

1 *This is the pre-peer reviewed version of the following article: Zanata, T.B. et al. (2017)*
2 *Global patterns of interaction specialization in bird–flower networks. J. Biogeography*
3 *2017: 1–20. doi: 10.1111/jbi.13045, which has been published in final form at*
4 *<http://onlinelibrary.wiley.com/doi/10.1111/jbi.13045/full>. This article may be used for non-*
5 *commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving."*

6 ORIGINAL ARTICLE

7 Global patterns of interaction specialization in bird-flower networks

8 Thais B. Zanata*, Bo Dalsgaard, Fernando C. Passos, Peter A. Cotton, James J.
9 Roper, Pietro K. Maruyama, Erich Fischer, Matthias Schleuning, Ana M. Martín
10 González, Jeferson Vizentin-Bugoni, Donald C. Franklin, Stefan Abrahamczyk,
11 Ruben Alárcon, Andréa C. Araujo, Francielle P. Araújo, Severino M. de Azevedo-
12 Junior, Andrea C. Baquero, Katrin Böhning-Gaese, Daniel W. Carstensen, Henrique
13 Chupil, Aline G. Coelho, Rogério R. Faria, David Hořák, Tanja T. Ingversen, Štěpán
14 Janeček, Glauco Kohler, Carlos Lara, Flor M. G. Las-Casas, Ariadna V. Lopes,
15 Adriana O. Machado, Caio G. Machado, Isabel C. Machado, María A. Maglianesi,
16 Tiago S. Malucelli, Jayasilan Mohd-Azlan, Alan C. Moura, Genilda M. Oliveira,
17 Paulo E. Oliveira, Juan Francisco Ornelas, Jan Riegert, Licléia C. Rodrigues, Liliana
18 Rosero-Lasprilla, Ana M. Rui, Marlies Sazima, Baptiste Schmid, Ondřej Sedláček,
19 Allan Timmermann, Maximilian G. R. Vollstädt, Zhiheng Wang, Stella Watts,
20 Carsten Rahbek and Isabela G. Varassin

21 Thais B. Zanata: Programa de Pós-Graduação em Ecologia e Conservação,
22 Universidade Federal do Paraná, Curitiba-PR, Brasil; Laboratório de Ecologia
23 Vegetal, Departamento de Botânica, Universidade Federal do Paraná, Centro
24 Politécnico, Cx. 19031, 81531-980, Curitiba-PR, Brasil; Center for Macroecology,
25 Evolution and Climate, Natural History Museum of Denmark, University of
26 Copenhagen, Universitetsparken 15, 2100 Copenhagen Ø, Denmark.

- 27 Bo Dalsgaard: Center for Macroecology, Evolution and Climate, Natural History
28 Museum of Denmark, University of Copenhagen, Universitetsparken 15, 2100
29 Copenhagen Ø, Denmark.
- 30 Fernando C. Passos: Laboratório de Biodiversidade, Conservação e Ecologia de
31 Animais Silvestres, Departamento de Zoologia, Universidade Federal do Paraná,
32 Caixa Postal 19020, 81531-980, Curitiba-PR, Brasil.
- 33 Peter A. Cotton: Marine Biology & Ecology Research Centre, University of
34 Plymouth, Plymouth PL4 8AA, United Kingdom.
- 35 James J. Roper: Graduate program in Ecosystem Ecology, Universidade Vila Velha
36 and Graduate program in Ecology and Conservation, Universidade Federal do Paraná,
37 Brasil.
- 38 Pietro K. Maruyama: Departamento de Biologia Vegetal, Instituto de Biologia,
39 Universidade Estadual de Campinas, Campinas-SP, Brasil.
- 40 Erich Fischer: Centro de Ciências Biológicas e da Saúde, Universidade Federal de
41 Mato Grosso do Sul, 79070-900 Campo Grande-MS, Brasil.
- 42 Matthias Schleuning: Senckenberg Biodiversity and Climate Research Centre (BiK-
43 F), Senckenberganlage 25, 60325 Frankfurt (Main), Germany.
- 44 Ana M. Martín González: Pacific Ecoinformatics and Computational Ecology Lab,
45 Berkeley, USA; Center for Macroecology, Evolution and Climate, Natural History
46 Museum of Denmark, University of Copenhagen, Universitetsparken 15, 2100
47 Copenhagen Ø, Denmark.
- 48 Jeferson Vizentin-Bugoni: Programa de Pós-Graduação em Ecologia, Universidade
49 Estadual de Campinas (Unicamp), Cx. Postal 6109, CEP: 13083-970, Campinas-SP,
50 Brasil.

- 51 Donald C. Franklin: Research Institute for Environment & Livelihoods, Charles
52 Darwin University, Darwin NT 0909, Australia.
- 53 Stefan Abrahamczyk: University of Bonn, Nees Institute for Biodiversity of Plants,
54 Meckenheimer Allee 170, 53115 Bonn, Germany; University of Zurich, Institute of
55 Systematic and Evolutionary Botany, Zollikerstr. 107, 8008 Zurich, Switzerland.
- 56 Ruben Alárcon: Biology Program, California State University Channel Islands,
57 Camarillo, CA 23012, USA.
- 58 Andréa C. Araujo: Laboratório de Ecologia, Centro de Ciências Biológicas e da
59 Saúde, Universidade Federal de Mato Grosso do Sul, 79070-900 Campo Grande-MS,
60 Brasil.
- 61 Francielle P. Araújo: Universidade Estadual do Rio Grande do Sul, São Francisco de
62 Paula-RS, Brasil.
- 63 Severino M. de Azevedo-Junior: Department of Biology, Rural Federal University of
64 Pernambuco, Recife-PE, Brasil.
- 65 Andrea C. Baquero: Center for Macroecology, Evolution and Climate, Natural
66 History Museum of Denmark, University of Copenhagen, Universitetsparken 15,
67 2100 Copenhagen Ø, Denmark.
- 68 Katrin Böhning-Gaese: Senckenberg Biodiversity and Climate Research Centre (BiK-
69 F), Senckenberganlage 25, Frankfurt am Main 60325, Germany; Institute for Ecology,
70 Evolution and Diversity, Goethe University Biologicum, Max-von-Laue Strasse 13,
71 Frankfurt am Main 60439, Germany.
- 72 Daniel W. Carstensen: Center for Macroecology, Evolution and Climate, Natural
73 History Museum of Denmark, University of Copenhagen, Universitetsparken 15,
74 2100 Copenhagen Ø, Denmark.

- 75 Henrique Chupil: Programa de Pós-Graduação em Zoologia - UFPR; Instituto de
76 Pesquisas Cananéia, Rua Tristão Lobo, 199, 11990-000, Cananéia-SP, Brasil.
- 77 Aline G. Coelho: Laboratório de Ornitologia, Departamento de Ciências Biológicas,
78 Universidade Estadual de Feira de Santana, Feira de Santana-BA, 44036-900, Brasil.
- 79 Rogério R. Faria: Campus de Aquidauana, Universidade Federal de Mato Grosso do
80 Sul, 79200-000, Aquidauana-MS, Brasil.
- 81 David Hořák: Department of Ecology, Faculty of Science, Charles University in
82 Prague, Vinicna 7, 128 43 Praha 2, Czech Republic.
- 83 Tanja Ingversen: Dybbøl Bygade 73, 6400 Sønderborg, Denmark.
- 84 Štěpán Janeček: Institute of Botany, Czech Academy of Sciences, Dukelská 135:
85 Třeboň 37901, Czech Republic; Department of Ecology, Faculty of Science, Charles
86 University in Prague, Viničná 7, 12844 Praha 2, Czech Republic.
- 87 Glauco Kohler: Instituto Nacional de Pesquisas da Amazônia, Av. André Araújo
88 2936, Petrópolis, 69080-971, Manaus-AM, Brasil.
- 89 Carlos Lara: Centro de Investigación en Ciencias Biológicas, Universidad Autónoma
90 de Tlaxcala, Km 10.5 Autopista Tlaxcala-San Martín Texmelucan, San Felipe
91 Ixtacuixtla, Tlaxcala 90120, México.
- 92 Flor M. G. Las-Casas: Laboratory of Ecology, Systematics and Evolution of Birds,
93 Federal University of Pernambuco, 50670-420, Recife-PE, Brasil.
- 94 Ariadna V. Lopes: Departamento de Botânica, Universidade Federal de Pernambuco,
95 50670-901, Recife-PE, Brasil.
- 96 Adriana O. Machado: Instituto de Biologia, Universidade Federal de Uberlândia,
97 Uberlândia-MG, Brasil.
- 98 Caio G. Machado: Laboratório de Ornitologia, Departamento de Ciências Biológicas,
99 Universidade Estadual de Feira de Santana, Feira de Santana-BA, 44036-900, Brasil.

- 100 Isabel C. Machado: Departamento de Botânica, Centro de Biociências, Universidade
101 Federal de Pernambuco, Recife-PE, 50372-970, Brasil.
- 102 María A. Maglianesi: Vicerrectoría de Investigación, Universidad Estatal a Distancia
103 (UNED), San José, Costa Rica and Senckenberg Biodiversity and Climate Research
104 Centre (BiK-F), Senckenberganlage 25, 60325 Frankfurt (Main), Germany.
- 105 Tiago S. Malucelli: Laboratório de Ecologia Vegetal, Departamento de Botânica,
106 Centro Politécnico, Cx. 19031, 81531-980, Curitiba-PR, Brasil.
- 107 Jayasilan Mohd-Azlan: Department of Zoology, Faculty of Resource Science and
108 Technology, Universiti Malaysia Sarawak, 94300 Kota Samarahan. Sarawak,
109 Malaysia; Research Institute for the Environment and Livelihoods, Charles Darwin
110 University, Ellengowan Drive, Northern Territory 0909, Australia.
- 111 Alan C. Moura: Laboratório de Ornitologia, Departamento de Ciências Biológicas,
112 Universidade Estadual de Feira de Santana, Feira de Santana-BA, 44036-900, Brasil.
- 113 Genilda M. Oliveira: Instituto Federal de Brasília, Campus Samambaia, Brasília-DF,
114 Brasil.
- 115 Paulo E. Oliveira: Instituto de Biologia, Universidade Federal de Uberlândia,
116 Uberlândia-MG, Brasil.
- 117 Juan Francisco Ornelas: Departamento de Biología Evolutiva, Instituto de Ecología,
118 AC, Carretera antigua a Coatepec No. 351, El Haya, Xalapa, Veracruz, 91070,
119 México.
- 120 Jan Riegert: Department of Zoology, Faculty of Science, University of South
121 Bohemia, Branišovská 1760, 370 05 České Budějovice, Czech Republic.
- 122 Licléia C. Rodrigues: Laboratório de Ornitologia, Departamento de Zoologia, ICB,
123 Universidade Federal de Minas Gerais. Caixa Postal 486, 31270-901, Belo Horizonte-
124 MG, Brasil.

- 125 Liliana Rosero-Lasprilla: Escuela de Ciencias Biológicas, Grupo de Investigación
126 Biología para la Conservación, Universidad Pedagógica y Tecnológica de Colombia,
127 Tunja, Boyacá, Colombia.
- 128 Ana M. Rui: Departamento de Ecologia, Zoologia e Genética, Instituto de Biologia,
129 Universidade Federal de Pelotas, Capão do Leão-RS, Brasil.
- 130 Marlies Sazima: Departamento de Biologia Vegetal, Instituto de Biologia,
131 Universidade Estadual de Campinas, Cx. Postal 6109, 13083-970, Campinas-SP,
132 Brasil.
- 133 Baptiste Schmid: Swiss Ornithological Institute, Seerose 1, 6204 Sempach,
134 Switzerland.
- 135 Ondřej Sedláček: Department of Ecology, Faculty of Science, Charles University in
136 Prague, Vinicna 7, 128 43 Praha 2, Czech Republic.
- 137 Allan Timmermann: Department of Bioscience, Aarhus University, Ny Munkegade
138 114, 8000 Aarhus C, Denmark.
- 139 Maximilian G. R. Vollstädt: Senckenberg Biodiversity and Climate Research Centre
140 (BiK-F), Senckenberganlage 25, Frankfurt am Main 60325, Germany; Institute for
141 Ecology, Evolution and Diversity, Goethe University Biologicum, Max-von-Laue
142 Strasse 13, Frankfurt am Main 60439, Germany.
- 143 Zhiheng Wang: Department of Ecology and Key Laboratory for Earth Surface
144 Processes of the Ministry of Education, College of Urban and Environmental
145 Sciences, Peking University, Beijing 100871, China.
- 146 Stella Watts: Landscape and Biodiversity Research Group, Department of
147 Environmental and Geographical Sciences, University of Northampton, Avenue
148 Campus, St George's Avenue, Northampton NN2 6JD, United Kingdom.

149 Carsten Rahbek: Center for Macroecology, Evolution and Climate, Natural History
150 Museum of Denmark, University of Copenhagen, Universitetsparken 15, 2100
151 Copenhagen Ø, Denmark.

152 Isabela G. Varassin: Laboratório de Ecologia Vegetal, Departamento de Botânica,
153 Universidade Federal do Paraná, Centro Politécnico, Cx. 19031, 81531-980, Curitiba-
154 PR, Brasil.

155 *Correspondence Thais B. Zanata: Laboratório de Ecologia Vegetal, Departamento de
156 Botânica, Universidade Federal do Paraná, Centro Politécnico. Cx. 19031, 81531-980.
157 Curitiba-PR, Brasil. thaisbzanata@gmail.com

158 Short running head Global patterns of bird-flower interactions

159 WORD COUNT

160 Abstract: 286

161 Main body text (including the abstract and references): 8,090

162 Estimative of the number of pages required by figures and tables: 3 ½ pages

163 ABSTRACT

164 Aim: Among the world's three major nectar-feeding bird taxa, hummingbirds are the
165 most phenotypically specialized for nectarivory, followed by sunbirds, while the
166 honeyeaters are the least phenotypically specialized taxa. We tested whether this
167 phenotypic specialization gradient is also found in the interaction patterns with their
168 floral resources.

169 Location: Americas, Africa, Asia and Oceania/Australia.

170 Methods: We compiled interaction networks between birds and floral resources for 79
171 hummingbird, nine sunbird and 33 honeyeater communities. Interaction specialization
172 was quantified through connectance (C), complementary specialization (H_2'), binary
173 (Q_B) and weighted modularity (Q), with both observed and null-model corrected
174 values. We compared interaction specialization among the three types of bird–flower
175 communities, both independently and while controlling for potential confounding
176 variables, such as plant species richness, asymmetry, latitude, insularity, topography,
177 sampling methods and intensity.

178 Results: Hummingbird-flower networks were more specialized than honeyeater-
179 flower networks. Specifically, hummingbird-flower networks had a lower proportion
180 of realized interactions (lower C), decreased niche overlap (greater H_2') and greater
181 modularity (greater Q_B). However, we found no significant differences between
182 hummingbird– and sunbird–flower networks, nor between sunbird– and honeyeater–
183 flower networks.

184 Main conclusions: As expected, hummingbirds and their floral resources have greater
185 interaction specialization than honeyeaters, possibly because of greater phenotypic
186 specialization and greater floral resource richness in the New World. Interaction
187 specialization in sunbird–flower communities was similar to both hummingbird–
188 flower and honeyeater–flower communities. This may either be due to the relatively
189 small number of sunbird–flower networks available, or because sunbird–flower
190 communities share features of both hummingbird–flower communities (specialized
191 floral shapes) and honeyeater–flower communities (fewer floral resources). These
192 results suggest a link between interaction specialization and both phenotypic
193 specialization and floral resource richness within bird–flower communities at a global
194 scale.

195 Keywords: Honeyeaters, hummingbirds, modularity, niche partitioning, plant-animal
196 interactions, ornithophily, specialization, sunbirds.

197 INTRODUCTION

198 Specialization is of major importance in ecology and occurs at all levels, from the
199 individual to the community (Devictor *et al.*, 2010). The origin and evolution of
200 specialization are important to understand species interactions (Futuyma & Moreno,
201 1988), such as plant-animal interactions involved in pollination (Waser *et al.*, 1996;
202 Johnson & Steiner, 2000). Birds include the most abundant and speciose vertebrate
203 pollinators, with flower visitation reported in more than 50 families (Cronk & Ojeda,
204 2008). Of these families, three are highly specialized for nectarivory: in the New
205 World (NW), hummingbirds (Apodiformes, Trochilidae, 363 species) found
206 throughout the Americas, and in the Old World (OW), sunbirds (Passeriformes,
207 Nectariniidae, 132 species), in Africa, Asia and Oceania/Australia, and honeyeaters
208 (Passeriformes, Meliphagidae, 175 species), in Asia and Oceania/Australia (Stiles,
209 1981; Fleming & Muchhala, 2008; del Hoyo *et al.*, 2016), with a limited distribution
210 overlap between these two OW families (Barker *et al.*, 2002). These three families
211 contain most of the specialized nectar-feeding bird species, and are an example of
212 convergent evolution, as they have independently evolved adaptations associated with
213 nectarivory (Prum *et al.*, 2015). This pattern of evolution has generated interest in
214 understanding the differences and similarities in the morphology and ecology of these
215 nectar-feeding birds and their floral resources (Pyke, 1980; Stiles, 1981; Collins &
216 Paton, 1989; Fleming, 2005; Cronk & Ojeda, 2008; Fleming & Muchhala, 2008).

217 Despite the convergent evolution between these families, they vary in the
218 extent of phenotypic specialization towards pollination (*sensu* Ollerton *et al.*, 2007).

219 Hummingbirds are the most phenotypically specialized for nectarivory, followed by
220 sunbirds, with honeyeaters as the most phenotypically generalized taxa (Stiles, 1981;
221 Fleming & Muchhala, 2008). Bill length in hummingbird communities is more
222 variable than in OW communities (Fleming & Muchhala, 2008), which may likewise
223 facilitate a finer resource partitioning among hummingbird species (Abrahamczyk &
224 Kessler, 2010; Maglianesi *et al.*, 2014). Greater phenotypic specialization of
225 hummingbirds is also manifest by their small size, which is one of their adaptations
226 that allows for hovering flight (Pyke, 1980). Hovering is the prevalent mode of flower
227 foraging among hummingbirds (Collins & Paton, 1989), with perching being
228 predominant in the typically heavier sunbirds and honeyeaters (Pyke, 1980; Fleming
229 & Muchhala, 2008; but see Janeček *et al.*, 2011; Wester, 2013). Small size and
230 hovering flight are likely to have favoured the diversification of hummingbird-
231 pollinated plant species, because the evolutionary transition from small and delicate
232 insect-pollinated to hummingbird-pollinated species was probably relatively simple
233 (Castellanos *et al.*, 2003; Thomson & Wilson, 2008). This greater diversification of
234 floral resources may have promoted a greater interaction specialization in NW
235 communities (Dalsgaard *et al.*, 2011).

236 Sunbirds and the plants they visit are thought to be the second-most
237 phenotypically specialized community. Sunbirds have bills, tongues and digestive
238 tracts that are better adapted to nectar-feeding than those of honeyeaters, the least
239 specialized group (Stiles, 1981). Also, flowers visited by sunbirds, as in
240 hummingbird-pollinated species, tend to have tubular or gullet shapes, while
241 honeyeaters tend to visit flowers with less restrictive morphologies (Stiles, 1981;
242 Fleming & Muchhala, 2008). Although phenotypic specialization of nectar-feeding
243 birds and their floral resources clearly decreases from specialized hummingbird to

244 sunbird and then generalized honeyeater communities (Stiles, 1981; Fleming &
245 Muchhala, 2008), it remains unclear whether the interaction specialization of bird-
246 flower communities would reflect the same phenotypic specialization gradient.

247 Species interaction patterns, such as bird-flower interactions, can be
248 investigated by a network approach. This approach can reveal emergent properties at
249 the community-level that are not apparent in pairwise interactions (Bascompte &
250 Jordano, 2007). Some of the main network metrics that quantify interaction
251 specialization at the community-level are connectance, complementary specialization
252 and modularity (Blüthgen, 2010). Community-level specialization quantified by these
253 metrics is associated with the concept of ecological specialization (*sensu* Ollerton *et*
254 *al.*, 2007) and the realized Eltonian niche (Devictor *et al.*, 2010), where interactions
255 are treated as one dimension of the ecological niche and the degree of interaction
256 specialization represents niche partitioning among species (Blüthgen, 2010).

257 Because of the observed phenotypic specialization in the three types of bird-
258 flower communities, we predicted the following interaction specialization gradient:
259 hummingbird-flower > sunbird-flower > honeyeater-flower. We compiled a dataset of
260 121 networks, and tested the differences of bird-flower interaction specialization
261 between these three bird families, both independently and while controlling for
262 potential confounding variables such as plant species richness, asymmetry, latitude,
263 insularity, topography, and sampling methods and intensity.

264 MATERIALS AND METHODS

265 Bird-flower interaction networks

266 We gathered published and unpublished data on flower visitation by hummingbirds,
267 sunbirds and honeyeaters sampled at the community-level in a locality (data source

268 and geographical information of each network are available in the Appendix of
269 Supporting Information: Tables S1–S2). For each study interaction lists between bird
270 and plant species were transformed into adjacency matrices, with birds as columns
271 and plants as rows. In these matrices, flower visits by birds were represented in binary
272 networks by their absence (0) or occurrence (1), or in weighted networks by their
273 absence (0) or their interaction frequency (Bascompte & Jordano, 2007). Interaction
274 frequency represents the number of observations of birds either visiting or carrying
275 pollen from a given plant species. We included interactions only of the nectar-feeding
276 specialist families: Meliphagidae, Nectariniidae and Trochilidae (Stiles, 1981;
277 Fleming & Muchhala, 2008; del Hoyo *et al.*, 2016). We excluded interactions with
278 known exotic plant species and illegitimate interactions, when the bird did not contact
279 the floral reproductive structures, for example, piercing the corolla to reach the nectar.
280 These interactions were excluded because they are unlikely to involve bird-flower
281 evolutionary relationships. Information about interaction legitimacy was unavailable
282 in four studies that were used in the analyses (Pettet, 1977; Collins & Rebelo, 1987;
283 Brooker *et al.*, 1990; Wester, 2013), in which case we assumed that all interactions
284 were legitimate. We used databases of Flora of the West Indies
285 (botany.si.edu/antilles/WestIndies/query.cfm), Brazilian Flora Checklist
286 (floradobrasil.jbrj.gov.br) and Tropicos (www.tropicos.org) to classify plant species
287 as exotic.

288 We compiled a total of 121 bird-flower interaction networks, of which 79
289 described hummingbird-, 9 sunbird- and 33 honeyeater-flower interactions.
290 Interaction frequency was available for 67 (85%) hummingbirds (HU), 5 (55%)
291 sunbirds (SU) and 23 (70%) honeyeaters (HO) networks. Bird species richness ranged
292 from 2 to 24 in the HU, 2 to 13 in the SU and 2 to 12 in the HO networks, while plant

293 species richness ranged from 2 to 65 in the HU, 2 to 26 in the SU and 2 to 39 in the
294 HO networks (for detailed values of each network, see Table S2).

295 Measuring specialization of interaction networks

296 To quantify interaction specialization, we used two binary metrics, connectance (C)
297 and binary modularity (Q_B), and two weighted metrics, complementary specialization
298 (H_2') and weighted modularity (Q). These metrics range from 0 to 1, where the most
299 generalized network has a value of 0 and the most specialized network has a value of
300 1 (H_2' , Q_B , Q), with the inverse for connectance (C).

301 Connectance is defined as the proportion of observed pairwise interactions
302 relative to the total number of possible interactions in the community, where the total
303 number of possible interactions is calculated as the richness of visited plant species
304 multiplied by the richness of nectar-feeding birds (Jordano, 1987; Blüthgen, 2010).
305 Complementary specialization is derived from two-dimensional Shannon entropy, and
306 quantifies the niche partitioning among species considering partner availability,
307 defined by the marginal totals in the interaction matrix, and so measures the
308 exclusiveness of interactions (Blüthgen *et al.*, 2006; Martín González *et al.*, 2015).
309 Finally, modularity is a network pattern that emerges when some species interact
310 strongly with each other but less so with the remaining species, thereby creating
311 strongly-connected sub-groups within a less connected network (Olesen *et al.*, 2007;
312 Dormann & Strauss, 2014; Maruyama *et al.*, 2014). Binary modularity was measured
313 using the Barber metric (Barber, 2007), with simulated annealing as the search
314 algorithm in the MODULAR software (Marquitti *et al.*, 2014). Weighted modularity
315 was calculated with the standard specifications of the QuanBiMo algorithm and using
316 the greatest modularity value after five independent runs (Dormann & Strauss, 2014;

317 Schleuning *et al.*, 2014). Connectance, complementary specialization and weighted
318 modularity were calculated with the *bipartite* package (Dormann *et al.*, 2008) in R (R
319 Core Team, 2017). Although metric values were correlated (Pearson's $r > 0.8$, $p <$
320 0.05 for spatial and non-spatial correlation), we analysed all metrics separately
321 because they can describe complementary patterns of interaction specialization
322 (Martín González *et al.*, 2015).

323 Null-model corrections of network metrics

324 Network metrics are often influenced by species richness and sampling effort. Thus,
325 null models were proposed to control for these effects (Schleuning *et al.*, 2014;
326 Martín González *et al.*, 2015; Dalsgaard *et al.*, 2017) The idea behind the use of null
327 models is to calculate deviations between observed values and null-model
328 expectations, assuming random species interactions, while controlling network
329 properties that may be related to species richness and sampling effort (Dalsgaard *et*
330 *al.*, 2017). We used Patefield's algorithm (Patefield, 1981) to generate randomized
331 networks, an algorithm commonly used in geographical analyses of interaction
332 networks (Dalsgaard *et al.*, 2017). This algorithm constrains network size
333 (representing species richness) and the interaction matrix marginal totals (the sum of
334 interaction frequencies of each bird and plant species, which may be a consequence of
335 species abundance or sampling effort; Dormann *et al.*, 2009). Patefield algorithm
336 requires interaction frequency to generate randomized networks, and so we only use
337 null-model corrections on weighted networks. Thus, sample size was larger for
338 observed connectance and binary modularity than null-model corrected connectance
339 and binary modularity (see above; Table S2). For each of the observed networks, we
340 generated 1,000 randomized networks to estimate connectance and complementary

341 specialization and 100 to estimate binary and weighted modularity. We used fewer
342 randomizations for modularity metrics because their calculation requires time-
343 consuming algorithms (Olesen *et al.*, 2007; Schleuning *et al.*, 2014; Sebastián-
344 González *et al.*, 2015). For each of the randomized networks, we calculated the
345 network metrics following the same procedure as adopted for the observed networks
346 (see above). To quantify how the observed network values depart from the null
347 expectation, we calculated the null-model corrected values, by subtracting the
348 observed metric value from the mean value across all randomized networks (Δ -
349 transformation; Schleuning *et al.*, 2014; Martín González *et al.*, 2015; Dalsgaard *et*
350 *al.*, 2017).

351 Comparing hummingbird, sunbird and honeyeater interaction specialization
352 We compared observed and null-model corrected metrics of hummingbird-, sunbird-
353 and honeyeater-flower networks, testing for differences of interaction specialization
354 between the three types of communities. First, for data with equal variances, we
355 compared them using one-way ANOVA with Tukey's multiple comparison test, and
356 for data with unequal variances we used Kruskal-Wallis test with Dunn's multiple
357 comparison test. Analysis and graphs were plotted in GraphPad Prism 6.0 (GraphPad
358 Software, La Jolla, California, USA; Morgan, 1998). Second, we compared
359 interaction specialization between the three types of bird-flower communities while
360 controlling for potentially confounding variables (see below), using linear multiple
361 regression models and corrected Akaike Information Criterion (AICc). We used the
362 threshold of $\Delta\text{AICc} \leq 2$ to identify minimum adequate models (MAM; Burnham &
363 Anderson, 2002).

364 In the linear models, the three types of bird-flower communities were assigned
365 as a categorical variable with three levels (hummingbirds, sunbirds and honeyeaters).
366 Nine potentially confounding variables were also included in the models: 1) plant
367 species richness (\log_{10} transformed), included because species-rich communities are
368 expected to have greater specialization (Dalsgaard *et al.*, 2011; Martín González *et*
369 *al.*, 2015; but see Vázquez & Stevens, 2004; Schleuning *et al.*, 2012); 2) asymmetry
370 (\log_{10} transformed), described as the ratio between bird and plant species richness and
371 included because connectance decreases when asymmetry increases (Blüthgen *et al.*,
372 2006); 3) absolute latitude, because several studies have found greater network
373 specialization towards the tropics (Olesen & Jordano, 2002; Dalsgaard *et al.*, 2011;
374 but see Vázquez & Stevens, 2004; Schleuning *et al.*, 2012); 4) insularity, where
375 mainland communities were classified as “0” and island communities as “1” and
376 included in the models because insular communities are expected to be more
377 generalized than mainland communities (Martín González *et al.*, 2015; Traveset *et al.*,
378 2015); 5) topography (square root transformed), defined as the elevational range of
379 the sampled localities calculated in 1×1 km grid cells within a concentric distance of
380 10 km from each sampled locality; 6) duration of each study (\log_{10} transformed),
381 based on the number of sampling months; 7) the method used to record species
382 interactions, included because it may influence network structure (Ramírez-Burbano
383 *et al.*, 2017), where focal observations were classified as “0” and sampling pollen
384 loads on visiting birds as “1”; 8) sampling coverage, where “1” represents studies that
385 sampled the supposed entire communities of bird and plant species, and “0”
386 represents studies that sampled only a subset of the community (for example, studies
387 focusing on ornithophilous plant species or a given plant family); and 9) sampling
388 intensity (\log_{10} transformed), calculated as the ratio between the square root of the

389 total number of interactions and the square root of the product of the number of bird
390 and plant species in the network (Schleuning *et al.*, 2012; Dalsgaard *et al.*, 2017). As
391 interaction frequency is required to quantify sampling intensity, we were only able to
392 estimate this variable on the weighted networks (Table S2).

393 Model selection was performed using the *dredge* function in *MuMIn* package
394 (Barton, 2016). We reported the standardized coefficients for an averaged model
395 (AVM) and the importance ($\sum w_i$) of each predictor variable measured across all
396 models (Burnham & Anderson, 2002). We used an importance threshold value of \geq
397 0.80 to identify relevant predictor variables (Sebastián-González *et al.*, 2015). When
398 the bird-flower community variable was selected in the MAM, we used partial
399 regressions to detect the total and individual variation explained by this variable. The
400 differences between the three types of bird-flower communities were tested by Tukey
401 contrasts for general linear hypothesis, using the *glht* function in *multcomp* package
402 (Hothorn *et al.*, 2008). We considered multicollinearity to be absent when the
403 variance inflation factor (VIF) or the generalized variance inflation factor (GVIF) was
404 ≤ 10 (Hair *et al.*, 2009), both indices were measured using the *vif* function in *car*
405 package (Fox & Weisberg, 2011). We checked for positive spatial autocorrelation in
406 the residuals of the MAM with the lowest $\Delta AICc$ computing Moran's *I* in 14-equally
407 spaced distance classes and applying a permutation test with 10,000 iterations, using
408 the *correlog* function in *pgirmess* package (Giraudoux, 2016). Initial analyses found
409 that interaction specialization was associated with the method of recording
410 interactions (Tables 1–2). Therefore, we checked the consistency of our results by
411 repeating all analyses using networks sampled only through focal observations
412 (Tables S3–S4). Analyses were conducted using R (R Core Team, 2017).

413 To determine how our sample of networks spanned the global nectar-feeding
414 bird species richness gradient in each bird family, we compared the cumulative
415 frequency distribution of bird species richness in grid cells across the global
416 distribution and the richness in the grid cells containing the sampled networks.
417 Comparisons were done using a two-sample Kolmogorov-Smirnov test in GraphPad
418 Prism 6.0 (Morgan, 1998). The global richness dataset was based on presence-absence
419 data for Trochilidae, Nectariniidae and Meliphagidae at the spatial resolution of 1×1
420 latitudinal-longitudinal degree grid following Rahbek & Graves (2001). This global
421 richness dataset was also used to build the richness maps in ArcMap 9.2 (ESRI, 2009;
422 Figs. 2–3).

423 RESULTS

424 Hummingbird-flower networks had lower connectance than sunbird- and honeyeater-
425 flower networks. Furthermore, hummingbird-flower networks had greater
426 complementary specialization and modularity than honeyeater-flower networks.
427 However, no differences were found between the complementary specialization and
428 modularity of hummingbird- and sunbird-flower networks, and neither between
429 sunbird- and honeyeater-flower networks. Likewise, using null-model corrected
430 values, a similar pattern of interaction specialization was observed (Fig. 1).
431 Additionally, when potentially confounding variables were included in the linear
432 models, hummingbird-flower networks still had lower connectance than sunbird- and
433 honeyeater-flower networks and greater complementary specialization and null-model
434 corrected binary modularity than honeyeater-flower networks. Moreover, no
435 differences were found between complementary specialization and binary modularity
436 of hummingbird- and sunbird-flower networks, and neither between sunbird- and

437 honeyeater-flower networks. Only weighted modularity did not differ between the
438 networks of these bird-flower communities when including potentially confounding
439 variables (Tables 1–2). Hummingbird communities visited more plant species than
440 sunbird and honeyeater communities (Kruskal-Wallis test: $K = 28.32$, $p < 0.001$;
441 Dunn's multiple comparison tests: hummingbirds $>$ sunbirds = honeyeaters).

442 Several of the confounding variables were associated with estimates of
443 interaction specialization (Tables 1–2). Notably, specialization increased with plant
444 species richness for both observed and null-model corrected metrics (Fig. S1).
445 Moreover, communities with greater bird than plant species richness exhibited greater
446 specialization, with lower observed connectance, but not in null-model corrected
447 connectance, where the pattern was reversed. On islands, networks were less
448 specialized, with lower observed and null-model corrected weighted modularity.
449 Sampling also influenced specialization, with greater specialization detected in
450 networks sampled by focal observations (Tables 1–2). Nonetheless, restricting the
451 analysis to networks sampled through focal observations, the most important predictor
452 variables ($\sum w_i \geq 0.80$) were the same as in the complete dataset, with the same pattern
453 described above for interaction specialization between the bird-flower communities
454 (Tables S3–S4). Intensity of sampling affected interaction specialization, with
455 decreased null-model corrected connectance and weighted modularity when sampling
456 intensity was high (Tables 1–2).

457 Hummingbird-flower networks were geographically widely distributed, but
458 with some parts of North America and the Amazon region being poorly sampled.
459 Also, when the cumulative frequency distribution of nectar-feeding bird species
460 richness was compared between the global and the sampled localities, we found that
461 species-poor communities were disproportionately less sampled than species-rich

462 hummingbird communities (Fig. S2). In the Old World, in contrast, network
463 distributions were more restricted and some species-rich regions, especially in Central
464 Africa, India, Southeast Asia and Southeast Australia, had few or no studies of bird-
465 flower interactions at the community-level (Figs. 2–3). Nonetheless, the included
466 networks encompassed well the global bird species richness gradient in sunbird and
467 honeyeater communities (Fig. S2).

468 DISCUSSION

469 We found that New World (NW) hummingbird-flower interaction networks are more
470 specialized than Old World (OW) honeyeater-flower networks, as predicted. Notably,
471 hummingbird-flower networks have fewer realized interactions, lower niche overlap
472 and greater binary modularity, as compared to honeyeater-flower networks.
473 Interaction specialization of sunbird-flower networks, however, was similar to both
474 hummingbird- and honeyeater-flower networks, contrarily to the prediction that they
475 would have intermediate values (see below).

476 The greater overall specialization between hummingbirds and their floral
477 resources indicates that their interactions are more species-restricted than the
478 interactions of honeyeaters with their flowers (Blüthgen, 2010). This greater
479 interaction partitioning in hummingbird networks may be a consequence of the
480 greater variation in bill length among hummingbirds than honeyeaters, as well as the
481 greater richness of bird-pollinated plant species in the NW (Fleming, 2005;
482 Abrahamczyk & Kessler, 2015; Higgins *et al.*, 2016). Hummingbird bill morphology
483 in combination with corolla morphology may play a key role in constraining
484 interactions via morphological mismatching (Cotton, 1998a; Temeles *et al.*, 2002;
485 Vizenin-Bugoni *et al.*, 2014). Indeed, in most hummingbird-flower communities,

486 there is a subset of flowers with long, curved corollas visited by one or few long and
487 curve-billed birds (Feinsinger & Colwell, 1978; Maglianesi *et al.*, 2014). Thus,
488 increased range of bill and corolla lengths in hummingbird-flower networks may
489 contribute to reduced niche overlap and increased community-level specialization
490 (Cotton, 1998b; Abrahamczyk & Kessler, 2010; Maruyama *et al.*, 2014; Maglianesi *et*
491 *al.*, 2015). A greater specialization in hummingbird-flower networks could also be
492 due to a greater spatio-temporal floral resource predictability (Fleming & Muchhala,
493 2008). NW annual flowering cycles tend to be more predictable (Bawa *et al.*, 2003)
494 than, for instance, the supra-annual mass flowering in South Asian forests (Sakai,
495 2002). Additionally, flowers are more diverse and abundant in the understory of NW
496 in comparison to OW forests (LaFrankie *et al.*, 2006). This greater diversity may
497 create new interaction opportunities for hummingbirds, resulting in greater niche
498 partitioning in the NW than in the OW networks. Conversely, the lower specialization
499 of honeyeater communities, compared to hummingbird communities, is likely to be
500 due to the much less variable bill length and corolla shapes in those communities
501 (Ford & Paton, 1977), particularly in northern Australia where most of the flowers
502 visited by honeyeaters have an open or cup-shaped corolla that is morphologically
503 accessible to several bird species (Ford *et al.*, 1979; Franklin & Noske, 2000). Hence,
504 more uniform bill lengths and more generalized corolla shapes among honeyeater-
505 flower communities may result in lower interaction specialization, when compared to
506 hummingbird-flower networks. Honeyeaters also tend to have broader dietary
507 preferences in general, feeding on other resources, such as fruits, insects and lerp
508 more frequently than hummingbirds do (Pyke, 1980; Higgins *et al.*, 2016). Although
509 hummingbirds also forage for insects as a source of protein (Stiles, 1995). These
510 diversified feeding habits of honeyeaters may decrease competition for nectar

511 resources, producing the more generalized interactions with flowers demonstrated
512 here (though see Dalsgaard *et al.*, 2017 for an opposed example on frugivorous birds
513 networks). The greatest overall specialization in hummingbird networks implies in
514 narrower niche overlaps of interactions. If combined to species turnover across
515 continental scales, this greater specialization may imply in a larger spatial variability
516 of interactions, resulting in a larger spatial β -diversity of interactions (Trojelsgaard *et*
517 *al.*, 2015) for hummingbird networks. Additionally, temporal variation of resources
518 spanning across the entire year in NW communities (Bawa *et al.*, 2003) may also
519 cause a temporal variation in interactions, resulting in a larger temporal β -diversity of
520 interactions.

521 The similarity between sunbird-flower networks and the other two bird-flower
522 communities is likely to be due two reasons: First, relatively few sunbird-flower
523 networks were available, resulting in wider confidence intervals (Fig. 1). Perhaps
524 more networks would have reduced that variability and made it possible to detect
525 differences between sunbird-flower networks and hummingbird- and honeyeater-
526 flower networks, respectively. Second, sunbird-flower communities are ecologically
527 similar to both hummingbird- and honeyeater-flower communities, and indeed have
528 intermediate interaction specialization levels (Fig. 1). For instance, although sunbirds
529 are considered less phenotypically specialized for nectar-feeding than hummingbirds
530 (Stiles, 1981), the flowers they visit may have rather restrictive morphologies, with
531 tubular and gullet shapes, similar to those of hummingbird-pollinated flowers
532 (Fleming & Muchhala, 2008). This greater morphological restriction of flowers may
533 decrease niche overlap among species, as tubular and gullet corolla shapes may be
534 inaccessible to some species of the nectar-feeding bird community (Pettet, 1977;
535 Temeles *et al.*, 2002). This morphological mismatch in both communities may

536 produce the similar interaction specialization of sunbird- and hummingbird-flower
537 networks that we detected in this study. Additionally, some sunbirds may have
538 specialized feeding behaviours, similar to hummingbirds, of hovering and traplining
539 while visiting flowers (Padyšáková & Janeček, 2016). However, this is not consistent
540 with the observation that interaction specialization was also similar between sunbird-
541 and honeyeater-flower communities. The specialization similarity of honeyeater and
542 sunbird communities may be related to their lower floral resource richness in
543 comparison to hummingbird communities, as demonstrated in this study. This lower
544 resource diversity may increase niche overlap, producing the more generalized
545 feeding-niches found in OW networks. However, we suggest that it is likely that a
546 larger sample size of sunbird-flower networks would have resolved these possibilities
547 in favour of our first explanation because the sunbird interaction specialization tends
548 to be intermediate between those of hummingbirds and honeyeaters (Fig. 1).

549 The correlation between plant species richness and interaction specialization
550 may be because niche availability is greater in species-rich than species-poor
551 communities, thereby promoting greater niche partitioning among species (Dalsgaard
552 *et al.*, 2011; Martín González *et al.*, 2015; Sebastián-González *et al.*, 2015; but see
553 Schleuning *et al.*, 2012). The greater generalization of interactions of insular
554 compared to mainland networks may be a consequence of their species-poor
555 communities (MacArthur & Wilson, 1963; Dalsgaard *et al.*, 2009; Abrahamczyk *et*
556 *al.*, 2015), but may also be due to the tendency of generalist species having greater
557 establishment success on islands than specialist species (Olesen *et al.*, 2002;
558 Maldonado *et al.*, 2013; Traveset *et al.*, 2015). Moreover, at least for hummingbirds,
559 generalized interactions on islands may have been influenced by their rather recent
560 colonization (McGuire *et al.*, 2014; Abrahamczyk *et al.*, 2015) and a greater level of

561 strong and periodic disturbances in islands in comparison to mainland communities,
562 which may favour generalized over specialized bird species (Dalsgaard *et al.*, 2009).

563 In conclusion, we confirmed that interactions are more specialized in
564 hummingbird- than in honeyeater-flower networks, but we were unable to show that
565 sunbird-flower networks differ from those of hummingbirds and honeyeaters. The
566 increased interaction specialization in the hummingbird-flower networks may be a
567 consequence of their greater floral resource richness and phenotypic specialization, in
568 contrast to honeyeater-flower communities (Stiles, 1981; Fleming & Muchhala,
569 2008). These results suggest that there is a potential link between phenotypic
570 specialization and floral resource richness with interaction specialization among
571 nectarivorous bird-flower communities across global scales.

572 ACKNOWLEDGEMENTS

573 We thank Marco A. R. Mello and two anonymous reviewers for their comments on a
574 previous version of the manuscript; Louis A. Hansen and Bjørn Hermansen for their
575 help in building the bird species richness map; André C. Guaraldo for his help with
576 the analysis. CAPES Foundation supported TBZ (PDSE scholarship proc. 8105/2014-
577 6), JVB, AGC, RRF, AVL, LCR and LRL; CNPq supported FCP, EF, ACA, AVL,
578 CGM, ICM, LCR, LRL, MSa and IGV; Czech Science Foundation supported DH, SJ
579 and OS (project n. 14-36098G); PAC was supported by the British Ornithologists'
580 Union and Wolfson College, University of Oxford; PKM by FAPESP (proc.
581 2015/21457-4); ACA and RRF by FUNDECT; ACB by Oticon Fonden Denmark;
582 DWC and TTJ by The Danish Council for Independent Research Natural Sciences;
583 TTJ by F-HF, KHF, TFNS, University of Aarhus; CL by the CACyPI-Uatx-2016GK;
584 AVL by FACEPE and OeAD; CGM by FAPESB; MAM by the CONICIT, MICIT,

585 CCT and UNED from Costa Rica, OTS, DAAD, and the research-funding program
586 “LOEWE” of Hesse’s Ministry of Higher Education, Research, and the Arts
587 (Germany); JMA thanks to Michael J. Lawes, RIEL, DCC, NTDP&I, RAAF,
588 P&WCNT, and NTP&W; LRL by FAEP, Unicamp and Ministerio del Medio
589 Ambiente de Colombia; ZW by National Natural Science Foundation of China; AGC
590 thanks to Reserva Ecológica Michelin; GK thanks Valentim Casett, owner of the
591 Prima Luna Reserve; TBZ, BD, AMMG and CR thank the Danish National Research
592 Foundation for its support of the Center for Macroecology, Evolution and Climate.

593 REFERENCES

- 594 Abrahamczyk, S. & Kessler, M. (2010) Hummingbird diversity, food niche
595 characters, and assemblage composition along a latitudinal precipitation gradient
596 in the Bolivian lowlands. *Journal of Ornithology*, 151, 615–625.
- 597 Abrahamczyk S. & Kessler M. (2015) Morphological and behavioural adaptations to
598 feed on nectar: how feeding ecology determines the diversity and composition of
599 hummingbird assemblages. *Journal of Ornithology*, 156, 333–347.
- 600 Abrahamczyk, S., Souto-Vilarós, D., McGuire, J.A., & Renner, S.S. (2015) Diversity
601 and clade ages of West Indian hummingbirds and the largest plant clades
602 dependent on them: a 5-9 Myr young mutualistic system. *Biological Journal of*
603 *the Linnean Society*, 114, 848–859.
- 604 Barber, M.J. (2007) Modularity and community detection in bipartite networks.
605 *Physical Review E*, 76, 1-11.
- 606 Barker, F.K., Barrowclough, G.F., & Groth, J.G. (2002) A phylogenetic hypothesis
607 for passerine birds: taxonomic and biogeographic implications of an analysis of
608 nuclear DNA sequence data. *Proceedings of the Royal Society of London B*, 269,

- 609 295–308.
- 610 Barton, K. (2016) MuMIn: Multi-Model Inference - R package. *R Foundation for*
611 *Statistical Computing*.
- 612 Bascompte, J. & Jordano, P. (2007) Plant-animal mutualistic networks: the
613 architecture of biodiversity. *Annual Review of Ecology, Evolution and*
614 *Systematics*, 38, 567–593.
- 615 Bawa, K.S., Kang, H., & Grayum, M.H. (2003) Relationships among the time,
616 frequency, and duration of flowering in tropical rain forest trees. *American*
617 *Journal of Botany*, 90, 877–887.
- 618 Blüthgen, N. (2010) Why network analysis is often disconnected from community
619 ecology: a critique and an ecologist's guide. *Basic and Applied Ecology*, 11,
620 185–195.
- 621 Blüthgen, N., Menzel, F., & Blüthgen, N. (2006) Measuring specialization in species
622 interaction networks. *BMC Ecology*, 6, 1–12.
- 623 Brooker, M., Braithwaite, R., & Estbergs, J. (1990) Foraging ecology of some
624 insectivorous and nectarivorous species of birds in forests and woodlands of the
625 wet-dry tropics of Australia. *Emu*, 90, 215–230.
- 626 Burnham, K.P. & Anderson, D.R. (2002) *Model selection and multimodel inference:*
627 *a practical information-theoretic approach*. Springer, New York.
- 628 Castellanos, M.C., Wilson, P., & Thomson, J.D. (2003) Pollen transfer by
629 hummingbirds and bumblebees, and the divergence of pollination modes in
630 Penstemon. *Evolution; international journal of organic evolution*, 57, 2742–
631 2752.
- 632 Collins, B.G. & Paton, D.C. (1989) Consequences of differences in body mass, wing
633 length and leg morphology for nectar-feeding birds. *Australian Journal of*

- 634 *Ecology*, 14, 269–289.
- 635 Collins, B.G. & Rebelo, T. (1987) Pollination biology of the Proteaceae in Australia
636 and southern Africa. *Austral Ecology*, 12, 387–421.
- 637 Cotton, P.A. (1998a) Coevolution in an Amazonian hummingbird-plant community.
638 *Ibis*, 140, 639–646.
- 639 Cotton, P.A. (1998b) The hummingbird community of a lowland Amazonian
640 rainforest. *Ibis*, 140, 512–521.
- 641 Cronk, Q. & Ojeda, I. (2008) Bird-pollinated flowers in an evolutionary and
642 molecular context. *Journal of Experimental Botany*, 59, 715–727.
- 643 Dalsgaard, B., Magård, E., Fjeldså, J., Martín González, A.M., Rahbek, C., Olesen,
644 J.M., Ollerton, J., Alarcón, R., Araujo, A.C., Cotton, P.A., Lara, C., Machado,
645 C.G., Sazima, I., Sazima, M., Timmermann, A., Watts, S., Sandel, B.,
646 Sutherland, W.J., & Svenning, J.C. (2011) Specialization in plant-hummingbird
647 networks is associated with species richness, contemporary precipitation and
648 quaternary climate-change velocity. *PLoS ONE*, 6, e25891.
- 649 Dalsgaard, B., Martín González, A.M., Olesen, J.M., Ollerton, J., Timmermann, A.,
650 Andersen, L.H., & Tossas, A.G. (2009) Plant-hummingbird interactions in the
651 West Indies: floral specialisation gradients associated with environment and
652 hummingbird size. *Oecologia*, 159, 757–766.
- 653 Dalsgaard, B., Schleuning, M., Maruyama, P.K., Dehling, D.M., Sonne, J., Vizentin-
654 Bugoni, J., Zanata, T.B., Fjeldså, J., Böhning-Gaese, K., & Rahbek, C. (2017)
655 Opposed latitudinal patterns of network-derived and dietary specialization in
656 avian plant-frugivore interaction systems. *Ecography*, doi: 10.1111/ecog.02604.
- 657 Devictor, V., Clavel, J., Julliard, R., Lavergne, S., Mouillot, D., Thuiller, W., Venail,
658 P., Villéger, S., & Mouquet, N. (2010) Defining and measuring ecological

- 659 specialization. *Journal of Applied Ecology*, 47, 15–25.
- 660 Dormann, C.F., Frund, J., Bluthgen, N., & Gruber, B. (2009) Indices, graphs and null
661 models: analyzing bipartite ecological networks. *The Open Ecology Journal*, 2,
662 7–24.
- 663 Dormann, C.F., Gruber, B., & Fründ, J. (2008) Introducing the bipartite package:
664 analysing ecological networks. *R News*, 8, 8–11.
- 665 Dormann, C.F. & Strauss, R. (2014) A method for detecting modules in quantitative
666 bipartite networks. *Methods in Ecology and Evolution*, 5, 90–98.
- 667 ESRI (2009) Environmental Systems Resource Institute. ArcMap 9.2. .
- 668 Feinsinger, P. & Colwell, R.K. (1978) Community organization among Neotropical
669 nectar-feeding birds. *American Zoologist*, 18, 779–795.
- 670 Fleming, T.H. (2005) The relationship between species richness of vertebrate
671 mutualists and their food plants in tropical and subtropical communities differs
672 among hemispheres. *Oikos*, 111, 556–562.
- 673 Fleming, T.H. & Muchhala, N. (2008) Nectar-feeding bird and bat niches in two
674 worlds: pantropical comparisons of vertebrate pollination systems. *Journal of*
675 *Biogeography*, 35, 764–780.
- 676 Ford, H.A. & Paton, D.C. (1977) The comparative ecology of ten species of
677 honeyeaters in South Australia. *Australian Journal of Ecology*, 2, 399–407.
- 678 Ford, H.A., Paton, D.C., & Forde, N. (1979) Birds as pollinators of Australian plants.
679 *New Zealand Journal of Botany*, 17, 509–519.
- 680 Fox, J. & Weisberg, S. (2011) *An R Companion to Applied Regression*. SAGE,
681 Thousand Oaks, CA.
- 682 Franklin, D.C. & Noske, R.A. (2000) Nectar sources used by birds in monsoonal
683 north-western Australia: a regional survey. *Australian Journal of Botany*, 48,

- 684 461–474.
- 685 Futuyma, D.J. & Moreno, G. (1988) The evolution of ecological specialization.
686 *Annual Review of Ecology and Systematics*, 19, 207–233.
- 687 Giraudoux, P. (2016) pgirmess: Data Analysis in Ecology - R package. *R Foundation*
688 *for Statistical Computing*.
- 689 Hair, J.F., Black, W.C., & Babin, B.J. (2009) *Multivariate Data Analysis: A Global*
690 *Perspective*. Prentice Hall, Upper Saddle River.
- 691 Higgins, P., Christidis, L., Ford, H., & Bonan, A. (2016) Honeyeaters (Meliphagidae).
692 *Handbook of the Birds of the World Alive* (ed. by J. del Hoyo, A. Elliott, J.
693 Sargatal, D.A. Christie, and E. Juana), retrieved from
694 <http://www.hbw.com/node/52353>. Lynx Edicions, Barcelona.
- 695 Hothorn, T., Bretz, F., & Westfall, P. (2008) Simultaneous inference in general
696 parametric models. *Biometrical Journal*, 50, 346–363.
- 697 del Hoyo, J., Elliot, A., Sargatal, J., Christie, D.A., & Juana, E. (2016) *Handbook of*
698 *the Birds of the World Alive*. Lynx Edicions, Barcelona.
- 699 Janeček, Š., Patačová, E., Bartoš, M., Padyšáková, E., Spitzer, L., & Tropek, R.
700 (2011) Hovering sunbirds in the Old World: occasional behaviour or
701 evolutionary trend? *Oikos*, 120, 178–183.
- 702 Johnson, S.D. & Steiner, K.E. (2000) Generalization versus specialization in plant
703 pollination systems. *Trends in Ecology and Evolution*, 15, 140–143.
- 704 Jordano, P. (1987) Patterns of mutualistic interactions in pollination and seed
705 dispersal: connectance, dependence asymmetries, and coevolution. *The American*
706 *Naturalist*, 129, 657–677.
- 707 LaFrankie, J. V., Ashton, P.S., Chuyong, G.B., Co, L., Condit, R., Davies, S.J.,
708 Foster, R., Hubbell, S.P., Kenfack, D., Lagunzad, D., Losos, E.C., Nor, N.S.M.,

- 709 Tan, S., Thomas, D.W., Valencia, R., & Villa, G. (2006) Contrasting structure
710 and composition of the understory in species-rich tropical rain forests. *Ecology*,
711 87, 2298–2305.
- 712 MacArthur, R.H. & Wilson, E.O. (1963) An equilibrium theory of insular
713 zoogeography. *Evolution*, 17, 373–387.
- 714 Maglianesi, M.A., Bluthgen, N., Bohning-Gaese, K., & Schleuning, M. (2014)
715 Morphological traits determine specialization and resource use in plant-
716 hummingbird networks in the neotropics. *Ecology*, 95, 3325–3334.
- 717 Maglianesi, M.A., Blüthgen, N., Böhning-Gaese, K., & Schleuning, M. (2015)
718 Functional structure and specialization in three tropical plant-hummingbird
719 interaction networks across an elevational gradient in Costa Rica. *Ecography*, 38,
720 1119–1128.
- 721 Maldonado, M.B., Lomáscolo, S.B., & Vázquez, D.P. (2013) The importance of
722 pollinator generalization and abundance for the reproductive success of a
723 generalist plant. *PLoS ONE*, 8, e75482.
- 724 Marquitti, F.M.D., Guimarães, P.R., Pires, M.M., & Bittencourt, L.F. (2014)
725 MODULAR: software for the autonomous computation of modularity in large
726 network sets. *Ecography*, 37, 221–224.
- 727 Martín González, A.M., Dalsgaard, B., Nogués-Bravo, D., Graham, C.H., Schleuning,
728 M., Maruyama, P.K., Abrahamczyk, S., Alarcón, R., Araujo, A.C., Araújo, F.P.,
729 Azevedo, S.M., Baquero, A.C., Cotton, P.A., Ingversen, T.T., Kohler, G., Lara,
730 C., Las-Casas, F.M.G., Machado, A.O., Machado, C.G., Maglianesi, M.A.,
731 McGuire, J.A., Moura, A.C., Oliveira, G.M., Oliveira, P.E., Ornelas, J.F.,
732 Rodrigues, L.D.C., Rosero-Lasprilla, L., Rui, A.M., Sazima, M., Timmermann,
733 A., Varassin, I.G., Vizentin-Bugoni, J., Wang, Z., Watts, S., Rahbek, C., &

- 734 Martinez, N.D. (2015) The macroecology of phylogenetically structured
735 hummingbird-plant networks. *Global Ecology and Biogeography*, 24, 1212–
736 1224.
- 737 Maruyama, P.K., Vizentin-Bugoni, J., Oliveira, G.M., Oliveira, P.E., & Dalsgaard, B.
738 (2014) Morphological and spatio-temporal mismatches shape a Neotropical
739 savanna plant-hummingbird network. *Biotropica*, 46, 740–747.
- 740 McGuire, J.A., Witt, C.C., Remsen, J.V., Corl, A., Rabosky, D.L., Altshuler, D.L., &
741 Dudley, R. (2014) Molecular phylogenetics and the diversification of
742 hummingbirds. *Current Biology*, 24, 910–916.
- 743 Morgan, W.T. (1998) A review of eight statistics software package for general use.
744 *The American Statistician*, 52, 70–82.
- 745 Olesen, J.M., Bascompte, J., Dupont, Y.L., & Jordano, P. (2007) The modularity of
746 pollination networks. *Proceedings of the National Academy of Sciences of the*
747 *United States of America*, 104, 19891–19896.
- 748 Olesen, J.M., Eklidsen, L.I., & Venkatasamy, S. (2002) Invasion of pollination
749 networks on oceanic islands: importance of invader complexes and endemic
750 super generalists. *Diversity and Distributions*, 8, 181–192.
- 751 Olesen, J.M. & Jordano, P. (2002) Geographic patterns in plant-pollinator mutualistic
752 networks. *Ecology*, 83, 2416–2424.
- 753 Ollerton, J., Killick, A., Lamborn, E., Watts, S., & Whiston, M. (2007) Multiple
754 meanings and modes: on the many ways to be a generalist flower. *Taxon*, 56,
755 717–728.
- 756 Padyšáková, E. & Janeček, Š. (2016) Sunbird hovering behavior is determined by
757 both the forager and resource plant. *Biotropica*, 48, 687–693.
- 758 Patefield, W.M. (1981) Algorithm AS 159: an efficient method of generating random

- 759 R × C tables with given row and column totals. *Journal of the Royal Statistical*
760 *Society. Series C (Applied Statistics)*, 30, 91–97.
- 761 Pettet, A. (1977) Seasonal changes in nectar-feeding by birds at Zaria, Nigeria. *Ibis*,
762 119, 291–308.
- 763 Prum, R.O., Berv, J.S., Dornburg, A., Field, D.J., Townsend, J.P., Moriarty Lemmon,
764 E., & Lemmon, A.R. (2015) A comprehensive phylogeny of birds (Aves) using
765 targeted next-generation DNA sequencing. *Nature*, 526, 569–573.
- 766 Pyke, G.H. (1980) The foraging behaviour of Australian honeyeaters: a review and
767 some comparisons with hummingbirds. *Australian Journal of Ecology*, 5, 343–
768 369.
- 769 R Core Team (2017) *R: A language and environment for statistical computing*. R
770 Foundation for Statistical Computing, Vienna, Austria.
- 771 Rahbek, C. & Graves, G.R. (2001) Multiscale assessment of patterns of avian species
772 richness. *Proceedings of the National Academy of Sciences of the United States*
773 *of America*, 98, 4534–4539.
- 774 Ramírez-Burbano, M.B., Stiles, G., González, C., Amorim, F., Dalsgaard, B., &
775 Maruyama, P.K. (2017) The role of the endemic and critically endangered
776 Colorful Puffleg *Eriocnemis mirabilis* in plant-hummingbird networks of the
777 Colombian Andes. *Biotropica*, in press.
- 778 Sakai, S. (2002) General flowering in lowland mixed dipterocarp forests of south-east
779 Asia. *Biological Journal of the Linnean Society*, 75, 233–247.
- 780 Schleuning, M., Fründ, J., Klein, A.M., Abrahamczyk, S., Alarcón, R., Albrecht, M.,
781 Andersson, G.K.S., Bazarian, S., Böhning-Gaese, K., Bommarco, R., Dalsgaard,
782 B., Dehling, D.M., Gotlieb, A., Hagen, M., Hickler, T., Holzschuh, A., Kaiser-
783 Bunbury, C.N., Kreft, H., Morris, R.J., Sandel, B., Sutherland, W.J., Svenning,

- 784 J.C., Tschardtke, T., Watts, S., Weiner, C.N., Werner, M., Williams, N.M.,
785 Winqvist, C., Dormann, C.F., & Blüthgen, N. (2012) Specialization of
786 mutualistic interaction networks decreases toward tropical latitudes. *Current*
787 *Biology*, 22, 1925–1931.
- 788 Schleuning, M., Ingmann, L., Strauß, R., Fritz, S.A., Dalsgaard, B., Dehling, D.M.,
789 Plein, M., Saavedra, F., Sandel, B., Svenning, J.C., Böhning-Gaese, K., &
790 Dormann, C.F. (2014) Ecological, historical and evolutionary determinants of
791 modularity in weighted seed-dispersal networks. *Ecology Letters*, 17, 454–463.
- 792 Sebastián-González, E., Dalsgaard, B., Sandel, B., & Guimarães, Jr. P.R. (2015)
793 Macroecological trends in nestedness and modularity of seed-dispersal networks:
794 human impact matters. *Global Ecology and Biogeography*, 24, 293–303.
- 795 Stiles, F.G. (1981) Geographical aspects of bird-flower coevolution, with particular
796 reference to Central America. *Annals of the Missouri Botanical Garden*, 68,
797 323–351.
- 798 Stiles, F.G. (1995) Behavioral, ecological and morphological correlates of foraging
799 for arthropods by the hummingbirds of a tropical wet forest. *The Condor*, 97,
800 853–878.
- 801 Temeles, E.J., Linhart, Y.B., Masonjones, M., & Masonjones, H.D. (2002) The role of
802 flower width in hummingbird bill length-flower length relationships. *Biotropica*,
803 34, 68–80.
- 804 Thomson, J.D. & Wilson, P. (2008) Explaining evolutionary shifts between bee and
805 hummingbird pollination: convergence, divergence, and directionality.
806 *International Journal of Plant Sciences*, 169, 23–38.
- 807 Traveset, A., Olesen, J.M., Nogales, M., Vargas, P., Jaramillo, P., Antolín, E., Trigo,
808 M.M., & Heleno, R. (2015) Bird-flower visitation networks in the Galápagos

- 809 unveil a widespread interaction release. *Nature Communications*, 6, 6376.
- 810 Trojelsgaard, K., Jordano, P., Carstensen, D.W., & Olesen, J.M. (2015) Geographical
811 variation in mutualistic networks: similarity, turnover and partner fidelity.
812 *Proceedings of the Royal Society B: Biological Sciences*, 282, 20142925.
- 813 Vázquez, D.P. & Stevens, R.D. (2004) The latitudinal gradient in niche breadth:
814 concepts and evidence. *The American Naturalist*, 164, 1–19.
- 815 Vizentin-Bugoni, J., Maruyama, P.K., & Sazima, M. (2014) Processes entangling
816 interactions in communities: forbidden links are more important than abundance
817 in a hummingbird-plant network. *Proceedings of the Royal Society B: Biological*
818 *Sciences*, 281, 1–8.
- 819 Waser, N.M., Chittka, L., Price, M. V, Williams, N.M., & Ollerton, J. (1996)
820 Generalization in pollination systems, and why it matters. *Ecology*, 77, 1043–
821 1060.
- 822 Wester, P. (2013) Sunbirds hover at flowers of *Salvia* and *Lycium*. *Ostrich: Journal of*
823 *African Ornithology*, 84, 27–32.

824 SUPPORTING INFORMATION

825 Supporting information can be found in the online version of this article:

826 Appendix S1 – Data source (Table S1), geographical and sampling details of each
827 study used to build the interaction networks of bird-flower communities (Table S2).

828 Appendix S2 – Multiple linear regression models of bird-flower interaction networks
829 sampled through focal observation (Table S3–S4).

830 Appendix S3 – Linear regressions between network metrics and the plant species
831 richness (Fig. S1) and two-sample Kolmogorov-Smirnov test comparing the global

832 and sampled cumulative species richness frequency of the three bird families (Fig.
833 S2).

834 BIOSKETCH

835 Thais B. Zanata is an ecologist interested in mutualistic interactions, specifically in
836 the effects of morphology, evolution and climate on interaction patterns across broad
837 geographical scales. This working group, a collaboration among researchers that study
838 bird-flower interactions in different continents, was established to test the ideas
839 proposed by Fleming & Muchhala (2008). Authors contributions: TBZ, BD and IGV
840 conceived the ideas; BD, PAC, JJR, PKM, EF, MSc, AMMG, JVB, DCF, SA, RA,
841 ACA, FPA, SMAJ, ACB, KBG, DWC, HC, AGC, RRF, DH, TTI, SJ, GK, CL,
842 FMGLC, AVL, AOM, CMG, ICM, MAM, TSM, JMA, ACM, GMO, PEO, JFO, JR,
843 LCR, LRL, AMR, MSa, BS, OS, AT, MGRV, ZW, SW, CR and IGV collected the
844 data and helped with discussion; TBZ analysed the data; and TBZ, BD, FCP, PAC,
845 JJR, PKM, EF, MSc, AMMG, JVB, DCF and IGV led the writing. All authors read
846 and approved the final version of the manuscript.

847 Editor: Holger Kreft

Table 1 – Multiple linear regression models predicting observed (OBS) and null-model corrected (Δ) connectance (C) and complementary specialization (H_2') of bird-flower interaction networks. Connectance (C) is the realized proportion of possible interactions in a community while complementary specialization (H_2') measures niche overlap among species. Predictor variables are described in the Material and Methods. Numbers in bold are predictor variables whose importance (Σw_i) is ≥ 0.80 . A dash indicates that the predictor variable was absent from the MAM. Numbers in parenthesis are the sample size used in each analysis.

		Connectance (C)						Complementary specialization (H_2')					
		C_{OBS} (121)			ΔC (94)			$H_2'_{OBS}$ (94)			$\Delta H_2'$ (94)		
		Σw_i	AVM	MAM ^a	Σw_i	AVM	MAM ^b	Σw_i	AVM	MAM ^c	Σw_i	AVM	MAM ^d
Predictor variables	Bird-flower community	1.00			1.00			0.96			0.98		
	Hummingbirds		-0.11	-0.11 (A)		-0.11	-0.12 (A)		0.16	0.13 (A)		0.17	0.17 (A)
	Sunbirds		0.02	0.03 (B)		0.09	0.09 (B)		0.05	0.01 (AB)		0.08	0.10 (AB)
	Honeyeaters		1.01	1.00 (B)		0.16	0.17 (B)		0.11	0.11 (B)		-0.04	-0.04 (B)
	Plant species richness	1.00	-0.55	-0.54	1.00	-0.23	-0.24	0.92	0.18	0.17	1.00	0.22	0.22
	Asymmetry	1.00	-0.33	-0.32	1.00	0.19	0.19	0.28	-0.03	–	0.31	-0.07	–

Insularity	0.63	0.07	0.07	0.24	0.02	–	0.45	-0.08	–	0.33	-0.05	–
Topography	0.23	0.01	–	0.29	-0.01	–	0.43	0.03	0.03	0.38	0.03	–
Latitude	0.36	0.01	–	0.37	0.01	–	0.42	-0.02	-0.03	0.27	-0.01	–
Sampling method	0.94	0.14	0.15	0.98	0.19	0.20	0.97	-0.36	-0.33	0.99	-0.38	-0.39
Duration	0.25	-0.01	–	0.30	-0.02	–	0.26	0.02	–	0.29	0.02	–
Sampling coverage	0.23	-0.01	–	0.21	-0.01	–	0.25	0.03	–	0.23	0.02	–
Sampling intensity				1.00	-0.17	-0.17	0.57	-0.12	–	0.25	0.03	–
AICc			-146.96			-168.06			-58.51			-64.07
R ² adjusted			0.69			0.50			0.34			0.38
R ² total Bird-flower community			0.32			0.29			0.21			0.25
R ² only Bird-flower community			0.04			0.16			0.04			0.08
Moran's <i>I</i>			≤0.04 ^{NS}			≤0.13 ^{NS}			≤0.06 ^{NS}			≤0.08 ^{NS}

AICc – corrected Akaike's Information Criterion; AVM – standardized coefficients of the averaged model measured across all models; MAM – standardized coefficients of the minimum adequate model with the lowest Δ AICc; R² adjusted – variation explained by the minimum adequate model with the lowest Δ AICc; R² only Bird-flower community – adjusted unique variation explained by the difference between the three types of bird-

flower communities; $R^2_{\text{total Bird-flower community}}$ – adjusted total variation explained by the difference between the three types of bird-flower communities ; Σw_i – importance of each predictor variable measured across all models; $^{NS}p > 0.05$; number of models with $\Delta AICc \leq 2$: a - three; b - four; c - eleven; d - six. For all models with $\Delta AICc \leq 2$, the predictor variable that represents the difference between the three types of bird-flower communities was selected. Letters next to the standardized coefficients represent the group that each bird-flower community belongs, based on Tukey contrasts for general linear hypotheses. Multicollinearity between predictor variables is absent, as generalized variance inflation factor (GVIF) ≤ 1.72 .

Table 2 – Multiple linear regression models predicting observed (OBS) and null-model corrected (Δ) binary (Q_B) and weighted modularity (Q) of bird-flower interaction networks. Modularity is a network metric that detects preferentially interacting subsets of species within the community. Predictor variables are described in the Material and Methods. Numbers in bold are predictor variables whose importance (Σw_i) is ≥ 0.80 . A dash indicates that the predictor variable was absent from the MAM. Numbers in parenthesis are the sample size used in each analysis.

		Binary modularity (Q_B)						Weighted modularity (Q)					
		Q_{B-OBS} (121)			ΔQ_B (94)			Q_{OBS} (94)			ΔQ (94)		
		Σw_i	AVM	MAM ^a	Σw_i	AVM	MAM ^b	Σw_i	AVM	MAM ^c	Σw_i	AVM	MAM ^d
Predictor variables	Bird-flower community	0.99			1.00			0.28			0.27		
	Hummingbirds		0.07	0.07 (A)		0.08	0.08 (A)		0.06	–		0.05	–
	Sunbirds		-0.01	-0.01 (AB)		0.01	0.01 (AB)		0.06	–		0.05	–
	Honeyeaters		0.12	0.09 (B)		-0.06	-0.02 (B)		0.12	–		-0.07	–
	Plant species richness	1.00	0.14	0.17	1.00	0.12	0.08	1.00	0.21	0.21	1.00	0.26	0.23
	Asymmetry	0.57	0.08	0.09	0.64	-0.06	–	0.24	-0.03	–	0.60	-0.09	–
	Insularity	0.43	-0.04	–	0.71	-0.04	-0.05	0.99	-0.11	-0.11	0.82	-0.08	-0.09

Topography	0.33	0.01		0.63	0.01	0.02	0.74	0.03	0.03	0.68	0.02	0.03
Latitude	0.52	-0.01	–	0.60	-0.01	-0.01	0.23	-0.01	–	0.22	-0.01	–
Sampling method	0.63	-0.08	-0.08	1.00	-0.15	-0.14	0.91	-0.21	-0.20	0.95	-0.21	-0.20
Duration	0.25	-0.01	–	0.22	-0.01	–	0.23	-0.01	–	0.27	0.01	–
Sampling coverage	0.24	0.01	–	0.27	0.02	–	0.22	0.01	–	0.22	0.01	–
Sampling intensity				0.39	0.03	–	1.00	-0.15	-0.15	0.25	0.02	–
AICc			-211.75			-241.09			-138.50			-154.64
R ² adjusted			0.35			0.55			0.52			0.49
R ² _{total Bird-flower community}			0.23			0.34			–			–
R ² _{only Bird-flower community}			0.06			0.09			–			–
Moran's <i>I</i>			≤0.06 ^{NS}			≤0.17 ^{NS}			≤0.07 ^{NS}			≤0.06 ^{NS}

AICc – Akaike's Information Criterion corrected; AVM – standardized coefficients of the averaged model measured across all models; MAM – standardized coefficients of the minimum adequate model with the lowest Δ AICc; R² adjusted – variation explained by the minimum adequate model with the lowest Δ AICc; R²_{only Bird-flower community} – adjusted unique variation explained by the difference between the three types of bird-flower communities; R²_{total Bird-flower community} – adjusted total variation explained by the difference between the three types of bird-flower

communities; Σw_i - importance of each predictor variable measured across all models; $^{NS}p > 0.05$; number of models with $\Delta AICc \leq 2$: a - eleven; b - nine; c - one; d - five. Only for binary modularity, all models with $\Delta AICc \leq 2$ the predictor variable that represents the difference between the three types of bird-flower communities was selected. Letters next to the standardized coefficients represent the group that each bird-flower community belongs, based on Tukey contrasts for general linear hypotheses. Multicollinearity between predictor variables is absent in binary modularity models, as generalized variance inflation factor (GVIF) ≤ 1.60 and in weighted modularity models, as variance inflation factor (VIF) ≤ 2.30 .

848 Figure legends

849 Fig. 1 – Comparison of the observed (OBS, first column) and null-model corrected
850 (Δ , second column) metrics of bird-flower interaction networks of the three types of
851 bird communities (hummingbirds, sunbirds and honeyeaters). Connectance (C) is the
852 realized proportion of possible interactions in a community, complementary
853 specialization (H_2') measures niche overlap among species and modularity (Q_B and Q)
854 detects preferentially interacting subsets of species within the community. Observed
855 connectance and binary modularity have a sample size of 121 networks, while the
856 other metrics have a sample size of 94 networks. Dots represent each network and
857 lines indicate mean and confidence interval at 95% confidence level. Letters represent
858 the group that each bird-flower community belongs, based on Tukey's or Dunn's
859 multiple comparison tests.

860 Fig. 2 – Specialization of bird-flower interaction networks in hummingbird, sunbird
861 and honeyeater communities, measured by observed connectance (C_{OBS}), which
862 describes the realized proportion of possible interactions in a community, and
863 observed complementary specialization ($H_2'_{OBS}$), which calculates the niche overlap
864 among species. Observed connectance has a sample size of 121 networks, while
865 complementary specialization has a sample size of 94 networks. The species richness
866 of hummingbirds, sunbirds and honeyeaters are represented in grey shades,
867 intensifying with an increase in species richness. Points were moved slightly to
868 improve visualization.

869 Fig. 3 – Specialization of bird-flower interaction networks in in hummingbird, sunbird
870 and honeyeater communities, measured by observed binary (Q_{B-OBS}) and weighted

871 modularity (Q_{OBS}). Modularity is a network metric that detects preferentially
872 interacting subsets of species within the community. Observed binary modularity has
873 a sample size of 121 networks, while weighted modularity has a sample size of 94
874 networks. The species richness of hummingbirds, sunbirds and honeyeaters are
875 represented in grey shades, intensifying with an increase in species richness. Points
876 were moved slightly to improve visualization.





