK-393 fibroblasts (51). TAAR1 mRNA expression in resting T cells increases in response to METH administration (52). METH increases intracellular cAMP levels in human astrocytes whereas TAAR1 knockout cells have significantly reduced cAMP levels in response to METH administration (53). Interestingly, TAAR1 knockout mice show no significant difference in body weight, temperature, locomotor activity and other behaviours compared to wild-type mice; however increased firing rate of dopaminergic and serotoninergic neurons are noted (54-56). Conversely, TAAR1 transgenic mice show increased sensitivity to METH. RO5203648, a selective TAAR1 agonist, alleviates METH-induced neurochemical effects in rats, including hyperactivity, psychomotor effects and addiction (57-59).

4. Sympathetic and parasympathetic regulation of the immune system

Sympathetic and parasympathetic nervous systems play an important role in regulating the immune system. The sympathetic nervous system is involved in stress-induced remodelling of lymph

node innervation; increased norepinephrine and epinephrine levels inhibit immune cell functions and promote intestinal inflammation (60-62). The parasympathetic nervous system has an anti-inflammatory role via activation of the cholinergic anti-inflammatory pathway (63, 64). Acetylcholine decreases the production of pro-inflammatory cytokines such as TNF- α by human macrophages through nicotinic receptors (65). Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxins and intestinal inflammation (63, 66). The vagus nerve also indirectly modulates immune activity of the spleen through connections with the splenic sympathetic nerve (67, 68). However, the effects of METH on the activity of sympathetic and vagus nerves and their modulation of systemic and local immune responses have not been studied.

5. The effects of METH on the Gut-Brain axis

The rapid and sustained release of norepinephrine following METH use results in arterial vasoconstriction, leading to tachycardia and hypertension. Similar effects can also be seen in the mesenteric vessels of the gut, leading to acute intestinal ischemia (69, 70). In METH users, the most common effects of gastrointestinal (GI) vasoconstriction and bowel ischemia include abdominal or stomach cramping, severe constipation and/or diarrhoea and tissue dehydration. In some cases, loss of blood flow to GI muscles leads to severe, potentially fatal conditions such as paralytic ileus (Fig. 2) (71). Potential consequences of paralytic ileus include severe infection, tissue death (gangrene), perforation of the intestinal wall and serious disruptions in the levels of electrolytes. In severe cases, bowel infarction can lead to development of septic shock with multiple organ failure (70).

Bowel ischemia is associated with increased intestinal permeability, oxidative and nitrosative stress. Several findings suggest that dysfunction of the intestinal mucosal barrier leading to increased intestinal permeability plays an important role in the pathophysiology of anxiety, stress, depression, cognitive decline, chronic fatigue and eating and sleep disorders. All of these are common in METH users (Fig 1, 2).

Disruption of the gut wall integrity, damage to intestinal epithelial cells and derangement of tight junctions leads to the leakage of macromolecules, microbial products, and microbiota from the intestinal lumen into the circulation, mesenteric lymph nodes, spleen and liver (72). With the concurrent increase in BBB permeability following METH use, these gut-derived components have the ability to enter the brain (73). Extensive release of dopamine and norepinephrine stimulates growth of bacteria which may also influence neural activity in stress responsive brain areas (74). Therefore the intestinal microbiota may act as a mediator in the communication between the gut and the brain (75). However, the mechanisms underlying METH-induced increases in intestinal

permeability and damage to the GI tract leading to systemic immune response and neuropsychiatric disorders are not clear.

Increases in intestinal permeability may be due to the inhibition of GI motility observed in METH users (71). In the gut, dopamine and norepinephrine act on receptors of the enteric nervous system resulting in decreased bowel contractility, intestinal smooth muscle tone and alteration of the migratory motor complex (76-78). METH-mediated release of neurotransmitters might also lead to generation of oxidative stress molecules, including ROS and reactive nitrogen species (RNS) which can cause damage and death of enteric neurons and subsequent GI dysfunction.

Recent advances in research have described the importance of gut microbiota in many neuropsychiatric conditions including autism, anxiety, depression, eating and sleep disorders. Current evidence suggests that multiple mechanisms, including immune, endocrine and neuroendocrine pathways, may be involved in gut microbiota-to-brain signalling and that the brain can in turn alter microbial composition and behaviour via the enteric nervous system (79-81). However, changes in the gut microbiota after METH use and the interplay between intestinal microbiota, immune response and neuropsychiatric manifestations associated with METH use have not been studied (Fig 2)

6. METH and its effects on the Immune System

The human immune system has a profound influence on the brain. Increasing evidence shows that there are numerous interactions between the nervous and immune systems (82). The immune system also plays an important role in the pathogenesis of neuropsychiatric disorders including cognitive decline, anxiety, mood changes and depressive states as well as increased attention, decreased fatigue and euphoria rush (80, 83, 84), which are associated with METH use (Fig. 3).

6.1. The effects of METH on Susceptibility to Infection

Chronic METH use and lack of hygiene leads to alteration in primary physical barriers and increases the occurrence of skin infections (85). METH use also increases the risk of chronic infections such as methicillin-resistant *Staphylococcus aureus* (MRSA), human immunodeficiency virus (HIV), hepatitis and sexually transmitted diseases (3).

In the presence of METH, the number of macrophages, NK, DC, monocytes and granulocytes are reduced, further contributing to the increased susceptibility to infections (86, 87). High METH dose induces apoptotic death in rat thymic and splenic lymphocytes and produces severe immunosuppression, which could also contribute to the higher rate of infections observed in chronic

METH users (86, 88). METH also changes the cytokine response to retroviral infection in rodents (89, 90).

6.2. The effects of METH on Inflammation and inflammatory markers

Pro-inflammatory cytokines (IL-1, IL-6, IL-8, TNF- α) have been implicated in damaging and destroying existing neurons leading to the neurobiological manifestations of different mental states. Indeed, METH use results in IL-6 and IL-8 production by neuronal cells, leading to myelin degeneration in mice (3). Similarly, mice treated with METH show increased expression of pro-inflammatory cytokines (IL-1 β) for up to 3 weeks in brain regions (91). METH-related cell activation is seen in astrocytes and leads to excessive secretion of inflammatory cytokines such as IL-6 and IL-8 inducing inflammation (86, 92), as well as enhancing expression of chemokines and chemokine receptors such as CXCR4 and CCR5 in the brain (93). In addition, METH induces a pro-inflammatory profile of *in vitro* cultured macrophages, by upregulating TNF- α , IL-8, CXCL16, CXCL1 and downregulating CCL7 (disrupting toll-like receptor 9 (TLR-9) signaling pathway) (94). Suppression of TLR-9 indicates that suppressed innate immune responses may ensue in METH users.

6.3. METH-associated Immune cell changes

Immune cell mediated neuro-inflammation and neurodegeneration is induced by METH use. METH is a weak base and alkalizes the acidic organelles within macrophages, leading to impaired phagocytosis, antigen processing and presentation (95). As a consequence, this can lead to a reduction of pathogen uptake and processing, increasing infections. Immunological factors such as cytokines, chemokines and adhesion molecules are linked with neuronal degeneration as well as neuropsychiatric complications (96, 97).

METH modifies a number of immune cell (natural killer (NK) cells, dendritic cells (DC), monocytes, macrophages and granulocytes) activities, leading to immunosuppression (86). In addition, METH affects antigen presenting cells (APCs) in the brain (microglia and astrocytes) and leads to increased secretion of pro-inflammatory cytokines (IL-1, IL-6, IL-8), interferons and TNF- α (98). Murine models show that METH modifies thymic and splenic cellularity, in turn altering peripheral T lymphocyte populations (97). Furthermore, METH suppresses adaptive immunity by altering T cell populations, specifically the CD4+/CD8+ T-cell ratio (97, 99).

Microglia and astrocytes usually perform compensatory actions during brain injury and protect the brain as excess neuro-inflammation leads to damage. However, METH activates G-protein receptors and initiates signalling of the Akt/NF-kB pathway to increase cell proliferation and cytokine secretion (IL-6 and IL-8) in astrocytes. The effect of METH-related IL-6 and IL-8 expression in

astrocytes is reduced in the presence of 2-methyl-6-(phenylethynyl)-pyridine (MPEP), an antagonist of metabotropic glutamate receptor 5 (mGlu5) (100). Furthermore, HIV-1 envelope protein gp120 acts synergistically with METH to further increase IL-6 expression (101).

METH activates the Sigma-1 receptor which in turn activates NF-κB via SRC/ERK, thus increasing high mobility group box-1 (HMGB1) gene expression in astrocytes (102). This promotes cell proliferation and migration. HMGB1 also regulates gene transcription and acts an inflammatory mediator by activating TLR-4 (103). Activated immune cells such as macrophages and monocytes also increase the expression of HMGB1 during inflammation (104, 105).

METH also increases glutamate release from a number of brain regions such as the striatum, cerebral cortex and hippocampus (106, 107). The phosphorylation of PI3/Akt molecules via glutamate receptor engagement leads to the activation of transcription factor NF-κB and ultimately facilitates inflammation, neurotoxicity and apoptosis (108). Chronic METH exposure affects monoaminergic neurons by the loss of DAT, SERT and vesicular monoamine transporter type-2 (VMAT-2) in striatum and central gray matter of rat brain (109).

6.4. The effects of METH on the expression of death receptor PD-1 and its ligand PD-L1

Programmed cell death-1 ligand (PD-L1), is a transmembrane protein that plays a major role in suppressing the immune system. T cells express the receptor PD-1 and upon interaction with PD-L1 inhibitory signals are triggered resulting in T cell apoptosis. Cancer cells that express high levels of PD-L1 as a mechanism for immune evasion are associated with poor prognosis in patients (110-112). In inflammatory disorders the expression of PD-L1 is reduced leading to activation of T cells. Thus, PD-L1 is important in regulating immune responses. The level of PD-L1 and PD-1 expression in human brain cells under normal physiological conditions is low. However, activated neuro-immune cells such as astrocytes, microglia, T cells, B cells, macrophages, DC and non-immune cells (endothelial and epithelial cells) appear to have increased in expression (113).

The expression of PD-L1 is elevated in brain endothelial cells, on macrophages and microglia, following METH exposure (113). PD-1 signaling attenuates phosphorylation of protein kinase C (PKC), necessary for the activation of NF-kB and for production of IL-2 (114), thus METH exposure inhibits T cell activation. Overexpression of PD-1 and PD-L1 following METH exposure in macrophages may also suppress immunity by altering antigen presentation (113).

Conversely, reductions in PD-1 and PD-L1 expression are noted in astrocytes following METH exposure (113). Inhibition of PD-1/PD-L1 expression in astrocytes stimulates overproduction of inflammatory cytokines such as interleukins and leads to inflammation and neuronal damage. However, whether METH-related reduction of PD-1/PD-L1 expression in astrocytes alters PKC

activation or regulates the production of NF κ B and proinflammatory mediators such as IL-1, IL-6, IL-8 and TNF- α to cause the neuronal damage by inflammation are not clear and therefore warrants further investigation.

7. Conclusion and future prospects

Recent advances in research have described the importance of gut microbiota in many neuropsychiatric conditions including autism, anxiety, depression, eating and sleep disorders. Current evidence suggests that multiple mechanisms, including immune, endocrine and neurocrine pathways, may be involved in gut-to-brain signalling and that the brain can in turn alter microbial composition and behaviour of the gut via the enteric nervous system (79-81). Though it is apparent that METH use alters numerous aspects of the nervous, immune and gastrointestinal systems, changes in gut microbiota following METH use and the interplay between intestinal microbiota, immune response and neuropsychiatric manifestations associated with METH use are yet to be studied.

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References

- 1. Hauer P. Systemic affects of methamphetamine use. S D Med. 2010;63(8):285-287.
- 2. Rommel N, Rohleder NH, Wagenpfeil S, Haertel-Petri R, Kesting MR. Evaluation of methamphetamine-associated socioeconomic status and addictive behaviors, and their impact on oral health. Addict Behav. 2015;50:182-187.
- 3. Salamanca SA, Sorrentino EE, Nosanchuk JD, Martinez LR. Impact of methamphetamine on infection and immunity. Front Neurosci. 2014;8:445.
- 4. Defalque RJ, Wright AJ. Methamphetamine for Hitler's Germany: 1937 to 1945. Bulletin of anesthesia history. 2011;29(2):21-24, 32.
- 5. Cartier J, Farabee D, Prendergast ML. Methamphetamine use, self-reported violent crime, and recidivism among offenders in California who abuse substances. J Interpers Violence. 2006;21(4):435-445.
- 6. McGuinness T. Methamphetamine abuse. Am J Nurs. 2006;106(12):54-59.
- 7. Rutkowski BA. California Substance Abuse Research Consortium, September 2005: update on recent methamphetamine trends in four California regions. J Psychoactive Drugs. 2006;Suppl 3:369-375.
- 8. Roche A, McEntee A, Fischer J, Kostadinov V. Methamphetamine use in Australia. In.: National Centre for Education and Training on Addiction (NCETA), Flinders University; 2015.
- 9. Kirkpatrick MG, Haney M, Vosburg SK, Comer SD, Foltin RW, Hart CL. Methamphetamine self-administration by humans subjected to abrupt shift and sleep schedule changes. Psychopharmacology (Berl). 2009;203(4):771-780.

- 10. Mendelson J, Rawson R, Newton T, Galloway G, de Wit H, Dewey SL, Hart CL, Epstein DH. Treatment of methamphetamine dependence. Mayo Clin Proc. 2008;83(3):369-370; author reply 370-361.
- 11. Perez AY, Kirkpatrick MG, Gunderson EW, Marrone G, Silver R, Foltin RW, Hart CL. Residual effects of intranasal methamphetamine on sleep, mood, and performance. Drug Alcohol Depend. 2008;94(1-3):258-262.
- 12. Padilla R, Ritter AV. Meth mouth: methamphetamine and oral health. J Esthet Restor Dent. 2008;20(2):148-149.
- 13. Shaner JW, Kimmes N, Saini T, Edwards P. "Meth mouth": rampant caries in methamphetamine abusers. AIDS Patient Care STDS. 2006;20(3):146-150.
- 14. Williams N, Covington JS, 3rd. Methamphetamine and meth mouth: an overview. J Tenn Dent Assoc. 2006;86(4):32-35.
- 15. Ruda-Kucerova J, Amchova P, Babinska Z, Dusek L, Micale V, Sulcova A. Sex Differences in the Reinstatement of Methamphetamine Seeking after Forced Abstinence in Sprague-Dawley Rats. Front Psychiatry. 2015;6:91.
- 16. Du J, Quan M, Zhuang W, Zhong N, Jiang H, Kennedy DN, Harrington A, Ziedonis D, Fan X, Zhao M. Hippocampal volume reduction in female but not male recent abstinent methamphetamine users. Behav Brain Res. 2015;289:78-83.
- 17. Hser YI, Evans E, Huang YC. Treatment outcomes among women and men methamphetamine abusers in California. J Subst Abuse Treat. 2005;28(1):77-85.
- 18. Park M, Kim HJ, Lim B, Wylegala A, Toborek M. Methamphetamine-induced occludin endocytosis is mediated by the Arp2/3 complex-regulated actin rearrangement. J Biol Chem. 2013;288(46):33324-33334.
- 19. Kiyatkin EA, Sharma HS. Permeability of the blood-brain barrier depends on brain temperature. Neuroscience. 2009;161(3):926-939.
- 20. Ji X, Liu W, Xie K, Liu W, Qu Y, Chao X, Chen T, Zhou J, Fei Z. Beneficial effects of hydrogen gas in a rat model of traumatic brain injury via reducing oxidative stress. Brain Res. 2010;1354:196-205.
- 21. Price TO, Ercal N, Nakaoke R, Banks WA. HIV-1 viral proteins gp120 and Tat induce oxidative stress in brain endothelial cells. Brain Res. 2005;1045(1-2):57-63.
- 22. Sharma HS, Sharma A. Nanoparticles aggravate heat stress induced cognitive deficits, blood-brain barrier disruption, edema formation and brain pathology. Prog Brain Res. 2007;162:245-273.
- 23. Zehendner CM, Librizzi L, Hedrich J, Bauer NM, Angamo EA, de Curtis M, Luhmann HJ. Moderate hypoxia followed by reoxygenation results in blood-brain barrier breakdown via oxidative stress-dependent tight-junction protein disruption. PLoS One. 2013;8(12):e82823.
- 24. Kermode AG, Thompson AJ, Tofts P, MacManus DG, Kendall BE, Kingsley DP, Moseley IF, Rudge P, McDonald WI. Breakdown of the blood-brain barrier precedes symptoms and other MRI signs of new lesions in multiple sclerosis. Pathogenetic and clinical implications. Brain. 1990;113 (Pt 5):1477-1489.
- 25. Zhu R, Yang T, Kobeissy F, Mouhieddine TH, Raad M, Nokkari A, Gold MS, Wang KK, Mechref Y. The Effect of Chronic Methamphetamine Exposure on the Hippocampal and Olfactory Bulb Neuroproteomes of Rats. PloS one. 2016;11(4):e0151034.
- 26. Mahajan SD, Aalinkeel R, Sykes DE, Reynolds JL, Bindukumar B, Adal A, Qi M, Toh J, Xu G, Prasad PN, Schwartz SA. Methamphetamine alters blood brain barrier permeability via the modulation of tight junction expression: Implication for HIV-1 neuropathogenesis in the context of drug abuse. Brain Res. 2008;1203:133-148.
- 27. Ramirez SH, Potula R, Fan S, Eidem T, Papugani A, Reichenbach N, Dykstra H, Weksler BB, Romero IA, Couraud PO, Persidsky Y. Methamphetamine disrupts blood-brain barrier function by induction of oxidative stress in brain endothelial cells. J Cereb Blood Flow Metab. 2009;29(12):1933-1945.

- 28. Bemeur C, Desjardins P, Butterworth RF. Evidence for oxidative/nitrosative stress in the pathogenesis of hepatic encephalopathy. Metab Brain Dis. 2010;25(1):3-9.
- 29. Rodrigo R, Erceg S, Felipo V. Neurons exposed to ammonia reproduce the differential alteration in nitric oxide modulation of guanylate cyclase in the cerebellum and cortex of patients with liver cirrhosis. Neurobiol Dis. 2005;19(1-2):150-161.
- 30. Northrop NA, Halpin LE, Yamamoto BK. Peripheral ammonia and blood brain barrier structure and function after methamphetamine. Neuropharmacology. 2016;107:18-26.
- 31. Martins T, Baptista S, Goncalves J, Leal E, Milhazes N, Borges F, Ribeiro CF, Quintela O, Lendoiro E, Lopez-Rivadulla M, Ambrosio AF, Silva AP. Methamphetamine transiently increases the blood-brain barrier permeability in the hippocampus: role of tight junction proteins and matrix metalloproteinase-9. Brain Res. 2011;1411:28-40.
- 32. Parikh NU, Aalinkeel R, Reynolds JL, Nair BB, Sykes DE, Mammen MJ, Schwartz SA, Mahajan SD. Galectin-1 suppresses methamphetamine induced neuroinflammation in human brain microvascular endothelial cells: Neuroprotective role in maintaining blood brain barrier integrity. Brain Res. 2015;1624:175-187.
- 33. Rusyniak DE. Neurologic manifestations of chronic methamphetamine abuse. Neurologic clinics. 2011;29(3):641-655.
- 34. NIH. DrugFacts: Methamphetamine. NIH. 2016
- 35. Metzger RR, Haughey HM, Wilkins DG, Gibb JW, Hanson GR, Fleckenstein AE. Methamphetamine-induced rapid decrease in dopamine transporter function: role of dopamine and hyperthermia. The Journal of pharmacology and experimental therapeutics. 2000;295(3):1077-1085.
- 36. Hong SJ, Zhang D, Zhang LH, Yang P, Wan J, Yu Y, Wang TH, Feng ZT, Li LH, Yew DT. Expression of dopamine transporter in the different cerebral regions of methamphetamine-dependent rats. Hum Exp Toxicol. 2015;34(7):707-717.
- 37. Yuan J, Lv R, Robert Brasic J, Han M, Liu X, Wang Y, Zhang G, Liu C, Li Y, Deng Y. Dopamine transporter dysfunction in Han Chinese people with chronic methamphetamine dependence after a short-term abstinence. Psychiatry Res. 2014;221(1):92-96.
- 38. Haughey HM, Fleckenstein AE, Metzger RR, Hanson GR. The effects of methamphetamine on serotonin transporter activity: role of dopamine and hyperthermia. J Neurochem. 2000;75(4):1608-1617.
- 39. Sogawa C, Sogawa N, Tagawa J, Fujino A, Ohyama K, Asanuma M, Funada M, Kitayama S. 5-Methoxy-N,N-diisopropyltryptamine (Foxy), a selective and high affinity inhibitor of serotonin transporter. Toxicol Lett. 2007;170(1):75-82.
- 40. Cass WA, Manning MW. Recovery of presynaptic dopaminergic functioning in rats treated with neurotoxic doses of methamphetamine. J Neurosci. 1999;19(17):7653-7660.
- 41. Allard PO, Rinne J, Marcusson JO. Dopamine uptake sites in Parkinson's disease and in dementia of the Alzheimer type. Brain Res. 1994;637(1-2):262-266.
- 42. Morgan DG, May PC, Finch CE. Dopamine and serotonin systems in human and rodent brain: effects of age and neurodegenerative disease. J Am Geriatr Soc. 1987;35(4):334-345.
- 43. Murray AM, Weihmueller FB, Marshall JF, Hurtig HI, Gottleib GL, Joyce JN. Damage to dopamine systems differs between Parkinson's disease and Alzheimer's disease with parkinsonism. Ann Neurol. 1995;37(3):300-312.
- 44. Lam VM, Espinoza S, Gerasimov AS, Gainetdinov RR, Salahpour A. In-vivo pharmacology of Trace-Amine Associated Receptor 1. Eur J Pharmacol. 2015;763(Pt B):136-142.
- 45. Miller GM. The emerging role of trace amine-associated receptor 1 in the functional regulation of monoamine transporters and dopaminergic activity. J Neurochem. 2011;116(2):164-176.
- 46. Borowsky B, Adham N, Jones KA, Raddatz R, Artymyshyn R, Ogozalek KL, Durkin MM, Lakhlani PP, Bonini JA, Pathirana S, Boyle N, Pu X, Kouranova E, Lichtblau H, Ochoa FY,

- Branchek TA, Gerald C. Trace amines: identification of a family of mammalian G protein-coupled receptors. Proc Natl Acad Sci U S A. 2001;98(16):8966-8971.
- 47. Miller GM. Avenues for the development of therapeutics that target trace amine associated receptor 1 (TAAR1). J Med Chem. 2012;55(5):1809-1814.
- 48. Maguire JJ, Parker WA, Foord SM, Bonner TI, Neubig RR, Davenport AP. International Union of Pharmacology. LXXII. Recommendations for trace amine receptor nomenclature. Pharmacol Rev. 2009;61(1):1-8.
- 49. Panas HN, Lynch LJ, Vallender EJ, Xie Z, Chen GL, Lynn SK, Scanlan TS, Miller GM. Normal thermoregulatory responses to 3-iodothyronamine, trace amines and amphetamine-like psychostimulants in trace amine associated receptor 1 knockout mice. J Neurosci Res. 2010;88(9):1962-1969.
- 50. Panas MW, Xie Z, Panas HN, Hoener MC, Vallender EJ, Miller GM. Trace amine associated receptor 1 signaling in activated lymphocytes. J Neuroimmune Pharmacol. 2012;7(4):866-876.
- 51. Reese EA, Bunzow JR, Arttamangkul S, Sonders MS, Grandy DK. Trace amine-associated receptor 1 displays species-dependent stereoselectivity for isomers of methamphetamine, amphetamine, and para-hydroxyamphetamine. J Pharmacol Exp Ther. 2007;321(1):178-186.
- 52. Sriram U, Haldar B, Cenna JM, Gofman L, Potula R. Methamphetamine mediates immune dysregulation in a murine model of chronic viral infection. Front Microbiol. 2015;6:793.
- 53. Cisneros IE, Ghorpade A. Methamphetamine and HIV-1-induced neurotoxicity: role of trace amine associated receptor 1 cAMP signaling in astrocytes. Neuropharmacology. 2014;85:499-507.
- 54. Bradaia A, Trube G, Stalder H, Norcross RD, Ozmen L, Wettstein JG, Pinard A, Buchy D, Gassmann M, Hoener MC, Bettler B. The selective antagonist EPPTB reveals TAAR1-mediated regulatory mechanisms in dopaminergic neurons of the mesolimbic system. Proc Natl Acad Sci U S A. 2009;106(47):20081-20086.
- 55. Leo D, Mus L, Espinoza S, Hoener MC, Sotnikova TD, Gainetdinov RR. Taar1-mediated modulation of presynaptic dopaminergic neurotransmission: role of D2 dopamine autoreceptors. Neuropharmacology. 2014;81:283-291.
- 56. Revel FG, Moreau JL, Gainetdinov RR, Bradaia A, Sotnikova TD, Mory R, Durkin S, Zbinden KG, Norcross R, Meyer CA, Metzler V, Chaboz S, Ozmen L, Trube G, Pouzet B, Bettler B, Caron MG, Wettstein JG, Hoener MC. TAAR1 activation modulates monoaminergic neurotransmission, preventing hyperdopaminergic and hypoglutamatergic activity. Proc Natl Acad Sci U S A. 2011;108(20):8485-8490.
- 57. Jing L, Li JX. Trace amine-associated receptor 1: A promising target for the treatment of psychostimulant addiction. Eur J Pharmacol. 2015;761:345-352.
- 58. Jing L, Zhang Y, Li JX. Effects of the trace amine associated receptor 1 agonist RO5263397 on abuse-related behavioral indices of methamphetamine in rats. Int J Neuropsychopharmacol. 2015;18(4).
- 59. Cotter R, Pei Y, Mus L, Harmeier A, Gainetdinov RR, Hoener MC, Canales JJ. The trace amine-associated receptor 1 modulates methamphetamine's neurochemical and behavioral effects. Front Neurosci. 2015;9:39.
- 60. Sloan EK, Capitanio JP, Cole SW. Stress-induced remodeling of lymphoid innervation. Brain Behav Immun. 2008;22(1):15-21.
- 61. Straub RH, Wiest R, Strauch UG, Harle P, Scholmerich J. The role of the sympathetic nervous system in intestinal inflammation. Gut. 2006;55(11):1640-1649.
- 62. Johnson JD, Campisi J, Sharkey CM, Kennedy SL, Nickerson M, Greenwood BN, Fleshner M. Catecholamines mediate stress-induced increases in peripheral and central inflammatory cytokines. Neuroscience. 2005;135(4):1295-1307.

- 63. Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, Wang H, Abumrad N, Eaton JW, Tracey KJ. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. Nature. 2000;405(6785):458-462.
- 64. Martelli D, McKinley MJ, McAllen RM. The cholinergic anti-inflammatory pathway: a critical review. Auton Neurosci. 2014;182:65-69.
- 65. Pavlov OV, Sel'kov SA, Lalayan DV, Arzhanova ON. Secretion of proinflammatory cytokines by villous chorion tissue in spontaneous abortion. Bulletin of experimental biology and medicine. 2003;135(4):377-379.
- 66. Bonaz BL, Bernstein CN. Brain-gut interactions in inflammatory bowel disease. Gastroenterology. 2013;144(1):36-49.
- 67. Matteoli G, Boeckxstaens GE. The vagal innervation of the gut and immune homeostasis. Gut. 2013;62(8):1214-1222.
- 68. Matteoli G, Gomez-Pinilla PJ, Nemethova A, Di Giovangiulio M, Cailotto C, van Bree SH, Michel K, Tracey KJ, Schemann M, Boesmans W, Vanden Berghe P, Boeckxstaens GE. A distinct vagal anti-inflammatory pathway modulates intestinal muscularis resident macrophages independent of the spleen. Gut. 2014;63(6):938-948.
- 69. Herr RD, Caravati EM. Acute transient ischemic colitis after oral methamphetamine ingestion. Am J Emerg Med. 1991;9(4):406-409.
- 70. Brannan TA, Soundararajan S, Houghton BL. Methamphetamine-associated shock with intestinal infarction. MedGenMed. 2004;6(4):6.
- 71. Carlson TL, Plackett TP, Gagliano RA, Jr., Smith RR. Methamphetamine-induced paralytic ileus. Hawaii J Med Public Health. 2012;71(2):44-45.
- 72. Suzuki T. Regulation of intestinal epithelial permeability by tight junctions. Cell Mol Life Sci. 2013;70(4):631-659.
- 73. Northrop NA, Yamamoto BK. Methamphetamine effects on blood-brain barrier structure and function. Front Neurosci. 2015;9:69.
- 74. Lyte M, Vulchanova L, Brown DR. Stress at the intestinal surface: catecholamines and mucosa-bacteria interactions. Cell Tissue Res. 2011;343(1):23-32.
- 75. Bercik P, Denou E, Collins J, Jackson W, Lu J, Jury J, Deng Y, Blennerhassett P, Macri J, McCoy KD, Verdu EF, Collins SM. The intestinal microbiota affect central levels of brain-derived neurotropic factor and behavior in mice. Gastroenterology. 2011;141(2):599-609, 609.e591-593.
- 76. Dive A, Foret F, Jamart J, Bulpa P, Installe E. Effect of dopamine on gastrointestinal motility during critical illness. Intensive Care Med. 2000;26(7):901-907.
- 77. Dunser MW, Hasibeder WR. Sympathetic overstimulation during critical illness: adverse effects of adrenergic stress. J Intensive Care Med. 2009;24(5):293-316.
- 78. Zizzo MG, Mule F, Mastropaolo M, Serio R. D1 receptors play a major role in the dopamine modulation of mouse ileum contractility. Pharmacol Res. 2010;61(5):371-378.
- 79. Carabotti M, Scirocco A, Maselli MA, Severi C. The gut-brain axis: interactions between enteric microbiota, central and enteric nervous systems. Annals of gastroenterology: quarterly publication of the Hellenic Society of Gastroenterology. 2015;28(2):203-209.
- 80. Fetissov SO, Dechelotte P. The new link between gut-brain axis and neuropsychiatric disorders. Current opinion in clinical nutrition and metabolic care. 2011;14(5):477-482.
- 81. Mayer EA, Tillisch K, Gupta A. Gut/brain axis and the microbiota. The Journal of clinical investigation. 2015;125(3):926-938.
- 82. Louveau A, Smirnov I, Keyes TJ, Eccles JD, Rouhani SJ, Peske JD, Derecki NC, Castle D, Mandell JW, Lee KS, Harris TH, Kipnis J. Structural and functional features of central nervous system lymphatic vessels. Nature. 2015;523(7560):337-341.
- 83. Petra AI, Panagiotidou S, Hatziagelaki E, Stewart JM, Conti P, Theoharides TC. Gut-Microbiota-Brain Axis and Its Effect on Neuropsychiatric Disorders With Suspected Immune Dysregulation. Clinical therapeutics. 2015;37(5):984-995.

- 84. Kerr D, Krishnan C, Pucak ML, Carmen J. The immune system and neuropsychiatric diseases. International review of psychiatry (Abingdon, England). 2005;17(6):443-449.
- 85. Rusyniak DE. Neurologic manifestations of chronic methamphetamine abuse. Psychiatr Clin North Am. 2013;36(2):261-275.
- 86. Harms R, Morsey B, Boyer CW, Fox HS, Sarvetnick N. Methamphetamine administration targets multiple immune subsets and induces phenotypic alterations suggestive of immunosuppression. PLoS One. 2012;7(12):e49897.
- 87. Saito M, Yamaguchi T, Kawata T, Ito H, Kanai T, Terada M, Yokosuka M, Saito TR. Effects of methamphetamine on cortisone concentration, NK cell activity and mitogen response of T-lymphocytes in female cynomolgus monkeys. Exp Anim. 2006;55(5):477-481.
- 88. Peerzada H, Gandhi JA, Guimaraes AJ, Nosanchuk JD, Martinez LR. Methamphetamine administration modifies leukocyte proliferation and cytokine production in murine tissues. Immunobiology. 2013;218(8):1063-1068.
- 89. Liang H, Wang X, Chen H, Song L, Ye L, Wang SH, Wang YJ, Zhou L, Ho WZ. Methamphetamine enhances HIV infection of macrophages. Am J Pathol. 2008;172(6):1617-1624.
- 90. Yu Q, Zhang D, Walston M, Zhang J, Liu Y, Watson RR. Chronic methamphetamine exposure alters immune function in normal and retrovirus-infected mice. Int Immunopharmacol. 2002;2(7):951-962.
- 91. Loftis JM, Janowsky A. Neuroimmune basis of methamphetamine toxicity. Int Rev Neurobiol. 2014;118:165-197.
- 92. Shah A, Silverstein PS, Singh DP, Kumar A. Involvement of metabotropic glutamate receptor 5, AKT/PI3K signaling and NF-kappaB pathway in methamphetamine-mediated increase in IL-6 and IL-8 expression in astrocytes. J Neuroinflammation. 2012;9:52.
- 93. Najera JA, Bustamante EA, Bortell N, Morsey B, Fox HS, Ravasi T, Marcondes MC. Methamphetamine abuse affects gene expression in brain-derived microglia of SIV-infected macaques to enhance inflammation and promote virus targets. BMC immunology. 2016;17(1):7.
- 94. Burns A, Ciborowski P. Acute exposure to methamphetamine alters TLR9-mediated cytokine expression in human macrophage. Immunobiology. 2016;221(2):199-207.
- 95. Talloczy Z, Martinez J, Joset D, Ray Y, Gacser A, Toussi S, Mizushima N, Nosanchuk JD, Goldstein H, Loike J, Sulzer D, Santambrogio L. Methamphetamine inhibits antigen processing, presentation, and phagocytosis. PLoS Pathog. 2008;4(2):e28.
- 96. House RV, Thomas PT, Bhargava HN. Comparison of immune functional parameters following in vitro exposure to natural and synthetic amphetamines. Immunopharmacol Immunotoxicol. 1994;16(1):1-21.
- 97. In SW, Son EW, Rhee DK, Pyo S. Methamphetamine administration produces immunomodulation in mice. J Toxicol Environ Health A. 2005;68(23-24):2133-2145.
- 98. Whitney NP, Eidem TM, Peng H, Huang Y, Zheng JC. Inflammation mediates varying effects in neurogenesis: relevance to the pathogenesis of brain injury and neurodegenerative disorders. J Neurochem. 2009;108(6):1343-1359.
- 99. Mahajan SD, Hu Z, Reynolds JL, Aalinkeel R, Schwartz SA, Nair MP. Methamphetamine modulates gene expression patterns in monocyte derived mature dendritic cells: implications for HIV-1 pathogenesis. Mol Diagn Ther. 2006;10(4):257-269.
- 100. Golembiowska K, Konieczny J, Wolfarth S, Ossowska K. Neuroprotective action of MPEP, a selective mGluR5 antagonist, in methamphetamine-induced dopaminergic neurotoxicity is associated with a decrease in dopamine outflow and inhibition of hyperthermia in rats. Neuropharmacology. 2003;45(4):484-492.
- 101. Shah A, Silverstein PS, Kumar S, Singh DP, Kumar A. Synergistic Cooperation between Methamphetamine and HIV-1 gsp120 through the P13K/Akt Pathway Induces IL-6 but not IL-8 Expression in Astrocytes. PLoS ONE. 2012;7(12):e52060.

- 102. Zhang Y, Lv X, Bai Y, Zhu X, Wu X, Chao J, Duan M, Buch S, Chen L, Yao H. Involvement of sigma-1 receptor in astrocyte activation induced by methamphetamine via up-regulation of its own expression. J Neuroinflammation. 2015;12:29.
- 103. Yang H, Hreggvidsdottir HS, Palmblad K, Wang H, Ochani M, Li J, Lu B, Chavan S, Rosas-Ballina M, Al-Abed Y, Akira S, Bierhaus A, Erlandsson-Harris H, Andersson U, Tracey KJ. A critical cysteine is required for HMGB1 binding to Toll-like receptor 4 and activation of macrophage cytokine release. Proc Natl Acad Sci U S A. 2010;107(26):11942-11947.
- 104. Bustin M. Regulation of DNA-dependent activities by the functional motifs of the high-mobility-group chromosomal proteins. Mol Cell Biol. 1999;19(8):5237-5246.
- 105. Klune JR, Dhupar R, Cardinal J, Billiar TR, Tsung A. HMGB1: endogenous danger signaling. Mol Med. 2008;14(7-8):476-484.
- 106. Quinton MS, Yamamoto BK. Neurotoxic effects of chronic restraint stress in the striatum of methamphetamine-exposed rats. Psychopharmacology (Berl). 2007;193(3):341-350.
- 107. Raudensky J, Yamamoto BK. Effects of chronic unpredictable stress and methamphetamine on hippocampal glutamate function. Brain Res. 2007;1135(1):129-135.
- 108. Cadet JL, Jayanthi S, Deng X. Speed kills: cellular and molecular bases of methamphetamine-induced nerve terminal degeneration and neuronal apoptosis. Faseb j. 2003;17(13):1775-1788.
- 109. Guilarte TR, Nihei MK, McGlothan JL, Howard AS. Methamphetamine-induced deficits of brain monoaminergic neuronal markers: distal axotomy or neuronal plasticity. Neuroscience. 2003;122(2):499-513.
- 110. Lin Y-M, Sung W-W, Hsieh M-J, Tsai S-C, Lai H-W, Yang S-M, Shen K-H, Chen M-K, Lee H, Yeh K-T, Chen C-J. High PD-L1 Expression Correlates with Metastasis and Poor Prognosis in Oral Squamous Cell Carcinoma. PLoS ONE. 2015;10(11):e0142656.
- 111. Kiyasu J, Miyoshi H, Hirata A, Arakawa F, Ichikawa A, Niino D, Sugita Y, Yufu Y, Choi I, Abe Y, Uike N, Nagafuji K, Okamura T, Akashi K, Takayanagi R, Shiratsuchi M, Ohshima K. Expression of programmed cell death ligand 1 is associated with poor overall survival in patients with diffuse large B-cell lymphoma. Blood. 2015;126(19):2193-2201.
- 112. He J, Hu Y, Hu M, Li B. Development of PD-1/PD-L1 Pathway in Tumor Immune Microenvironment and Treatment for Non-Small Cell Lung Cancer. Scientific Reports. 2015;5:13110.
- 113. Mishra V, Schuetz H, Haorah J. Differential induction of PD-1/PD-L1 in Neuroimmune cells by drug of abuse. International Journal of Physiology, Pathophysiology and Pharmacology. 2015;7(2):87-97.
- 114. Sheppard KA, Fitz LJ, Lee JM, Benander C, George JA, Wooters J, Qiu Y, Jussif JM, Carter LL, Wood CR, Chaudhary D. PD-1 inhibits T-cell receptor induced phosphorylation of the ZAP70/CD3zeta signalosome and downstream signaling to PKCtheta. FEBS Lett. 2004;574(1-3):37-41.

Figure 1. Schematic diagram of the complex signals activated by METH

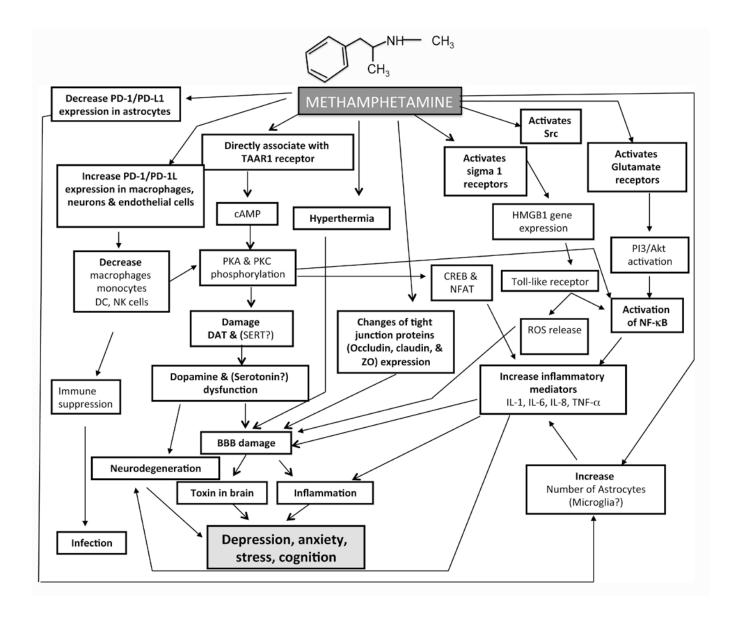


Figure 2. METH ($C_{10}H_{15}N$) and the gut-brain axis. Solid arrows are known effects of METH and dotted arrows are hypothesised effects

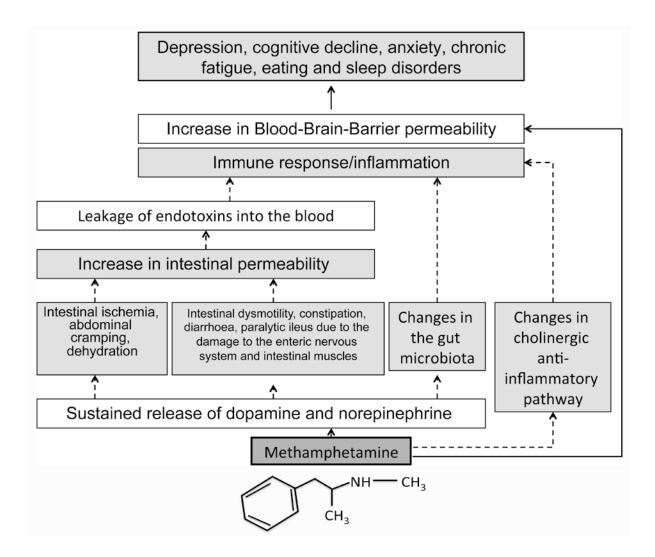


Fig. 3. Schematic diagram of the neuro-immunological affects of METH. Solid arrows are known effects of METH and dotted arrows are hypothesised effects

