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Rapid screening and identification of chemical hazards in surface and drinking water using high resolution mass spectrometry and a case-control filter

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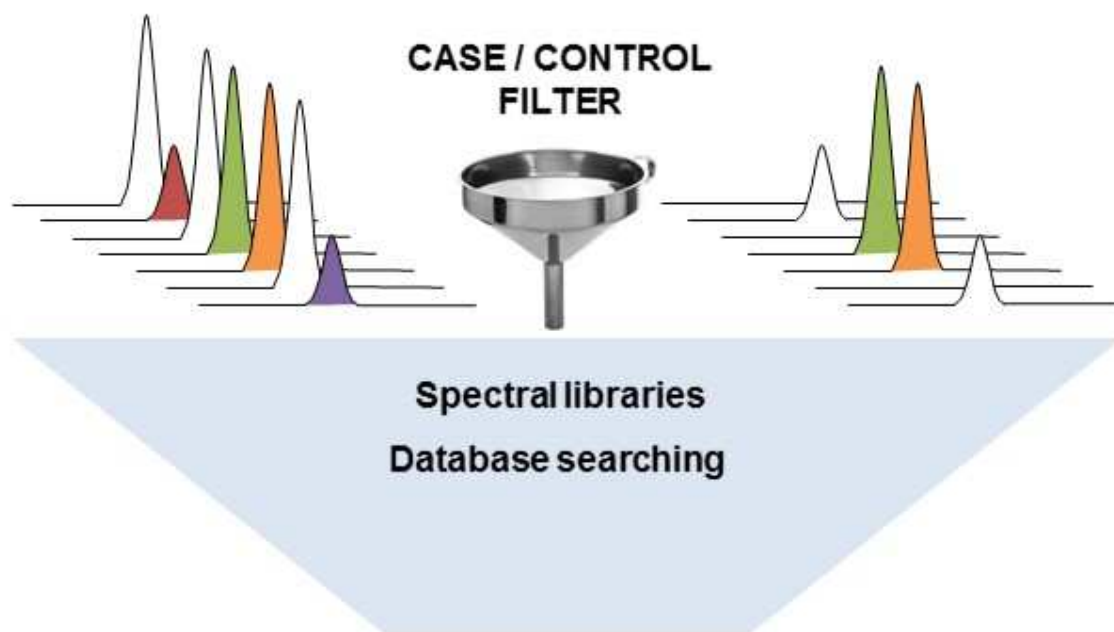
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1 **Rapid screening and identification of chemical hazards in surface and**
2 **drinking water using high resolution mass spectrometry and a case-control**
3 **filter**

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16

17 **ABSTRACT**

18 Access to clean, safe drinking water poses a serious challenge to regulators, and requires analytical
19 strategies capable of rapid screening and identification of potentially hazardous chemicals,
20 specifically in situations when threats to water quality or security require rapid investigations and
21 potential response. This study describes a fast and efficient chemical hazard screening strategy for
22 characterising trace levels of polar organic contaminants in water matrices, based on liquid
23 chromatography high resolution mass spectrometry with post-acquisition 'case-control' data
24 processing. This method allowed for a rapid response time of less than 24 hours for the screening of
25 target, suspect and non-target unknown chemicals via direct injection analysis, and a second, more
26 sensitive analysis option requiring sample pre-concentration. The method was validated by fortifying
27 samples with a range of pesticides, pharmaceuticals and personal care products (n=46); with >90%
28 of target compounds positively screened in samples at 1 ng mL⁻¹, and 46% at 0.1 ng mL⁻¹ when
29 analysed via direct injection. To simulate a contamination event samples were fortified with
30 compounds not present in the commercial library (designated 'non-target compounds'; fipronil and
31 fenitrothion), tentatively identified at 0.2 and 1 ng mL⁻¹, respectively; and a compound not included
32 in any known commercial library or public database (designated 'unknown' compounds; 8Cl⁻
33 perfluorooctanesulfonic acid), at 0.8 ng mL⁻¹. The method was applied to two 'real-case' scenarios:
34 (1) the assessment of drinking water safety during a high-profile event in Brisbane, Australia; and (2)
35 to screen treated, re-circulated drinking water and pre-treated (raw) water. The validated workflow

36 was effective for rapid prioritisation and screening of suspect and non-target potential hazards at
37 trace levels, and could be applied to a wide range of matrices and investigations where comparison
38 of organic contaminants between an affected and control site and or timeframe is warranted.

39

40 **Key words:** suspect screening, water monitoring, LC-QTOF, data reduction strategy, hazard
41 identification

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42 1. INTRODUCTION

43 The World Health Organization attributed an estimated 4.9 million deaths to management of, and
44 exposure to, known chemicals in 2004[1]. Due to the large number of new chemicals registered
45 every year, and the relatively small proportion of which are thoroughly tested, the potential risk to
46 biota and human health is largely unknown[2]. Sources of hazardous chemicals include chemical
47 manufacturers, service stations, hazardous materials waste sites, and common household products
48 Environment[3]. The relatively uncharacterised nature of hazardous chemicals poses a serious
49 challenge for regulators in charge of safeguarding human health and environmental wellbeing.

50
51 Historically, analytical methods used for aquatic monitoring typically cover only a small fraction of
52 known, target chemicals. This approach is limited in situations where an issue of concern is
53 identified, such as deliberate or accidental chemical spills, or extreme weather events (e.g. floods,
54 heavy rain or droughts that can generate contaminant concentration pulses of ecotoxicological
55 relevance to the aquatic environment) but the link to a specific chemical hazard is unclear[4].
56 Recently advances in high resolution mass spectrometry (HRMS) and data processing software has
57 seen a rise in non-target analytical strategies[5-13], and particularly suspect screening[14-16], to
58 address the need for analysis of an increasing number of analytes in complex mixtures. 'Non-target
59 analysis' refers to detection and tentative identification of analytes for which chemical reference
60 standards are unavailable. 'Suspect screening' is a form of non-target analysis whereby analytes are
61 identified on the basis of accurate mass, elemental composition and structure prediction, followed
62 by database or library searching. 'Unknown' non-target analysis is an unbiased approach, and is
63 usually performed after targeted and suspect screening. It involves different data filtering strategies
64 to reduce the size of the search space, followed by assignment of probable chemical formula based
65 on MS/MS fragmentation and other strategies[17]. Non-target analyses have been used to
66 investigate contaminants in waste[6, 14] and surface waters[10, 11, 16, 18, 19], foodstuffs[20-22],
67 and forensic applications[11, 23, 24], but have not yet been applied in response to time-critical
68 environmental hazard assessment.

69
70 There is a need for analytical strategies capable of rapid non-target and suspect screening for
71 identification of hazardous chemicals, specifically in situations where exposure is unknown or
72 involves complex chemical mixtures, and requires an immediate response. Data reduction strategies
73 based on comparison of 'case' samples (which have an outcome of interest or concern), and 'control'
74 samples (which do not have the observed outcome/concern) can be used to rapidly analyse the large
75 amount of data generated during screening experiments using HRMS. The case-control approach has

76 been successfully used in proteomics and metabolomics studies[25, 26], but currently has limited
77 use in environmental monitoring applications, including water quality testing. Briefly, a peak-finding
78 algorithm is used to identify molecular features across different samples, followed by case-control
79 comparison to identify suspect features for subsequent identification by searching against available
80 spectral libraries, and to eliminate any matrix-specific interferences. The combination of accurate
81 mass data and statistical evaluation of sample constituents allow for the rapid extraction and
82 prioritisation of the most important chemical suspects for further identification.

83

84 Here we present a new approach for the rapid identification of unknown polar chemical hazards in
85 water, based on the post-acquisition comparison of samples in a case-control setting. High
86 resolution quadrupole time-of-flight tandem mass spectrometry (QTOF-MS/MS) is used together
87 with “smart” data-mining software to (1) develop a rapid case-control screening method to identify
88 the presence of potential hazardous chemicals; (2) validate the method using fortified water samples
89 and simulate a contamination event; and (3) apply the screening strategy to raw and drinking water
90 samples in two independent ‘real-case’ scenarios.

91

92 **2. EXPERIMENTAL METHODS**

93 **2.1 Chemicals and standards**

94 A standard working solution of 46 model compounds was prepared in methanol at 1 mg/L
95 concentration (Table S1). A surrogate standard containing 12 labelled compounds at 1 mg/L was
96 added prior to sample extraction, and used to monitor method performance; an injection standard
97 of acetylsulfamethoxazole-d4 at 10 ng/mL was added prior to injection and used to monitor
98 instrument performance. Calibration standard solutions were prepared in 20% methanol. All
99 reagents and standards were high purity analytical grade (refer to Supplementary Material).

100

101 **2.2 Sample preparation**

102 Model chemicals were fortified in 1 L drinking water. Target chemicals (Table S1) were fortified at
103 concentrations of 10, 5, 1, 0.5 and 0.1 ng mL⁻¹. Non-target chemicals (Table 1) were fortified in
104 samples at levels of 10, 1 and 0.2 ng mL⁻¹, with the exception of 8 Cl⁻ PFOS, which was fortified at 40,
105 4 and 0.8 ng mL⁻¹. All samples underwent two treatments: (1) a 1 mL aliquot was sampled, filtered,
106 and analysed immediately via direct injection (i.e. with no sample preconcentration); and (2) 500 mL
107 was pre-concentrated via solid phase extraction (SPE) using 6 cc Oasis HLB cartridges (Waters) to
108 increase sensitivity. All samples were filtered post-extraction using a 0.22 µm PTFE syringe filter
109 (Phenomenex) and transferred to 1.5 mL glass vials prior to analysis by LC-MS/MS. A procedural

110 blank, instrumental blanks and calibration curves were included with each batch of samples for
111 quality assurance/control purposes. Quantification of target compounds was performed using
112 labelled standards. For further details refer to Supplementary Material.

113

114 **2.3 LC/MS-MS parameters**

115 Samples were analysed with a Shimadzu Nexera X2 UHPLC system equipped with a binary pump and
116 reverse-phase XDB-C18 analytical column, coupled to a 5600 TripleTOF mass spectrometer (SCIEX,
117 Melbourne, Australia) and equipped with electrospray ionization interface working in positive and
118 negative ionization modes. In negative mode chromatographic separation was achieved using a
119 reverse-phase XDB-C18 analytical column (4.6×50 mm, 1.8µm; Agilent Technologies, Santa Clara, CA)
120 maintained at 45°C, with a flow rate of 0.6 mL min⁻¹. Mobile phases consisted of: (A) methanol:water
121 (99:1, v/v); and (B) methanol:water (90:10, v/v); with 5mM ammonium acetate in both phases, with
122 a gradient ramp as follows: 0min, 10%B; 0.2min, 10%B; 6.50min, 100%B; 9.50min 100%B; 9.6min,
123 10%B followed by equilibration at initial conditions for 2.20min. In positive mode, separation was
124 achieved using a XDB-C18 column (2.1×100 mm, 1.8 µm; Agilent Technologies, Santa Clara, CA)
125 maintained at 50°C, with a flow rate of 0.4 mL min⁻¹. Different chromatographic columns were used
126 for positive and negative ionization modes that were selected following analytical column validation
127 tests (including injection volume tests). For all the compounds investigated chromatographic
128 performance including sensitivity and mass resolution for all analytes were very satisfactory in both
129 ionization modes and with both analytical columns. Mobile phases consisted of: (A) 100% ultrapure
130 water; (B) methanol, with 0.1% formic acid in both phases; with a gradient ramp as follows: 0min,
131 5%B; 0.2min, 5%B; 10.2min, 100%B; 14.7min 100%B; 14.9min, 10%B followed by equilibration at
132 initial conditions for 2.20min. Injection volume was 10 µL and 5 µL in negative and positive mode,
133 respectively. The ion source parameters were optimized as follows: source voltage, -4500 V and
134 5500 V for negative and positive ionization, respectively; temperature, 600°C; curtain gas, 35 L min⁻¹;
135 and ion source gas at 70 psi. High purity nitrogen was used as the nebulizer gas, curtain gas and
136 collision gas.

137

138 MS and MS/MS data was acquired in high-sensitivity mode with both data-dependent (information
139 dependent acquisition, IDA) and data independent (Sequential Window Acquisition of all Theoretical
140 fragment-ion spectra, SWATH) modes. Data was processed with PeakView®, MS Library and
141 MultiQuant software (SCIEX). Confirmation of target analytes was based on retention time
142 (±0.5min), accurate mass (mass error <5ppm; mass error score >80%), isotopic distribution (isotope
143 score >60%) and automatic MS/MS library searching (library score >70%). SWATH data was

144 processed both with and without product fragment ions to inform the search sequence. Extracted
145 ion chromatogram (XIC) parameters were set to >300, corresponding to a signal-to-noise ratio of
146 five, and XIC width was set to 0.01 Da. Confirmation of compounds not included *a priori* in the
147 analytical method (hereon in referred to as 'non-target compounds') and structural characterization
148 of compounds not found in spectral libraries (hereon in referred to as 'unknown' compounds) was
149 based on accurate mass (mass error <5 ppm), elemental composition assignment, isotopic pattern
150 distribution, ring and double bonds (RDB) factor and MS/MS spectrum interpretation. *In-silico*
151 fragmentation with a mass tolerance threshold <10 ppm was used for structural elucidation of
152 unknown analytes. See Supplementary Material for further description of instrument parameters,
153 data processing and compound identification.

154

155 **2.4 Suspect screening with case-control filter**

156 A sample collection and data processing strategy based on case and control samples was developed.
157 Control samples were collected prior to an *event*, and reflected historical or baseline chemical
158 composition. Case samples were collected during an *event*, and captured chemical composition
159 during the period of interest. Case and control samples were extracted and analysed simultaneously
160 to minimize variability. Suspect screening was performed by comparing experimental spectra with a
161 commercial MS/MS spectral library (SCIEX) containing ~2900 common aquatic organic micro-
162 pollutants representing structurally diverse chemical classes. To create the 'filter', the intensity ratio
163 of case to control was calculated for each suspect. For suspects where the peak intensity of case
164 relative to control was ≤ 3 , the suspect was 'filtered' out, as the difference between case and control
165 was deemed negligible. This strategy allowed for a rapid and substantial reduction in the number of
166 suspect masses investigated.

167

168 **2.5 Method Application**

169 *2.5.1 Rapid screening of drinking water to identify potential hazards during an event*

170 From 15-16 November 2014, meetings of the Group of Twenty (G20), which includes leaders from 20
171 major economies, took place in Brisbane, Australia. To test and safeguard the Brisbane city drinking
172 water, potable water samples were analysed using a case-control approach and reported daily.
173 Samples from a key water treatment plant were collected daily from 3-17 November 2014 and
174 represented the 'case' samples (n=2 per day; n=30 total). Samples were collected from the same
175 treatment plant prior to the event on 30 October 2014 to represent the 'control' sample (n=4).

176

177 *2.5.2 Risk management of treated drinking water samples*

178 To screen for chemicals of concern, pre- and post-treatment spot water samples were collected
179 from nine water treatment facilities in the South East Queensland region in Australia, from 7-10 April
180 2015. Sampling sites were supernatant ponds containing treated, re-circulated water (case), and pre-
181 treated (raw) water (control) ($n=2$ from each sampling site; $n=36$ total). Treated samples were
182 processed through conventional means of coagulation, flocculation, sedimentation and filtration.

183

184 3. RESULTS AND DISCUSSION

185 3.1 Validation of the analytical method

186 The analytical method was optimised and validated using a range of 46 compounds covering
187 different physico-chemical properties (Table S2), including 31 pesticides/herbicides and 15
188 pharmaceuticals and personal care products. Recovery was calculated by comparing samples
189 (drinking water from a dam, and ultrapure water; $n=5$ replicates) fortified pre- and post-extraction at
190 1 ng mL^{-1} , and ranged from 20 to 109 %. Linearity was established from 0.1 to 50 ng mL^{-1} using least-
191 squares regression, with $R^2 > 0.99$ in all cases. Method quantification limits (MQLs) were determined
192 as the concentration of the lowest standard that produced a peak signal ten times the background
193 noise, and ranged from 0.1 to 1 ng mL^{-1} . Intra- and inter-day precision were determined from
194 triplicate analyses of samples fortified at 5 ng mL^{-1} , and were $< 11 \%$ RSD (Table S2).

195

196 3.2 Method development for identification of chemical hazards

197 3.2.1 Data acquisition and filtering strategy

198 A conceptual framework for the analysis and identification of potentially hazardous chemicals is
199 presented in Figure 1. The strategy is divided into five components including sample preparation and
200 analysis (described in Experimental Methods); and data acquisition, data processing and compound
201 identification, discussed below.

202

203 Data was acquired in both IDA and SWATH acquisition modes, as described above and both methods
204 successfully screened and identified all 46 compounds, with similar performance. However, it is
205 important to note that for the compounds investigated via SWATH, relatively high concentrations
206 with minimal matrix effects (i.e. high signal-to-noise ratio) produced high quality MS and MS/MS
207 spectra, facilitating successful identification. In the case of lower concentration analytes, or more
208 complex matrices, this may not always be the case as non-selective fragmentation may hamper
209 identification[27]. Deconvolution of characteristic low m/z fragments typically used for identification
210 of environmental pollutants could be improved, and a second injection with IDA may be required for
211 tentative identification of some compounds (Figure 1). Additionally, because of the wider isolation

212 window in SWATH compared to IDA, more ions are transferred to the collision cell for
213 fragmentation, but some ions may still be missed. For example, no MS/MS was recorded for the
214 most intense peak (249.0244 m/z) of the non-target compound bisphenol S. In order to record all
215 product ions for all precursor ions, a wide pass MS^{All} approach would be required. For both IDA and
216 SWATH experiments the quality of mass spectra (and hence the closeness of the library match)
217 decreased with decreasing chemical concentration - at fortification levels of 10, 0.5 and 0.1 ng mL⁻¹,
218 positive library matches from direct injection samples were returned for 46, 42 and 29 compounds,
219 respectively.

220

221 *3.2.2 Rapid identification of target compounds*

222 With non-specific screening methods, it is essential to incorporate parameters that give the greatest
223 breadth and depth of coverage for compound identification, without comprising data quality.
224 Collision energy and declustering potential were optimized as 35±15 eV and ±80V (for negative and
225 positive ionisation modes), respectively, as these parameters gave the best results (high quality MS
226 and MS/MS) for the greatest number of compounds. Notwithstanding, these generic parameters did
227 not generate the best quality spectra for each individual analyte, and in some cases, it was necessary
228 to perform a second injection, increasing the collision energy to obtain adequate fragmentation for
229 subsequent identification.

230

231 Using a test solution of known composition, and percentage of detection fails as the endpoint,
232 software parameters were systematically varied and the results assessed to evaluate the reasons for
233 compound identification failures in each case. Failures were typically attributed to: low or no library
234 match; low or no isotope score; and high mass error scores. Greater failures in library matching were
235 observed at the lower concentration range (SI Figure S1). A full description of parameters used for
236 compound identification is included in the Supplementary Material. It is important to note that
237 identification of unknown chemicals can only be regarded as tentative until confirmed (retention
238 time, accurate mass, and mass spectra) with an analytical standard, and tentative identification is
239 subject to analyst bias and experience [5, 28].

240

241 **3.3 Chemical hazard identification under simulated conditions**

242 To test the capability of the developed approach, a contamination incident was simulated by
243 fortifying water samples with known contaminants, (case), with unperturbed drinking water serving
244 as the control. The level of difficulty and reliability of the identification approach increased from
245 scenario (a) to scenario (d), Figure 1, and is described in detail below:

246

247 *3.3.1 Scenario A: Identification of target chemicals*

248 The first scenario was directed at identifying target compounds that are present in both commercial
249 MS/MS libraries and for which chemical standards are available in a typical laboratory setting (or can
250 be easily acquired). Using the strategy proposed in Figure 1, identification of 46 fortified model
251 compounds in drinking water samples (Table S1) by direct injection was assessed using optimised
252 SWATH and IDA acquisition methods. Results showed $\geq 90\%$ of target chemicals were positively
253 matched against MS/MS library at 1 ng mL^{-1} , $\leq 86\%$ at 0.5 ng mL^{-1} and $\leq 59\%$ at 0.1 ng mL^{-1} . At 10 ng
254 mL^{-1} all target chemicals were positively library matched. When comparing SWATH data, processing
255 with and without the inclusion of fragment ions search function, data with fragment ion inclusion
256 slightly outperformed data without fragment ion inclusion (Figure S1).

257

258 *3.3.2 Scenario B: Suspect screening of chemicals*

259 The second scenario identified compounds that were present in a commercial MS/MS library, but for
260 which standards were not held by the investigating laboratory. The fungicide fipronil and insecticide
261 fenitrothion were chosen for this simulation. A positive identification and library match for fipronil
262 was generated for samples fortified at 0.2 ng mL^{-1} , and for fenitrothion at 1 ng mL^{-1} . Tentative
263 identification of fipronil was confirmed with mass error score (91.1), isotope score (96.6), combined
264 score (93.1), library score (100), and mass error (-0.9 ppm). An example for the identification of
265 fipronil with TOF-MS and TOF-MS/MS library match is provided in Figure 2. For fenitrothion at 1 ng
266 mL^{-1} , the relative intensity of case to control was less than 3, and thus the analyte would nominally
267 be excluded from further analysis. However, it is important to note that in a real incident scenario,
268 such as a chemical spill or toxic event, it is possible that contaminants would be present at much
269 higher concentrations.

270

271 *3.3.3 Scenario C: Identification of non target compounds*

272 The third scenario was aimed at identifying non target compounds, for which compound structures
273 were available via online chemical databases (e.g. ChemSpider, Pubchem), but not in commercial
274 spectral libraries. In this instance the flame-retardant tetrabromobisphenol A (TBBPA), the anti-
275 corrosive 4,4'-sulfonyldiphenol, and the plasticiser hexafluorobisphenol A (BPAF) were chosen for
276 the simulation, and fortified at 0.2 to 40 ng mL^{-1} . In this scenario an accurate mass was measured for
277 the chemicals, but no positive identification was possible because there was no spectral library
278 match available for these compounds. Therefore, further investigation was required.

279

280 An example of this process using bisphenol S is shown in Figure 3. Four theoretical elemental
281 compositions were generated based on accurate mass (M-H ion, 249.02344 m/z), and were
282 prioritised for further investigation by selecting compounds with the smallest mass error and the
283 highest MS/MS rank. Probable chemical structures were generated for the highest priority elemental
284 composition (C₁₂H₁₀O₄S) with the aid of ChemSpider. Each structure was fragmented *in silico*, and the
285 theoretical fragmentation pattern compared with the experimentally-derived MS/MS for
286 congruence. We note that a reference spectrum for bisphenol S is now available in MassBank (but
287 was not at the time of analysis), and could be used in place of *in silico* fragmentation pattern. This
288 step was critical as it allowed for tentative compound identification, despite no chemical standard
289 being available. Generally, when searching for unknown compounds, elemental compositions that
290 yield the highest number of results, and/or database entries that contain common chemical names
291 are likely to be primary suspects for tentative identification, but does not always yield a successful
292 result.

293

294 3.3.4 Scenario D: Identification unknown compounds

295 The final scenario was directed at 'unknown compounds', compounds that have not previously been
296 described in the literature, and are not listed in any chemical database (i.e. chemical structures are
297 not available). This may be the case for new chemical formulations, by-products or transformation
298 products formed from parent species, for example, and is an important new area of research, as in
299 some cases these secondary products are more toxic than the parent[29, 30]. For this scenario a
300 newly discovered and synthesised by-product of the perfluorinated flame retardant
301 perfluorooctanesulfonic acid (PFOS), 8Cl PFOS[31], was used for the simulation, and was detected at
302 the lowest fortification level (0.8 pg mL⁻¹).

303

304 Elucidation of the chemical structure of this compound is shown in Figure 4. The 'Formula Finder'
305 algorithm generated five possible elemental compositions. Evidence from the isotopic profile of MS
306 and MS/MS reflected the natural isotopic abundance of a single chlorine (MS; 514.8988 m/z and
307 514.8956 m/z, with 30% relative intensity), and single sulfur atom (MS/MS; 79.9598 m/z and
308 82.9628 m/z, with 4% relative intensity), respectively; and a mass defect >0.9 confirmed the
309 presence of fluorine[31, 32]. Together, this suggested C₈HClF₁₆O₃S as the most likely chemical
310 formula. Fragments at 329.9470, 279.9463 and 229.9489 m/z indicate sequential loss of [CF₂]⁻, and
311 ring/double bond factor of zero suggest a linear structure, as shown in Figure 4.

312

313 **Table 1. List of non-target and unknown compounds, their classification and properties.**

Compound name	Classification	Molecular formula	CAS #	Molecular weight	Speciation	Availability in chemical libraries
Tetrabromobisphenol A (TBBPA)	flame retardant	C ₁₅ H ₁₂ Br ₄ O ₂	79-94-7	540.75	negative	present in online database (e.g.ChemSpider)
4,4'-Sulfonyldiphenol (BPS)	anticorrosive	C ₁₂ H ₁₀ O ₄ S	80-09-1	249.02	negative	present in online database (e.g.ChemSpider)
Hexafluorobisphenol A (BPAF)	polymer applications	C ₁₅ H ₁₀ F ₆ O ₂	1478-61-1	335.05	negative	present in online database (e.g.ChemSpider)
Fipronil	fungicide	C ₁₂ H ₄ Cl ₂ F ₆ N ₄ OS	120068-37-3	434.93	negative	present in commercial chemical library
Fipronil-sulfone*	fungicide degradation products	C ₁₂ H ₄ Cl ₂ F ₆ N ₄ O ₂ S	120068-36-2	450.93	negative	present in commercial chemical library
Fenitrothion	phosphorothioate insecticide	C ₉ H ₁₂ NO ₅ PS	122-14-5	278.02	positive	present in commercial chemical library
Chlorinated PFOS (8 Cl-PFOS)	PFOS by-product, perfluorinated flame retardant	C ₈ H ₀ O ₃ F ₁₆ Cl	N/A	514.90	negative	Unknown, not present in any library

* Fipronil-sulfone was not fortified in sample but detected as the degradation product of fipronil

314

315 **3.4 Application of the method in real case studies**

316 *3.4.1 Case 1: Drinking water during an incident event*

317 During the high profile G20 summit of world leaders held in Brisbane in November 2014, local water
318 authorities were charged with ensuring the integrity of the city's drinking water supplies. A rapid,
319 sensitive and effective water screening approach was therefore required to identify any substances
320 that could pose a threat to consumers.

321

322 Immediately prior to and during the event, drinking water samples (case samples) were collected
323 and screened daily, with results reported at the end of each day. Eight hundred and twenty four
324 spectral features were isolated in the control sample collected prior to the event. Of these,
325 doxylamin, fludroxycortide, mirtazapine, and morphine were detected and tentatively identified via
326 direct injection; and atrazine, caffeine, DEET, fludroxycortide, valacyclovir, PFOA, and PFOS were
327 identified in SPE pre-concentrated samples. On average, up to six suspect detects were observed in
328 case samples. However, these were chemicals that were already present in the 'control' sample, but
329 were flagged due to higher intensity ratios (>3 ratio) in case samples, possibly the result of inter-
330 day variability in drinking water source and supply. We note that no suspicious compounds were
331 detected in samples over the G20 sampling period.

332

333 Observations from this sampling campaign highlight the importance of obtaining a control sample
334 with the exact matrix composition as the case. This allows for the most effective results, i.e.
335 optimum elimination of analytes of non-concern, and focusing of attention on a significantly reduced
336 number of analytes of potential concern. The combination of direct injection analysis and analysis of
337 pre-concentrated samples provided sensitive and comprehensive coverage.

338

339 *3.4.2 Case 2: Risk management of treated drinking water samples*

340 The case-control method was successfully applied in a water treatment risk management strategy
341 aimed at screening raw and treated/recirculated drinking water samples for identification of
342 potential hazards. Raw water samples represented the 'control' sample and treated/recirculated
343 water represented the 'case' samples to identify any hazards in treated water. The processing of the
344 samples was then reversed (i.e. treated water represented the 'control' samples while the raw water
345 samples represented the 'case') to evaluate water treatment removal efficiency. A small number of
346 non-target chemicals were detected in both raw and treated samples at one water treatment facility
347 (Table S3, Figure S2). These included the fungicide fipronil and its degradation product fipronil
348 sulfone; plasticizers dioctyl phthalate (DEHP) and dibutyl phthalate; and the albertoxin altenuene.

349 DEHP is the most widely used plasticiser, and was detected at five times higher levels in
350 treated/recirculated water samples, indicating possible addition of this chemical to drinking water
351 during the water treatment process.

352

353 **4. CONCLUSIONS**

354 Here we present a rapid, case-control screening approach to identify non-target and unknown
355 chemical hazards, with novel application to water samples in different scenarios. This case-control
356 strategy allowed us to focus our efforts on the identification of suspect compounds related to a case
357 (e.g. incident event), by filtering spectral features by intensity ratio, thereby dramatically reducing
358 data processing time. The methodology was successful in screening a large group of chemicals
359 (n=46) with various physio-chemical properties in drinking water, with >90% and >60% of model
360 compounds tentatively identified at ultra-trace levels of 0.5 and 0.1 ng mL⁻¹, respectively. The case-
361 control screening approach was efficient and accurate in reducing the complexity of processing
362 HRMS data to identify the pollutants of concern in pre- and post-treated drinking water, and surface
363 waters, and has potential for application in many other situations and matrices including food
364 contamination and biota.

365

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367 The authors gratefully acknowledge Vitukawalu Matanitobua, Kelvin O'Halloran, Andrew Banks and
368 Andrew Novic for field work and sample preparation; Chris Hodgkins for software support; and
369 funding support from the Australian Research Council (FF120100546; LE140100129) and the
370 Queensland Department of Health.

371

372 **SUPPLEMENTARY MATERIAL**

373 The Supplementary Material includes a detailed description of the experimental methods and
374 compound identification, list of the 46 model compounds used and their classification and
375 properties, validation parameters for the analytical method, and a list (and example) of the non-
376 target and unknown chemicals detected in raw and treated/recirculated drinking water at a water
377 treatment facility.

378

379 REFERENCES

- 380 1. Prüss-Ustün, A., et al., *Knowns and unknowns on burden of disease due to chemicals: a systematic*
381 *review*. Environmental Health, 2011. **10**: p. 9-9.
- 382 2. Muir, D.C. and P.H. Howard, *Are there other persistent organic pollutants? A challenge for*
383 *environmental chemists*. Environ Sci Technol, 2006. **40**(23): p. 7157-66.
- 384 3. European Environment Agency, E.E., *Hazardous substances in Europe's fresh and marine waters, in*
385 *EEA Technical Report 2011*.
- 386 4. Escher, B.I., et al., *Most Oxidative Stress Response In Water Samples Comes From Unknown Chemicals:*
387 *The Need For Effect-Based Water Quality Trigger Values*. Environmental Science & Technology, 2013.
388 **47**(13): p. 7002-7011.
- 389 5. Herrera-Lopez, S., et al., *Simultaneous screening of targeted and non-targeted contaminants using an*
390 *LC-QTOF-MS system and automated MS/MS library searching*. J Mass Spectrom, 2014. **49**(9): p. 878-
391 93.
- 392 6. Gomez, M.J., et al., *Rapid automated screening, identification and quantification of organic micro-*
393 *contaminants and their main transformation products in wastewater and river waters using liquid*
394 *chromatography-quadrupole-time-of-flight mass spectrometry with an accurate-mass database*. J
395 Chromatogr A, 2010. **1217**(45): p. 7038-54.
- 396 7. Gómez, M.J., et al., *Automatic Searching and Evaluation of Priority and Emerging Contaminants in*
397 *Wastewater and River Water by Stir Bar Sorptive Extraction followed by Comprehensive Two-*
398 *Dimensional Gas Chromatography-Time-of-Flight Mass Spectrometry*. Analytical Chemistry, 2011.
399 **83**(7): p. 2638-2647.
- 400 8. Gomez, M.J., et al., *Spatio-temporal evaluation of organic contaminants and their transformation*
401 *products along a river basin affected by urban, agricultural and industrial pollution*. Sci Total Environ,
402 2012. **420**: p. 134-45.
- 403 9. Krauss, M., H. Singer, and J. Hollender, *LC-high resolution MS in environmental analysis: from target*
404 *screening to the identification of unknowns* Analytical and Bioanalytical Chemistry, 2010. **397**(3): p.
405 943-951.
- 406 10. Ibáñez, M., et al., *Rapid non-target screening of organic pollutants in water by ultraperformance liquid*
407 *chromatography coupled to time-of-flight mass spectrometry*. TrAC Trends in Analytical Chemistry,
408 2008. **27**(5): p. 481-489.
- 409 11. Bletsou, A.A., et al., *Targeted and non-targeted liquid chromatography-mass spectrometric workflows*
410 *for identification of transformation products of emerging pollutants in the aquatic environment*. TrAC
411 Trends in Analytical Chemistry, 2015. **66** p. 32-44.
- 412 12. Schymanski, E.L., et al., *Non-target screening with high-resolution mass spectrometry: critical review*
413 *using a collaborative trial on water analysis*. Anal Bioanal Chem, 2015. **407**(21): p. 6237-55.
- 414 13. Baz-Lomba, J.A., M.J. Reid, and K.V. Thomas, *Target and suspect screening of psychoactive substances*
415 *in sewage-based samples by UHPLC-QTOF*. Anal Chim Acta, 2016. **31**(914): p. 81-90.
- 416 14. Gago-Ferrero, P., et al., *Extended Suspect and Non-Target Strategies to Characterize Emerging Polar*
417 *Organic Contaminants in Raw Wastewater with LC-HRMS/MS*. Environ. Sci. Technol., 2015. **49**(20): p.
418 12333-12341.
- 419 15. Pochodyloa, A.L. and D.E. Helbling, *Emerging investigators series: prioritization of suspect hits in a*
420 *sensitive suspect screening workflow for comprehensive micropollutant characterization in*
421 *environmental samples*. Environ. Sci.: Water Res. Technol., 2017. **3**: p. 54-65.
- 422 16. Badea, R., et al., *Suspect screening of large numbers of emerging contaminants in environmental*
423 *waters using artificial neural networks for chromatographic retention time prediction and high*
424 *resolution mass spectrometry data analysis*. Sci. Total Environ., 2015. **538**: p. 934-941.
- 425 17. Andra, S.S., et al., *Trends in the application of high-resolution mass spectrometry for human*
426 *biomonitoring: An analytical primer to studying the environmental chemical space of the human*
427 *exposome* Environment International 2017. **100**: p. 32-61.
- 428 18. Gomez, M.J., et al., *Photodegradation study of three dipyrone metabolites in various water systems:*
429 *identification and toxicity of their photodegradation products*. Water Res, 2008. **42**(10-11): p. 2698-
430 706.
- 431 19. Ruff, M., et al., *Quantitative target and systematic non-target analysis of polar organic micro-*
432 *pollutants along the river Rhine using high-resolution mass-spectrometry – Identification of unknown*
433 *sources and compounds*. Water Research, 2015. **87**: p. 145-154.

- 434 20. Gomez-Ramos, M.M., et al., *Liquid chromatography-high-resolution mass spectrometry for pesticide*
435 *residue analysis in fruit and vegetables: screening and quantitative studies*. J Chromatogr A, 2013.
436 **1287**: p. 24-37.
- 437 21. Gomez-Ramos, M.M., et al., *Screening of environmental contaminants in honey bee wax comb using*
438 *gas chromatography-high-resolution time-of-flight mass spectrometry*. Environ Sci Pollut Res Int,
439 2016. **23**(5): p. 4609-20.
- 440 22. He, Z., et al., *Wide-scope screening and quantification of 50 pesticides in wine by liquid*
441 *chromatography/quadrupole time-of-flight mass spectrometry combined with liquid*
442 *chromatography/quadrupole linear ion trap mass spectrometry*. Food Chem, 2016. **196**: p. 1248-55.
- 443 23. Fels, H., et al., *Liquid chromatography-quadrupole-time-of-flight mass spectrometry screening*
444 *procedure for urine samples in forensic casework compared to gas chromatography-mass*
445 *spectrometry*. Drug Test Anal, 2016.
- 446 24. Montesano, C., et al., *Broad Screening and Identification of Novel Psychoactive Substances in Plasma*
447 *by High-Performance Liquid Chromatography-High-Resolution Mass Spectrometry and Post-run*
448 *Library Matching*. J Anal Toxicol, 2016. **40**(7): p. 519-28.
- 449 25. Zhang, A., et al., *Metabolomics in diagnosis and biomarker discovery of colorectal cancer*. Cancer Lett,
450 2014. **345**(1): p. 17-20.
- 451 26. Mapstone, M., et al., *Plasma phospholipids identify antecedent memory impairment in older adults*.
452 Nat Med, 2014. **20**(4): p. 415-8.
- 453 27. Zhu, X., Y. Chen, and R. Subramanian, *Comparison of Information-Dependent Acquisition, SWATH, and*
454 *MSAll Techniques in metabolite Identification Study Employing Ultrahigh-Performance Liquid*
455 *Chromatography-Quadrupole Time of-Flight Mass Spectrometry*. Analytical Chemistry, 2014. **86**: p.
456 1202-1209.
- 457 28. Hug, C., et al., *Identification of novel micropollutants in wastewater by a combination of suspect and*
458 *nontarget screening*. Environ Pollut, 2014. **184**: p. 25-32.
- 459 29. Donner, E., et al., *Ecotoxicity of carbamazepine and its UV photolysis transformation products*. Sci
460 Total Environ, 2013. **443**: p. 870-6.
- 461 30. Escher, B.I. and K. Fenner, *Recent Advances in Environmental Risk Assessment of Transformation*
462 *Products*. Environmental Science & Technology, 2011. **45**(9): p. 3835-3847.
- 463 31. Rotander, A., et al., *Novel fluorinated surfactants tentatively identified in firefighters using liquid*
464 *chromatography quadrupole time-of-flight tandem mass spectrometry and a case-control approach*.
465 Environ Sci Technol, 2015. **49**(4): p. 2434-42.
- 466 32. Myers, A.L., et al., *Using mass defect plots as a discovery tool to identify novel fluoropolymer thermal*
467 *decomposition products*. J Mass Spectrom, 2014. **49**(4): p. 291-6.

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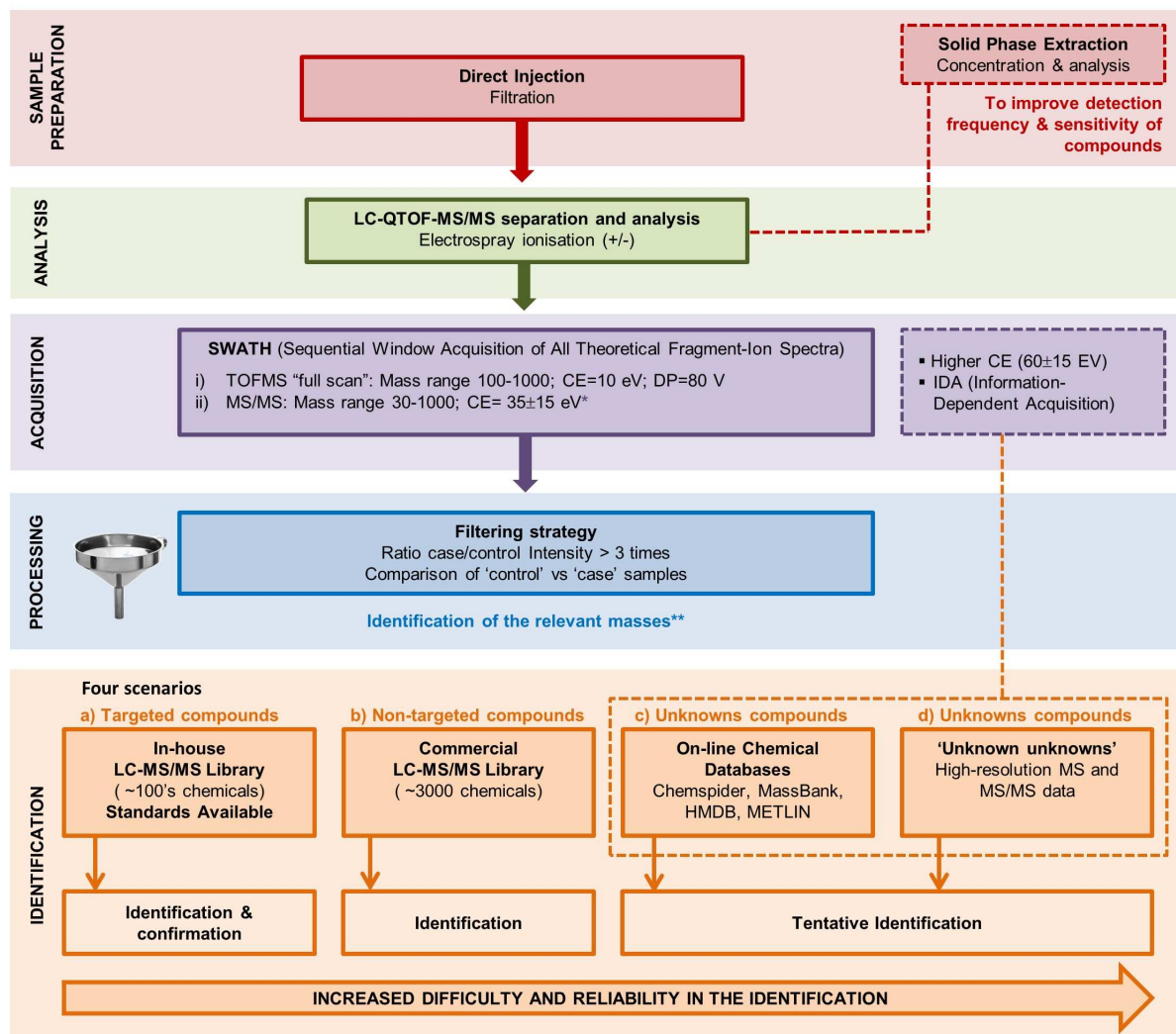
LIST OF FIGURES

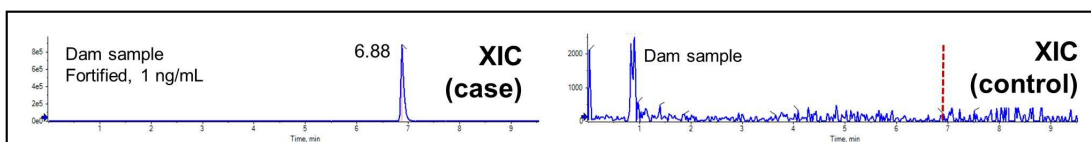
Figure 1. Conceptual framework and workflow for identification of contaminants based on case-control strategy

Figure 2. The identification of the fortified non-target compound fipronil using commercial library (SCIEX) showing a positive MS/MS library match.

Figure 3. Identification of the unknown compound bisphenol S (BPS) using a public chemical database (ChemSpider)

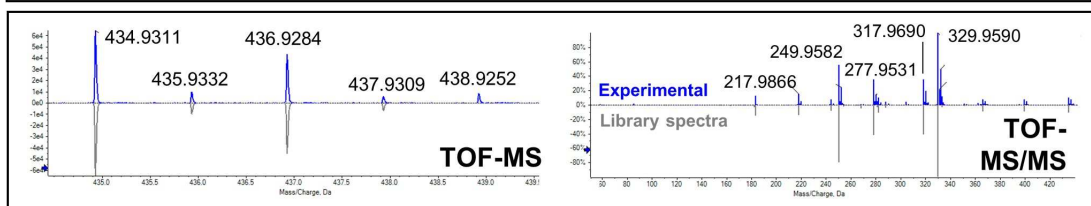
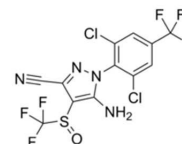
Figure 4. Identification of unknown-unknown compound (8Cl-PFOS) not previously recorded in any commercial library or on-line chemical database.



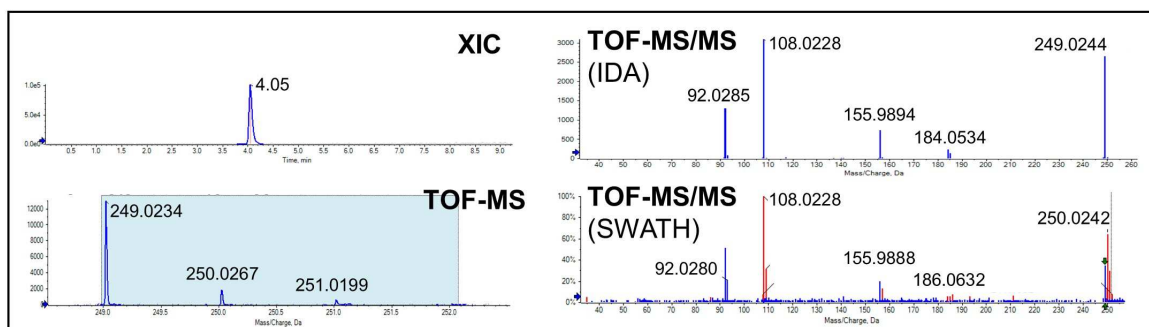

FILTERING RESULTS Ratio case/control = 3

Control Intensity	Intensity	Name	Formula	Mass (Da)	Extraction Mass (Da)	Isotope	Found At Mass (Da)	Mass Error Score	Isotope Score	Combined Score	Error (ppm)	Library Score
11436	1036814	Fipronil	C₁₂H₄Cl₂F₆N₄O₂S	435.93872	434.93144	0	434.93106	91.1	96.6	93.1	-0.9	100
3138	24178	Fipronil-sulfone	C ₁₂ H ₄ Cl ₂ F ₆ N ₄ O ₂ S	451.93362	450.92635	0	450.92627	98.4	83.8	81.7	-0.2	97.1
1100	114453			249.02292	249.02292	0	249.02344			23	2.1	0
1022	71035			282.98331	282.98331	0	282.98379			31.8	1.7	0
2479	227188			335.0509	335.0509	0	335.05108			35.1	0.5	0
1573	58660			369.01097	369.01097	0	369.01193			41	2.6	0
768	1036814			434.93114	434.93114	0	434.93083			48.3	-0.7	0
949	714460			436.92808	436.92808	0	436.92843			46.3	0.8	0
350	142121			438.92495	438.92495	0	438.92529			27.1	0.8	0
825	331005			514.89995	514.89995	0	514.90036			42.8	0.8	0
799	117551			516.89697	516.89697	0	516.89736			39.5	0.8	0

Proposed Structure:
Fipronil



Name	Exp. RT (min)	Elemental Composition	Theoretical mass (Da)	Exp. Mass (Da)	Mass Error (ppm)	Isotope Ratio Difference (%)	Mass Error Score	Isotope Score	Library Score	Combined Score
Fipronil	6.89	C ₁₂ H ₄ Cl ₂ F ₆ N ₄ O ₂ S	434.9311	434.93102	-1.0	9	90.3	87.2	96.1	90.5



ELEMENTAL COMPOSITION

Hit	Formula	m/z	RDB	ppm	MS Rank	MSMS Rank	MSMS Rank	Found
1	C7H10F4O3S	249.0214	1.0	8.0	3	7.3 (10)	1	NA/0
2	C5H10F3N2O4P	249.0258	1.0	-9.5	4	8.5 (11)	2	NA/2
3	C9H11FO5S	249.0238	4.0	-1.8	2	14.2 (11)	3	NA/1
4	C12H10O4S	249.0227	8.0	2.8	1	13.3 (10)	4	NA/45

MS Details | MSMS Details | Compound Details

Isotope cluster details Charge: -1

Peak Use m/z % Intensity Width

Elements from

Elements to C40 H60 N5 O10 S2 F20 P

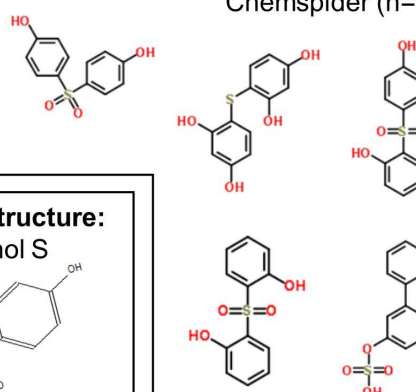
Mass tolerance (ppm) 10

Intensity tolerance (%) 10

#C/#heteroatoms greater than 0

DATABASE MATCHES

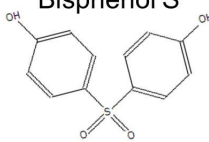
Chemspider (n=45)



IN SILICO FRAGMENTATION

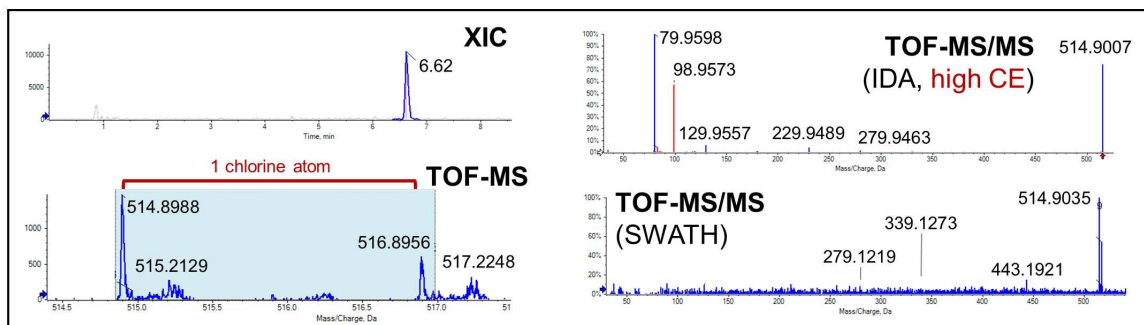
Fragment	m/z	Num H	Broken Bonds	Bond Closure	Rad.	Error (ppm)	Composition
92.0268	0	1	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	13.2	C6H4O-
93.0346	1	1	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	46.6	C6H5O-
155.9887	1	1	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	1.1	C6H4O3S-
249.0227	0	0	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	6.8	C12H9O4S-

Num. fragments: 4

Proposed structure:
Bisphenol S

Linked to MS/MS

Theoretical mass (Da)	Exp. mass (Da)	Proposed Formula	Mass error (ppm)	RDB	Database structures found	Fragments (relative abundance)	Elemental composition (error, ppm)
249.0227	249.0234	C12H10O4S	MS: 2.8 MS/MS: 13.3	8.0	45	108.0220 (100) 249.0244 (87) 155.9887 (25) 92.0268 (42) 184.0530	C6H4O2- (2.9) [M-H]- (6.9) C6H4O3S- (4.9) C6H4O- (13.2) C12H8O2- (2.3)



ELEMENTAL COMPOSITION

Hit	Formula	m/z	RDB	ppm	MS Rank	MSMS Rank	MSA Rank	Found
1	C8H4ClF11N2O5S2	514.9002	2.0	-2.7	1	7.6 (19)	2	NA/0
2	C9H8ClF10O5PS2	514.9007	1.0	-3.7	2	8.3 (26)	1	NA/0
3	C8HClF16O3S	514.9007	0.0	-3.6	3	5.8 (3)	3	NA/0
4	C10H6ClF8N2O5PS2	514.8944	5.0	8.6	4	17.4 (26)	4	NA/0
5	C10H3ClF13O3PS	514.8949	3.0	7.6	5	16.9 (15)	5	NA/0

MS Details MSMS Details Compound Details

Isotope cluster details Charge: -1

Elements from: C60 H50 N2 O5 S2 F40 P Cl

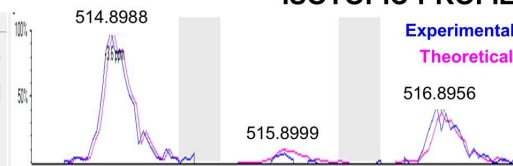
Elements to: C60 H50 N2 O5 S2 F40 P Cl

Mass tolerance (ppm): 10

Intensity tolerance (%): 10

#C/#heteroatoms greater than: 0

ISOTOPIC PROFILE



No database or library matches

IN SILICO FRAGMENTATION

Fragments	Peaks	m/z	Num H	Broken Bonds	Bond Closure	Rad.	Error (ppm)	Composition
		116.9724	2	2			12.6	C2 H Cl F3-
		116.9724	2	2			12.6	C2 H Cl F3-
		129.9542	0	1			11.7	C F2 O3 S-
		179.9510	0	1			15.8	C2 F4 O3 S-
		229.9478	0	1			4.8	C3 F6 O3 S-
		279.9446	0	1			6.3	C4 F8 O3 S-
		329.9414	0	1			17.1	C5 F10 O3 S-

Num. fragments: 10

Proposed structure: 8Cl-PFOS



PFOS with one fluorine atom substituted by one chlorine atom

Theoretical mass (Da)	Exp. mass (Da)	Proposed Formula	Mass error (ppm)	RDB	Database structures found	Fragments (relative abundance)	Elemental composition (error, ppm)
514.9007	514.8988	C8HCIF16O3S	MS: -3.6 MS/MS: 5.3	0.0	0	79.9591 (100) 514.9053 (74) 98.9565 (58) 129.9557 (6) 229.9489 (4) 279.9463 (2)	SO3- (21.1) [M-H]- (9.0) O3SF- (7.5) CF2O3S- (11.7) C3F6O3S- (4.8) C4F8O3S- (6.3)

- Non-target suspect screening of polar contaminants in water matrices
- 'Case-control' data processing to efficiently reduce HRMS data complexity
- Rapid, <24h response time to chemical hazard screening in real-life case studies
- >90% of target compounds (n=46) positively screened in samples at 1 µg/L

ACCEPTED MANUSCRIPT