



Microsatellite analysis of populations of the endangered tree Gomortega keule suggests pre-Columbian differentiation

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Keywords:	<i></i> Gomortega keule <i></i> , population genetics, fragmentation, genetic structure, megafaunal extinction, relict populations





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- 2 keule suggests pre-Columbian differentiation 3 ¹ Diego Muñoz-Concha * 4 ² Michael R Davey 5 ² Gracia Ribas 6 ^{2,3} Sean Mayes 7 8 9 ¹ Departamento de Ciencias Agrarias, Universidad Católica del Maule, Curicó, 10 Chile ² Plant and Crop Sciences Division, University of Nottingham, Loughborough, 11 LE12 5RD, UK 12 ³ Crops for the Future Research Centre, Jalan Broga, Semenyih, 43500 13 14 Selangor, Malaysia 15 Email: dmunoz@ucm.cl * Corresponding author. 16 17
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20 ABSTRACT

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22 Temperate forests have been affected extensively by human activities, resulting 23 in land cover changes and population fragmentation. However, these 24 anthropogenic effects can be superimposed onto the natural history of species, 25 making it difficult to determine which effect is more important for a particular 26 species. Gomortega keule is an endangered tree that is found in one of the 27 world's biodiversity hotspots in central-south Chile. Human activities have 28 significantly impacted on the original habitat in this region in recent years and 29 are commonly considered to be the main cause of the scarcity of this species. 30 However, aspects of the natural history of this every every every also help 31 explain its present-day genetic structure. Therefore, in this study, we undertook 32 microsatellite genotyping of the two southernmost populations of G. keule, 33 which are 7.5 km apart from each other and well isolated from other 34 populations. We found that there was genetic differentiation between these 35 populations, suggesting that they exhibited at least some differentiation before 36 becoming isolated, most likely before human activities first impacted the region 37 some two centuries ago. Molecular estimates of their divergence time supported 38 a more ancient differentiation of the populations than would be explained by 39 human activities alone. Therefore, it is possible that their isolation may have 40 followed the extinction of megafaunal seed dispersers around 12,000 years 41 before present in this region, as indicated by fruit characteristics, the absence of 42 recruitment by seedlings and the existence of clonal trees. 43

Keywords: *Gomortega keule*, population genetics, fragmentation, genetic
structure, megafaunal extinction, relict populations

48 INTRODUCTION

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Temperate forests have been significantly affected by human activities, particularly agriculture and forestry, resulting in land use changes and the fragmentation of existing species. However, the life history traits and natural history of species can also be key elements that affect population isolation and thus the population genetics of trees (Bacles and Jump 2011).

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56 The coastal forests of central-south Chile lie within one of the 25 biodiversity 57 hotspots on Earth (Myers et al. 2000). This region served as a refugium during 58 the Quaternary glacial period, with many species surviving as a result of the 59 temperatures being moderated by the ocean (Villagrán & Armesto 2005). Indeed, the distributions of several woody species are currently restricted to 60 61 latitudes 36°–40°, which has been attributed to this natural history (Villagrán & 62 Armesto 2005). However, over the last two centuries, this region has also been 63 extensively affected by fires, timber exploitation, land clearance and the 64 introduction of exotic forestry plantations (Armesto et al. 2010), and consequently the population fragmentation and reproductive isolation that is 65 66 observed in many native woody species in these forests is commonly attributed to anthropogenic disturbance. 67

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69 Gomortega keule (Mol.) Baillon is an endangered species that is found in 70 central-south Chile. It exhibits a distribution pattern of small, isolated 71 populations over a narrow geographical area near to the coast (Fig. 1) and 72 occurs in only one of the national protected areas in Chile. San Martín & 73 Sánchez (2000) provided details of 22 locations where the species exists, most

74 of which have less than 100 individuals. Wood extraction activities and land 75 clearance for agriculture occurred on a small scale before the arrival of 76 Europeans in central Chile (Armesto et al. 2010), and indigenous people had 77 some impact on the native forests, particularly in the main river basins. 78 However, it is unlikely that G. keule populations were strongly affected by 79 these activities because this every reen tree mainly grows in mountainous areas, 80 often in ravines and on pronounced slopes, where the impacts of indigenous 81 human populations were restricted. Over the last two centuries, however, the 82 areas in which G. keule occurs have been considerably affected by the logging 83 of trees and especially the setting of fires for land clearance (Serra et al. 1986; 84 Armesto et al. 2010), as well as the establishment of exotic timber plantations 85 in more recent years, which has led to a reduction in native forest cover from 86 21% in AD 1975 to 7% in AD 2000 in the region where the species occurs (Armesto et al. 2010). Furthermore, individual G. keule trees have been found 87 88 as stump shoots, indicating that the parental trees had been damaged or logged 89 (Serra et al. 1986) and suggesting the loss of individuals in recent decades. In a 90 genetic study of 11 populations using inter simple sequence repeat (ISSR) 91 markers, García-Gonzáles et al. (2008) concluded that human activities caused 92 the isolation of populations of G. keule. However, this assertion should be 93 considered with caution since this is a long-lived species that occurs in a habitat 94 that has experienced only relatively recent anthropogenic disturbance. 95

96 The ecology of *G. keule* has some interesting features. The flowers are 97 pollinated by syrphid flies (Lander et al. 2009). Although barochory has been 98 suggested (Le Quesne & Stark 2006), there have been no studies on seed 99 dispersion. Interestingly, the fruit are among the largest in the Chilean flora and 100 are edible. Janzen & Martin (1982) proposed the idea of 'evolutionary

101 anachronism' for several Central American species, such as Gymnocladus 102 dioicus (Zaya & Howe 2009), and this could also apply to Gomortega keule. If 103 this was the case, the fruit of G. keule would have been dispersed by animals 104 that once existed in central Chile, such as Paleolama, Antifer, Equus, 105 Milodontidae and Gomphoteriidae (Labarca & López 2006). Indeed, the edible 106 pulp and very hard endocarp that encloses the seed (Fig. 2) appear to fit the megafaunal dispersion model (Guimarães et al. 2008). Today, the fruit of these 107 108 trees rot on the forest floor in the absence of animals that are able to eat the 109 complete fruit and transport the seed. In some regions, domestic animals (cows, 110 pigs and goats) eat the flesh of the fruit but leave the seed. Although small seedlings can be found, they do not grow further, possibly due to foraging by 111 112 domestic or wild (rabbits, hares) animals, or because of the ecological 113 conditions that occur near settlements (humidity, light, litter alteration). Thus, 114 with the extinction of the megafauna in South America around 12,000 years 115 before present (yr BP) (Barnosky & Lindsey 2010), G. keule may have lost one 116 class of its natural seed dispersers, causing it to become confined to areas that 117 are close to the stands and populations where it remains today.

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119 In this study, we investigated whether the isolation of populations of G. keule 120 occurred much longer ago than could be explained by human activities in the 121 last 200 years. To do this, we evaluated the genetic differentiation of two 122 spatially close but distinct populations based on the notion that if these 123 populations showed significant genetic differentiation, a process of genetic 124 differentiation must have already been underway before any superimposed 125 human activities in recent centuries. This could argue for a more complex 126 situation than suggested by García-Gonzáles et al. (2008), with the megafaunal 127 extinction around 12,000 yr BP potentially contributing to this process.

128 129 The two southernmost populations of G. keule were selected for this study, 130 which are only 7.5 km apart and in the same valley, but nearly 70 km from the 131 next nearest population. Importantly, there is a clear geographical separation 132 between these populations today, with no G. keule trees between. While most 133 populations of G. keule are generally of a similar size to the study populations 134 (60-80 individuals over <1 hectare), we chose these particular populations 135 because they occur in an area with a more limited degree of recent 136 anthropogenic influence, being relatively distant from current human 137 settlements. By contrast, populations near big cities (e.g. Concepción) or 138 agricultural areas (northern area of distribution) are likely to have been affected 139 for a longer period and more deeply by human activities. 140 141 142 MATERIALS AND METHODS 143 144 Sample populations 145 Plant materials were obtained from the Maule and Bío-Bío regions of Chile 146 147 (Table 1) in AD 2007. In population A (Fig. 1; n = 75), six G. keule trees (trunk 148 diameters >20 cm) were growing from old stumps, while the remainder were 149 young trees or shoots (trunk diameters <10 cm) growing under the canopy in a 150 plantation of *Pinus radiata*. These young individuals were derived from the 151 stumps of old trees that had been cut before or at the time of plantation 152 establishment and so would be expected to have the same genotypes as the 153 original logged trees. No seedlings were seen during collection. By contrast, 154 population B (n = 63) mainly comprised very old trees, some of which were the

155	oldest representatives of this species we found (trunk diameters ca. 2 m). As for
156	population A, no seedlings were observed in association with this population.
157	Samples were collected from all of the G. keule trees that could be found in
158	each of these populations, with only one sample being collected from each
159	group of shoots or trunks derived from an individual tree stump in population
160	A. In addition, 14 individuals were sampled from a population in the northern
161	area of the species' distribution (190 km away from populations A and B) as an
162	outgroup (group C).
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164	Sample collection and DNA extraction
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166	Young leaves were collected from each tree or alternatively the cambium was
167	sampled if leaves could not be found for a particular tree. The collected tissues
168	were dried in sealable plastic jars containing self-indicating silica gel crystals,
169	which were renewed as required, and stored at -20°C.
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171	DNA extraction followed Lander et al. (2007), whereby 5 mg
172	diethyldithiocarbamic sodium salt, 10 mg PVP-40000 and 5 μ l β -
173	mercaptoethanol were added per 1 ml of cetyltrimethylammonium bromide
174	(CTAB) solution that had been pre-heated to 65°C. This solution (500 μ l) was
175	then added to each sample for DNA extraction.
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177	Microsatellite genotyping
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179	Each sample was genotyped using seven microsatellite primer pairs. Five of
180	these (Gk-1, Gk-30, Gk-31, Gk-35 and Gk-39) were developed by Lander et al.

181 (2007), while the remaining two (CS2 and CS8) were developed by S. Mayes

following the method of Haddrill et al. (2002). The procedure to develop primers CS2 and CS8 involved the digestion and PCR amplification of genomic DNA, hybridisation to filters with artificial SSR repeat oligonucleotides, elution, and amplification. Rather than using cloning, a proportion of the enriched library was pyrosequenced using a 1/16 run (non-titanium reagents; 454 Life Sciences, Connecticut, USA).

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To visualise the DNA, 1% agarose gels were prepared in 1× TAE buffer and ethidium bromide was added to a final concentration of 0.5 μ g/ml before casting the gels. The PCR products with fluorescently labelled primers were then visualised on 2% agarose gels in 0.5× TBE buffer that were stained with ethidium bromide to a final concentration of 0.1 μ g/ml.

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195 The forward primers were directly labelled with D2, D3 or D4 fluorescent dye 196 (WellRED; Sigma Aldrich, St. Louis, Missouri, USA). The optimum annealing 197 temperature for PCR for each labelled primer was then determined in a Px2 198 Thermal Cycler using an annealing temperature gradient from 45°C to 60°C 199 across the block. PCR for each sample was performed using a 20-µl reaction 200 volume containing 2 µl template DNA, 2.5 µl buffer, 0.2 µl deoxynucleotide 201 (dNTP), 0.05 µl bovine serum albumin (BSA; 10 µg/µl), 0.02 µl of each 202 forward (labelled) and reverse primer (100 μ M), 15 μ l autoclaved H₂O, and 0.2 203 µl Tag polymerase. Master mixes for multiple reactions were made wherever 204 possible. The reaction conditions were 94°C for 3 min, 30 cycles of 94°C for 1 205 min, 1 min at the specific annealing temperature for each primer, 72°C for 1 206 min, and final extension at 72°C for 20 min. The annealing temperatures were 207 60.9°C (Gk-1), 65.0°C (Gk-30, Gk-31 and Gk-35), 56.8°C (Gk-39) and 52.7°C°C (Gk-44, CS2 and CS8). 208

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210	A total of 152 genotypes were analysed for seven loci using capillary
211	electrophoresis (CEQ 8000 Genetic Analysis System; Beckman Coulter,
212	California, USA). The PCR products were pooled according to their different
213	fluorescent tags, with a maximum of three different coloured samples being
214	analysed per well.
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216	Microsatellite data analysis
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218	The peaks that were produced by capillary electrophoresis were thoroughly
219	examined for each locus and assigned to classes according to Amos et al.
220	(2007) using a software tool developed by the same authors (FlexiBin).
221	
222	For each sample, matrices were generated using allele size to score co-dominant
223	markers at each locus. Polymorphic loci (Laurentin 2009) were used to generate
224	unbiased estimates of heterozygosity (Nei 1978) and investigate population
225	structure.
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227	Population genetic analysis
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229	Samples that were considered to have originated from the same individual were
230	excluded from the population analysis. This included any trees that were both
231	<10 m apart and had identical genotypes in the binary matrix In total, 19
232	distinct genotypes were identified among 42 samples, resulting in 23 samples
233	(42 minus 19) being excluded. Of the remaining 129 samples that were
234	included in the population analysis, 62 were from population A, 53 from B and
235	14 from C, all of which were assessed for all seven loci.

The number of alleles per population and per locus, and the allelic richness and private allelic richness per population were calculated with HP-Rare (Kalinowski 2005), using the rarefaction method to account for unequal sample sizes. The observed (H_0) and expected (H_E) heterozygosity for each locus and population were then computed with Arlequin 3.5 (Excoffier et al. 2005) using formulas from Nei (1987), and values of H_0 and H_E for the combined populations were calculated with GDA using formulas from Nei (1978).

245 According to Laurentin (2009), co-dominant markers can be used to assess the 246 diversity between groups with methodologies based on allele frequencies, such 247 as Wright's F-statistics and Nei's parameters. Therefore, the inbreeding 248 coefficient (F_{IS}) for each locus and population was computed using FSTAT 249 (Goudet 1995) according to Weir & Cockerham (1984). The pairwise fixation 250 index (Rho_{ST} or ρ_{ST}) was calculated for each population using Genepop 4.0.5.3 251 (Rousset 2008), following which a second version of the fixation index (F_{ST}) 252 was calculated under the stepwise mutation model using Arlequin.

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254 Exact tests for deviations from Hardy-Weinberg equilibrium (HWE) were 255 performed with Arlequin and Genepop using the Markov chain method 256 (forecasted chain length: 1,000,000; dememorisation steps: 100,000). Linkage 257 disequilibrium was tested separately for each population using Arlequin and a 258 Bonferroni correction was applied to the *P* values. The frequency of null alleles 259 was calculated assuming simultaneous inbreeding for each locus and population 260 with the software INEst (Chybicki & Burczyk 2009) using either the population 261 or individual inbreeding model (1,000,000 iterations).

The similarity index of Nei & Li (1979) was used to evaluate the genetic relationship among individuals, and cluster analysis (UPGMA) and principal coordinate analysis (PCoA) with Euclidean distances were used to visualise these relationships, both of which are appropriate for this type of study (Laurentin 2009). PCoA makes no assumptions about the distribution of the data or population genetics and the Euclidean distance does not consider the common absence of an allele as a shared characteristic (Kloda et al. 2008).

270

Cluster analysis and PCoA were performed using the MVSP software (Multivariate Statistical Package version 3.13; Kovach Computing Services, Anglesey, UK). A scatter plot was generated with the two largest eigenvalues along the first two axes and a dendrogram was produced with UPGMA using Nei & Li's index (Nei & Li 1979) as a measure of similarity. All 152 samples, which included those samples that were considered to have originated from the same individual, were included in the PCoA and cluster analysis.

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Population structure was assessed using a Bayesian cluster analysis with the software Structure 2.3.4 (Pritchard et al. 2000). Analyses were performed using the admixture model with independent allele frequencies (20 runs), with burn-in and simulation lengths of 300,000 and 1,000,000 iterations, respectively. The optimal value for K was estimated by calculating the statistic ΔK (Evanno et al. 2005). These procedures were initially followed for all samples and then for all samples except group C in an independent set of runs.

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The divergence time of populations A and B was estimated with the software
IMa2 (Hey & Nielsen 2007), which estimates population-genetic parameters by
calculating posterior probabilities in a Bayesian sampling framework. A total of

290 100,000 genealogies were sampled to estimate the joint posterior probability 291 distribution of the divergence time parameter. Generation times of 25, 100 and 292 175 years were used, as suggested by Jones et al. (2013) for long-lived trees, 293 and a mutation rate of 0.000316 was used, as proposed by Tamaki et al. (2016) 294 for trees. More than 100 runs were made to adjust the priors (values of q = 20, 295 m = 500 and t = 2), allowing for burn-in periods of 127–340 million steps. For 296 each generation time, 13 final estimates were made using different seed 297 numbers, and the mean values of the divergence time (yr BP) and the lower and 298 upper bounds of the estimated 95% highest posterior density intervals were 299 calculated. 300 301 302 RESULTS 303 304 **Microsatellite genotyping** 305 306 The number of alleles that was found for each locus is presented in Table 2. All

seven markers were polymorphic (major allele frequency <0.95) according to

Laurentin (2009). The allelic richness and private allelic richness (after

rarefaction) were lower in populations A (2.19 and 0.34, respectively) and B

(2.13 and 0.24) than in C (2.63 and 1.61) (Table 2). Private alleles were found

for all loci and their occurrence was associated with the presence of rare alleles,

as many of them showed low frequencies. Although sample size can affect the

number of alleles when a sampling approach is used, to the best of our

knowledge populations A and B represented the entire populations.

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316 The level of heterozygosity that was identified in G. keule (Table 3) is similar 317 to or lower than that reported for other endangered trees (Tamaki et al. 2008; 318 Finger et al. 2011; Shepherd & Perrie 2011). The largest difference between H_0 319 and $H_{\rm E}$ (>0.2) occurred at locus Gk-31 for population A and locus Gk-35 for 320 group C (Table 3), with exact tests for deviation from HWE being significant 321 (Bonferroni corrected P < 0.002) at each of these loci. The inbreeding 322 coefficient for each locus and population ranged from -0.233 to 0.874 (Table 4), 323 with the highest values (F_{1S} >0.8) being observed for locus *Gk-31* in population 324 A and locus *Gk-35* in group C.

325

326 The pairwise population fixation index ρ_{ST} was 0.045 for A–B, 0.882 for A–C 327 and 0.882 for B-C, while F_{ST} was 0.061, 0.407 and 0.382 for the same pairwise 328 comparisons, respectively. Significant linkage disequilibrium after Bonferroni 329 correction was present in one pairwise comparison for population A and one for 330 population B. The estimated frequency of null alleles was significantly different from zero (P < 0.001) for locus *Gk-31* in population A using the population 331 332 inbreeding model (PIM) and the individual inbreeding model (IIM) with INEst 333 (Chybicki & Burczyk 2009). Similarly, the frequency was also significantly different from zero for locus Gk-35 in group C (P < 0.001 using PIM and P < 0.001334 335 0.01 using IIM). Although the presence of null alleles could have some 336 influence on the accuracy of the statistical analyses, it was not considered likely 337 that it had a major influence on the overall results.

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339 Population structure

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341 There was some genetic differentiation among the populations studied. 342 Moderate genetic differentiation ($F_{ST} = 0.051-0.150$; Yeh 2000) was found

343 between populations A and B ($F_{ST} = 0.061$), while very great genetic 344 differentiation ($F_{ST} > 0.25$; Yeh 2000) was found between populations A and C 345 $(F_{ST} = 0.41)$, and B and C $(F_{ST} = 0.38)$. 346 347 The PCoA analysis (Fig. 3) showed that there was some differentiation between 348 populations A and B, which is consistent with their geographical location: these 349 populations are only 7.5 km apart (Fig. 1) and each occupies <1 hectare. 350 351 The dendrogram (Fig. 4) supported the PCoA analysis, with all samples except 352 those from group C falling into a separate cluster, and samples from 353 populations A and B generally being grouped into two partially overlapping 354 clusters. 355 356 The Bayesian analysis with the software Structure (Fig. 5) appeared to confirm 357 the groups that were identified by the PCoA and the dendrogram analysis. The 358 graphical output of Structure for K = 2 indicated some degree of differentiation 359 between populations A and B. When the analysis included group C, the highest 360 ΔK corresponded to two clusters, placing populations A and B in the same cluster. However, the output for K = 3 also suggested some differentiation 361 362 between populations A and B that is biologically meaningful. 363 364 The estimated divergence times for populations A and B and the 95% highest 365 posterior density intervals were 890 (395-2908) yr BP for a generation time of 25 years, 3956 yr BP (1978-12,263) for 100 years and 6784 yr BP (3185-366

367 20,951) for 175 years.

368

370 371 DISCUSSION 372 373 Megafaunal dispersal and natural history of G. keule 374 375 It has been proposed that the seeds of G. keule require megafaunal dispersal 376 (Muñoz-Concha & Davey 2011), and so it is possible that populations A and B 377 have become increasingly isolated following the extinction of megafauna from 378 Chile, leading to their current level of differentiation. This idea that population 379 differentiation was already underway before human activities affected these 380 populations is supported by a number of observations from the present study. 381 The data suggest that populations A and B, which are geographically very 382 close, have a moderate level of genetic differentiation but are clearly more 383 closely related to each other than to group C, which could argue for the 384 previous existence of a more continuous population that is now fragmented. 385 Since populations A and B occur in the same valley, fragmentation as a result 386 of physical barriers alone can be ruled out. The minimum divergence times 387 calculated in the present study are greater than 200 yr BP (minimum 395 yr BP 388 with 95% confidence for a 25-year generation time), which would argue for

pre-existing differentiation before any superimposed effect of human activities.

The PCoA analysis showed a clearer separation of the populations than was found by García-Gonzáles et al. (2008), who used ISSR to analyse the genetic structure of 11 populations of *G. keule*, but only sampled some of the individuals from populations A and B (29 and 7, respectively). In addition to the larger number of individuals that were sampled in the present study, the use of co-dominant microsatellite markers improved the resolution of a number ofthe analyses and also allowed heterozygosity-based indices to be calculated.

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399 Differentiation between the examined populations of G. keule is demonstrated 400 by the fixation index values, which lie within the range of those assessed with 401 microsatellites for other endangered tree species, such as Magnolia stellata 402 (Tamaki et al. 2008), Dalbergia monticola (Andrianoelina et al. 2009), 403 Medusagyne oppositifolia (Finger et al. 2011) and Pseudopanax ferox 404 (Shepherd & Perrie 2011). Furthermore, similar levels of genetic differentiation 405 have also been observed among isolated populations of Myrtus nivellei, a relict 406 species that is currently experiencing geographic range contraction (Migliore et 407 al. 2013).

408

409 It has previously been argued that clonality plays a role in maintaining 410 unaltered genotypes over a long period of time (even thousands of years) in 411 other woody species with relict and isolated populations, as discussed by Backs 412 et al. (2015), Bradbury et al. (2016) and Migliore et al. (2013). Although there 413 have been no reported studies on the role of clonality as a mechanism to 414 maintain heterozgosity and prevent inbreeding, and no current evidence for the 415 age of clones or trees of G. keule, the ability to maintain a population through 416 off-shoots could be a key factor that explains the current level of variation in 417 these populations and so warrants further investigation.

418

The generation times of trees can be greatly extended through vegetative (clonal) reproduction. Indeed, clones of relict tree species have been dated back to 3000–11,000 yr BP in *Eucalyptus absitia* (Bradbury et al. 2016) and over 13,000 yr BP in *Quercus palmeri* (May et al. 2009). Therefore, since clones of 423 *G. keule* were observed in the present study and by Lander et al. (2010), it is
424 possible that this species has an extended generation time, making a generation
425 time of 25 years unlikely.

426

427 The estimated time for the divergence of populations A and B is over 800 yr 428 BP, placing the population differentiation process well before two centuries 429 ago. Therefore, the isolation of these two populations of G. keule was probably 430 well progressed before extensive anthropogenic disturbances first impacted the 431 area around 200 yr BP (Armesto et al. 2010), indicating that population 432 differentiation may have started in glacial times, well before human-mediated 433 fragmentation of the area. This situation was also discussed for the endemic tree 434 Nothofagus alessandrii by Torres-Díaz et al. (2007), who concluded that there 435 was probably no gene flow between populations of this species before they 436 were impacted by relatively recent human activities.

437

438 A long regeneration cycle suggests that the study populations have been 439 isolated for a long time. The rare occurrence or complete absence of successful 440 sexual regeneration as evidenced by a lack of seedling establishment in the G. 441 keule populations sampled has also been observed in other relict tree species 442 such as Myrtus nivellei (Migliore et al. 2013), Q. palmeri (May et al. 2009), Q. 443 hinckleyi (Backs et al. 2015) and E. absita (Bradbury et al. 2016). The failure of 444 seedlings to establish and develop into new trees may indicate that the effective 445 generation time is very long, but could also reflect the cessation of seedling recruitment due to the more recent introduction of grazing animals such as 446 447 hares.

449 The fruit of G. keule matches the definition of megafaunal fruits that was 450 introduced by Guimarães et al. (2008), being 4-7 cm in diameter and yellow, 451 with a small number of large seeds (Fig. 2). The species also matches the 452 predictions of those authors, showing clumped spatial patterns, reduced 453 geographical ranges and high levels of among-population structuring. Other 454 traits that suggest that G. keule may have megafaunal seed dispersal 455 characteristics include the lack of modern dispersal agents, large and well 456 protected seed, edible fruit pulp and a strong vegetative propagation ability, as 457 previously discussed for Gymnocladus dioicus by Zaya & Howe (2009). In 458 addition, the present-day rare occurrence or complete absence of recruitment by 459 seedlings and the isolated distribution of individuals and populations further 460 support the idea of megafaunal dispersal of G. keule.

461

A number of questions still need to be answered to better understand the
population dynamics of *G. keule*. These include the juvenility and longevity of
individuals, the ecological conditions that are required for seedling survival,
and the minimum viable population size.

466

467 Sampling strategy

468

Some of the samples that were found to be genetically identical were clearly shoots from an old tree that had since disappeared, as reported previously (San Martín & Donoso 1996). However, some trees of *G. keule* that were growing <2 m apart were found to be genetically different, allowing us to reject the assumption that close physical proximity indicates that individuals are clonally related. The intricate way in which the trees occupy space on the forest floor is also well illustrated by the case where a root (10 cm diameter) that was sampled from population B was found to be different from three individuals growing <7
m away but identical to a large tree growing 15 m away. Therefore, developing
an understanding of the spatial relationships within the population will be an
important aspect of any future conservation planning.

480

481 The findings of the present study support the idea that vegetative propagation 482 (i.e. regeneration by sprouting) is currently important in natural populations of 483 G. keule, as stated by San Martín & Donoso (1996). Some individuals that were 484 physically close to each other had the same genetic profile and so were 485 probably produced from a single tree. Although we did not undertake extensive 486 sampling of shoots that were very close together, the observation of shoots with 487 the same genetic profile may indicate that clonality has played an important role 488 in the recent survival of G. keule trees.

489

490 Clonal production is a relatively common event in trees with reduced 491 populations such as *Eucalyptus absita* (Bradbury et al. 2016) and *Myrtus* 492 *nivellei* (Migliore et al. 2013), with some remarkably old individuals being 493 found, as seen in *Quercus palmeri* (May et al. 2009) and *Q. hinckleyi* (Backs et 494 al. 2015). Since clonality effectively extends the reproductive cycle time by 495 maintaining the original genetic combinations, the estimated divergence times 496 for *G. keule* may be underestimates.

497

498 Implications for conservation

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500 Although the occurrence of small and increasingly isolated populations of *G*. 501 *keule* seems to have preceded human history in the area, anthropogenic effects

502 remain very important for the future conservation status of this species. It is

503 apparent that there has been extensive destruction of the habitat, populations 504 and trees of this species, as evidenced by the fact that most individuals are 505 recently re-grown trees derived from old tree stumps with signs of destruction 506 by humans (logging, fires).

507

508 Long-term isolation and genetic differentiation among populations may have 509 important implications for conservation management strategies for G. keule and 510 other species that occur in disjunct populations, particularly where the natural 511 dispersal agents are no longer present. Conservation efforts should first be 512 directed towards the protection of remnant populations and individuals, 513 including the exclusion of cattle and hares from protected zones, and the careful 514 management of light through canopy maintenance, particularly for individuals 515 that are re-growing in current or former forestry plantations. Given the very 516 limited levels of seedling survival at present, it will be difficult to maintain high 517 genetic diversity through natural seed production. Therefore, seeds or 518 vegetative material should be sampled from as many populations as possible to 519 maximise the amount of genetic diversity that is captured for immediate ex situ 520 conservation. However, the risks of outbreeding depression should also be 521 evaluated.

522

The level of population differentiation that was observed in *G. keule* may offer a model for the conservation of other Chilean species for which the dispersal agents may be extinct, particularly *Pitavia punctata* and *Jubaea chilensis*, with a need to sample from all extreme populations of each of these. The southern populations of *G. keule* are very important from an agricultural and forestry perspective, as each isolated population contains valuable genetic resources for

529	the future domestication and genetic breeding of the species, and may contain
530	genetic information that is not found among other extant populations.
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533	
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545	CONFLICT OF INTEREST
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749 Table 1 Geographical locations and altitudes of the Gomortega keule

750 populations sampled.

Population /group	Locality	Latitude South	Longitude West	Altitude (metres above sea level)	Number of individuals sampled
А	Predio Carmávida, Bosques Arauco S.A. (private forestry farm)	37° 41′	73° 18′	300	75
В	Predio Pino Huacho, Forestal Tierra Chilena Ltda. (private forestry farm)	37° 40′	73° 13′	450	63
C	Reserva Nacional Los Queules (national protected area)	35° 58′	72° 40′	500	11
C	Predio Ralbún, Forestal Celco S.A. (private forestry farm)	36° 03′	72° 38′	540	2

751

754 **Table 2** Number of alleles, allelic richness and private allelic richness per locus and population/group in *Gomortega*

755 keule.

T	Allele size		Number	of alleles		А	llelic richne	ess	Priva	te allelic ri	ichness
Locus	(bp)	А	В	С	Total	Α	В	С	Α	В	С
Gk-1	224-240	5	4	4	7	2.49	1.92	2.96	0.43	0.26	1.76
Gk-30	182-190	3	4	4	5	2.11	3.35	2.49	0.03	1.07	0.38
Gk-31	208-228	4	3	5	8	1.74	1.52	3.35	0.66	0.06	1.99
Gk-35	224–252	4	2	3	7	2.42	1.99	2.21	0.54	0.11	2.21
Gk-39	133-201	2	2	3	5	1.97	1.83	1.84	0.16	0.03	1.84
CS2	104–119	2	3	3	4	1.98	2.11	2.76	0.01	0.02	1.66
CS8	210-220	3	3	4	5	2.64	2.23	2.83	0.53	0.16	1.40
Mean		3.29	3.00	3.71	5.86	2.19	2.13	2.63	0.34	0.24	1.61

756

759

760 **Table 3** Observed (H_0) and expected (H_E) heterozygosity for each locus and

761 population/group of Gomortega keule.

Locus	I	A	В		С		All populations		
Locus	H_0	H _E	H ₀	$H_{\rm E}$	H ₀	$H_{\rm E}$	Ho	$H_{\rm E}$	
Gk-1	0.468	0.451	0.283	0.308	0.786	0.643	0.426	0.500	
Gk-30	0.339	0.391	0.735	0.730	0.500	0.585	0.519	0.616	
Gk-31	0.032	0.255	0.094	0.159	0.643	0.712	0.124	0.298	
Gk-35	0.468	0.512	0.566	0.503	0.071	0.500	0.465	0.614	
Gk-39	0.469	0.475	0.358	0.343	0.286	0.262	0.403	0.535	
CS2	0.500	0.498	0.491	0.523	0.571	0.648	0.504	0.569	
CS8	0.597	0.588	0.453	0.431	0.429	0.558	0.519	0.534	
Mean	0.410	0.453	0.426	0.428	0.469	0.558	0.423	0.524	

762

766

765 Table 4 Inbreeding coefficient (F_{IS}) for each locus and population/group of

Gomorteg	a keule.			
Locus	А	В	С	All
Gk-1	-0.036	0.083	-0.233	-0.033
Gk-30	0.134	-0.008	0.150	0.058
Gk-31	0.874	0.410	0.100	0.533
Gk-35	0.086	-0.126	0.862	0.083
Gk-39	0.014	-0.046	-0.095	-0.013
CS2	-0.005	0.063	0.122	0.039
CS8	-0.015	-0.050	0.239	0.002
All	0.095	0.006	0.164	0.069

767 F_{IS} values >0 indicate a deficit and <0 indicate an excess of heterozygotes.

768

772

Fig. 1 Geographic distribution of *Gomortega keule*. A, Map of Chile with an
enlargement of the collection area (arrow). B, Populations of *G. keule* (green
dots) and the sampled populations (yellow). The distance between populations
A and B is 7.5 km.

777

Fig. 2 Fruit of *Gomortega keule*. A, Fruit showing the endocarp that encloses
the seed. B, Representation of extinct megafauna (Gomphoteriidae) eating the
fruit of *G. keule*.

781

Fig. 3 Principal coordinate analysis for three populations of *Gomortega keule* using microsatellites for seven loci. Populations A (n = 75) and B (n = 63) are the southernmost populations of this species, while group C (n = 14) comprises individuals found in the northern area of the species' natural distribution. The graph was generated using MVSP 3.13 with the first two axes accounting for 30.5% of the variance (Axis 1, 19.4%; Axis 2, 11.1%).

788

Fig. 4 Phenetic dendrogram for populations of *Gomortega keule* generated by
cluster analysis with UPGMA using the index of Nei & Li (1979) and the
software MVSP 3.13.

792

Fig. 5 Structure analysis of samples of *Gomortega keule* excluding group C
showing two clusters (top); and of all samples showing two (middle) and three
(bottom) clusters. A Bayesian analysis was conducted using the software
Structure 2.3.4 with the admixture model.

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Fig. 1 Geographic distribution of Gomortega keule. A, Map of Chile with an enlargement of the collection area (arrow). B, Populations of G. keule (green dots) and the sampled populations (yellow). The distance between populations A and B is 7.5 km.

124x179mm (300 x 300 DPI)



Fig. 2 Fruit of Gomortega keule. A, Fruit showing the endocarp that encloses the seed. B, Representation of extinct megafauna (Gomphoteriidae) eating the fruit of G. keule.

78x39mm (300 x 300 DPI)



Fig. 3 Principal coordinate analysis for three populations of Gomortega keule using microsatellites for seven loci. Populations A (n = 75) and B (n = 63) are the southernmost populations of this species, while group C (n = 14) comprises individuals found in the northern area of the species' natural distribution. The graph was generated using MVSP 3.13 with the first two axes accounting for 30.5% of the variance (Axis 1, 19.4%; Axis 2, 11.1%).



URL: http://mc.manuscriptcentral.com/nzjb



Fig. 4 Phenetic dendrogram for populations of Gomortega keule generated by cluster analysis with UPGMA using the index of Nei & Li (1979) and the software MVSP 3.13.

44x37mm (300 x 300 DPI)



Fig. 5 Structure analysis of samples of Gomortega keule excluding group C showing two clusters (top); and of all samples showing two (middle) and three (bottom) clusters. A Bayesian analysis was conducted using the software Structure 2.3.4 with the admixture model.

58x52mm (300 x 300 DPI)

