## RESEARCH ARTICLE

# Core Promoter Short Tandem Repeats as Evolutionary Switch Codes for Primate Speciation 

MINA OHADI ${ }^{1 *}$, ELAHEH VALIPOUR ${ }^{1}$, SAEED GHADIMI-HADDADAN ${ }^{1}$, PEGAH NAMDAR-ALIGOODARZI ${ }^{1}$, ABOUZAR BAGHERI ${ }^{1}$, ALI KOWSARI ${ }^{2}$, MARYAM REZAZADEH ${ }^{1}$, HOSSEIN DARVISH ${ }^{3}$, and SOMAYEH KAZEMINASAB ${ }^{1}$<br>${ }_{2}^{1}$ Genetics Research Center, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran<br>${ }_{3}^{2}$ Stem Cell Research Center, Golestan University of Medical Science, Gorgan, Iran<br>${ }^{3}$ Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Alteration in gene expression levels underlies many of the phenotypic differences across species. Because of their highly mutable nature, proximity to the +1 transcription start site (TSS), and the emerging evidence of functional impact on gene expression, core promoter short tandem repeats (STRs) may be considered an ideal source of variation across species. In a genome-scale analysis of the entire Homo sapiens protein-coding genes, we have previously identified core promoters with at least one STR of $\geq 6$-repeats, with possible selective advantage in this species. In the current study, we performed reverse analysis of the entire Homo sapiens orthologous genes in mouse in the Ensembl database, in order to identify conserved STRs that have shrunk as an evolutionary advantage to humans. Two protocols were used to minimize ascertainment bias. Firstly, two species sharing a more recent ancestor with Homo sapiens (i.e. Pan troglodytes and Gorilla gorilla gorilla) were also included in the study. Secondly, four non-primate species encompassing the major orders across Mammals, including Scandentia, Laurasiatheria, Afrotheria, and Xenarthra were analyzed as out-groups. We introduce STR evolutionary events specifically identical in primates (i.e. Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla) vs. non-primate out-groups. The average frequency of the identically shared STR motifs across those primates ranged between 0.00005 and 0.06 . The identified genes are involved in important evolutionary and developmental processes, such as normal craniofacial development (TFAP2B), regulation of cell shape (PALMD), learning and long-term memory (RGS14), nervous system development (GFRA2), embryonic limb morphogenesis (PBX2), and forebrain development (APAF1). We provide evidence of core promoter STRs as evolutionary switch codes for primate speciation, and the first instance of identity-by-descent for those motifs at the interspecies level. Am. J. Primatol. 77:34-43, 2015. © 2014 Wiley Periodicals, Inc.

Key words: short tandem repeat; core promoter; primate; non-primate; evolution

## INTRODUCTION

Once considered "junk" DNA, short tandem repeats (STRs) are now believed to play a significant role in genome evolution by creating and maintaining quantitative genetic variation [Heidari et al., 2012; Iglesias et al., 2004; Jansen et al., 2012; King et al., 2006; Valipour et al., 2013]. Tandem repeats located in coding regions may increase the evolvability of proteins [Fondon \& Garner, 2004; Gemayel et al., 2010; Verstrepen et al., 2005]. Non-coding genes such as microRNAs are also influenced by purifying selection in STRs [Trivedi \& Hancock, 2012]. Repeats in the cis-regulatory regions are implicated in quantitative genetics and complex traits [Donaldson et al., 2008; Hammock \& Young, 2005]. Cisregulatory STRs are also suggested to be involved in

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evolutionary processes in Homo sapiens [Rockman \& Wray, 2002].

Vinces and co-workers have shown that genes driven by repeat-containing promoters show significantly higher rates of transcriptional divergence, and variations in repeat length result in changes in expression and local nucleosome positioning [Vinces et al., 2009]. In their experiments, replacement of tandem repeats with various sequences of the same size did not restore normal gene expression. Borel and co-workers have recently reported that in comparison with single nucleotide polymorphisms (SNPs), promoter tandem repeat variation is of more effect as causative cis expression quantitative trait loci for protein-coding gene expression in the case of CSTB gene [Borel et al., 2012]. Because of their combinatory and repeat potential, STRs may be considered a more efficient source of evolutionary codes for speciation in comparison with SNPs. STRs, by virtue of their special mutational and functional qualities have a major role in generating the genetic variation underlying adaptive evolution [Jansen et al., 2012; Kashi \& King, 2006; Mohammadparast et al., 2014]. Selection could shape STRs into "tuning knobs" that facilitate evolutionary adaptation by increasing adjustability [King et al., 2006]. This possibility is consistent with numerous examples of evolutionary conservation of STRs in genes with neurological, neurodevelopmental, and embryogenic function [Bolton et al., 2013; Darvish et al., 2013; King, 2012; Valipour et al., 2013; Zarif Yeganeh et al., 2009, 2010].

Because of their proximity to the +1 transcription start site (TSS), core promoter STRs may have a bona fide effect on phenotypic variation across species. The majority of the Homo sapiens core promoter STRs are conserved across evolution [Ohadi et al., 2012]. In a similar finding, it has recently been shown that STRs near the TSSs of genes (most of them involved in development) are often highly conserved, and that distance from a STR to the nearest TSS is a good predictor of the STR conservation score [Sawaya et al., 2012]. In a recent finding by Li et al. [2012] a short genomic di-nucleotide repeat structure in the promoter of ECE. 1 constitutes a novel and functional core promoter element, coincides with Homo sapiens evolution, and contributes to the pathogenesis of Alzheimer's disease. This genomic region is conserved between Homo sapiens and Pan troglodytes. The Homo sapiens SOX5, GABRA3, and MECOM genes provide further examples of functional core promoter STRs [Heidari et al., 2012; Valipour et al., 2013].

In a genome-scale analysis of the entire human protein-coding core promoters annotated in the GeneCards database, we have recently catalogued the Homo sapiens core promoter STRs with exceptional repeat numbers of $\geq 6$-repeats [Ohadi et al., 2012], which comprise less than $2 \%$ of the genes. The
idea was that the STRs that have reached that length may be of selective advantage, and therefore of prime importance for a pilot research of this kind. At the top of that list, the PAXBP1 gene contains the longest identified STR in a protein-coding core promoter. Indeed, this core promoter is a functional complex of multiple consecutive CT-STRs that has been exceptionally expanded in primates and not in any nonprimate order [Mohammadparast et al., 2014]. Among primates, this STR reaches maximum length and complexity in Homo sapiens. PAXBP1 function is indispensible for the recruitment of Pax3 and Pax7 [Diao et al., 2012], which in turn, are involved in the development of normal craniofacial features [Liu et al., 2012; Murdoch et al., 2012; Paternoster et al., 2012], and spine morphogenesis [Guerreiro et al., 2013], properties that are differentially distinct in primates vs. non-primates.

In a reverse analysis, in the current study, we screen the entire Homo sapiens orthologous genes in mouse, based on the Ensembl database, in order to identify core promoter STRs that have been possibly directionally contracted/evolved in the process of primate evolution. We introduce several STR evolutionary switches that are specific to primates, and may be important for the emergence and evolution of this order.

## METHODS

## Bioinformatics

The bioinformatics analyses were performed in two steps. In step I, the entire human protein-coding orthologous genes in mouse were analyzed for the density and repeat numbers of different classes of core promoter ( -120 to +1 of TSS) STRs using the Ensembl database (http://asia.ensembl.org/index. html ). This interval was screened for the presence of STRs using the Microsatellite Repeats Finder at the following link: http://biophp.org/minitools/microsatellite_repeats_finder/demo.php. STRs of $\geq 6$-repeats were then analyzed in Homo sapiens and Pan troglodytes, to detect conserved STRs across the three species. We have previously investigated the other end of the spectrum (i.e. STRs which are exceptionally long in human) [Ohadi et al., 2012]. That study covered STRs of potential interest in mouse, which were less than the exceptional length ( $<6$-repeats), and had been expanded in human. In Step II, we analyzed another primate species sharing an ancestor with Homo sapiens (i.e. Gorilla gorilla gorilla), and four species belonging to four major non-primate orders across mammals, including Scandentia, Laurasiatheria, Afrotheria, and Xenarthra, in order to examine specificity of STR evolutionary patterns between primates and non-primates.

The list of Homo sapiens genes were based on the annotated genes in the GeneCards database
(http://www.genecards.org/index.shtml). The sequences were retrieved from the Ensembl database. The following species (in parentheses) were used for the major orders across Mammals, primates (Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla), Rodents (Mouse), Scandentia (Tree Shrew), Laurasiatheria (Dog), Afrotheria (Elephant), and Xenarthra (Armadillo). The following datasets were used for the species analyzed in this study: Homo sapiens: February 2009 Homo sapiens high coverage assembly GRCh37 (GCA_000001405.13) from the Genome Reference Consortium; Pan troglodytes: Ensembl Pan troglodytes Version 2.1.4 (February 2011) of the Pan troglodytes genome assembly (known as Pan_troglodytes-2.1.4 or CHIMP2.1.4); Gorilla gorilla gorilla: Assembly gorGor3.1, December 2009, Database vs. 74.31; Mouse: the Genome assembly: GRCm38 (GCA_000001635.3); Tree shrew: Assembly tupBel1, June 2006, Database vs. 74.1, Dog: Assembly CanFam3.1 (GCA_000002285.2), Database vs. 74.31; Elephant: Assembly Loxafr3.0, July 2009, Dataset version 74.3, Armadillo: Assembly Dasnov3.0, December 2011, Dataset version 74.3.

Mouse core promoters containing at least one STR of $\geq 6$-repeats were compared with the corresponding sequences (i.e. -120 to +1 ) in Homo sapiens, Pan troglodytes, Gorilla gorilla gorilla, and the four out-group species, following sequence alignment of the corresponding transcripts, using the ClusalW2 version 4 software (http://www.ebi.ac.uk/ Tools/msa/clustalw2). ClustalW2 is a general purpose multiple sequence alignment program for nucleotides or proteins. The process of alignment was performed using the default settings (the main parameters are the gap opening penalty, and the gap extension penalty) of the software.

Core promoter STRs with $\geq 6$-repeats include less than $2 \%$ of the protein-coding genes [Ohadi et al., 2012], and are hereafter designated as "exceptionally long." The "STR formula" designates classes of STRs, and their repeat numbers based on the Ensembl data sets. The term "conserved" refers to the site of occurrence, and the class of STR. The term "identical" refers to the class of STR, and the number of repeats (STR formula).

The statistical analysis for the frequency of the STR motifs was based on the average frequency of those motifs at the interval between -120 and +1 of TSS, based on the Ensembl database, in the primates, H. sapiens $(n=19,000)$ Gorilla gorilla ( $n=2,000$ ), and Pan troglodytes, $(n=2,000$ ), where " $n$ " represents the number of genes screened in each species.

This research adhered to the legal requirements and protocols of the University of Social Welfare and Rehabilitation Sciences, Tehran, Iran. Also, this research adhered to the American Society of Primatologists principles for the ethical treatment of primates.

## RESULTS

Step I: Conserved Core Promoter STRs Across Mouse, Pan troglodytes, and Homo sapiens

Exceptionally long mouse core promoter STRs were observed in 309 genes (Suppl. 1). In 210 of those genes, STRs or the corresponding transcripts were non-existent in Homo sapiens. Among the remaining genes ( $n=99$ ) (Table I), STRs were conserved in Homo sapiens and mouse, of which, in 48 genes, those STRs were also conserved in Pan troglodytes (Table I). Di-nucleotide STRs were among the most conserved across the three species. For example, in the gene, GLT25D2, CA-repeats ranging from 17 to 24 repeats were conserved in mouse, Pan troglodytes, and Homo sapiens. In the gene, $R N F 130$, CG-repeats ranging from 6- to 7-repeats are conserved in the three species.

Tri-nucleotide STRs were also conserved to a lesser extent, of which combinations of GCG and GCC were among the common conserved motifs. For example, in the gene, $M B D 6$, repeats of GCG were conserved across the three species.

## Step IIA: Identical Evolution of Certain STRs in Primates (i.e. Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla) vs. Out-groups Belonging to the Major Nonprimate Orders Across Mammals

Within the conserved STRs among mouse, Pan troglodytes, and Homo sapiens, in 37 genes the evolutionary pattern of STRs was identical in Pan troglodytes and Homo sapiens vs. mouse (Table II). In order to examine the specificity of this evolutionary pattern, we analyzed another primate species sharing an ancestor with humans (i.e. Gorilla gorilla gorilla), and four species belonging to the major nonprimate orders across mammals. In 21 out of the 37 genes, STRs were identical across Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla vs. the nonprimate out-groups (shaded areas in Table II). In the remaining 16 genes, STRs were identical, specifically between Homo sapiens and Pan troglodytes (Table II). The average frequency of the identically shared STR formulas across Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla ranged between 0.00005 and 0.06 (Table III). The following examples are STR evolutionary events that are identical in Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla, and no non-primates studied. In the gene, STAP1, TG/23 in mouse is split to two TG-STRs ranging from 7 to 12 repeats in the three primates. In the gene, PALMD, TC/29 CA/4 in mouse is shrunk to TC/3 CA/4 in the three primates. In another remarkable example, in the gene, $R G S 14, \mathrm{GT} / 20$ is split into several identical GT-repeats ranging from 3- to 6repeats in the three primates. In the gene, $N O V$, the STR formula in mouse is CA/17, which has been
TABLE I. Step I: Evolutionary Analysis of the Mouse Exceptionally Long Core Promoter STRs ( $\geq$ 6-Repeats) in Pan troglodytes and Homo sapiens ${ }^{\text {a }}$

TABLE I. Continued

| Gene symbol | Mouse transcript ID | Mouse STR formula |  |  |  |  |  | Pan troglodytes transcript ID | Pan troglodytes STR formula |  |  |  |  | Homo sapiens transcript ID |  | Homo sapiens STR formula |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| RAB33B | ENSMUST00000054387 | tc/7 | cgc/3 |  |  |  |  | ENSPTRT00000030587 | tc/3 | ga/3 |  |  |  | ENST00000305626 | ga/3 | tc/3 |  |  |  |  |  |
| PTAR1 | ENSMUST00000099560 | $\mathrm{gc} / 7$ |  |  |  |  |  | ENSPTRT00000074570 | cg/6 | $\mathrm{gc} / 3$ | cg/3 |  |  | ENST00000340434 | $\mathrm{gc} / 3$ | cg/9 |  |  |  |  |  |
| ACVR1B | ENSMUST00000000544 | $\mathrm{gc} / 7$ | ggag/5 | gc/4 |  |  |  | ENSPTRT00000009165 | gc/8 |  |  |  |  | ENST00000257963 | cg/3 | gc/4 |  |  |  |  |  |
| GFRA2 | ENSMUST00000022699 | ag/7 | ggga/3 | $\mathrm{ga} / 3$ |  |  |  | ENSPTRT00000037137 | $\mathrm{ga} / 3$ | ag/4 |  |  |  | ENST00000400782 | ga/3 | ga/3 |  |  |  |  |  |
| TFAP2B | ENSMUST00000027059 | gt/7 | at/4 | gcgc/3 |  |  |  | ENSPTRT00000033774 | $\mathrm{ta} / 3$ | at/4 | tg/4 |  |  | ENST00000393655 | $\operatorname{tg} / 3$ | ta/3 | $\mathrm{ta} / 3$ | $\operatorname{tg} / 4$ |  |  |  |
| CHST1 | ENSMUST00000065797 | gc/7 |  |  |  |  |  | ${ }^{\text {b }}$ |  |  |  |  |  | ENST00000308064 | gc/4 |  |  |  |  |  |  |
| STAG2 | ENSMUST00000069619 | ccetc/6 | cetc/3 | ct/3 | ccct/4 |  |  | b |  |  |  |  |  | ENST00000371144 | tcce/3 | cctc/3 | ag/3 |  |  |  |  |
| PLEKHA5 | ENSMUST00000032357 | tcge/6 | cg/3 | $\mathrm{gc} / 3$ |  |  |  | c |  |  |  |  |  | ENST00000299275 | cg/3 | $\mathrm{gc} / 3$ | $\mathrm{gc} / 3$ | ggc/5 |  |  |  |
| SETD2 | ENSMUST00000153838 | ggag/6 | ga/3 | $\mathrm{ga} / 3$ | gcgg/3 | ccag/4 | gcc/3 | b |  |  |  |  |  | ENST00000409792 | $\mathrm{gcc} / 10$ | cetce/3 |  |  |  |  |  |
| IQGAP2 | ENSMUST00000068603 | gcgg/6 | cgag/4 |  |  |  |  | ENSPTRT00000031522 | gc/4 | ggc/3 |  |  |  | ENST00000274364 | gc/4 |  |  |  |  |  |  |
| LIMD1 | ENSMUST00000026269 | ccgce/6 | ccgce/5 |  |  |  |  | ENSPTRT00000027735 | ac/26 |  |  |  |  | ENST00000273317 | ac/33 |  |  |  |  |  |  |
| PRKCH | ENSMUST00000021527 | tgc/6 |  |  |  |  |  | ENSPTRT0000001175 2 | $\mathrm{gcg} / 3$ |  |  |  |  | ENST00000332981 | $\mathrm{gcg} / 3$ |  |  |  |  |  |  |
| RAP1B | ENSMUST00000064667 | gcg/6 |  |  |  |  |  | ENSPTRT00000009547 | gc/3 | cg/3 |  |  |  | ENST00000250559 | gc/3 |  |  |  |  |  |  |
| PBX2 | ENSMUST00000038149 | cgc/6 |  |  |  |  |  | ENSPTRT00000033293 | tc/4 | $\mathrm{ct} / 3$ | cctc/3 |  |  | ENST00000453487 | $\mathrm{ct} / 3$ | ct 3 | cctc/3 |  |  |  |  |
| TTLL11 | ENSMUST00000028248 | ggc/6 | cgg/3 | $\mathrm{gcg} / 4$ |  |  |  | b |  |  |  |  |  | ENST00000373776 | $\mathrm{gcg} / 3$ |  |  |  |  |  |  |
| CASP3 | ENSMUST00000093517 | tgc/6 |  |  |  |  |  | ENSPTRT00000061788 | cggg/3 |  |  |  |  | ENST00000308394 | cggg/3 |  |  |  |  |  |  |
| MLL5 | ENSMUST00000115128 | gc/6 | gggc/3 | gcc/5 |  |  |  | ENSPTRT00000036230 | gcc/7 | gce/4 |  |  |  | ENST00000334877 | gcc/7 | gce/4 |  |  |  |  |  |
| MEX3C | ENSMUST00000091852 | ccg/6 | gc/3 | gc/3 |  |  |  | ENSPTRT00000018406 | gce/3 |  |  |  |  | ENST00000406189 | gce/5 | gc/3 |  |  |  |  |  |
| AMIGO1 | ENSMUST00000106656 | cgg/6 | $\mathrm{ac} / 3$ | cg/3 | cgg/3 | $\mathrm{gc} / 5$ |  | ENSPTRT00000001979 | ac/3 | cg/3 | cgg/3 | gc/5 |  | ENST00000369864 | cg/3 | cgg/3 | $\mathrm{gc} / 5$ |  |  |  |  |
| ANKRD13B | ENSMUST00000092892 | ccg/6 | ccg/3 | $\mathrm{gc} / 3$ |  |  |  | c |  |  |  |  |  | ENST00000394859 | ccg/4 |  |  |  |  |  |  |
| ANKRD17 | ENSMUST00000081914 | gcg/6 | ct/4 | cect/3 | ccct/3 |  |  | ENSPTRT00000030072 | ccet/4 | gcg/5 |  |  |  | ENST00000358602 | ccet/4 | gcg/6 |  |  |  |  |  |
| GOLGA4 | ENSMUST00000084820 | cgc/6 |  |  |  |  |  | ENSPTRT00000046942 | gc/3 |  |  |  |  | ENST00000361924 | gc/3 |  |  |  |  |  |  |
| ABI2 | ENSMUST00000052332 | agg/6 | ctc/3 | gag/3 |  |  |  | ENSPTRT00000023794 | cg/3 | cgc/3 |  |  |  | ENST00000261017 | $\mathrm{gc} / 3$ |  |  |  |  |  |  |
| KIAA0368 | ENSMUST00000102889 | tce/6 |  |  |  |  |  | ENSPTRT00000039289 | $\mathrm{ta} / 3$ |  |  |  |  | ENST00000259335 | $\mathrm{ta} / 3$ |  |  |  |  |  |  |
| ZBTB2 | ENSMUST00000100078 | gcg/6 | ct/3 |  |  |  |  | ENSPTRT00000034541 | ct/3 |  |  |  |  | ENST00000325144 | tc/3 | ct/3 | ccg/7 |  |  |  |  |
| GSDMD | ENSMUST00000023238 | gga/6 |  |  |  |  |  | ENSPTRT00000038201 | cg/3 |  |  |  |  | ENST00000262580 | cg/3 |  |  |  |  |  |  |
| TMPRSS2 | ENSMUST00000000395 | ctc/6 | tc/3 |  |  |  |  | ENSPTRT00000025975 | ccgece/3 |  |  |  |  | ENST00000497881 | ccgece/3 |  |  |  |  |  |  |
| ATXN1 | ENSMUST00000167708 | agg/6 | ga/4 | gagg/3 |  |  |  | ENSPTRT00000065472 | 0 |  |  |  |  | ENST00000450222 | gagc/5 | ga/3 | gagg/4 | gggag/3 | gga/7 |  |  |
| ERI1 | ENSMUST00000033927 | cg/6 | ct/3 | ccg/3 |  |  |  | b ${ }^{\text {b }}$ |  |  |  |  |  | ENST00000519292 | gc/3 |  |  |  |  |  |  |
| SRCIN1 | ENSMUST00000107596 | ac/6 | cg/3 | $\mathrm{gc} / 3$ | ac/3 | cg/3 | cg/3 | c |  |  |  |  |  | ASMPATCHT00000002241 | cg/4 | cg/5 | ac/3 | cg/5 | ac/4 | ac/4 |  |
| RNF130 | ENSMUST00000054684 | gc/6 |  |  |  |  |  | ENSPTRT00000032588 | cg/7 |  |  |  |  | ENST00000522208 | cg/6 | ccg/3 |  |  |  |  |  |
| RBMS2 | ENSMUST00000092033 | ct/6 | tc/3 |  |  |  |  | ENSPTRT00000009384 | cctc/3 | tc/4 |  |  |  | ENST00000262031 | cctc/3 | tc/4 |  |  |  |  |  |
| PRR7 | ENSMUST00000046533 | gc/6 |  |  |  |  |  | b |  |  |  |  |  | ENST00000323249 | cg/5 | gc/4 |  |  |  |  |  |
| TRPC5 | ENSMUST00000040184 | ga/6 |  |  |  |  |  | b |  |  |  |  |  | ENST00000262839 | tc/12 | tc/18 | ac/10 |  |  |  |  |
| TSPAN5 | ENSMUST00000029800 | gc/6 | ag/3 |  |  |  |  | ENSPTRT00000030330 | gc/6 |  |  |  |  | ENST00000305798 | cg/5 | $\mathrm{gc} / 3$ |  |  |  |  |  |
| LY9 | ENSMUST00000068878 | ca/6 | ac/3 | ca/3 | ca/3 |  |  | ENSPTRT00000002849 | ac/5 | ca/5 |  |  |  | ENST00000368041 | ac/5 | $\mathrm{ca} / 5$ |  |  |  |  |  |
| HTRA4 | ENSMUST00000084031 | gt/6 | at/3 |  |  |  |  | ENSPTRT00000037383 | $\mathrm{gt} / 3$ |  |  |  |  | ENST00000302495 | gt/3 |  |  |  |  |  |  |
| APAF1 | ENSMUST00000020157 | gc/6 | $\mathrm{gcg} / 3$ |  |  |  |  | ENSPTRT00000048551 | gc/3 | gt/4 |  |  |  | ENST00000333991 | gc/3 | gt/4 |  |  |  |  |  |
| FAM168B | ENSMUST00000047534 | gc/6 | gc/3 |  |  |  |  | c |  |  |  |  |  | ENST00000409185 | $\mathrm{gc} / 3$ | $\mathrm{gc} / 3$ |  |  |  |  |  |
| AATF | ENSMUST00000018841 | gc/6 |  |  |  |  |  | ENSPTRT00000016671 | cg/3 | gc/3 |  |  |  | ENST00000225402 | cg/3 | gc/3 |  |  |  |  |  |
| ENOX1 | ENSMUST00000022589 | gc/6 | gc/3 | cg/3 |  |  |  | ENSPTRT00000010726 | tc/3 | ct/3 | gc/5 |  |  | ENST00000261488 | gc/5 | $\mathrm{gc} / 3$ | cg/3 |  |  |  |  |
| UCMA | ENSMUST00000027978 | tc/6 |  |  |  |  |  | ENSPTRT00000004215 | cct/3 | ct/4 |  |  |  | ENST00000378681 | cct/3 | ct/4 |  |  |  |  |  |
| XYLT1 | ENSMUST00000032892 | cg/6 | cct/3 |  |  |  |  | ENSPTRT00000014408 | ggc/6 | gcg/3 | agg/4 | ggc/7 |  | ENST00000261381 | cgg/4 |  |  |  |  |  |  |
| TMPRSS2 | ENSMUST00000000395 | ctc/6 | tc/3 |  |  |  |  | ENSPTRT00000025975 | ccgcce/3 |  |  |  |  | ENST00000497881 | ccgece/3 |  |  |  |  |  |  |
| PDE2A | ENSMUST00000166652 | ct/6 | ct/4 | tce/3 |  |  |  | ENSPTRT00000007523 | tc/3 | $\mathrm{gt} / 3$ | ct/5 | $\mathrm{ct} / 3 \mathrm{ct} / 3 \mathrm{ct} / 4$ | tcc/3 | ENST00000334456 | ct/3 | ct/5 | ct/3 | ct/3 | ct/3 | ct/4 | tcc/3 |
| NHLH1 | ENSMUST00000059794 | gt/6 | $\mathrm{ag} / 3$ | at/4 | ca/3 | gt/4 |  | ENSPTRT00000002834 | gt/7 | at/4 |  |  |  | ENST00000302101 | gt/6 | at/4 |  |  |  |  |  |
| RNF19B | ENSMUST00000168461 | cgg/5 |  |  |  |  |  | ENSPTRT00000045611 | cgg/3 |  |  |  |  | ENST00000356990 | cgg/3 |  |  |  |  |  |  |

Transcript not found
Ortholog not found.
TABLE II. Step II: Mouse Conserved Exceptionally Long Core Promoter STRs That Have Been Evolved Identically in Primates (Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla), vs. Non-primate Out-groups ${ }^{\text {a,b }}$

${ }^{\mathrm{a}}$ Gene order is based on the length of the STRs in mouse. The STR formula designates classes of STRs and their repeat numbers based on the Ensembl data sets (See Bioinformatics section).
${ }^{\mathrm{b}}$ Shaded areas represent identical STR formulas.
${ }^{\mathrm{e}}$ An exceedingly rare STR, ggegg/3, with the frequency of 0.0001 , emerges in Gorilla gorilla gorilla, and is evolved to GGCGG/3 GC/3, in Homo sapiens and Pan troglodytes.

TABLE III. Frequency of the STR Motifs Identically Evolved in Three Primates; Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla

| Gene symbol | STR formula shared by primates |  |  |  |  | STR motif frequency ${ }^{\text {a }}$ |  |  |  |  | Gene ontology |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| SLC39A8 | cg/4 |  |  |  |  | 0.005 |  |  |  |  | Metal ion transmembrane transporter activity |
| PALMD | ct/3 | $\mathrm{ac} / 4$ |  |  |  | 0.01 | 0.001 |  |  |  | Regulation of cell shape |
| STAP1 | $\operatorname{tg} / 9$ | $\operatorname{tg} / 7$ |  |  |  | 0.0001 | 0.0005 |  |  |  | Signal transduction |
| RGS14 | gt/6 | $\mathrm{gt} / 3$ | gt/5 | gt/4 | ct/3 | 0.0005 | 0.01 | 0.002 | 0.005 | 0.01 | Learning and long-term memory |
| NOV | ca/4 | 0.002 |  |  |  |  |  |  |  |  | Regulation of cell growth and gene expression |
| CHD3 | gag/13 | cge/3 | tgg/3 | ggcg/3 |  | 0.00005 | 0.0001 | 0.001 | 0.0001 |  | Regulation of transcription |
| C2 | $\operatorname{tg} / 3$ | ac/3 |  |  |  | 0.01 | 0.005 |  |  |  | Regulation of complement activation |
| TRIM8 | gga/4 | gc/4 |  |  |  | 0.001 | 0.02 |  |  |  | Protein homodimerization activity |
| SELL | ag/3 |  |  |  |  | 0.01 |  |  |  |  | Regulation of immune response |
| SPO11 | gt/3 |  |  |  |  | 0.01 |  |  |  |  | Spermatid development and female gamete generation |
| PLAG1 | cgg/9 |  |  |  |  | 0.0001 |  |  |  |  | Regulation of transcription |
| SLC3A1 | ca/3 |  |  |  |  | 0.01 |  |  |  |  | Basic amino acid transport |
| VKORC1L1 | ggc/4 | ggc/4 | gcg/6 |  |  | 0.003 | 0.003 | 0.0002 |  |  | Integral to membrane |
| IKZF2 | tc/4 | tcc/3 |  |  |  | 0.009 | 0.006 |  |  |  | Positive regulation of transcription from RNA polymerase II promoter |
| SH3BP1 | cg/3 | ga/3 | ga/3 |  |  | 0.06 | 0.009 | 0.009 |  |  | Signal transduction |
| CRYM | ggcgg/3 | gc/3 |  |  |  | 0.0001 | 0.06 |  |  |  | Negative regulation of transcription from RNA polymerase II promoter |
| RAB33B | ga/3 | tc/3 |  |  |  | 0.009 | 0.01 |  |  |  | Autophagy |
| GFRA2 | ga/3 | $\mathrm{ga} / 3$ |  |  |  | 0.009 | 0.009 |  |  |  | Nervous system development |
| TFAP2B | $\operatorname{tg} / 3$ | ta/3 | ta/3 | $\operatorname{tg} / 4$ |  | 0.01 | 0.006 | 0.006 | 0.005 |  | Skin development |
| PRKCH | gcg/3 |  |  |  |  | 0.0003 |  |  |  |  | Positive regulation of glial cell proliferation |
| PBX2 | ct/3 | ct/3 | cctc/3 |  |  | 0.01 | 0.01 | 0.0002 |  |  | Embryonic limb morphogenesis |
| CASP3 | cggg/3 |  |  |  |  | 0.0002 |  |  |  |  | Heart development |
| MLL5 | gcc/7 | gcc/4 |  |  |  | 0.0005 | 0.005 |  |  |  | DNA methylation |
| ANKRD17 | ccet/4 | gcg/6 |  |  |  | 0.0001 | 0.0001 |  |  |  | Regulation of smooth muscle cell differentiation |
| GOLGA4 | $\mathrm{gc} / 3$ |  |  |  |  | 0.06 |  |  |  |  | Protein targeting to Golgi |
| KIAA0368 | ta/3 |  |  |  |  | 0.006 |  |  |  |  | May play a role in ERAD and other enhanced proteolysis |
| GSDMD | cg/3 |  |  |  |  | 0.06 |  |  |  |  | Cellular response to extracellular stimulus |
| TMPRSS2 | ccgcce/3 |  |  |  |  | 0.00005 |  |  |  |  | Proteolysis |
| RBMS2 | cctc/3 | tc/4 |  |  |  | 0.0002 | 0.009 |  |  |  | RNA processing |
| LY9 | ac/5 | ca/5 |  |  |  | 0.0003 | 0.001 |  |  |  | Cell adhesion |
| HTRA4 | $\mathrm{gt} / 3$ |  |  |  |  | 0.01 |  |  |  |  | Endopeptidase activity |
| APAF1 | $\mathrm{gc} / 3$ | gt/4 |  |  |  | 0.06 | 0.005 |  |  |  | Nervous system development, forebrain development |
| AATF | cg/3 | gc/3 |  |  |  | 0.06 | 0.06 |  |  |  | Regulation of mitotic cell cycle |
| UCMA | cct/3 | ct/4 |  |  |  | 0.006 | 0.009 |  |  |  | Regulation of osteoblast differentiation |
| PDE2A | ct/3 | ct/5 | ct/3 | ct/3 | ct/3 | 0.01 | 0.002 | 0.01 | 0.01 | 0.01 | Negative regulation of transcription from RNA polymerase II promoter |
|  | ct/4 | tcc/3 |  |  |  | 0.01 | 0.009 |  |  |  |  |
| NHLH1 | gt/6 | at/4 |  |  |  | 0.002 | 0.001 |  |  |  | Regulation of transcription |
| RNF19B | cgg/3 |  |  |  |  | 0.01 |  |  |  |  | Involved in the cytolytic activity of natural killer cells and cytotoxic T-cells |

[^1]shrunk to CA/4 in Homo sapiens, Pan troglodytes, and Gorilla gorilla gorilla. In the gene, TRIM8, CTCC/9 is evolved to GGA/4 GC/4 in the three primates. In the gene, $S E L L, \mathrm{AG} / 9 \mathrm{GA} / 5$, in mouse is shrunk to GA/3. Shrinkage of GT/9 in the gene, $S P O 11$, to GT/3 in the three primates is another example of directional shrinkage of core promoter STRs. In the gene, SLC3A1, CA/8 in mouse is contracted to CA/3. In the gene, $I K Z F 2$, combinatory AGG/7 GAG/4 TC/4 is evolved to TCC/3 TC/4. In the gene, $L y 9$, the following formula in mouse, $\mathrm{CA} / 6 \mathrm{CA} / 3 \mathrm{CA} / 3 \mathrm{CA} / 3$, is evolved to $\mathrm{AC} / 5 \mathrm{CA} / 5$ in the three primates. In another remarkable observation, in the gene, NHLH1, the following STR formula in mouse, GT/6 AG/3 AT/4 CA/ 3 GT/4, is contracted to GT/6 AT/4 in the three primates. In the gene, TMPRSS2, the following formula in mouse, CTC/6, is evolved to CCGCCC/3. Of note, the latter evolutionary event might have risen by point mutations rather than slippage mispairing, within the CTCCTC motif in mouse, including a T- to C- and C- to G-, which gives rise to the motif, CCGCCC.

## Step IIB: Identical Evolution of Certain STRs in Homo sapiens and Pan troglodytes vs. Gorilla gorilla gorilla, and the Out-groups Belonging to the Major Non-primate Orders Across Mammals

Within the genes in which core promoter STRs had been differentially evolved in primates vs. nonprimates, we observed differential evolution of certain STRs in Homo sapiens and Pan troglodytes vs. Gorilla gorilla gorilla and non-primates (Table II). For example, in the gene, CHD3, the STR formula for multiple STRs is identical in Homo sapiens and Pan troglodytes. In the gene, PLAG1, the following STR formula in mouse, GCG/8 GGC/3, is evolved to GGC/9 in Homo sapiens and Pan troglodytes, only. In the gene, $S H 3 B P 1$, GA/ $7 \mathrm{GA} / 3$ is replaced by CG/3 GA/3 GA/3 in Homo sapiens and Pan troglodytes. The core promoter of the $A A T F$ gene contains a STR of CG/6 in mouse, whereas this STR is split into GC/3 GC/3, specifically in Pan troglodytes and Homo sapiens. In the gene, $M L L 5$, the following formula in mouse, GCC/6 GGGC/3 GCC/5, is evolved to GCC/7 GCC/4 in Homo sapiens and Pan troglodytes. In the gene, RNF19B, GGC/6 in mouse is shrunk to GGC/3 in both Homo sapiens and Pan troglodytes. In the gene, ANKRD17, the following formula: GCG/6 CT/4 CCCT/3 CCCT/3 is evolved to CCCT/4 GCG/4, specifically in Pan troglodytes and Homo sapiens. In the gene, VKORC1L1, GGC/7 GCG/3 in mouse is evolved to GGC/4 GGC/4 GCG/6 specifically in Homo sapiens and Pan troglodytes.

In the gene, CRYM, an exceedingly rare STR, GGCGG/3, with the frequency of 0.0001 , emerges in Gorilla gorilla gorilla, and is evolved to GGCGG/3 GC/3, in Homo sapiens and Pan troglodytes.

A human-specific STR formula was observed in the gene, TFAP2B, in which an extra GT/3 was observed in Homo sapiens, and not in Pan troglodytes, and Gorilla gorilla gorilla. This specificity occurred on a background of conserved STRs in the three primates.

## DISCUSSION

Alteration in the levels of gene expression is considered to be a mechanism for phenotypic differences across species [Brawand et al., 2011; Esmaeil-zadeh-Gharehdaghi et al., 2011; Mohammadparast et al., 2014; Wang \& Rekaya, 2009]. We have recently reported core promoter STRs that are expanded in Homo sapiens, and proposed that interspecies variation in gene expression levels may, at least in part, be attributed to the core promoter STRs [Ohadi et al., 2012]. This notion is further supported by the functional role of core promoter STRs to modulate gene expression [Heidari et al., 2012; Li et al., 2012; Valipour et al., 2013]. In a reverse study, here we provide results of mouse orthologous gene core promoters that contain exceptionally long STRs, and have been conserved across evolution, shrinkage of which may be linked with the primate evolution.

In eukaryotes, the "core promoter" interval can vary from one gene to the other in respect with structure and length [Darvish et al., 2011; Moshonov et al., 2008; Sandelin et al., 2007; Vinces et al., 2009]. This interval contains the basic and crucial transcription factor (TF) binding sites for constitutive gene expression. For TATA-containing promoters ( $24 \%$ of the protein-coding genes) [Yang et al., 2007], this region normally includes -40 to +1 of TSS. For TATA-less promoters, this interval can be extended [Darvish et al., 2011; Singh et al., 2012]. As a pilot study, the interval -120 to +1 is chosen in the current study, which needs to be extended in the future studies.

The proximity of the core promoter STRs to the +1 TSS may be an evolutionary constrain on those STRs to evolve to a certain length, which may be species-specific. The idea of "exceptional expansion" of the STRs ( $<2 \%$ of the genes) proposes that selective/adaptive expansion of certain STRs may be linked with speciation in certain species, and not the others. STRs that have reached that length may be of more potential importance for a preliminary research of this kind. Along that notion, shrinkage of those STRs may also be linked with speciation. Data from the mouse genome have confirmed the abundance of STRs but have also revealed impressive differences with human [Mouse Genome Sequencing Consortium, 2002]. If identical search criteria are used, the mouse genome proves to be repeat-rich with two- to three-fold more STRs than human. Moreover, STRs are longer in mice than in humans.

Our current results provide core promoter STRs that are exceptionally long in mouse, and have been shrunk in primates. Surveys of variability of homologous STR loci among species may be subject to ascertainment bias for STR length where STR loci isolated in one species tend to be longer than homologous loci in related species [Vowles \& Amos, 2006]. A life cycle of creation and degeneration may be considered for STRs, which undergo neutral evolution [Buschiazzo \& Gemmel, 2006]. Two protocols were used in the current study to minimize ascertainment bias. Firstly, two primates sharing closer ancestors (i.e. chimpanzee and gorilla) were included in the study. Secondly, four species encompassing major orders across mammals were also included as out-group. In 37 genes, we observed identical evolution of core promoter STR formulas in primates vs. out-groups. The STR motifs identically shared were of the frequencies ranging from 0.00005 to 0.06 . In 16 of the 37 genes, STRs were specifically identical between Homo sapiens and Pan troglodytes, pointing to the possible role of those STRs in the emergence of a common ancestor shared by the two species. In the remaining 21 genes, the STR formula was also identical in Gorilla gorilla gorilla, indicating a possible role for those STRs in the emergence of a more distant ancestor, which could have led to the common ancestor of great apes. "Specific" and "identical" co-occurrence of those STR formulas (rare frequencies of $<0.01$ for the majority of the shared motifs) (Table III) in three primates whose ancestors have diverged four to eight million years ago, supports an evolutionary role for those STRs as "switch codes" for primate speciation, and excludes the notion of ascertainment bias. In other words, this is the first support of identity-by-descent for STRs at the interspecies level.

Remarkably, in the gene, TFAP2B, an extra GT/3 was observed in Homo sapiens, and not in Pan troglodytes and Gorilla gorilla gorilla. This specificity occurred on a background of identical STRs in the three primates. Remarkably, mutations in the $T F A P 2 B$ gene result in craniofacial malformations in human [Milunsky et al., 2008].

We observed three exceptions in the differential evolution of the STRs from Step II in primates vs. non-primates. The first and second exceptions were observed in the genes, $R A B 33 B$, and $R B M S 2$, in which combination of STRs was also conserved in one non-primate species, Armadillo, and the third exception was in the gene, KIAA0368, in which TA/3 was conserved in one non-primate species, Dog. Gene Ontology for the genes listed (Table III) revealed important evolutionary and developmental functions for those genes, such as regulation of cell shape (PALMD), learning and long-term memory (RGS14), nervous system development (GFRA2), embryonic limb morphogenesis ( $P B X 2$ ), and forebrain development (APAF1).

Beyond a certain length, STRs become the binding site for TFs. For example, a minimum of 3repeats is required for the CA-, GC-, AT-, CT, GCG, and GCC-repeats to become binding sites for TFs, where further repeats add additional TFs (http://asp. ii.uib.no:8090/cgi-bin/CONSITE/consite).

We recognize that our discovery of the instances of possible functional changes does not constitute proof, but rather provide direction for further study of the importance of the listed STRs in primate evolution, development, and disease.

## REFERENCES

Bolton KA, Ross JP, Grice DM, et al. 2013. STaRRRT: a table of short tandem repeats in regulatory regions of the human genome. BMC Genomics 14:795.
Borel C, Migliavacca E, Letourneau A, et al. 2012. Tandem repeat sequence variation as causative Cis-eQTLs for protein-coding gene expression variation: the case of CSTB. Human Mutation 33:1302-1309.
Brawand D, Soumillon M, Necsulea A., et al. 2011. The evolution of gene expression levels in mammalian organs. Nature 478:343-348.
Buschiazzo E, Gemmel NJ. 2006. The rise, fall and renaissance of microsatellites in eukaryotic genomes. BioEssays 28:1040-1050.
Darvish H, Heidari A, Hosseinkhani S, et al. 2013. Biased homozygous haplotypes across the human caveolin 1 upstream purine complex in Parkinson's disease. Journal of Molecular Neuroscience 51:389-393.
Darvish H, Nabi MO, Firouzabadi SG, et al. 2011. Exceptional human core promoter nucleotide compositions. Gene 475:79-86.
Diao Y, Guo X, Li Y, et al. 2012. Pax3/7BP is a Pax7- and Pax3binding protein that regulates the proliferation of muscle precursor cells by an epigenetic mechanism. Cell Stem Cell 11:231-241.
Donaldson ZR, Kondrashov FA, Putnam A, et al. 2008. Evolution of a behavior-linked microsatellite-containing element in the 5' flanking region of the primate AVPR1A gene. BMC Evolutionary Biology 8:180.
Esmaeilzadeh-Gharehdaghi E, Banan M, Farashi S, et al. 2011. Support for down-tuning of the calreticulin gene in the process of human evolution. Progress in Neuropsychopharmacology and Biological Psychiatry 35:1770-1773.
Fondon JW III, Garner HR. 2004. Molecular origins of rapid and continuous morphological evolution. Proceedings of the National Academy of Science United States of America 101:18058.
Gemayel R, Vinces MD, Legendre M, Verstrepen KJ. 2010. Variable tandem repeats accelerate evolution of coding and regulatory sequences. Annual Review of Genetics 44:445477. Review.

Guerreiro I, Nunes A, Woltering JM, et al. 2013. Role of a polymorphism in a Hox/Pax-responsive enhancer in the evolution of the vertebrate spine. Proceedings of the National Academy of Science United States of America 110:10682-10686.
Hammock EAD, Young LJ. 2005. Microsatellite instability generates diversity in brain and sociobehavioral traits. Science 308:1630-1634.
Heidari A, Nariman Saleh Fam Z, Esmaeilzadeh-Gharehdaghi E, et al. 2012. Core promoter STRs: novel mechanism for inter-individual variation in gene expression in humans. Gene 492:195-198.
Iglesias AR, Kindlund E, Tammi M, Wadelius C. 2004. Some microsatellites may act as novel polymorphic cis-regulatory
elements through transcription factor binding. Gene 341:149-165.
Jansen A, Gemayel R, Verstrepen KJ. 2012. Unstable microsatellite repeats facilitate rapid evolution of coding and regulatory sequences. Genome Dynamics 7:108-125. Review.
Kashi Y, King DG. 2006. Simple sequence repeats as advantageous mutators in evolution. Trends in Genetics 22:253-259.
King DG. 2012. Evolution of simple sequence repeats as mutable sites. Advances in Experimental Medical Biology 769:10-25.
King DG, Trifonov EN, Kashi Y. 2006. Chapter 4. Tuning knobs in the genome: evolution of simple sequence repeats by indirect selection In: Caporale LH, editor. The implicit genome. UK: Oxford Press.
Li Y, Seidel K, Marschall P, et al. 2012. A polymorphic microsatellite repeat within the ECE-1c promoter is involved in transcriptional start site determination, human evolution, and Alzheimer's disease. Journal of Neuroscience 32:16807-16820.
Liu F, van der Lijn F, Schurmann C, Zhu G, Mallar Chakravarty M. 2012. A genome-wide association study identifies five loci influencing facial morphology in Europeans. PLoS ONE 8:e1002932.
Milunsky JM, Maher TA, Zhao G, et al. 2008. TFAP2A mutations result in branchio-oculo-facial syndrome. American Journal of Human Genetics 82:1171-1177.
Mohammadparast S, Bayat H, Biglarian A, Ohadi M. 2014. Exceptional expansion and conservation of the CT-STR complex in the core promoter of PAXBP1 in primates. American Journal of Primatology doi: 10.1002/ajp.22266.
Moshonov S, Elfakess R, Golan-Mashiach M, Sinvani H, Dikstein R. 2008. Links between core promoter and basic gene features influence gene expression. BMC Genomics 9:92.
Mouse Genome Sequencing Consortium. 2002. Initial sequencing and comparative analysis of the mouse genome. Nature 420:520-562.
Murdoch M, DelConte C, García-Castro MI. 2012. Pax7 lineage contributions to the mammalian neural crest. PLoS ONE 7: e41089.
Ohadi M, Mohammadparast S, Darvish H. 2012. Evolutionary trend of exceptionally long human core promoter short tandem repeats. Gene 507:61-67.
Paternoster L, Zhurov AI, Toma AM, Kemp JP, Pourcain BS. 2012. Genome-wide association study of three-dimensional facial morphology identifies a variant in Pax3 associated with nasion position. American Journal of Human Genetics 90:478-485.
Rockman MV, Wray GA. 2002. Abundant raw material for cisregulatory evolution in humans. Molecular Biology and Evolution 19:1991-2004.

Sandelin A, Carninci P, Lenhard B, et al. 2007. Mammalian RNA polymerase II core promoters: insights from genomewide studies Nature Reviews Genetics 8:424-436.
Sawaya SM, Lennon D, Buschiazzo E, Gemmell N, Minin VN. 2012. Measuring microsatellite conservation in mammalian evolution with a phylogenetic birth-death model. Genome Biology and Evolution 4:636-647.
Singh DP, Bhargavan B, Chhunchha B, et al. 2012. Transcriptional protein Sp 1 regulates LEDGF transcription by directly interacting with its cis-elements in GC-rich region of TATA-less gene promoter. PLoS ONE 7:e37012.
Trivedi S, Hancock JM. 2012. Low microsatellite frequencies in neuron and brain expressed microRNAs. Gene 508: 73-77.
Valipour E, Kowsari A, Bayat H, et al. 2013. Polymorphic core promoter GA-repeats alter gene expression of the early embryonic developmental genes. Gene 531:175-179.
Verstrepen KJ, Jansen A, Lewitter F, Fink GR. 2005. Intragenic tandem repeats generate functional variability. Nature Genetics 37:986.
Vinces MD, Legendre M, Caldara M, Hagihara M, Verstrepen KJ. 2009. Unstable tandem repeats in promoters confer transcriptional evolvability. Science 324:1213-1216.
Vowles EJ, Amos W. 2006. Quantifying ascertainment bias and species-specific length differences in human and chimpanzee microsatellites using genome sequences. Molecular Biology Evolution 23:598-607.
Wang Y, Rekaya R. 2009. A comprehensive analysis of gene expression evolution between humans and mice. Evolutionary Bioinformatics Online 5:81-90.
Yang C, Bolotin E, Jiang T, Sladek FM, Martinez E. 2007. Prevalence of the initiator over the TATA box in human and yeast genes and identification of DNA motifs enriched in human TATA-less core promoters. Gene 389:52-65.
ZarifYeganeh M, Ghaffarpour M, Farhud DD, et al. 2009. Skew in the human caveolin 1 gene upstream purine complex homozygote haplotype compartment in multiple sclerosis. Journal of Neuroimmunology 216:103-107.
Zarif Yeganeh M, Mirabzadeh A, Khorram Khorshid HR, et al. 2010. Novel extreme homozygote haplotypes at the human caveolin 1 gene upstream purine complex in sporadic Alzheimer's disease. American Journal of Medical Genetics B Neuropsychiatric Genetics 153B:347-349.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.


[^0]:    Abbreviations: SNP, single nucleotide polymorphism; STR, short tandem repeat; TF, transcription factor; TSS, transcription start site.

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    *Correspondence to: Mina Ohadi, Genetics Research Center, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran. E-mail: ohadi.mina@yahoo.com
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[^1]:    ${ }^{\text {a }}$ The average frequency of STR motifs in the -120 to +1 TSS is presented for each motif. If the motif has been repeated more than once in a STR formula, the frequency has been re-stated as well.

