# Utah State University DigitalCommons@USU

All Graduate Theses and Dissertations

**Graduate Studies** 

5-1990

# Autecological Characteristics of Chrysopogon aucheri and Cymbopogon jwarancusa, Dominant Rangeland Grasses in Baluchistan

Mohammad Saleem Utah State University

Follow this and additional works at: https://digitalcommons.usu.edu/etd

Part of the Plant Sciences Commons

## **Recommended Citation**

Saleem, Mohammad, "Autecological Characteristics of Chrysopogon aucheri and Cymbopogon jwarancusa, Dominant Rangeland Grasses in Baluchistan" (1990). *All Graduate Theses and Dissertations*. 6464.

https://digitalcommons.usu.edu/etd/6464

This Dissertation is brought to you for free and open access by the Graduate Studies at DigitalCommons@USU. It has been accepted for inclusion in All Graduate Theses and Dissertations by an authorized administrator of DigitalCommons@USU. For more information, please contact digitalcommons@usu.edu.



## AUTECOLOGICAL CHARACTERISTICS OF <u>CHRYSOPOGON</u>

## AUCHERI AND CYMBOPOGON JWARANCUSA, DOMINANT

## RANGELAND GRASSES IN BALUCHISTAN

by

Mohammad Saleem

## A dissertation submitted in partial fulfillment of the requirements for the degree

of

## DOCTOR OF PHILOSOPHY

in

Range Science

Approved:

UTAH STATE UNIVERSITY Logan, Utah

## DEDICATION

To the memory of my father, the late Salim Khan. To my mother, Syeeda-Lo, and my brother, Mohammad Shah, for their love and support, and to my teachers, for their labor and encouragement.

#### ACKNOWLEDGEMENTS

A very special thanks is extended to Dr. Christopher A. Call for his friendly advice, patience, thoughtful guidance, and encouragement that provided the inspiration and culmination of this dissertation. I thank the Pakistan Agriculture Research Council, the Food and Agricultural Organization (FAO), and the Baluchistan Forest Department for their financial support of myself and my family.

Acknowledgement is made of the constructive comments and guidance of Dr. Dick Fisher, Dr. Doug Johnson, Dr. Dave Pyke, Dr. Fred Provenza, and Dr. William Campbell of my graduate committee. I thank them for their support in fulfilling this assignment. I thank Dr. Ben Norton for his initial help and encouragement in this research. Dr. Donald Sisson, and Jane Post were very helpful with statistical analyses.

Sincere appreciation is extended to Diana Thimmes for preparing the final draft of this manuscript.

I am grateful for the affection and moral support of Suzann Winn and her family; Lucy Thompson, International Programs; and Teri Price, Range Science Department. Appreciation is also extended to the many graduate students from Pakistan who from time to time helped me with this study. I am indebted to the families of International Muslim Students for arranging food during the months of Ramazan during my stay. Finally I extend a special acknowledgement to my mother, brother, and sister for their love and support and to my wife and children for their love.

### Mohammad Saleem

## TABLE OF CONTENTS

																		Page
DEDIC	ATION			•	•	•	•	•										ii
ACKNO	WLEDGEMENTS	• • •																iii
LIST	OF TABLES .			•				•										vii
LIST	OF FIGURES																	viii
ABSTR	ACT							• •										xv
CHAPT	ER																	
I.	INTRODUCT	ION																1
	Objective	s.					• •											3
II.	ECOLOGY O JWARANCUS	F <u>CH</u> A:	RYS GEF	SOF RMI	POG NA	<u>on</u> TI	<u>AU</u> ON	JCH RF	IER		ANI SE	<u>o</u> c	CYN	<u>1B0</u>	OPO	DGO	<u>NC</u>	5
	Summary .										•			•	•		•	5
	Introduct Materials	ion and	Me	eth	od	s	• •	:	•	•	•	•	•	•	•	•	:	5 7
	Results . Discussion	n .	•	•	•	•	•••	•	:	•	•	•	•	•	•	•	•	9 11
III.	ECOLOGY O	F <u>CH</u>	RYS	SOP	OG	<u>on</u>	AU	ICH	ER	IZ	ANI		CYM	IBC	PC	G	DN	
	JWARANCUS	<u>A</u> : SI	EEC	LI	NG	DI	EVE	LO	PM	ENT	C	•	•	•	•	•	•	18
	Summary . Introduct:	 ion	•	•	•	•	• •	•	•	•	•	•	•	•	•	•	•	18
	Materials	and	Me	th	od	s.							:			:	•	22
	Results .	• •		•		• •												24
	Discussion	n .	•	•	•	• •	• •	•	•	•	•	•	•	•	•	•		33
IV.	ECOLOGY OF JWARANCUSA:	CHRY		PO	<u>GO1</u>		AUC	HE D	RI DFI	AN	ID	CY	MB	OF	200	ON	I	
	RESPONSE .	• •	•	•	•	• •		•	•	•	•	•	•					40
	Summary .																	40
	Introductio	n.	•		• •	• •	•	•	•		•	•	•	•	•			41
	Results a	ind M	let	noo	ds	•	•	٠	•	•	•	•	•	•	•	•	•	44
	Discussion	•••	•	:	• •	•	•	•	•	•	•	•	•	•	•	•	•	4/
		2 C C C C C C C C C C C C C C C C C C C			-		-	-	-	-	-	-	-		-			55

v

## CHAPTER

CHAPIER												Page
V. SYNTHESIS	•	•	•	•	•	•						68
LITERATURE CITED				•								72
APPENDIX	•	•	•	•								79
VITA												82

## LIST OF TABLES

Table		Page
1.	Cumulative germination (%) of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) seeds as influenced by six alternating temperature regimes. Germination values are not adjusted for seed viability	10
2.	Cumulative germination (%) of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) seeds as influenced by six alternating temperature regimes. Germination values are adjusted for seed viability	12
3.	Mean germination time (days) of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) as influenced by six alternating temperature regimes	13
4.	Mean of proportion (% by weight <u>+</u> SD) of different root types at different stages of seedling development for <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwarancusa</u> (Cy) .	34
5.	Mean monthly maximum and minimum temperature and monthly precipitation data for Quetta, Baluchistan	80
6.	Heavy defoliation (85 % removal) regimes for monocultures and 50:50 mixtures of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwarancusa</u> (Cy) .	81

vii

#### Figure Page Grass seedling root morphology (from Newman 1. and Moser 1988) . . . . . . . . . . . . 21 . . 2. Mean Haun stage (leaf development) of Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) seedlings in relation to days after emergence and cumulative growing degree days (CGDD). Values for species at each time interval with different letters are significantly different $(LSD_{05} = 2.45, n = 10)$ . . . . . . . . . . . . 25 Mean tiller number of Chrysopogon aucheri (Ch) 3. and Cymbopogon jwarancusa (Cy) seedlings in relation to days after emergence and cumulative growing degree days (CGDD). Values for species at each time interval with different letters are significantly different (LSD<sub>05</sub> = 0.87, n = 10). 26 Mean total shoot (A) and root (B) dry weight (mg) 4. and shoot/root ratio (C) of Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) in relation to time (days) after emergence with different letters are significantly different $(LSD_{05} = 23.6 \text{ for A}, 19.4 \text{ for B}, \text{ and } 0.63 \text{ for}$ 28 $C; n = 10) \dots \dots$ Mean primary root number (main root and rootlets 5. > 2 mm) (A), root length (cm) (B), and root dry weight (mg) (C) of Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) in relation to time (days) after emergence. Values for species at each time interval with different letters are significantly different (LSD<sub>.05</sub> = 15.5 for A, 35.9 for B, 30 Mean seminal root number (main root and rootlets 6. > 2 mm) (A), root length (cm) (B), and root dry weight (mg) (C) of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and Cymbopogon jwarancusa (Cy) in relation to time (days) after emergence. Values for species at each time period with different letters are significantly different $(LSD_{05} =$

23.3 for A, 44.9 for B, and 4.2 for C; n = 10). 31

- 8. Defoliation treatments for <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> grown in monocultutre and mixture. 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; 46
- 10. Mean number of tillers per plant for <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) without clipping. Values for species with different letters within monoculture or mixture at each time interval are significantly different (LSD<sub>05</sub> = 5.8; n = 8). . 49

ix

Page

- Mean number of tillers per plant for Chrysopogon 11. aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): (A) 0 clip (control); (B) 1 clip = standing crop clipped at 32 weeks and regrowth at 44 weeks (final harvest) after emergence; (C) 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; (D) 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters within monoculture or mixture at each time interval are significantly different [LSD<sub>05</sub> (A, B, C, and D) = 10.1 for 32 weeks, 12.0 for 36 weeks, and 12.8 for 40 and 44 weeks; n = 4].... 51
- Mean shoot dry weight (g) of 12. Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): (A) 0 clip = (control) = standing crop clipped at 44 weeks (final harvest) after emergence; (B) 1 clip = standing crop clipped at 32 weeks and regrowth at 44 weeks after emergence; (C) 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, and 44 weeks after emergence; (D) 3 clip = standing crop clipped at 32 weeks and regrowth at 36, 40 and 44 weeks after emergence. Values with different letters within monoculture or mixture at each time interval are significantly different [LSD.05 (A, B, and C and D) = 0.61 for 32 weeks, 0.36 for 36 weeks, 0.22 for 40 weeks, and 0.35 for 52

Page

- Above-ground dry weight (g) (cumulative shoot 13. dry weight above 3-cm stubble height and crown dry weight below 3-cm) from 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes: 0 clip (control) = standing crop above 3-cm and crown below 3-cm clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth and crown clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks, and regrowth clipped at 36 weeks, and regrowth and crown clipped at 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks, regrowth clipped at 36 and 40 weeks, and regrowth and crown clipped at 44 weeks after emergence. Above 3-cm and below 3-cm dry weight values for species with different letters within monoculture or mixture under each clipping regime are significantly different  $(LSD_{05} = 1.7 \text{ for above})$ 3-cm, and .88 for below 3-cm; n = 4) . . . .
  - Mean root dry weight (g) harvested from 14. 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks, and regrowth clipped at 44 weeks after emergence, 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40 and 44 weeks after emergence. Values for species with different letters within monoculture or mixture under each clipping regime are significantly different (LSD<sub>05</sub> = 1.7; 55 . . . . . . .

Page

- 15. Mean percent crude protein (A) and percent in vitro digestible dry matter (% IVDMD) (B) for Chrysopogon aucheri (Ch) and Cymbopogon <u>iwarancusa</u> (Cy) plants grown in monoculture under different clipping regimes (3-cm stubble height): 1 clip = initial standing crop clipped at 32 weeks after emergence; 2 clip = regrowth clipped at 36 weeks after emergence; 3 clip = regrowth clipped again at 40 weeks after emergence; and F clip = final regrowth clipped at 44 weeks after emergence. Values for species with different letters under each clipping regime are significantly different (LSD<sub>05</sub> = 2.0 for A, 5.3 for B; n = 4) . . . . . . . . .
- 16. Mean number of tillers per plant for Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represent a treatment where the species indicated is clipped to a 3-cm stubble height and the associate species is not clipped): (A) 0 clip (control), (B) 1 clip = standing crop of either species clipped at 32 weeks after emergence; (C) 2 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 weeks after emergence; and (D) 3 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 and 40 weeks after emergence. Values for species with different letters at each time interval are significantly different  $(LSD_{05} (A, B, C \text{ and } D) = 10.0 \text{ for } 32$ weeks, 12.0 for 36 weeks and 12.8 for 40 and 44

57

Page

Mean shoot dry weight (g) of 17. Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): (A) 1 clip= standing crop of either species clipped at 32 weeks and regrowth clipped at 44 weeks (final harvest) after emergence; (B) 2 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 and 40 weeks after emergence; and (C) 3 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36, 40 and 44 weeks after emergence. Values for species with different letters at each time interval are significantly different [LSD<sub>05</sub> (A, B, and C) = 0.61 for 32 weeks, 0.36 for 36 weeks, 0.22 for 40 weeks, and 0.53 for 44 weeks; n = 4] . . . . . . . . . . . . . . . . . . .

18. Above-ground dry weight (g) (cumulative shoot dry weight above 3-cm stubble height and below 3-cm) from 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): 0 clip (control) = standing crop above 3-cm and crown below 3-cm clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks, and regrowth and crown clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks, regrowth clipped at 36 weeks, and regrowth and crown clipped at 44 weeks after emergence. Values for species above 3-cm and below 3-cm with different letters under each clipping regime are significantly different (LSD<sub>.05</sub> = 1.7 for above 3-cm, and .88 for below 3-cm; n = 4) 

Page

58

Mean root dry weight (g) harvested from 19. 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, and 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36,40 and 44 weeks after emergence. Values for species with different letters under each clipping regimes are significantly different (LSD<sub>.05</sub> = 1.70; n = 4) . 61

Page

#### ABSTRACT

Autecological Characteristics of <u>Chrysopogon</u> <u>aucheri</u> and <u>Cymbopogon jwarancusa</u>, Dominant Rangeland Grasses in Baluchistan

by

Mohammad Saleem, Doctor of Philosophy Utah State University, 1990

Major Professor: Dr. Christopher A. Call Department: Range Science

Controlled environment experiments were designed to study the germination, seedling development, and defoliation responses of <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> to better understand their autecology and potential use in range improvement programs in Baluchistan.

In experiment 1, <u>Cymbopogon jwarancusa</u> had greater seed fill and viability than <u>Chrysopogon aucheri</u>. When incubated at six different alternating temperature regimes, seeds of <u>Cymbopogon jwarancusa</u> had greater cumulative germination at five temperature regimes and faster germination at the colder temperature regimes than <u>Chrysopogon aucheri</u>.

In experiment 2, seedling shoot and root development was characterized at 15-day intervals over a 60-day

period. Seedlings of both species had a "panicoid" type seedling morphology. Chrysopogon aucheri and Cymbopogon warancusa developed comparable numbers of leaves and tillers per plant during the 60-day period. Chrysopogon aucheri had a greater number, length, and dry weight of primary and seminal roots than Cymbopogon jwarancusa at 30 and 60 days, respectively. Adventitious root length was also higher for Chrysopogon aucheri than Cymbopogon jwarancusa at 60 days. Seedlings of both species had similar shoot:root ratios and relative growth rates. In experiment 3, seedlings of both species were planted in monocultures and in a 50:50 mixtures. Defoliation treatments, implemented 32 weeks after emergence, included: equally clipping all plants of both species zero, one, two, or three times (at 4-week intervals) in monoculture and mixture; and clipping one species zero, one, two, or three times (at 4-week intervals) without clipping the associated species in mixture. Both species remained vegetative and did not differ in leaf and tiller development until about 32 weeks after emergence. During later growth, <u>Chrysopogon aucheri</u> reproduced while Cymbopogon jwarancusa remained vegetative. Cymbopogon jwarancusa produced more tillers on control plants and defoliated plants (mainly in monoculture). At lower frequencies of defoliation Chrysopogon aucheri produced more shoot and root biomass than Cymbopogon jwarancusa

xvi

(mainly in mixture). In 50:50 mixtures when one species was defoliated and the other not, both species were comparable in shoot dry weight; however, <u>Chrysopopgon</u> <u>aucheri</u> was superior to <u>Cymbopogon jwarancusa</u> in root dry weight at all defoliation regimes. The initial standing crop and subsequent regrowth of <u>Chrysopogon aucheri</u> were comparable or higher in crude protein and digestibility than <u>Cymbopogon jwarancusa</u>.

(99 pages)

#### CHAPTER I

#### INTRODUCTION

Baluchistan, the largest province of Pakistan, has a land area of 347,193 km<sup>2</sup> that comprises 43% of the total area of the country. The climate is arid to semiarid, with annual precipitation ranging from 50 mm in the western part of the province to 450 mm in the northeastern part (Baluchistan Agriculture Department 1985). Maximum and minimum temperatures range from 50 °C in summer to -10 °C in winter. The length of the spring and summer growing season depends upon the pattern and occurrence of precipitation, which can be quite variable (Baluchistan Forest Department 1986). Topographically, the entire province has small mountain ranges and upland valleys with elevations ranging from 1,500 to 2,438 m. Soils are mostly skeletal and are derived from limestone, sandstone, and shale (FAO 1981). Rangeland comprises 94% of the land area of Baluchistan. Cymbopogon and Chrysopogon grasslands constitute 50 % of the range area of the province (Baluchistan Forest Department 1986).

Historically, <u>Cymbopogon</u> and <u>Chrysopogon</u> grasslands originated from woodland ecosystems as a result of deforestation, shifting cultivation, and burning (Singh et al. 1985). They are maintained at a subclimax stage (Clements 1928) by repeated burning and grazing. These grasslands are the major source of forage for local and nomadic herds of livestock (sheep, goats, camels, and cattle) and are grazed continuously either season-long or year-long. Overgrazing has reduced the canopy cover, increased soil erosion potential, and allowed the invasion of undesirable species (Debadgahoa and Shankarnarayan 1973). Rangelands now produce 10 to 50 % less than their potential (Khan 1971), which ranges from less than 30 kg/ha per year to 280 kg/ha per year (FAO 1981).

Chrysopogon aucheri and Cymbopogon jwarancusa, the dominant, perennial, C<sub>4</sub> bunchgrasses on Baluchistan grasslands, have been found growing on a wide variety of soils over a wide range of elevations. <u>Chrysopogon aucheri</u> is readily grazed, whereas <u>Cymbopogon jwarancusa</u> is grazed only after the former has been heavily grazed. The low palatability of <u>Cymbopogon jwarancusa</u> has been related to the oil content in its leaves (West Pakistan Forest Department 1960). As the grasslands deteriorate under heavy grazing pressure, <u>Chrysopogon aucheri</u> is gradually replaced by <u>Cymbopogon jwarancusa</u>, which in turn is gradually replaced by the relatively less palatable shrub, <u>Artemisia maritima</u>.

#### Objectives

Little is known about the ecology of these grasslands (Khan 1971), and few range improvement programs have been initiated in Baluchistan (Baluchistan Forest Department 1986). To better understand plant requirements for "prudent" grazing, one must understand the morphology, physiology, and long-term competitive abilities of these forage grasses (West 1968, Caldwell 1984). Because these grasses have grown in close association and reproduced under continuous grazing pressure for centuries, one may hypothesize that these two grass species are similar in germination, seedling establishment, defoliation response, and nutritional quality (except for oil content) when grown under similar growing conditions.

Due to a lack of understanding about the autecology of <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u>, studies were designed to investigate their germination, seedling establishment, defoliation response, and nutritional quality. Chapter II examines the germination responses of the two range grasses under different alternating temperature regimes. Chapter III addresses comparative seedling establishment under similar controlled growth conditions. Chapter IV characterizes morphological development, defoliation response, nutritional status, and

digestibility of these two bunchgrass species. Chapter V integrates the findings of the experiments and provides suggestions for management applications and future research.

#### CHAPTER II

# ECOLOGY OF <u>CHRYSOPOGON</u> <u>AUCHERI</u> AND <u>CYMBOPOGON</u> <u>JWARANCUSA</u>: GERMINATION RESPONSE

#### Summary

The ability of Cymbopogon jwarancuas to displace Chrysopogon aucheri on Baluchistan grasslands may be related to recruitment potential in addition to differences in palatability. A controlled environment study was designed to investigate the effects of different temperature regimes on germination responses of these two dominant range grasses. Cumulative germination and rate of germination (mean germination time) were evaluated at six alternating temperature regimes (5/10, 5/15, 5/20, 10/20, 10/25 and 10/30 °C,12 h night/12 h day) representing possible temperatures in western Baluchistan during the recruitment period (March - May). In addition to having greater seed fill and viability, Cymbopogon jwarancusa had greater and more rapid germination than Chrysopogon aucheri at a wider range of temperatures, especially cooler temperatures.

## Introduction

Plant establishment by seedling recruitment, the dominant form of regeneration for most species on rangelands, is only successful when plant requirements for seed germination, seedling development, and subsequent growth are matched with microenvironmental factors of the seedbed (Grubb 1977, Harper 1977). Variation in seed germination between species and between ecotypes of the same species have been attributed to differences in seed development, dormancy mechanisms, seed size, distribution of safe sites and differential responses to temperature, water, light, and gas exchange conditions (Koller 1972, Mayer and Poljakoff-Mayber 1982, Peart 1979, 1984, Fowler 1988).

Limited information is available on the germination requirements of Chrysopogon aucheri and Cymbopogon jwarancusa in Pakistan and of related C, species in other regions of the world. Production of viable, germinable seed has been related to inflorescence development in Chrysopogon aucheri (Hussain et al. 1980), and in Cymbopogon jwarancusa, Cymbopogon parkeri, and Cymbopogon oliveri (Ahmed et al. 1978). Ahmed et al. (1978) also observed that Cymbopogon jwarancusa had the highest cumulative germination and most rapid germination rate of the three Cymbopogon species. In Australia, Mott (1978) found that high temperature storage enhanced the germination of Chrysopogon fallax and Chrysopogon latifolius. Rai et al. (1980) observed that older (stored) seed of Chrysopogon fulvus had a higher germination capacity than freshly harvested seed, indicating an after-

ripening effect. Ghosh and Chatterjee (1981) used several chemical and physical treatments to break the dormancy of seed of <u>Cymbopogon flexuous</u> and <u>Cymbopogon maritime</u>. Primary dormancy was relieved by exposure to continuous light and by treatment with gibberellic acid, kinetin, or potassium nitrate after a period of storage. Secondary dormancy was relived by exposure to low temperature.

The germination studies with <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> did not simulate the range of temperature and moisture conditions to which seeds of both species are exposed during the germination/recruitment period (March - May) of the growing season in western Baluchistan. The objective of this study was to investigate the effects of different alternating temperature regimes on germination responses of seeds of <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> collected in the Quetta Region of Baluchistan.

### Materials and Methods

Seeds (caryopsis and attached lemma and palea) of <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> were collected from three protected sites near Quetta, Baluchistan during June and July, 1987. Prior to germination trials in 1988, seeds of both species from the three sites were tested for a caryopsis, and for viability. Five replicates of 100 randomly selected seeds from each

site were examined for a caryopsis by removing the lemma and palea. Four replicates of 50 caryopsis from each site were placed in a 1 % triphenyl tetrazolium chloride solution for 24 h at 22 °C in complete darkness to determine viability (Grabe 1970). Percent viability was determined by evaluating intensity of staining and staining patterns under a 10 x lens. Due to low seed fill and viability of <u>Chrysopogon aucheri</u>, seed from the three sites were combined into one lot. Seed of <u>Cymbopogon jwarancusa</u> were also combined into one lot.

In a controlled environment, four replicates of 50 filled seeds of each species were exposed to alternating temperature regimes (12 h night/12 h day, light intensity of 250  $\mu$  mol. m<sup>-2</sup>. sec<sup>-1</sup> during day period) of 5/10, 5/15, 5/20, 10/20, 10/25, and 10/30 °C, which simulated possible temperature regimes on rangelands near Quetta, Baluchistan from March through May (Table 5 in Appendix). Seeds were placed on two layers of Whatman No.1 filter paper (saturated with distilled water when necessary) in Petri dishes. Petri dishes were wrapped in polyethylene film to reduce evaporation losses and stabilize relative humidity. A seed was considered germinated when it had a radicle greater than 2 mm. Germinated seeds were counted and removed from Petri dishes every day over a 25-day period, and cumulative germination data were reported as a percentage of the total number of filled seeds in each

dish. Germination rates were estimated by calculating mean germination time, i.e. the mean time in days taken for nondormant, for viable seeds to germinate (Ellis and Roberts 1978).

The experiment was arranged in a completely randomized design with four replications per treatment for each species. Cumulative germination percentage and mean germination time data were subjected to analysis of variance, and means were separated by Fisher's least significant difference test (P < 0.05 level of significance). Cumulative germination percentage data were transformed prior to analysis using an arcsine transformation.

## Results

Considerable variability in caryopsis fill, viability, and germination was observed between the two species. Chrysopogon aucheri and Cymbopogon jwarancusa respectively had means of 25 (SD  $\pm$  4.4) and 47 % (SD  $\pm$  2.3) for seed fill and means of 6 (SD  $\pm$  1.6) and 87 % (SD  $\pm$  2) for seed viability. Germination did not occur in both species at the coldest temperature regime (5/10 °C) (Table 1). Cymbopogon jwarancusa had limited germination at 5/15 °C while Chrysopogon aucheri did not initiate germination until 5/20 °C. Germination increased with increasing temperature for both species, reaching a maximum value at

Table 1. Cumulative germination (%) of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) seeds as influenced by six alternating temperature regimes. Germination values are not adjusted for seed viability.

Temperature	Germinat	ion (%)
regime (°C)	Ch	Су
5/10	0.0 a x	0.0 e x
5/15	0.0 a x	12.0 d y
5/20	2.0 a x	63.0 c y
10/20	5.5 a x	86.5 a y
10/25	0.9 a x	65.5 c y
10/30	4.9 a x	74.0 b y

Mean in the same column followed by different letters (a through e) and in the same row followed by different letters (x through y) are significantly different (  $LSD_{.05} = 7.2$ ).

10/20 °C. Germination for both species declined as temperatures increased from 10/20 to 10/25 °C and then increased as temperatures increased from 10/25 to 10/30 °C. <u>Cymbopogon jwarancusa</u> germination was significantly greater than that of <u>Chrysopogon aucheri</u> at all temperature regimes ranging from 5/15 to 10/30 °C when differences in viability were not considered in the calculation of cumulative germination. However, when the low viability (6%) of <u>Chrysopogon aucheri</u> seeds was considered, cumulative germination was not significantly different between the two species at the optimum temperature range (10/20 °C) and at 10/30 °C (Table 2). Almost all the viable seeds of each species germinated at 10/20 °C.

Mean germination time was significantly slower for both species at colder temperature regimes (5/15 °C for <u>Cymbopogon jwarancusa</u> and 5/20 °C for <u>Chrysopogon aucheri</u>) (Table 3). Mean germination time of <u>Cymbopogon jwarancusa</u> seeds was slightly faster than that of <u>Chrysopogon aucheri</u> seeds at temperature regimes with the greatest cumulative germination (5/20 to 10/30 °C); however, this difference was not significant.

## Discussion

Differences in one or several environmental factors (temperature, moisture, light, oxygen, etc.) can result in

Table 2. Cumulative germination (%) of <u>Chrysopoqon</u> <u>aucheri</u> (Ch) and <u>Cymbopoqon jwarancusa</u> (Cy) seeds as influenced by six alternating temperature regimes. Germination values are adjusted for seed viability.

Temperature	Ge	<u>Germination (</u>								
regime (°C)	Ch	ı	C	У						
 					_					
5/10	0.0	d x	0.0	е	x					
5/15	0.0	d x	13.8	d	х					
5/20	33.3	bх	72.4	с	У					
10/20	91.6	a x	99.4	a	x					
10/25	16.6	сх	75.3	С	У					
10/30	83.3	a x	85.1	b	x					

Means in the same column followed by different letters (a through e) and in the same row followed by different letters (x through y) are significantly different (LSD<sub>.05</sub>=9.5).

Table 3. Mean germination time (days) of <u>Chrysopogon</u>

<u>aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) as influenced by six alternating temperature regimes.

Temperature	<u>Mean germination t</u>	<u>ime (days)</u>
regime (°C)	Ch	Су
and the second	and the second state of th	
5/10		
5/15		22.1 a x
5/20	17.6 a x	9.8 b y
10/20	10.3 b x	7.7 b x
10/25	9.0 b x	5.0 b x
10/30	9.1 b x	5.5 b x

Means in the same column followed by different letters (a through b) and in the same row followed by different letters (x through y) are significantly different (LSD<sub>.05</sub> = 7).

significantly different germination responses of wildland grass species (Ellern and Tadmor 1967, Dwyer and Wolde-Yohannis 1972, Young et al. 1973). Day-night temperature alterations are the normal condition in the field and are required for appreciable germination in many species (Koller 1972). If one temperature is within the upper or lower inhibitory range, the inhibitory effect may be nullified by the other alternating temperature (Gulliver and Hydecker 1973). In the colder temperature regimes, (5/10 to 5/20 °C) the 5 °C low temperature appeared to be inhibitory to the germination of Cymbopogon jwarancusa until the upper temperature reached 15 °C (5/15 °C temperature regime) and to the germination of Chrysopogon aucheri until the upper temperature reached 20 °C (5/20 °C temperature regime) (Tables 1 and 2). Cumulative germination increased significantly for Cymbopogon jwarancusa as the upper temperature reached 20 °C (5/20 °C temperature regime). Thus, considerable germination can occur in cold fluctuating temperature regimes if the soil surface temperature approaches 20 °C during the daytime and moisture is available.

The decline in cumulative germination in the 10/25 and 10/30 °C temperature regimes (Table 1) and the slight

increase in mean germination time in the 10/30 °C temperature regime for both species may be attributed to seed variability. The low viability (6%) of filled <u>Chrysopogon aucheri</u> seed made it difficult to consistently place a representative number of viable seeds in Petri dishes for the different germination treatments.

The small amount of literature available on the germination characteristics of <u>Chrysopogon</u> and <u>Cymbopogon</u> species from Pakistan and various range regions supports the findings of this study. <u>Cymbopogon</u> species appear to have higher germinability than <u>Chrysopogon</u> species under a variety of environmental conditions (Mott 1978, Rai et al. 1980, Ghosh and Chatterjee 1981). In one study (Ahmed et al. 1978), <u>Cymbopogon jwarancusa</u> had 99 % germination after 14 days in a 20 - 25 °C temperature regime.

The production of viable, germinable seed by <u>Cymbopogon jwarancusa</u> and <u>Chrysopogon aucheri</u> in Pakistan has been primarily related to inflorescence development. Ahmed et al. (1978) indicated that a relatively high percentage (> 50 %) of sterile florets contributed to low viable seed set (32 % viable seed) in a population of <u>Cymbopogon jwarancusa</u>. Hussain et al. (1980) reported that <u>Chrysopogon aucheri</u> had 60 % sterile florets, and considered this sterility value to be a reliable indicator of poor seed set and germination in this species. In both field nursery studies (Ahmed et al. 1978, Hussain et al.

1980), floret sterility was determined morphologically by quantifying the presence of male, female, bisexual, or barren florets. Female florets were absent in both <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u>; therefore, only bisexual florets were considered capable of seed set. Since nursery growing conditions were relatively more favorable than the field conditions, floret sterility and seed set were probably more influenced by genetic rather than environmental factors.

Seed fill, viability, and germination data from this study generally support the findings of Ahmed et al. (1978) and Hussain et al. (1980). Cymbopogon jwarancusa and Chrysopogon aucheri had 47 and 25 % caryopsis fill, respectively, and filled seeds had 87 and 6 % viability, respectively. It appears that Chrysopogon aucheri has an inherently lower potential for producing viable, germinable seed than Cymbopogon jwarancusa. Based upon seed fill, and viability data, if 1,000 randomly selected seeds of each species were sown under optimum conditions, only 15 Chrysopogon aucheri seeds would have the potential to germinate whereas 409 Cymbopogon jwarancusa seeds could potentially germinate.

The ability of <u>Cymbopogon jwarancusa</u> to displace <u>Chrysopogon aucheri</u> on Baluchistan grasslands has been primarily related to differences in palatability (West Pakistan Forest Department 1960). In addition to being

less palatable than <u>Chrysopogon aucheri</u>, it appears that <u>Cymbopogon jwarancusa</u> also has an advantage in recruitment potential. Results demonstrate that <u>Cymbopogon jwarancusa</u> seed not only have higher seed fill, and viability, they also have the capacity for greater germination than <u>Chrysopogon aucheri</u> over a wide range of alternating temperatures (Tables 1 and 3). This may allow <u>Cymbopogon</u> <u>jwarancusa</u> to germinate in a wide variety of seedbed microsites with different temperature regimes when transient soil moisture is available, especially during cooler temperatures in March (Table 5 in Appendix).

and adventitious root development between sendlings of Chrysonogon mucheri and Cymborogon iwarancuss at 15, 30, 45, and 60 days after emergence. In general, seedlings of both species were comparable in terms of shoot and root development over the 60-day growing period. Relative growth rates of total seedling biomass were 0.061 and 0.065 mg. mg<sup>-1</sup>, day<sup>-1</sup>, respectively, for <u>Chrysopogon Aucheri</u> and <u>Cymborogon iwarancuss</u>. Both species showed evidence of subcoleoptile internois elongation and subcoleoptils internode root development characteristic of "panicold" type seedlings.

#### CHAPTER III

## ECOLOGY OF <u>CHRYSOPOGON AUCHERI</u> AND <u>CYMBOPOGON</u> JWARANCUSA: SEEDLING DEVELOPMENT

#### Summary

A description of shoot and root morphology is essential for understanding the seedling establishment process in grasses. A controlled environment study, using an 80 % washed sand and 20 % loam soil growing medium, was conducted to determine differences in leaf and tiller development, and primary, subcoleoptile internode, seminal, and adventitious root development between seedlings of Chrysopogon aucheri and Cymbopogon jwarancusa at 15, 30, 45, and 60 days after emergence. In general, seedlings of both species were comparable in terms of shoot and root development over the 60-day growing period. Relative growth rates of total seedling biomass were 0.061 and 0.068 mg. mg<sup>-1</sup>. day<sup>-1</sup>, respectively, for <u>Chrysopogon aucheri</u> and Cymbopogon jwarancusa. Both species showed evidence of subcoleoptile internode elongation and subcoleoptile internode root development characteristic of "panicoid" type seedlings.
### Introduction

Germination and seedling development are critical stages in the establishment of perennial grasses on arid and semiarid rangelands. Frequently, competitive advantages gained during the seedling stage are maintained in the mature plant stage (Coyne and Bradford 1985). High success in seedling establishment is often associated with rapid root and shoot growth, a robust growth habit, and resistance to environmental stress (McKell 1972). In dry regions, rapid seedling root elongation allows roots to grow along the descending moisture front in subsurface soils (Milthorpe 1950, Harris 1967, Buckly 1982, Simanton and Jordan 1986).

Seedling development and morphology have been used to classify grasses and to provide a basis for understanding the establishment processes in grasses. Hoshikawa (1969) classified 219 species from 88 genera into six seedling types based on root morphology and observed that nearly all species of the same genus were of the same seedling type. More recently, Newman and Moser (1988) described the seedling root morphology of nine cool-season ( $C_3$ ) and nine warm-season ( $C_4$ ) perennial forage grasses commonly used in the northern USA. In both studies (Hoshikawa 1969, Newman and Moser 1988), most of the warm-season grasses

("panicoid" seedling type) had an elongated subcoleoptile internode with subcoleoptile internode root development (Fig. 1). The cool-season grasses ("festucoid" seedling type) had little or no subcoleoptile internode elongation and had seminal root development (Fig. 1). Adventitious roots originate at the base of the coleoptile, which is at the depth of seeding for festucoid seedlings and is placed near the soil surface by subcoleoptile internode elongation for panicoid seedlings. Hoshikawa (1969) examined one <u>Cymbopogon</u> species (species unknown) and found it was a panicoid-type seedling.

Dry surface soil prevents the development of adventitious roots, which become the major root system of established plants in many panicoid grasses (Tischler and Voigt 1987). Primary and subcoleoptile internode roots cannot supply sufficient water to keep up with the increasing evaporation demand of the growing shoot (Hyder et al. 1971, Wilson et al. 1976), resulting in seedling mortality and unsuccessful seedlings.

Information is lacking on seedling development and morphology of dominant  $C_4$  forage grasses in Baluchistan. This study was designed to characterize the seedling root and shoot development of <u>Chrysopogon aucheri</u> and <u>Cymbopogon</u> <u>jwarancusa</u> in an effort to better understand their establishment requirements.



Figure 1. Grass seedling root morphology (from Newman and Moser, 1988).

### Materials and Methods

Chrysopogon aucheri and Cymbopogon jwarancusa seeds, collected near Quetta, Baluchistan in summer 1987, were stored at 20 °C and 40 % relative humidity until May, 1989 for this experiment. Due to the low and slow germination of <u>Chrysopogon aucheri</u>, approximately 10,000 seeds were sown on two layers of paper towels in aluminum trays (16 x 33 x 2 cm) to obtain 60 germinated seeds at the same time. Ten days later, 100 seeds of Cymbopogon jwarancusa were sown on two layers of Whatman No. 1 filter paper in Petri dishes to obtain 60 germinated seeds at the same time. Paper media in trays and dishes were saturated with distilled water when necessary, and trays and dishes were covered with polyethylene film to reduce evaporative losses and stabilize relative humidity. Trays and dishes were kept in a growth room with a night/day temperature regime of 2)/25 °C and a 12-h photoperiod. A light intensity of 500  $\mu$  mol. m<sup>-2</sup>. sec<sup>-1</sup> (photosynthetically active radiation) was maintained during the daytime period.

Germinated seeds (radicle approximately 2 mm in length) were transplanted (1.5 cm deep) into pots (6.5 cm diameter, 25 cm deep) filled with 80 % washed sand and 20 % loam soil (v/v), simulating soil conditions in the field near Quetta. The pots were placed in a growth room under the previously described environmental conditions. Maximum and minimum temperatures were recorded daily for the calculation of growing degree days (Wilkins et al. 1984) accumulated after seedling emergence. For the first 15 days after transplanting, pots were watered to field capacity with distilled water every third day. After this 15-day period, pots were watered to field capacity with distilled water or 1/4 strength Hoagland solution (Hoagland and Arnon 1950) on an alternating basis every third day. Seedlings were thinned to one per pot after reaching the second leaf-stage 9 to 10 days after emergence.

Seedlings were destructively harvested at 15, 30, 45, and 60 days after emergence to observe root and shoot development. Seedlings from each pot were carefully washed to remove adhering soil and nearly all roots were retained. Root morphology was assessed according to the root system identification model of Newman and Moser (1988). Root measurements included: primary root number, length, and dry weight; seminal root number, length, and dry weight; and adventitious root number, length, and dry weight. Roots and rootlets exceeding 2 mm in length were counted as separate roots. Root and shoot samples were dried at 60 °C for 48 h to determine dry weights. Seedling shoot development was quantified by recording tiller number and Haun (1973) index every other day throughout the experiment. Tillers were labeled with different color

rings, and numbered according to the order of leaves on the main stem. Each new leaf on its first appearance at a growing point was allotted "0" Haun stage and it's subsequent growth was divided into decimal fractions that on each subsequent count accumulated to make one unit when there appeared another leaf. In a similar manner, the growth units were summed to provide the Haun stage per seedling on each harvest date. Seeding shoot development was related to cumulative growing degree days after seedling emergence.

The experiment was arranged in a split plot design with harvest date as the main plot and species as the sub plot. There were 10 replications (pots) for each species at each harvest date. Seedling development data were subjected to repeated measures analysis of variance, and mean comparisons were made using Fisher's least significant difference test (LSD) at the P < 0.05 level.

### Results

Chrysopogon aucheri and Cymbopogon jwarancusa had similar leaf and tiller development over the 60-day growing period (Figs. 2 and 3). By the last harvest, Chrysopogon aucheri and Cymbopogon jwarancusa had developed 11 leaves and 9 leaves per seedling, respectively. Seedlings of both species had 4 to 5 leaves per tiller.



Figure 2. Mean Haun stage (leaf development) of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwarancusa</u> (Cy) seedlings in relation to days after emergence and cumulative growing degree days (CGDD). Values for species at each time interval with different letters are significantly different (LSD<sub>.05</sub> = 2.45, n = 10).



Figure 3. Mean tiller number of <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwarancusa</u> (Cy) seedlings in relation to days after emergence and cumulative growing degree days (CGDD). Values for species at each time interval with different letters are significantly different (LSD<sub>.05</sub> = 0.87, n = 10).

Seedlings of both species initiated their first tillers by 45 days after emergence (Fig. 3). By 60 days after emergence, 5 of 10 <u>Chrysopogon aucheri</u> and 3 of 10 <u>Cymbopogon jwarancusa</u> seedlings were initiating a second tiller.

Seedlings of both species were comparable in terms of shoot and root dry weight and shoot:root ratio over the 60day growing period (Fig. 4 A, B, and C). Relative growth rates of total seedling dry weight for the 15 to 60 day post-emergence period were similar and quite low for <u>Chrysopogon aucheri</u> (0.061 mg. mg<sup>-1</sup>. day<sup>-1</sup>) and <u>Cymbopogon</u> <u>jwarancusa</u> (0.068 mg. mg<sup>-1</sup>. day<sup>-1</sup>).

Chrysopogon aucheri seedlings were generally comparable to Cymbopogon jwarancusa seedlings in juvenile (primary and seminal roots) and adventitious root development. Seedlings of each species developed 1 main primary root, 1 to 2 main seminal roots, and 5 to 7 main adventitious roots. Total root numbers shown in figures (Figs. 5 A, 6 A, and 7 A) are the sum of the main roots and rootlets (> 2 mm in length) for each root category. By 15 days post-emergence, <u>Chrysopogon aucheri</u> seedlings had a



Figure 4. Mean total shoot (A) and root (B) dry weight (mg) and shoot/root ratio (C) of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopoton jwarancusa</u> (Cy) in relation to time (days) after emergence. Values for species at each time interval with different letters are significantly different (LSD = 23.6 for A, 19.4 for B, and 0.63 for C; n = 10).

significantly greater number and dry weight of primary roots than Cymbopogon jwarancusa seedlings 30 days after emergence (Fig. 5 A, B and C). Differences in primary root development between species lessened over the remaining 30 days of the experiment as seminal and adventitious roots developed. Seedlings of both species initiated seminal roots 15 days after emergence. Seminal roots of Chrysopoqon aucheri were greater in number, length and dry weight than Cymbopogon jwarancusa at 60 days post-emergence (Fig. 6 A, B, and C). As with seminal roots, adventitious roots developed slowly over the first 30 days of the experiment (Fig. 7 A, B, and C). Seedlings of both species initiated adventitious roots 15 days after emergence. By 60 days post-emergence, Chrysopogon aucheri seedlings also had significantly greater adventitious root length than Cymbopogon jwarancusa seedlings (Fig. 7 B).

Subcoleoptile elongation was observed on 4 of 10 seedlings of each species at the 15 day harvest interval. Subcoleoptile internodes extended coleoptile nodes 5 to 15 mm above the scutelar node. Only one seedling of each species had 1 to 2 very small ( 2 mm long) roots on a subcoleoptile internode.



Figure 5. Mean primary root number (main root and rootlets > 2mm) (A), root length (cm) (B), and root dry weight (mg) (C) of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) seedlings in relation to time (days) after emergence. Values for species at each time interval with different letters are significantly different (LSD = 15.5 for A, 35.9 for B, and 2.0 for C; n = 10).



Figure 6. Mean seminal root number (main root and rootlets > 2mm) (A), root length (cm) (B), and root dry weight (mg) (C) of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) seedlings in relation to time (days) after emergence. Values for species at each time period with different letters are significantly different (LSD \_ 05 = 22.3 for A, 44.9 for B, and 4.2 for C; n = 10).



Figure 7. Mean adventitious root number (main root and rootlets
> 2 mm) (A), root length (cm) (B), and root dry weight (mg)
(C) of Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy)
seedlings in relation to time (days) after emergence.
Values for species at each time interval with different
letters are significantly different (LSD = 45.5 for A,
64.5 for B, and 16.6 for C; n = 10).

When expressed as a proportion (% by weight) of total seedling root dry weight, both species partitioned most resources toward primary root development during the first 30 days of growth and toward adventitious root development 30 to 60 days after emergence (Table 4). Biomass partitioning to the different root types was comparable for both species at all harvest intervals except 60 days after emergence, where <u>Chrysopogon aucheri</u> seedlings had a greater proportion of seminal root biomass than <u>Cymbopogon</u> <u>jwarancusa</u> seedlings.

### Discussion

When differences between species occurred, <u>Chrysopogon aucheri</u> seedlings were generally more vigorous than <u>Cymbopogon jwarancusa</u> seedlings under the controlled environmental conditions of this experiment. Seedling vigor in grasses is not characterized by a single attribute, but a combination of attributes such as large seed size, rapid germination, rapid root and shoot growth, tillering ability, and resistance to stress (McKell 1972, Coyne and Bradford 1985). Results from the germination response experiment (Chapter II) demonstrated that <u>Cymbopogon jwarancusa</u> seeds had greater cumulative germination and a more rapid rate of germination than <u>Chrysopogon aucheri</u> seeds. However, <u>Chrysopogon aucheri</u>

# Table 4. Mean proportion (% by weight ± SD) of different root types at different stages of seedling development for <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy).

Species and root type	Days after emergence			
	15	30	45	60
Ch		%-		
primary	83 <u>+</u> 31	51 <u>+</u> 8.8	19 <u>+</u> 16.8	8 <u>+</u> 6.9
subcoleoptile internode	e T*	0	0	0
seminal	Т	17 ± 13	19 <u>+</u> 13.2	29 <u>+</u> 8.6
adventitious	17 <u>+</u> 31	32 <u>+</u> 9.3	62 <u>+</u> 15.2	63 <u>+</u> 9.4
	100	100	100	100
Су				
primary	98 <u>+</u> 7.6	47 <u>+</u> 15	14 <u>+</u> 10.4	10 <u>+</u> 3.6
subcoleoptile internode	e T	0	0	0
seminal	Т	11 ± 10	11 ± 7.3	12 <u>+</u> 4.5
adventitious	2 <u>+</u> 7.6	42 <u>+</u> 11.3	75 <u>+</u> 16.3	78 <u>+</u> 7.0
	100	100	100	100

\* T = trace amount of biomass too small to contribute to overall root biomass.

seeds contained larger caryopses (4.5 + .4 mm long, 0. 97 + .04 mg mass) than those of Cymbopogon jwarancusa (1.6 + .3 mm long, 0.47 + .03 mg mass) indicating that Chrysopogon aucheri seedlings may have the potential to emerge from greater soil depths and have greater resources for initial seedling growth (McKell 1972, Fenner 1983). Once germinated, Chrysopogon aucheri seedlings were comparable to Cymbopogon jwarancusa seedlings in all the measured shoot and root parameters. However, at 30 and 60 days postemergence, primary and seminal root number, length, and dry weight were respectively significantly greater for Chrysopogon aucheri seedlings, as was adventitious root length at 60 days post-emergence. Rapid root elongation is a key characteristic for successful establishment in arid and semiarid areas where surface soils can dry quickly after a precipitation event (McKell 1972, Plummer 1943).

High relative growth rates (increase in terms of biomass invested) and low shoot:root ratios are generally considered to be important survival and environmental adaptations of seedlings (McKell 1972). In this study, both <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> seedlings had comparably low relative growth rates and initially high shoot:root ratios, indicating that establishment problems could arise in stressful environments. However, Coyne and Bradford (1985) observed that high relative growth rates were not necessarily desirable traits for the establishment of 17 C4 perennial grasses grown under limiting watering regimes in a controlled environment. Species with highest growth efficiencies at the start of sampling were the ones which had the greatest decline in efficiency by the end of the 51-day growing period. The mean relative growth rate of total seedling biomass for the 17 grasses was 0.133 g.  $g^{-1}$ . day<sup>-1</sup>. Simanton and Jordan (1986) reported that seedlings of Bouteloua curtipendula, a C<sub>4</sub> grass, had low shoot:root ratios, rapid germination, and rapid root elongation, yet had poor establishment in areas of the Southwestern U.S. with less than 190 mm summer rainfall and when subsurface soil moisture is low. In contrast, Eragrostis lehmanniana X Eragrostis trichophora, another C<sub>4</sub> grass with higher shoot:root ratios, slower germination rates, and slower seminal root elongation had more successful establishment on similar semiarid rangelands.

Even though leaf development was not significantly different for seedlings of both species, <u>Chrysopogon</u> <u>aucheri</u> seedlings reached the third leaf stage about 15 days after emergence, whereas <u>Cymbopogon jwarancusa</u> reached the third leaf stage about 18 days after emergence. Newman and Moser (1988) reported that seedlings of several warmseason (C<sub>4</sub>) grasses, including <u>Bouteloua curtipendula</u>, <u>B</u>. gracilis, Eragrostis tricodes, Andropogon gerardii var.

gerardii, Schizachyrium scoprium, Bothriochola caucasia, and Panicum virgatum, reached the third leaf stage 15 to 24 days after emergence in a controlled environment study, and that members of the Andropogoneae tribe reached the third leaf stage 3 to 8 days earlier than other warm-season grass species. Chrysopogon aucheri and Cymbopogon jwarancusa, both members of the Andropogoneae tribe, followed a similar pattern of leaf development, in terms of reaching the third leaf stage when adventitious root development typically occurs (Newman and Moser 1988). Rapid leaf and tiller development have also been associated with greater length, number, and order of branching of juvenile (primary and secondary roots) and adventitious roots in seedlings of Bouteloua curtipendula (Wilson and Briske 1979) and Bromus tectorum (Aquirre 1989). The sooner seedling roots increase their depth and volume of soil penetration, the greater is the probability of successful establishment in arid and semiarid regions as moisture becomes limiting in upper soil layers (Plummer 1943, Cook 1980, Buckly 1982, Aquirre 1989).

Successful establishment of grass seedlings requires formation of adventitious roots. The primary root, under normal conditions, is a short-lived structure (Tischler and Voigt 1987), and seminal roots and the subcoleoptile internode do not have sufficient xylem diameters to support water transport to maturing seedlings (Hyder et al. 1971,

Wilson et al. 1976). Subcoleoptile internode roots probably contribute less to the water economy of the shoot than primary or seminal roots (Tischler and Voigt 1987). Under the favorable environmental conditions of this study, adventitious roots initiated on seedlings of both species by 15 days after emergence, and comprised the greatest proportion of root biomass by 45 days after emergence. AS with other C, grasses with panicoid type seedlings, the subcoleoptile internode of Chrysopogon aucheri and Cymbopogon jwarancusa seedlings usually elevates the coleoptile to the soil surface, and adventitious roots typically develop close to the soil surface (Tischler and Voigt 1987). This type of morphology can be detrimental to seedling establishment in regions with limited precipitation, because adventitious roots may develop in the harsh environment associated with the soil surface, and primary and seminal roots cannot meet the water requirements of seedling leaves (Hyder et al. 1971).

Results from this controlled environment study may vary considerably from results obtained under field conditions. Temperature, moisture, and nutrient conditions were considerably more favorable in the laboratory than in the field near Quetta, and most importantly, seedlings of the more palatable <u>Chrysopogon aucheri</u> were not subjected to defoliation. Seedlings of both species have been observed at protected sites near Quetta and other regions

of Baluchistan (Saleem, personal observation). Cymbopogon jwarancusa may have a greater potential for germination than Chrysopogon aucheri, but once germination has occurred the fewer Chrysopogon aucheri seedlings may have equal or greater vigor and establishment success on areas protected from grazing.

#### CHAPTER IV

# ECOLOGY OF <u>CHRYSOPOGON AUCHERI</u> AND <u>CYMBOPOGON</u> JWARANCUSA: MORPHOLOGY AND DEFOLIATION RESPONSE

### Summary

Little is known about the defoliation responses of the palatable grass Chrysopogon aucheri, and the co-occurring unpalatable grass, Cymbopogon jwarancusa, under managed and unmanaged conditions on Baluchistan rangelands. Both species were grown in monoculture and in a 50:50 mixture in an 11-month (44-week) greenhouse study. Defoliation treatments were implemented when plants were 32 weeks old; and consisted of: equally clipping (3-cm stubble height) plants in monoculture and mixture zero, one, two, or three times at 4-week intervals ( 32, 36, and 40 weeks after emergence), and clipping (3-cm stubble height) one species in mixture zero, one, two, or three times at 4-week intervals (32, 36, and 40 weeks after emergence) without clipping the associate species. The final harvest of all plants in every defoliation treatment occurred at 44 weeks after emergence. Response to defoliation was measured in terms of leaf and tiller development, shoot and root biomass production, and nutritional quality and digestibility. Plants of both species had similar patterns of leaf and tiller development until defoliation treatments

Cymbopogon jwarancusa produced more were implemented. tillers per plant than Chrysopogon aucheri when both species were equally defoliated one, two, or three times in monoculture and equally defoliated three times in mixture. Most Chrysopogon aucheri plants developed inflorescences by 32 weeks after emergence, whereas all Cymbopogon jwarancusa plants remained vegetative throughout the experiment. Chrysopogon aucheri had greater shoot and root biomass than Cymbopogon jwarancusa in mixture when plants were equally defoliated zero, one, or two times, whereas shoot and root biomass were comparable under the same defoliation regimes in monoculture, and when equally defoliated three times in monoculture or mixture. When one species was defoliated zero, 1, 2 or 3 times and the associated species was not defoliated, shoot biomass was comparable for both species while Chrysopogon aucheri had greater root biomass than Cymbopogon jwarancusa. Chrysopogon aucheri had similar or higher crude protein content and % in vitro digestible dry matter when compared to Cymbopogon jwarancusa. Chrysopogon aucheri may not decrease in mixed Chrysopogon - Cymbopogon communities if the frequency and intensity of defoliation are controlled more closely as in this experiment.

### Introduction

Several factors, including the morphology, physiology, and palatability of plants, the type of herbivore, the

intensity, frequency, and timing of defoliation, and competition from surrounding plants, can differentially shape the defoliation responses of plants (Menke and Trilica 1981, Archer and Tieszen 1986, Wangoi and Hansen 1987). High rates of refoliation and tillering, high shoot:root and vegetative:reproductive ratios, and late elevation of apical meristems are associated with increased grazing tolerance (Dahl and Hyder 1977, Richards 1984, Archer and Tieszen 1986, Briske 1986). Milthorpe and Davidson (1966) reported that regrowth following defoliation in many grasses was first limited by carbohydrate reserves, then by photosynthesis, and later on by nutrient uptake. However, more recently, Richards and Caldwell (1985) demonstrated that regrowth in two Agropyron burchgrass species was most influenced by meristematic activity and photosynthesis and not by stored carbohydrate reserves. Plant and shard physical and chemical characteristics, such as growth stage, leaf:stem ratio, availability and distribution of plant parts, and nutrient and fiber content, influence the palatability of forage plant (Holm and Elloit 1980, Minson 1982, Hodgson 1982). These factors influence the bite size, intake, and digestibility of forage (Hodgson 1982). Plant secondary compounds, including essential oils, can greatly reduce palatability. Even though considered a nutritious forage, Cymbopogon jwarancusa is not readily grazed because of a

high essential oil content, comprised primarily of piperidine (Chopra et al. 1956, Saeed et al. 1978).

In Baluchistan, <u>Chrysopogon - Cymbopogon</u> grasslands are grazed continuously by a variety of herbivores, including cattle, and mixed herds of sheep, goats, horses, and camels. These herbivores vary in their grazing behavior and plant preference (Wangoi and Hansen 1987), which makes it difficult to speculate about patterns of defoliation. However, the continuous use of this scarce vegetation resource results in intensive, frequent defoliation of available forage species throughout time during the growing season.

Plant response to defoliation varies considerably with the level of competition from surrounding plants (Mueggler 1972). Highly preferred plants like <u>Chrysopogon aucheri</u> are often more frequently and intensively defoliated than less palatable species such as <u>Cymbopogon jwarancusa</u> and <u>Artemisia maritima</u>. Over time, reductions in above- and below-ground biomass of <u>Chrysopogon aucheri</u> may allow the encroachment of associated, less palatable species. Little is known about the growth and development and defoliation responses of <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> in a competitive environment. This study was designed to investigate the morphological characteristics of both species, and their responses to different defoliation regimes in a competitive environment.

## Materials and Methods

This experiment was conducted in a greenhouse under natural light conditions in Logan, Utah from November 1988 to August 1989. Temperatures ranged from 11 to 23 °C in winter months and 13 to 34 °C in summer months. Two-monthold seedlings of each species were transplanted (November 1988) into pots (28 cm diameter X 36 cm height) in monoculture and in a 50:50 mixture. Monocultures had 4 plants per pot and mixtures had 2 plants per species (arranged alternate to each other) per pot, giving a density equivalent to 15 plants/m<sup>2</sup>. The soil medium, composed of 80 % washed sand and 20 % loam soil, simulated the dominant soil texture on rangelands in Baluchistan. Pots were maintained at 50 % of field capacity (determined by pressure plate analysis) throughout the study. Plants were alternately watered (every 3-5 days) with distilled water and 1/4 - strength Hoagland solution (Hoagland and Arnon 1950).

Defoliation treatments, implemented when plants were 32 weeks old, consisted of: equally clipping all plants in monoculture and mixture for zero, one, two, or three times at 4-week intervals (32, 36, and 40 weeks after emergence); and clipping one species in mixture zero, one, two, or

three times at 4-week intervals (32, 36, and 40 weeks after emergence) without clipping the associated species in the same pot (Fig. 8). The final harvest of all plants in every defoliation treatment occurred at 44 weeks after emergence. Defoliation treatments were based upon field observations where <u>Chrysopogon aucheri</u> plants are intensively defoliated several times before <u>Cymbopogon</u> <u>jwarancusa</u> plants are defoliated. Plants were clipped to a 3-cm stubble height at each clipping interval. This stubble height simulated a heavy intensity of defoliation (85 % removal of initial standing crop).

Control plants (0 clipping) were monitored for leaf and tiller development at 4-week intervals from week 12 to week 44 (end of experiment). Leaf development of the main stem and subsequent tillers was determined by using the technique of Haun (1973). Each new leaf was identified with a permanent color mark and each new tiller was marked by a different colored ring at its base. Total plant Haun stage was obtained by summing the Haun stage of the main stem and the other tillers as they appeared over time. Prior to the initial clipping at 32 weeks, all live and dead tillers were counted on all plants in monoculture and mixture. After the initial clipping, the main stem and a secondary and tertiary tiller on each plant were marked and monitored for survival and leaf development for the remainder of the experiment.

0 NY Frequency of clipping 11/1 N W 111 3 WV V 32 36 40 44

46

Weeks after emergence

Figure 8. Defoliation treatments for <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> grown in monoculture and mixture. 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; and 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Shoot biomass was determined at appropriate clipping intervals for defoliated plants, and shoot and root biomass were determined at the end of the experiment for defoliated and control plants. Shoot and root biomass samples were oven dried at 65 °C for 48 h prior to weighing. Roots of the two species were intermingled in the mixture pots, but were relatively easy to separate because roots of <u>Chrysopogon aucheri</u> are dark brown while roots of <u>Cymbopogon jwarancusa</u> are pale yellow. Shoot samples from the monoculture treatments were analyzed for % dry matter, % organic matter (Harris 1970), % in vitro digestible dry matter (Goto and Minson 1977), and % crude protein (Hatch et al. 1985).

The experiment was a factorial with 2 species and 20 clipping treatments with four replications (pots) per clipping treatment (see Table 6 in Appendix). Data were subjected to analysis of variance and treatment means were separated by Fisher's least significant difference test (LSD) at the P <0.05 level of significance.

### Results

Without defoliation, both species had a similar pattern of leaf and tiller development in monoculture or mixture from 12 to 28 weeks after emergence (Figs. 9 and 10). <u>Cymbopogon jwarancusa</u> produced more tillers per plant



Weeks after emergence

Figure 9. Mean whole plant Haun stage for <u>Chrysopogon</u> <u>aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwarancusa</u> (Cy) plants grown in monoculture (Mon) and mixture (Mix) without clipping. Values for species with different letters within monoculture or mixture at each time interval are significantly different (LSD = 14.4; n = 8).



Weeks after emergence

Figure 10. Mean number of tillers per plant for <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in monoculture (Mon) and mixture (Mix) without clipping. Values for species with different letters within monoculture or mixture at each time interval are significantly different (LSD  $_{.05}$  = 5.8; n = 8). than <u>Chrysopogon aucheri</u> at all sampling dates after 36 weeks when both species were equally defoliated zero, one, two, or three times in monoculture (Fig. 11 A, B, C and D), and at all sampling dates when both species were equally defoliated three times in the mixture (Fig. 11 D).

<u>Chrysopogon aucheri</u> plants elongated apical meristems on main stems as early as 24 weeks after emergence. By week 32 when defoliation treatments were implemented, 144 of 192 main stems and 307 of 3075 secondary tillers on <u>Chrysopogon aucheri</u> plants had developed inflorescences. <u>Chrysopogon aucheri</u> plants defoliated one time (32 weeks after emergence) in monoculture or mixture entered the boot stage by the second clipping (36 weeks after emergence) but did not show signs of floral development after the second and third clippings (40 weeks after emergence). <u>Cymbopogon</u> <u>jwarancusa</u> plants remained vegetative regardless of the defoliation regime.

<u>Chrysopogon aucheri</u> had more shoot dry weight than <u>Cymbopogon jwarancusa</u> in mixture when plants were equally defoliated zero, one, or two times (Fig. 12 A, B, and C). Shoot dry weight was comparable for both species in monoculture under the same defoliation regimes (except for week 32 of the 3 clip treatment) and in mixture when plants were equally defoliated three times (Fig. 12 A, B, C, and D). Cumulative shoot dry weight (sum of initial standing crop and regrowth above 3-cm stubble height)

Figure 11. Mean number of tillers per plant for <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): (A) 0 clip (control); (B) 1 clip = standing crop clipped at 32 weeks and regrowth at 44 weeks (final harvest) after emergence; (C) 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; (D) 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters within monoculture or mixture at each time interval are significantly different [LSD<sub>.05</sub> (A, B, C, and D) = 10.1 for 32 weeks, 12.0 for 36 weeks, and 12.8 for 40 and 44 weeks; n = 4].



Figure 12. Mean shoot dry weight (g) of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): (A) 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; (B) 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; (C) 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; (D) 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters within monoculture or mixture at each time interval are significantly different [LSD<sub>.05</sub> (A, B, C, and D) = 0.61 for 32 weeks, 0.36 for 36 weeks, 0.22 for 40 weeks, and 0.35 for 44 weeks; n = 4].



Weeks after emergence
(Fig. 13), crown dry weight (below 3-cm stubble height) (Fig. 13), and root dry weight (Fig. 14) at the end of the experiment were greater for <u>Chrysopogon aucheri</u> than <u>Cymbopogon jwarancusa</u> in mixture when plants were equally defoliated zero, one, or two times. Cumulative shoot dry weight, crown dry weight, and root dry weight were similar for both species in monoculture under the same defoliation regimes, and in mixture and monoculture when plants were equally defoliated three times (Fig. 13 and 14).

Percent crude protein was comparable for both species in monoculture when the standing crop was clipped initially (1st clipping), and regrowth was clipped 4 weeks later (2nd clipping); however crude protein was greater for <u>Chrysopogon aucheri</u> than <u>Cymbopogon jwarancusa</u> when regrowth was clipped at subsequent 4-week intervals (3rd and final clippings) (Fig. 15 A). Digestibility of the standing crop at the initial clipping (1st clipping) and regrowth at the 3rd clipping was higher for <u>Chrysopogon</u> than <u>Cymbopogon jwarancusa</u>, and comparable for regrowth of both species at the 2nd and final clippings (Fig. 15 B).

In the 50:50 mixture when one species was defoliated zero, one, two or three times and the associated species was not defoliated, both species generally produced comparable numbers of tillers (Fig. 16 A, B, C, and D) and shoot dry weight (Fig. 17 A, B, and C) per plant. At week 40, <u>Cymbopogon jwarancusa</u> produced more tillers per plant



Above-ground dry weight (g) (cumulative shoot dry weight Figure 13. above 3-cm stubble height and crown dry weight below 3-cm) from 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes: 0 clip (control) = standing crop above 3-cm and crown below 3-cm clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth and crown clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks, and regrowth clipped at 36 weeks, and regrowth and crown clipped at 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks, regrowth clipped at 36 and 40 weeks and regrowth and crown clipped at 44 weeks after emergence. Above 3-cm and below 3-cm dry weight values for species with different letters within monoculture or mixture under each clipping regime are significantly different (LSD  $_{05}$  = 1.7 for above 3-cm and .88 for below 3-cm; n = 4).



Figure 14. Mean root dry weight (g) harvested from 44-week-old <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in monoculture (Mon) and mixture (Mix) under different clipping regimes (3-cm stubble height): 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters within monoculture or mixture under each clipping regime are significantly different (LSD<sub>.05</sub> = 1.7; n = 4).



Figure 15. Mean percent crude protein (A) and percent in vitro digestible dry matter (% IVDMD) (B) for <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in monoculture under different clipping regimes (3-cm stubble height): 1 clip = initial standing crop clipped at 32 weeks after emergence; 2 clip = regrowth clipped at 36 weeks after emergence; 3 clip = regrowth clipped again at 40 weeks after emergence; and F clip = final regrowth clipped at 44 weeks after emergence. Values for species with different letters under each clipping regime are significantly different (LSD 05 = 2.0 for A, 5.3 for B: n = 4).

Figure 16.

Mean number of tillers per plant for <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon jwarancusa</u> (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): (A) 0 clip (control); (B) 1 clip = standing crop of either species clipped at 32 weeks after emergence; (C) 2 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 weeks after emergence; and (D) 3 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 and 40 weeks after emergence. Values for species with different letters at each time interval are significantly different [LSD (A, B, C, and D) = 10.0 for 32 weeks, 12.0 for 36 weeks, and 12.8 for 40 and 44 weeks; n = 4].





.

Weeks after emergence

Mean shoot dry weight (g) of Chrysopogon aucheri (Ch) and Figure 17. Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): (A) 1 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 44 weeks (final harvest) after emergence; (B) 2 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; and (C) 3 clip = standing crop of either species clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters at each time interval are significantly different  $[LSD_{.05} (A, B, and C) = 0.61$  for 32 weeks, 0.36 for 36 weeks, 0.22 for 40 weeks, and 0.53 for 44 weeks; n = 4].

than <u>Chrysopogon aucheri</u> when plants were defoliated two times (Fig. 16 C); and at week 32, <u>Chrysopogon aucheri</u> produced more shoot dry weight than <u>Cymbopogon jwarancusa</u> when plants were defoliated three times (Fig. 17 C). Cumulative shoot dry weight was greater for <u>Chrysopogon aucheri</u> than <u>Cymbopogon jwarancusa</u> when plants remained undefoliated or were clipped one time (Fig. 18), while crown dry weight and root dry weight were greater for <u>Chrysopogon</u> <u>aucheri</u> than <u>Cymbopogon jwarancusa</u> when plants were clipped zero, one two, or three times (Figs. 18 and 19).

# Discussion

High tiller production, delayed elevation of apical meristems, and high vegetative:reproductive stem ratios are morphological characteristics associated with grazing tolerance (Branson 1953, Dahl and Hyder 1977, Richards and Caldwell 1985, Archer and Tieszen 1986, and Briske 1986). As in the 8-week seedling development experiment (Chapter III), <u>Chrysopogon aucheri</u> and <u>Cymbopogon jwarancusa</u> produced comparable numbers of leaves and tillers per plant during the early part of this experiment (up to 28 weeks after emergence). As plants matured (36 to 44 weeks after emergence), <u>Cymbopogon jwarancusa</u> produced more tillers per plant in monoculture than <u>Chrysopogon aucheri</u> under the different defoliation regimes. By the time defoliation treatments were implemented at 32 weeks after emergence,



Above-ground dry weight (g) (cumulative shoot dry weight Figure 18. above 3-cm stubble height and below 3-cm) from 44-week-old Chrysopogon aucheri (Ch) and Cymbopogon jwarancusa (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): 0 clip (control) = standing crop above 3-cm and crown below 3-cm clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth and crown clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks, regrowth clipped at 36 weeks, and regrowth and crown clipped at 44 weeks after emergence; and 3 clip = standing crop clipped at 32 weeks, regrowth clipped at 36 and 40 weeks, and regrowth and crown clipped at 44 weeks after emergence. Values for species above 3-cm and below 3-cm with different letters under each clipping regime are significantly different (LSD  $_{05}$  = 1.7 for above 3-cm and .88 for below 3-cm; n = 4).





Mean root dry weight (g) harvested from 44-week-old <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon Jwarancusa</u> (Cy) plants grown in mixture under different clipping regimes (each value represents a treatment where the species indicated is clipped to a 3-cm stubble height and the associated species is not clipped): 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; and 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence. Values for species with different letters under each clipping regime are significantly different (LSD<sub>.05</sub> = 1.70; n = 4).

most of the main stems and a small proportion of secondary tillers on <u>Chrysopogon aucheri</u> plants had elevated apical meristems and developed inflorescences. Leaf growth normally ceases on reproductive stems (Langer 1972, Dahl and Hyder 1977), and tillering from axillary buds can be slow following defoliation when water and nutrients are limiting (Branson 1953, Hyder 1972). However, water and nutrient augmentation in this controlled environment study may have allowed for greater tiller development following the defoliation of flowering Chrysopogon aucheri plants than would be found under limiting field conditions. Chrysopogon aucheri might have remained in a vegetative state longer if it had been defoliated before apical meristem elevation (Langer 1972). Cymbopogon jwarancusa never elevated apical meristems, and continued to produce new tillers and extend partially defoliated leaves from intercalary meristems.

Several factors, including plant age, plant size, the accumulation of certain metabolites, temperature, light intensity, and photoperiod are involved in the transformation from the vegetative to the reproductive state in most grasses (Langer 1972). Mature plants of both species develop flowers and set seed during the same time period (March to May) under field conditions in Baluchistan (Cope 1982; Saleem, personal observation). However, no information is available on floral development of young

plants of either species. In this greenhouse study in Logan (41 N latitude), Chrysopogon aucheri initiated flowering as early as 24 weeks after emergence in March when the day length is similar (approximately 12 h) to that in Quetta, Baluchistan (30 N latitude) in March (List 1948). Chrysopogon aucheri continued to develop inflorescences up to 32 weeks after emergence in May when the day length in Logan is up to 55 minutes longer than in Quetta (List 1948). Cymbopogon jwarancusa plants of similar age may not have flowered at the same time as the Chrysopogon aucheri in the greenhouse because they may require more time in a vegetative stage before floral induction can occur, or they may be more sensitive to variation in day length (Dahl and Hyder 1977). Certain species may detect day-length variations of less than 1 h that can delay or prohibit flowering (Dahl and Hyder 1977).

Plants having high reproductive to vegetative ratios can be easily removed from communities by excessive grazing (Branson, 1953, and Hyder 1972). Dahl and Hyder (1977) reported that several warm-season grasses were vulnerable to defoliation (i.e. removal of elevated apical meristem) when three-fourths to two-thirds of their stems were reproductive. In this study only 20 % of the stems of undefoliated <u>Chrysopogon aucheri</u> plants were reproductive. Therefore, <u>Chrysopogon aucheri</u> may not be that susceptible to grazing, even though some tillers elevate apical meristems.

The impact of defoliation on a plant varies, depending on the intensity, frequency, pattern, and timing of tissue removal (Branson 1953, Dahl and Hyder 1977, Archer and Tieszen 1986, Kalmbacher and Martin 1988). Kanodia et al. (1981) found that increasing the clipping height from 5 to 15 cm above the soil surface and increasing the clipping interval from 10 to 60 days increased aboveground biomass production of Chrysopogon fulvus in India. In the present study, clipping at a 3-cm stubble height at 4-week intervals did not decrease tiller development and did not result in significant decreases in aboveground biomass production for both species. This is not the case on unmanaged rangelands in Baluchistan where frequent, intensive grazing at any time during the growing season can decrease or eliminate Chrysopogon aucheri in grassland communities. Even though controlled environment conditions do not fully represent field conditions in Baluchistan, plant responses to the different defoliation treatments indicate that Chrysopogon aucheri could be maintained in communities when frequency and intensity of grazing are regulated.

Clipping does not completely mimic herbivore because of the manner in which plant parts are selected and removed from the plant. The pattern of defoliation greatly affects

the canopy structure and microenvironment of the plant, which influence defoliation responses (McNaughton 1986, Caldwell et al. 1983, Gold and Caldwell 1990, Wallace 1990). In the present study, however, the uniform pattern of defoliation may simulate grazing more closely than in other clipping studies because the scarcity of vegetation on Baluchistan rangelands may not allow grazing animals to be very selective (FAO 1981).

Plant response to defoliation also varies with the level of competition from surrounding plants. Pemadasa and Amarasinghe (1982), working in a Cymbopogon - Themeda grassland in Sri Lanka, found that clipping reduced the competitive ability of Themeda trimula and resulted in greater biomass production in associated Cymbopogon species. In the present study, when both species were defoliated equally and less frequently (zero, one, or two clippings) Chrysopogon aucheri produced more shoot and root biomass than Cymbopogon jwarancusa in mixture. However, under a higher frequency of defoliation (3 clippings) the competitive advantage of Chrysopogon aucheri was not evident in mixture, as both species produced comparable amounts of shoot and root biomass. When one species was defoliated zero to three times in mixture and the associated species was not defoliated, Chrysopogon aucheri was comparable to Cymbopogon jwarancusa in tiller development and shoot and root biomass production, and

superior in root biomass production. Even though growing conditions were more favorable in the greenhouse than in the field in Baluchistan, results indicate that <u>Chrysopogon</u> <u>aucheri</u> could coexist with <u>Cymbopogon jwarancusa</u> in the field if the intensity and frequency of defoliation by livestock was managed more effectively.

<u>Chrysopogon aucheri</u> may be similar to species such as <u>Schizachyrium scoparium, Bouteloua curtipendula</u>, and <u>Koeleria cristata</u>, which are tolerant of moderate grazing, but still decrease in grazed communities because of their relatively high palatabilities (Dahl and Hyder 1977). Palatability generally decreases with age in many grasses, particularly when plants go reproductive (Hodgson 1982). Despite going reproductive, <u>Chrysopogon aucheri</u> was comparable or higher (depending upon sampling date) in crude protein content and digestibility than <u>Cymbopogon</u> <u>jwarancusa</u> when frequently defoliated. However, the presence of essential oils in the foliage of <u>Cymbopogon</u> <u>jwarancusa</u> overrides nutritional differences when comparing the relative palatability of the two species.

Although not measured in this study, the essential oil content of <u>Cymbopogon jwarancusa</u> is regarded as an avoidance mechanism (Briske 1986) that greatly reduces the frequency and intensity of grazing of this species under unmanaged conditions (West Pakistan Forest Department 1960). Chopra et al. (1956) demonstrated that apart from

being an essential oil-bearing plant, <u>Cymbopogon jwarancusa</u> was a nutritious grass. Similarly, Ghosh and Mathur (1962) indicated that after the extraction of essential oils, <u>Cymbopogon flexuous</u> had a higher palatability than wheat or rice straw. More research is required under grazing conditions in the field to more accurately determine the effects of grazing tolerance and grazing avoidance on <u>Chrysopogon aucheri</u> - <u>Cymbopogon jwarancusa</u> interactions and to develop guidelines for more 'prudent' grazing.

# CHAPTER V

## SYNTHESIS

Under field conditions in Baluchistan, continuous season-long or year-long grazing by livestock (sheep, goats, camels, and cattle) has led to the gradual replacement of <u>Chrysopogon aucheri</u> by <u>Cymbopogon</u> <u>jwarancusa</u>. Species interactions have been primarily related to different grazing responses, i. e. the more palatable <u>Chrysopogon aucheri</u> being defoliated more intensively and frequently than the less palatable <u>Cymbopogon jwarancusa</u>. Three experiments were conducted in controlled environments to determine how germination, seedling development, and defoliation tolerance influence the growth and development of these two species.

<u>Cymbopogon jwarancusa</u> was superior to <u>Chrysopogon</u> <u>aucheri</u> in the germination phase in experiment 1. <u>Cymbopogon jwarancusa</u> had more filled seeds with higher viability than <u>Chrysopogon aucheri</u>. <u>Cymbopogon jwarancusa</u> had significantly higher cumulative germination in five of six alternating temperature regimes and a faster rate of germination in colder temperature regimes when both species started germination. The temperature regimes represent field temperatures during the normal recruitment period in Baluchistan. Germination at colder temperature regimes may allow <u>Cymbopogon jwarancusa</u> to have better recruitment than <u>Chrysopogon aucheri</u> under field conditions when soil moisture is more readily available.

In general, seedlings of both species were comparable in terms of shoot and root development over a 60-day growing period in experiment 2. When differences occurred, <u>Chrysopogon aucheri</u> seedlings were more vigorous than seedlings of <u>Cymbopogon jwarancusa</u>. <u>Chrysopogon aucheri</u> developed a greater number, length, and dry weight of primary and seminal roots than <u>Cymbopogon jwarancusa</u> at 30 and 60 days after emergence, respectively. <u>Chrysopogon</u> <u>aucheri</u> also had greater adventitious root length at 60 days after emergence than <u>Cymbopogon jwarancusa</u>. Superiority in root elongation may allow <u>Chrysopogon</u> <u>aucheri</u> seedlings the advantage of better survival in the field than <u>Cymbopogon jwarancusa</u> seedlings provided that <u>Chrysopogon aucheri</u> is not grazed at the seedling stage.

Rapid adventitious root development on seedlings of both species in a controlled environment indicates high potential for successful establishment. However, elongation of the subcoleoptile internode and root development on the subcoleoptile internode indicates that both species have "panicoid" type seedlings. Panicoid seedlings can have establishment problems in arid regions where low moisture near the soil surface may limit adventitious root development.

Both species had comparable leaf and tiller development as long as they remained in a vegetative state without defoliation. However, under the conditions of experiment 3 most of the main stems of <u>Chrysopogon aucheri</u> elevated apical meristems as early as 24 weeks after emergence and flowered before the first defoliation event 32 weeks after emergence.

Defoliation responses of the two species varied in experiment 3, depending upon the frequency of defoliation and whether a species was grown in mixture or monoculture. Cymbopogon jwarancusa produced more tillers per plant than Chrysopogon aucheri when both species were equally defoliated one, two, or three times in monoculture, where as Chrysopogon aucheri produced greater shoot and root biomass than Cymbopogon jwarancusa when both species were equally defoliated one, or two times in mixture. When one species was defoliated one, two, or three times and the associated species was not defoliated, shoot biomass was comparable for both species but Chrysopogon aucheri had greater root biomass than Cymbopogon jwarancusa. Thus, it is difficult to claim that one species has a distinct advantage over the other under the conditions of this study.

Results from these controlled environment experiments cannot be directly interpolated to the field conditions in Baluchistan, but they do provide some insights about the

growth and development and interaction of Chrysopogon aucheri and Cymbopogon jwarancusa. If a Chrysopogon-Cymbopogon grassland was protected from grazing, it appears that Cymbopogon jwarancusa would have greater recruitment potential than Chrysopogon aucheri. However, once seeds of both species germinated, it appears that seedlings and more mature plants of Chrysopogon aucheri would be comparable in growth and development to those of Cymbopogon jwarancusa. Despite its higher palatability, Chrysopogon aucheri should be able to co-exist with Cymbopogon jwarancusa if frequency and intensity of defoliation are carefully controlled. More research needs to be conducted to more fully understand the relationship between Chrysopogon aucheri and Cymbopogon jwarancusa under field conditions. Specific research needs include: floral development and seed set, seedling recruitment, adventitious root development on seedlings under water-limited conditions, quantification of oil content of Cymbopogon jwarancusa at different growth stages, and monitoring the intensity and frequency of defoliation of grasses under different livestock grazing systems.

### LITERATURE CITED

- Aguirre L. 1989. Early seedling development in three range grasses as influenced by temperature, drought, and competition. Ph.D. Dissertation, Utah State University, Logan, Utah.
- Ahmed, M., T. Hussain, and A. Hussain. 1978. Studies on some range grasses of Pakistan. Pak. J. For. 28:7-12.
- Archer, S., and L. L. Tieszen. 1986. Plant response to defoliation: Hierarchical consideration, p. 45-59. <u>In:</u> O. Gudmundsson (ed), Grazing research at northern latitudes. Plenum Publ. Corp., New York.
- Baluchistan Agriculture Department. 1985. Baluchistan agriculture statistics. Baluchistan Agriculture Department, Quetta, Pakistan.
- Baluchistan Forest Department. 1986. Forest Dept., Govt. of Baluchistan Record 1950-86.
- Branson, F. A. 1953. Two new factors affecting resistance of grasses to grazing. J. Range Manage. 6:165-171.
- Briske, D. D. 1986. Plant responses to defoliation and morphological considerations and allocation priorities, p. 425- 427. <u>In:</u> P.J. Joss, P. W. Lynch and O. B. Williams (eds), Rangelands: A resource under seige. Proc. 2nd Int. Rangel. Cong. Aust. Acad. Sci., Canberra.
- Buckly, R. C. 1982. Seed size and seedling establishment in tropical arid dune plants. Biotropica 14:314-15.
- Caldwell, M. M. 1984. Plant requirement for prudent grazing, p. 117-152. <u>In:</u> Developing strategies for rangeland management. National Research Council/Nat. Acad. of Sci. Westview Press, Boulder, Co.
- Caldwell, M. M., T. J. Dean, R. S. Nowak, R. S. Dzurec, and J. H. Richards. 1983. Bunchgrass architecture, light interception, and water-use efficiency: Assessment by fiber optic point quadrate and gas exchange. Oecologia 59:178-184.

- Chopra, R. N., K. L. Handa., L. D. Kapoor, and T. Sing. 1956. Nutritive value of grasses of Jammu and Kashmir. Ind. J. Agric. Sci. 26:415-57.
- Clements, F. E. 1928. Plant succession and indicators. Hafner Publ. Co., NY.
- Cook, R. E. 1980. Germination and size dependent mortality in <u>Viola blanda</u>. Oecologia 47:115-117.
- Cope, T. A. 1982. Flora of Pakistan. No. 143 Poaceae. National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan.
- Coyne, P. I., and J. A. Bradford. 1985. Morphology and growth in seedlings of several C4, perennial grasses. J. Range Manage. 38:504-512.
- Dahl, B. E., and D. N. Hyder 1977. Developmental morphology and management implications, p. 257-290. <u>In:</u> R. E. Sosebee (ed), Rangeland Plant Physiology. Soc. for Range Manage., Denver, Colorado.
- Debadgahoa, P. M., and K. A. Shankarnarayan. 1973. The grass cover of India. Ind. Counc. Agric. Res., New Delhi, India.
- Dwyer, D. D., and K. Wolde-Yohannis. 1972. Germination, emergence, water use, and production of Russian thistle (Salsola kali L.) Agro. J. 64:52-55.
- Ellern, S. J., and N. H. Tadmor. 1967. Germination of range plant seeds at alternating temperatures. J. Range Manage. 20:72-77.
- Ellis, R. H., and E. H. Roberts, 1978. Towards a rational basis for testing seed quality, p. 605-636. <u>In:</u> P. P. Hebblethwaite (ed), Seed production. Butterworths, London, England.
- Fenner, M. 1983. Relationships between seed weight, ash content, and seedling growth in twenty-four species of Compsitae. New Phytol. 95:697-706.
- Food and Agriculture Organization. 1981. FAO report: Assistance to rangelands and livestock development survey in Baluchistan. FAO Technical Cooperation Program, Pakistan 0107.

- Fowler, N. L. 1988. What is a safe site? : Neighbor, litter, germination date, and patch effects. Ecology 69:947-961.
- Ghosh, M. L., and S. K. Chatterjee. 1981. Growth and development physiology of <u>Cymbopogon</u> in the Gangetic plains of Bengal. Indian perfumer, 25:11-18.
- Ghosh, S. N., and M. L. Mathur. 1962. Studies on the palatability, chemical composition, digestibility, and nutritive value of spent lemon grass (Cymbopogon flexuosus). Ind. J. Res. Sci. 32:22-26.
- Gold, W. G., and M. M. Caldwell 1990. The effects of the spatial patten of defoliation on regrowth of a tussock grass: III photosynthesis, canopy structure and light interception. Oecologia 82:12-17.
- Goto, I., and D. J. Minson. 1977. Prediction of the dry matter digestibility of Tropical grasses using a pepsin-cellulase assay. Anim. Feed Sci. Tech. 2:247-253.
- Grabe, D. F. (ed.). 1970. Tetrazolium testing handbook for agricultural seeds. Contribution No. 29 to the handbook on seed testing. Assoc. of Seed Analysts. Lansing, Mich.
- Grubb, P. J. 1977. The maintenance of species-richness in plant communities: The importance of regeneration niche. Biol. Rev. 52, 107-45.
- Gulliver, R. L., and W. Hydecker. 1973. Establishment of seedlings in changeable environments, p. 433-462. <u>In:</u>
  W. Hydecker (ed), Seed ecology. Penn. State Univ. Press, Univ. Park, Penn.
- Harper, J. L. 1977. Population biology of plants. Academic Press, London, England.
- Harris, G. A. 1967. Some competition relationships between <u>Agropyron spicatum</u> and <u>Bromus tectorum</u>. Ecol. Monogr. 37:89-111.
- Harris, L. E. 1970. Nutrition research techniques for domestic and wild animals, Vol. 1. Agri-services, Logan, Utah.
- Hatch. C. C., S. V. Brayton, and A. B. Kopelove. 1985. A powerful Kjeldahl nitrogen method using peroxymonosulfuric acid. J. Agric. Food Chem. 33:1117-1123.

Haun, J. R. 1973. Visual quantification of wheat development. Agron. J. 65:116-119.

- Hoagland, D. R., and D. I. Arnon. 1950. The water-culture method for growing plants with out soil. Calif. Agric. Exp. Stn. Cir. 347. Univ. Calif., Davis, Calif.
- Hodgson, J. 1982. Influence of sward characteristics on diet selection and herbage intake by the grazing animal, p. 153-166. <u>In</u> J. B. Hacker (ed), Nutritional limits to animal production on pastures. Commonwealth Agr. Bureaux Pub., Farnham Royal, Slough, UK.
- Holm, A. M. R., and G. J. Elloit. 1980. Seasonal changes in the nutritive value of some native pasture species in northwestern Australia. Aust. Range. J. 2:175-182.
- Hoshikawa, K. 1969. Underground organs of the seedlings and the systematics of Gramineae. Bot. Gaz. 130:192-203.
- Hussain, A., T. Hussain, and M. Ahmed. 1980. Sex distribution and male sterility in some range grasses of Pakistan. Pak. J. Agri. Res. 1:77-80.
- Hyder, D. N. 1972. Defoliation in relation to vegetative growth, p. 304-317. <u>In:</u> V. B. Youngner, and C. M. McKell (eds), Biology and utilization of grasses. Academic Press, New York.
- Hyder, D. N., A. C. Everson, and R. E. Bement. 1971. Seedling morphology and seeding failures with blue grama. J. Range Manage. 24:287-297.
- Kalmbacher, R. S., and F. G. Martin. 1988. Effects of defoliation frequency and N-P-K fertilization on maidencane. J. Range Manage. 41:235-238.
- Kanodia, K. C., P. Rai, K. A. Shankarnarayan, and R. Kumar. 1981. Response of defoliation management-cum manuring on quantity and quality of Dhwalu grass. Ind. J. of Agron. 26:251-256.
- Khan, Ch. M. A. 1971. Tour note of Dr. M. Anwar on range resources of Baluchistan for the month of September. Tour note memo. 11 p.
- Koller, D. 1972. Environmental control of seed germination, p. 1-101. <u>In:</u> T. T. Kozlowski (ed), Seed biology. Vol. II. Germination control, metabolism, and pathology. Academic Press, New York.

- Langer, R. H. M. 1972. How grasses grow. Edward Arnold, Lincoln College, New Zealand.
- List, J. R. 1948. Smithsonian meteorological tables. Smithsonian Institution Press, Washington, D. C.
- Mayer, A. M., and A. Poljakoff-Mayber. 1982. The germination of seeds. 3rd Ed. Pergamon Press, New York.
- McKell, C. M. 1972. Seedling vigor and seedling establishment, p. 74-89. <u>In:</u> V. B. Youngner, and C. M. McKell (eds), The biology and utilization of grasses. Academic Press, New York.
- McNaughton, S. J. 1986. On plants and herbivores. Amer. Nature 128:765-770.
- Menke, J. W., and M. J. Trilica. 1981. Carbohydrate reserves, phenology, and growth cycles of nine Colorado range species. J. Range Manage. 34:269-277.
- Milthorpe, F. L. 1950. Changes in the drought resistance of wheat seedlings during germination. Ann. Bot. (London) 14:79.
- Milthorpe, F. L., and J. L. Davidson. 1966. Physiological aspects of regrowth cycles in grasses, p. 244-255. <u>In:</u> F. L. Milthorpe, and J. D. Ivins (eds), The growth of cereals and grasses. Butterworths, London, England.
- Minson, D. J. 1982. Effects of chemical and physical composition of herbage eaten upon intake, p. 167-182, <u>In:</u> J. B. Hacker (ed), Nutritional limits to animal production on pastures. Commonwealth Agr. Bureaux Pub., Farnham Royal, Slough, UK.
- Mott, J. J. 1978. Dormancy and germination in five native grass species from savannah woodland communities of the Northern Territory. Aust. J. Bot. 26:621-631.
- Mueggler, W. F. 1972. Influence of competition on the response of bluebunch wheat grass to clipping. J. Range Manage. 25:88-92.
- Newman, P. R., and L. W. Moser. 1988. Seedling root development and morphology of cool-season and warmseason grasses. Crop Sci. 28:148-151.

- Peart, M. H. 1979. Experiments on the biological significance of the morphology of seed dispersal units in grasses. J. Ecol. 67; 843-863.
- Peart, M. H. 1984. The effects of morphology, orientation and position of grass diaspores on seedling survival. J. Ecol. 72:437-453.
- Pemadasa, M. A., and L. Amarasinghe. 1982. The ecology of a montane grassland in Sri Lanka. V. Interference in populations of four major grasses. J. Eco. 70:731-744.
- Plummer, A. P. 1943. The germination and early seedling development of twelve range grasses. Amer. Soc. Agron. J. 35:19-34.
- Rai, P., P. S. Pathak., and R. D. Roy. 1980. Germination of range grasses and legumes under moisture stress conditions. Ind. J. Ecol. 7:224-228.
- Richards, J. H. 1984. Root growth response to defoliation in two <u>Agropyron</u> bunch grasses: Field observations with an improved root periscope. Oecologia (Berlin) 64:21-25.
- Richards, J. H., and Caldwell 1985. Soluble carbohydrates, concurrent photosynthesis and efficiency in regrowth following defoliation: A field study with <u>Agropyron</u> species. J. Appl. Ecol. 22:907-920.
- Saeed, T., P. J. Sandra, and M. J. E. Verzel. 1978. Constituents of the essential oil of <u>Cymbopogon</u> <u>iwarancusa</u>. Phytochem. 17:1433-1434.
- Simanton, J. R., and G. Jordan. 1986. Early root and seedling shoot elongation of selected warm-season perennial grasses. J. Range Manage. 39:63 - 67.
- Singh, J. S., Y. Hanxi, and P. E. Sajise. 1985. Structural and functional aspects of Indian and southeast Asian Savanna ecosystems, p. 34-51. <u>In:</u> J. C. Tothill and J. J. Mott (eds), Ecology and management of the world's Savannas. Australian Academy of Science, Canberra.
- Tischler, C. R., and P. W. Voigt. 1987. Seedling morphology and anatomy of rangeland plant species, p. 5-11. <u>In:</u> G. W. Frasier and R. A. Evans (eds), Proceedings of Symposium "Seed and Seedbed Ecology of Rangeland Plants". USDA-ARS, Washington, D. C.

- Wallace, L. L. 1990. Comparative photosynthesis responses of bigbluestem to clipping versus grazing. J. Range Manage. 43:58-60.
- Wangoi, M., and R. M. Hansen. 1987. Seasonal diets of camels, cattle, sheep, and goats in a common range in eastern Africa. J. Range Manage. 40:76-79.
- West, N. E. 1968. Outline for autecological studies of grasses. J. Range Manage. 21:102-105.
- West Pakistan Forest Department. 1960. Range plants handbook for the Quetta-Kalat Region. Government Press, Lahore, Pakistan.
- Wilkins, D. E., B. L. Klepper, and P. E. Rasmussen. 1984. Grain stubble effect on winter wheat seedling development. USDA-ARS, Western Region, Pendleton, OR. Paper No. 84- 1514.
- Wilson, A. M., and D. D. Briske. 1979. Seminal and adventitious root growth of blue grama seedlings on the central plains. J. Range Manage. 32:209-213.
- Wilson, A. M., D.N. Hyder, and D. D. Briske. 1976. Drought resistance characteristics of blue grama seedlings. Agron. J. 68:479-484.
- Young, J. A., R. A. Evans, and B. L. Kay. 1973. Temperature requirements for seed germination in an annual-type rangeland community. Agron. J. 65:656-659.

APPENDIX

Table 5. Mean monthly maximum and minimum temperature and monthly precipitation data for Quetta, Baluchistan.

Month	Max. temp.(°C)	Min. temp. (°C)	Rainfall mm
Jan	10.8	- 2.8	38.8
Feb	13.4	- 1.2	63.7
Mar	18.4	3.1	42.4
Apr	24.7	7.3	12.4
May	30.4	10.8	6.3
Jun	34.2	14.2	1.3
Jul	35.4	18.9	18.3
Aug	34.3	16.4	4.3
Sep	31.2	9.8	0.5
Oct	25.0	3.2	1.3
Nov	18.0	- 1.8	5.6
Dec	13.0	- 3.7	22.9

# Table 6. Heavy defoliation (85% removal) regimes for monocultures and 50:50 mixtures of <u>Chrysopogon aucheri</u> (Ch) and <u>Cymbopogon</u> <u>jwaracusa</u> (Cy).

Sequence of defoliation	<u>Chrysopogon<sup>1</sup></u> (monoculture)	<u>Chrysopogon Cymbopogon<sup>1</sup></u> (50:50)	<u>Cymbopogon</u> <sup>1</sup> (monoculture)
No defoliation $(0)^2$	Ch (0)	Ch (0) & Cy (0)	Су (0)
Initial (1)	Ch (1)	Ch (1) & Cy (0) Ch (0) & Cy (1) Ch (1) & Cy (1)	Cy (1)
Initial (1) + regrowth (2)	Ch (2)	Ch (2) & Cy (0) Ch (0) & Cy (2) Ch (2) & Cy (2) Ch (2) & Cy (1) Ch (1) & Cy (2)	Су (2)
Initial (1) + regrowth (2) + regrowth (3)	Ch (3)	Ch (3) & Cy (0) Ch (0) & Cy (3) Ch (3) & Cy (3) Ch (3) & Cy (1) Ch (1) & Cy (3) Ch (3) & Cy (2) Ch (2) & Cy (3)	Су (3)

<sup>1</sup>Four plants per pot; four replications (pots) per treatment.

<sup>2</sup>Numbers in parentheses represent defoliation frequencies of the initial standing crop and regrowth; i.e. 0 clip (control) = standing crop clipped at 44 weeks (final harvest) after emergence; 1 clip = standing crop clipped at 32 weeks and regrowth clipped at 44 weeks after emergence; 2 clip = standing crop clipped at 32 weeks and regrowth clipped at 36 and 44 weeks after emergence; and 3 clip = standing crop clipped at 32 weeks and regrowth clipped at 36, 40, and 44 weeks after emergence.

### VITA

### Mohammad Saleem

Candidate for the Degree of

#### Doctor of Philosophy

Dissertation: Autecological characteristics of <u>Chrysopogon</u> <u>aucheri</u> and <u>Cymbopogon jwarancusa</u>, dominant rangeland grasses in Balucistan.

Major Field: Range Science

Biographical Information:

- Personal Data: Born at Village Ahmadidargah, District Zhob, Baluchistan (Pakistan), February 2, 1950, Son of Salim Khan and Syeeda-Lo.
- Education: Received Intermediate and Secondary Education from Govt. Intermediate College Fortsandeman in 1969; received a Bachelor of Science in Forestry from University of Peshawar, September, 1974; received a Master of Science in Forestry from University of Peshawar September, 1981; completed requirements for a Doctor of Philosophy in Range Science at Utah State University, June 1990.
- Professional Experience: 1990---Divisional Forest Officer, Forest Department Baluchistan (Pakistan); 1986 - 1990 Graduate Fellow, Department of Range Science, Utah State University, Logan, Utah; 1974 - 1986 Range Forest Officer and Divisional Forest Officer, Forest Department, Baluchistan (Pakistan).