# Design and Synthesis of Inhibitors of Hypoxia Inducible Factor-1-mediated Functions 

Lingyun Yang<br>applejing1277@gmail.com

Follow this and additional works at: http://scholarworks.gsu.edu/chemistry_theses

## Recommended Citation

Yang, Lingyun, "Design and Synthesis of Inhibitors of Hypoxia Inducible Factor-1-mediated Functions." Thesis, Georgia State University, 2017.
http://scholarworks.gsu.edu/chemistry_theses/104

# DESIGN AND SYNTHESIS OF INHIBITORS OF HYPOXIA INDUCIBLE FACTOR-1MEDIATED FUNCTIONS 

by

## LINGYUN YANG

## Under the Direction of Professor Binghe Wang


#### Abstract

Hypoxia Inducible Factors (HIFs) are very important transcription factors that can respond to low oxygen concentrations in the cellular environment. Inhibition of HIF's transcriptional activity represents a promising approach to new anticancer compounds. Herein, we describe the design and synthesis of a series of HIF-1 inhibitors. Evaluation of these inhibitors using a cellbased luciferase assay led to the discovery compounds with sub-micromolar potency.


# DESIGN AND SYNTHESIS OF INHIBITORS OF HYPOXIA INDUCIBLE FACTOR-1MEDIATED FUNCTIONS 

by

## LINGYUN YANG

A Thesis Submitted in Partial Fulfillment of the Requirements for the Degree of Master of Science in the College of Arts and Sciences<br>Georgia State University

Copyright by Lingyun Yang 2017

# DESIGN AND SYNTHESIS OF INHIBITORS OF HYPOXIA INDUCIBLE FACTOR-1MEDIATED FUNCTIONS 

by

## LINGYUN YANG

## DEDICATION

I would like to dedicate this work to my family, especially my parents Xiaoling Chen and Siping Yang. Thanks for their supporting and encouragement through my whole graduate school studying. My mum never questions all decisions, which I have made about academic. I also want to thank my all friends, who always stay around me with love and kindness. Finally, I would like to thank my boyfriend Weiqing Chen, his patient and love gives me the power during this process. I dedicate this work to you.

## ACKNOWLEDGEMENTS

I would like to acknowledge to all people who help me during this work. Without them, I could not complete the whole work. First, I would like to thank Dr. Binghe Wang. During my past three years in his lab, he always put his encouragement and supporting on me even I have got multiple failure. He never gave me up and doubted me on my research experiments. I will forever remember the moments that I worked in his lab. I also would like to thank my committee members, Dr. Donald Hamelberg and Dr. Suri.S Iyer. Their comments to me have pushed me to become better in future career. I would also want to thank Dr. Al Baumstark who taught me not only the knowledge of chemistry but also the wisdom of life. He let me know that chemistry and life can integrate perfectly, and he pushed me to continue on as a chemist. I would like to thank Dr. Haishi Cao who brought me to chemistry field, and he let me know how interesting chemistry is and told me that I can utilize the knowledge to help others in career.

Secondly, I would like to thank Dr. Jalisa Holmes Ferguson for helping me with this project. She is not only my mentor but also my important friend. Without her help, I cannot finish all that I have alone. She taught me how to operate the reaction professionally and how to become a profession chemist. Thirdly, I would like to thank Dr. Chaofeng Dai for bringing me to this project and teaching me the techniques.

I would like to thank all my lab members for supporting and helping me in my past years, and I am so grateful that they always stand by me from the beginning no matter what happens. To Bingchen who works nearby my hood, thank you for giving me lots of tips on my research and experiments. To Zhixiang, thank you for being my friend and you were always here when I need you and gave me the help for the knowledgement. To Vayou who graduated with me at same time, thank you for being there when I was struggling; I would miss the conversation with
you about chemistry and everything else. To Kim and Manjusha, thank you for cheering me up from the beginning. To Zhengnan, thank you for helping me relax when I feel upset, thank you very much. To Dr. Kaili Ji, you were like my sister, thank you for teaching me all biology knowledge; I would remember our friendship forever. To Yueqing, thank you for helping me on operating equipment. To all my friend and lab members I forgot to mention, thank you for being with me and supporting me, you are my hidden treasure and a part of my life. I will forever appreciate all you did for me. Thank you very much.

I would like to thank all my friends who are around me. You gave me the strength and made me to go forward throughout my graduated studying. To Jie, thank you for keeping me happy when I was in bad mood, and you are like my sister. Thank you very much. To Porla and Steve, you are my sweet neighbor. Thank you for taking care of me in many aspects when I was too busy to focus in life. To fan, you are like my big brother. You always supported me to do the things that I like. Thank you for being patient and giving me many life suggestions. Thank you all my friends who I forgot to mention, and this work is for you. Thank you very much.

## TABLE OF CONTENTS

ACKNOWLEDGEMENTS ..... v
LIST OF TABLES ..... ix
LIST OF SCHEMES ..... xi
LIST OF ABBREVIATIONS ..... xii
1 INTRODUCTION ..... 1
1.1 Purpose of the Study ..... 1
1.2 Hypoxia Inducible Factor-1 Pathway ..... 1
1.3 HIF-1 Inhibitors ..... 3
1.4 HIF-1 Inhibitors Classes ..... 5
2 CHEMISTRY DESIGN AND EXPERIMENTS ..... 7
2.1 Design ..... 7
2.2 Chemistry ..... 8
3 BIOLOGY RESULT ..... 15
3.1 Results for Class I Analogues ..... 15
3.2 Results for Class II Analogues ..... 16
3.3 Results for Class III Analogues. ..... 18
3.4 Results for Class IV Analogues ..... 19
3.5 Results for Class V Analogues ..... 20
4 CONCLUSIONS AND FUTURE WORK ..... 21
4.1 Structure Activity Relationship Study ..... 21
4.2 Future Work ..... 22
5 EXPERIMENTAL ..... 22
REFERENCES ..... 37
APPENDICES ..... 41

## LIST OF TABLES

Table 1: Class I Analogues ..... 16
Table 2: Class II Analogues ..... 17
Table 3: Class III Analogues ..... 18
Table 4: Class IV Analogues ..... 20
Table 5: Class V Analogues ..... 21

## LIST OF FIGURES

Figure 1: Chemical Structure of AFP-464 ..... 3
Figure 2: Structure and $\mathrm{IC}_{50}$ value of KCN 1 ..... 4
Figure 3: Structure and $\mathrm{IC}_{50}$ value of SRIV-64b ..... 4
Figure 4: Manassantin A and B ..... 6
Figure 5: Lead compound structure and $\mathrm{IC}_{50}$ value ..... 6
Figure 6: Analogues Designed ..... 7

## LIST OF SCHEMES

Scheme 1: Synthesis of ortho-phenolic ether benzhydrol analogues ..... 9
Scheme 2: Synthesis of para-phenolic ether benzhydrol analogues 9a-9c ..... 10
Scheme 3: Synthesis of para-phenolic ether benzhydrol analogues $\mathbf{9 d}{ }^{\#}-\mathbf{9 g}$ * ..... 11
Scheme 4: Synthesis of meta-phenolic ether benzhydrol analogues 17a-17c ..... 12
Scheme 5: Synthesis of meta-phenolic ether benzhydrol analogues 17d and 17e* ..... 12
Scheme 6: Synthesis of ortho-benzyl ether benzhydrol analogues ..... 13
Scheme 7: Synthesis of tri-substituted phenolic ether benzhydrol analogues ..... 14

## LIST OF ABBREVIATIONS

| HIF-1 | Hypoxia Inducible Factor 1 |
| :---: | :---: |
| UPP | Ubiquition proteasome pathway |
| ODD | Oxygen dependent degradation domain |
| PHD | Prolyl hydroxylase dioxygenase |
| pVHL | Von Hippel Lindau tumor suppressor prptein |
| HRE | Hypoxia-responsive element |
| VEGF | Vascular epidermal growth factor |
| EPO | Erythropoietin |
| DMF | Dimethylformamide |
| $\mathrm{K}_{2} \mathrm{CO}_{3}$ | Potassium carbonate |
| $n-B u L i$ | $n$-Butyllithium |
| THF | Tetrahydrofuran |
| r.t. | Room temperature |
| $\mathrm{Boc}_{2} \mathrm{O}$ | Di-tert-butyl dicarbonate |
| TsCI | 4-Toluenesulfonyl chloride |
| $\mathbf{N E t}_{3}$ | Triethylamine |
| NaH | Sodium hydride |
| $\mathrm{NH}_{4} \mathrm{Cl}$ | Ammonium chloride |
| Luc | Luciferase |
| SAR | Structure activity relationship |
| $\mathrm{CDCl}_{3}$ | Deuterated chloroform |
| $\mathbf{M g S O}_{4}$ | Magnesium sulfate |

## 1 INTRODUCTION

### 1.1 Purpose of the Study

A tumor is an abnormal growth of body tissue. Malignant tumors or cancer represent a major threat to human health. Based on the GLOBOCAN database, ${ }^{[1]}$ tens of millions of people are diagnosed with cancer around the world each year, and more than half of them die from it. Cancer is generally considered a name for a class of diseases with varying characteristics. As a result, treatment outcome varies and a cure is an elusive goal. ${ }^{[2]}$ Although there are debates as to whether one can truly summarize certain traits as hallmarks of cancer, ${ }^{[3-4]}$ one thing is true that malignant solid tumors are often accompanied by a state of hypoxic, or low oxygen, conditions, presumably due to their rapid growth and inadequate vascularization. ${ }^{[5]}$ Such hypoxic states activate the expression of genes responsible for malignancy, aggressiveness, metastasis, and treatment-refractory properties. ${ }^{[6]}$

### 1.2 Hypoxia Inducible Factor-1 Pathway

Hypoxia Inducible Factors (HIF) are a family of transcription factors that regulate hypoxia-driven gene expression. ${ }^{[7-8]}$ HIFs can be activated under hypoxic conditions and induce target genes that regulate adaptive biological processes such as cell motility, anaerobic metabolism and angiogenesis. ${ }^{[9]} \mathrm{HIF}-1$ is the chief regulator in the response of growing tumor to hypoxia. ${ }^{[10]}$ HIF-1 is a heterodimeric protein complex composed of $\alpha$ and $\beta$ subunits, and HIF-1a plays a role as an oxygen-sensitive transcriptional activator while HIF-1 $\beta$ is constitutively expressed in the cell nucleus. ${ }^{[11-}$ ${ }^{13]}$ In the presence of oxygen, HIF-1a can be destroyed rapidly through the ubiquition-
proteasome pathway (UPP). ${ }^{[14]} \mathrm{HIF}$-1a subunit is dihydroxylated at key proline residues within the oxygen dependent degradation domain (ODD) of HIF-1a. During this process, PHD2, a member of the prolyl hydroxylase family, along with 2-oxoglutarate, oxygen and $\mathrm{Fe}(\mathrm{II})$ are required to promote the hydroxylation of HIF-1a. ${ }^{[15-17]}$ Following this process, HIF-1a can bind to the von Hippel Lindau tumor suppressor protein ( pVHL ) which can polyubiquitinate HIF-1a due to its E3-ubiquitin ligase activity, subsequently signaling degradation via the ubiquition-proteasome pathway (UPP). ${ }^{[18]}$ Under hypoxic conditions, PHD2 activity is inhibited because of the reduction of oxygen, and HIF-1a accumulates rapidly and forms the HIF-1 dimer with HIF-1 $\beta$, regulating the transcription of many genes in a cell. ${ }^{[19]}$ With the help of cofactor p300, HIF-1 binds to the hypoxia-responsive element (HRE) sequence of DNA, and promotes the expression of a number of target genes such as vascular epidermal growth factor (VEGF) and erythropoietin (EPO), which help hypoxic cells survive. ${ }^{[20-22]}$ Solid tumors need an increased blood supply to grow and spread to other organs and regions of the body, and overexpression of VEGF is able to develop the enhanced blood supply for them. ${ }^{[23]}$ EPO has a role of controlling erythropoiesis or red blood cell production, and it can be used to treat anemia from kidney failure or cancer treatment. ${ }^{[24]}$ Over expression of EPO is a factor that exhibits an anti-apoptotic action on numerous cells, including malignant ones. ${ }^{[25]}$

### 1.3 HIF-1 Inhibitors

Recently, many anti-cancer compounds have been developed to inhibit the HIF pathway. ${ }^{[26]}$ Some of them have been applied in the clinic successfully. The 2016 Laskar award was given to three scientists: William Kaelin, Jr., Peter Ratcliffe, and Gregg Semenza, for their seminal contributions in oxygen sensing and associated biological implications, further demonstrating the importance of this area. Inhibition of HIF-1 can be achieved in many ways, including decreased HIF-1a mRNA levels, decreased HIF-1a protein synthesis, increased HIF-1a degradation, decreased HIF subunit heterodimerization, decreased HIF binding to DNA, and decreased HIF transcription activity. ${ }^{[27]}$ Many drugs are in clinical trials to treat cancer by inhibition of HIF-1, and they exhibited molecular mechanisms of inhibiting HIF-1 at different steps. ${ }^{[28]}$ For example, the prodrug AFP-464 (Figure 1) is already applied in phase I cancer trials, and it almost blocks HIF-1a protein expression completely and partially inhibits HIF-1a mRNA expression by decreasing both the stability and translation of HIF-1amRNA. ${ }^{[29]}$


AFP-464
Figure 1: Chemical Structure of AFP-464

Earlier, KCN1 (Figure 2) was discovered as a potent HIF-1 inhibitor without any signficant toxicity. ${ }^{[9]}$


Figure 2: Structure and $\mathrm{IC}_{50}$ value of KCN 1

Based on KCN1, a new compound SRIV-64b (Figure 3) was designed and synthesized by our group. Compared to $\mathrm{KCN} 1, \mathrm{IC}_{50}$ of $0.59 \mu \mathrm{M}$ (Figure 2), SRIV-64b was determined to have a lower $\mathrm{IC}_{50}$ value and was considered the most potent HIF-1 inhibitor of this series synthesized previously. ${ }^{[9]}$


Figure 3: Structure and $\mathrm{IC}_{50}$ value of SRIV-64b

### 1.4 HIF-1 Inhibitors Classes

During the past several years, more than 200 compounds have been synthesized by our group. However, only less than half of the compounds in the library can be considered as potent HIF inhibitors. In order to find more patent HIF inhibitors, new scaffolds were needed.

Natural products manassantin A and manassantin B (Figure 4), isolated from Saururus cernuus have been shown to inhibit HIF-1 in vitro at nanomolar concentrations. ${ }^{[30-32]}$ With these natural products and our earlier HIF-1 inhibitors in mind, ${ }^{[9]}$ we were interested in designing "hybrid" compounds as a new scaffold of potential HIF-1 inhibitors. As a result, lead compound 2 (Figure 5) was developed with the goal of achieving manassantin-like activity against HIF, while having a more facile synthetic route than manassantins. Compound 2 was synthesized and shown as a potent HIF inhibitor, and successfully demonstrated HIF inhibition in a luciferase assay in glioblastoma cells. Our goal was to design new analogues by modifying lead compound 2 and to test them for HIF inhibitory activity. Based on substituents at different positions on the phenyl ring, five classes of compounds were designed and synthesized (Figure 6).


Manassantin $\mathrm{A}: \mathrm{R}_{1}=\mathrm{R}_{2}=-\mathrm{OCH}_{3}$ Manassantin $\mathrm{B}: \mathrm{R}_{1}=\mathrm{R}_{2}=-\mathrm{OCH}_{2} \mathrm{O}-$

1

Figure 4: Manassantin A and B


2

$$
I C_{50}=0.58 \mu \mathrm{M}
$$

Figure 5: Lead compound structure and $\mathrm{IC}_{50}$ value
R.


Class III: Meta-phenolic ether benzhydrol analogues


Class II:
Para-phenolic ether benzhydrol analogues
$\checkmark$



Class I:
Ortho-phenolic ether benzhydrol analogues


Class IV:
Ortho-phenylethan ether benzhydrol analogues

Figure 6: Analogues Designed

## 2 CHEMISTRY DESIGN AND EXPERIMENTS

This work includes compounds synthesized by Jalisa Ferguson* and Marquis Griffin*.
12a, 14d, 9d, 9g, 19b, 17e and 22a were synthezied by Dr. Jalisa Holmes Ferguson*.
14a and 9a were synthesized by Marquis Griffin*.

### 2.1 Design

In Class I, we are interested in exploring the effect of having an ortho-substitution on the phenyl ring of region A, while maintaining the methyl-protected catechol structure on the right. In doing so, we expect that the torsional angle defined by

O (hydroxyl)-C-C-C to be slightly perturbed by alkoxy substitution at the ortho position. In this class of compounds, we designed six analogues, and compared their different structures for structure activity relationship study.

In Class II, we are interested in examining the effect of para-subtitutions. In this case, the torsional angle defined by O (hydroxyl)-C-C-C is not expected to be perturbed. However, the various substitutions would allow us to explore the favorable and unfavorable interactions between the binding pocket and the para-substituents of the phenyl ring. In addition, such substitutions may also perturb the electronic properties of the phenyl ring and to some degree the hydrophobicity of the moiety. The various substituents at the para-position may also allow us to explore additional functional group interactions. In Class III, we are interested in examining the effect of substituents at the meta-position while keeping the protected catechol the same on the right side. Similarly, class III can be compared with class II. In Class IV and V, we are interested in changing the protected catechol to protected pyragol, while sampling various substituents on the phenyl ring on the left side. Below are the results of synthesis and biological evaluation.

### 2.2 Chemistry

2.2.1 Class I: Ortho-phenolic ether benzhydrol analogues.

Class I analogues were synthesized in two steps: nucleophilic substitution followed by lithium-halogen exchange with subsequent addition to aldehyde. 2Hydroxybenzaldehyde (3) was reacted with the corresponding bromide to generate
compounds (4a-f). Compounds (6a-f) were generated using a lithium-halogen exchange reaction of bromide of 5 using $n$-butyllithium. The aryllithium generated was used in situ in the subsequent addition reaction with aldehydes 4a-f, yielding final compounds 6a-f.

Scheme 1: Synthesis of ortho-phenolic ether benzhydrol analogues.

$R=$ allyl (4a, 6a), isopropyl (4b, 6b), propyl (4c, 6c), hexyl (4d, 6d), methylcyclohexyl (4e, 6e), and benzyl (4f, 6f).

Reagents and Conditions: (a) $\mathrm{RBr}, \mathrm{K}_{2} \mathrm{CO}_{3}, \mathrm{DMF}, 78{ }^{\circ} \mathrm{C}$, overnight, $97-99 \%$ yield. (b) $n$ - BuLi , THF, $-78{ }^{\circ} \mathrm{C}, 1.5$ hours; (c) 4, 2 hours, $55-69 \%$ yield.
2.2.2 Class II: Para-phenolic ether benzhydrol analogues

Synthesis of Class II analogues was achieved in either 2 or 4 steps. In Scheme 2, the intermediates 8a-c were prepared by reaction between 4-hydroxybenzaldehyde (7)
and bromide compounds. Next, lithium-halogen exchange of 5 using $n$-BuLi, followed by addition of aldehyde 8, afforded the final compounds 9a-c. In Scheme 3, for analogue 9g, di-tert-butyl dicarbonate was reacted with 2-(piperidin-4-yl)ethan-1-ol 10b under aqueous conditions to yield intermediate 11b. Tosylation of 11b with 4methylbenzenesulfonyl chloride under mild conditions yielded 12*. Next, the intermediates $14 \mathbf{a}^{\#}$-d were prepared by reaction between 4-bromophenol (13) and compounds $\mathbf{1 2}^{*}$ or bromide compounds. Lastly, lithium-halogen exchange of 14 using $n$-BuLi, followed by addition of 3,4-dimethoxybenzaldehyde, afforded the final compounds $9 \mathrm{~d}^{\text {\# }}-\mathbf{g}$ * .

Scheme 2: Synthesis of Para-phenolic ether benzhydrol analogues 9a-9c.

$R=$ isobutyl (8a, 9a), hexyl (8b, 9b), methylcyclohexyl (8c, 9c).

Reagents and Conditions: (a) $\mathrm{K}_{2} \mathrm{CO}_{3}, \mathrm{RBr}, \mathrm{DMF}, 78^{\circ} \mathrm{C}$, overnight, $95-97 \%$ yield. (b) $n$-BuLi, THF, $-78^{\circ} \mathrm{C}, 1.5$ hours; (c) 8a-c, 2 hours, $54-71 \%$ yield.

Scheme 3: Synthesis of Para-phenolic ether benzhydrol analogues $\mathbf{9 d} \mathbf{d}^{\boldsymbol{\#}} \mathbf{- 9} \mathrm{g}^{\text {* }}$


$\mathrm{n}=2$ (10b, 11b, 12*)
R=allyl (14a\#, 9d"), isopropyl (14b, 9e), propyl (14c, 9f), (1-(tert-butoxycarbonyl)piperidin-4-yl)ethyl (14d*, 9g*).

Reagents and Conditions: (a) $\mathrm{Boc}_{2} \mathrm{O}, \mathrm{H}_{2} \mathrm{O}$, room temperature, overnight, $96 \%$ yield. (b) TsCl , $\mathrm{NEt}_{3}$, THF, room temperature, overnight, $86 \%$ yield. (c) $\mathrm{K}_{2} \mathrm{CO}_{3}, 12$; RBr , DMF, $90^{\circ} \mathrm{C}, 5$ hours, 95-97\% yield. (d) n-BuLi, THF, $-78^{\circ} \mathrm{C}$, 1.5 hours; (e) 3,4-dimethoxybenzaldehyde, 2 hours, 54 $71 \%$ yield.
2.2.3 Class III: Meta-phenolic ether benzhydrol analogues

Class III analogues were synthesized as described in Schemes 4 and 5. In Scheme 4, the analogues were synthesized by alkylation of 3-hydroxybenzaldehyde (15) using the corresponding bromide to give intermediates 16a-b. Next, lithiumhalogen exchange of 5 using $n$-BuLi, followed by addition of aldehyde 16, afforded the final compounds 17a-b. In Scheme 5, for analogues 17c-e, the intermediates 19a-c
were prepared by reaction between 3-bromophenol (18) and bromide compounds. Next, analogues 17c-e were synthesized by lithium-halogen exchange as described above.

Scheme 4: Synthesis of Meta-phenolic ether benzhydrol analogues 17a-17c.

$R=$ isobutyl (16a, 17a), isopropyl (16b, 17b), propyl (16c, 17c).

Reagents and Conditions: (a) $\mathrm{K}_{2} \mathrm{CO}_{3}$, DMF, $78{ }^{\circ} \mathrm{C}$, overnight, $92-96 \%$ yield. (b) $n$-BuLi, THF, $78^{\circ} \mathrm{C}, 1.5$ hours; (c) $\mathbf{1 6 a - c}, 2$ hours, $67-78 \%$ yield.

Scheme 5: Synthesis of Meta-phenolic ether benzhydrol analogues 17d and 17e*.

$R=\operatorname{allyl}(19 a, 17 d)$, hexyl (19b, 17e)*.

Reagents and Conditions: (a) $\mathrm{K}_{2} \mathrm{CO}_{3}$, DMF, $90^{\circ} \mathrm{C}$, 5 hours, $93-97 \%$ yield. (b) $n$-BuLi, THF, -78 ${ }^{\circ} \mathrm{C}$, 1.5 hours; (c) 3,4-dimethoxybenzaldehyde, 2 hours, $63-69 \%$ yield.
2.2.4 Class IV: Ortho-benzyl ether benzhydrol analogues.

Synthesis of Class IV analogues was achieved in either 2 or 3 steps. For analogues 22b and 22c, intermediates 11a-b were prepared by the same method as described above (Scheme 3). Benzylic ether intermediates 21 were synthesized by $O$ alkylation with sodium hydride as the base and 2-bromobenzyl bromide (20). Final analogues 22a-c were synthesized using lithium-halogen exchange to generate the arylitihium, followed by addition to trimethoxyphenyl aldehyde.

Scheme 6: Synthesis of ortho-benzyl ether benzhydrol analogues.


10
11


20



21




22

R
$n=1(10 a, 11 a), 2(10 b, 11 b)$
R=phenyl (21a, 22a*), (1-(tert-butoxycarbonyl)piperidin-4-yl)methyl (21b, 22b), (1-(tert-butoxycarbonyl)piperidin-4-yl)ethyl (21c, 22c).

Reagents and Conditions: (a) $\mathrm{Boc}_{2} \mathrm{O}, \mathrm{H}_{2} \mathrm{O}$, room temperature, overnight, $98-99 \%$ yield. (b) ROH; 11a-11b, NaH, DMF, $0^{\circ} \mathrm{C}$-rt, overnight, $96-98 \%$ yield. (c) $n$-BuLi, THF, $-78{ }^{\circ} \mathrm{C}, 1.5$ hours; (d) 3,4,5-trimethoxybenzaldehyde, 2 hours, 59-71\% yield.
2.2.5 Class V: Tri-substituted phenolic ether benzhydrol analogues.

Class V analogues were synthesized in two steps. 4-Bromo-2-methylphenol (23) was reacted with bromide compounds to form the ether intermediates 24a-f via nucleophilic substitution. Next, analogues 25a-f were synthesized using lithiumhalogen exchange and addition to trimethoxyphenyl aldehyde.

Scheme 7: Synthesis of tri-substituted phenolic ether benzhydrol analogues.

$R=$ allyl (24a, 25a), isobutyl (24b, 25b), isopropyl (24c, 25c), propyl (24d, 25c), hexyl (24e, 25e), methylcyclohexyl (24f, 25f).

Reagents and Conditions: (a) RBr, $\mathrm{K}_{2} \mathrm{CO}_{3}, \mathrm{DMF}, 78^{\circ} \mathrm{C}$, overnight, $95 \%-99 \%$ yield. (b) $n$-BuLi, THF, $-78{ }^{\circ} \mathrm{C}, 1.5$ hours; (c) 3,4,5-trimethoxybenzaldehyde, 2 hours, $59 \%-65 \%$ yield.

## 3 BIOLOGY RESULT

All analogues synthesized herein were subsequently tested for their HIF-1 inhibition activity in a luciferase reporter assay performed by our collaborators at Emory University's Winship Cancer Institute, Dr. Erwin Van Meir. This luciferase reporter assay is a tool used to measure expression of gene expression activated by HIF. In this study, the $\mathrm{IC}_{50}$ value of each analogue was tested using the HRE of VEGF gene with a luciferase reporter gene. The firefly luciferase reporter gene were encoded by the HIFresponsive luciferase under the control of a promoter and tandem repeats of the hypoxia transcriptional response element (HRE). ${ }^{[33]} \mathrm{IC}_{50}$ values over $10 \mu \mathrm{M}$ for the analogues were considered not potent enough for further evaluation. Therefore, analogues with less than $1 \mu \mathrm{M}$ of $\mathrm{IC}_{50}$ were considered to have sufficient HIF inhibitor activity for further pursual.

### 3.1 Results for Class I Analogues

Compounds $\mathbf{6 a}, \mathbf{6 b}, \mathbf{6 c}, \mathbf{6 d}, \mathbf{6 e}$, and $\mathbf{6 f}$ in class I were evaluated for their abilities to inhibit the HIF-mediated transcription activity used in the luciferase assay as described above. It was found that the best compound showed $\mathrm{IC}_{50}$ of about $3.8 \mu \mathrm{M}$ (6c), while several had $\mathrm{IC}_{50}$ values of over $10 \mu \mathrm{M}$. Compared to compound $\mathbf{2}$, with $\mathrm{IC}_{50}$ of $0.58 \mu \mathrm{M}$, the ortho-modified $\mathbf{6 c}$ was six-fold less potent. Such results suggest that modifications at the ortho-position likely twist the molecule due to the disturbance of $\pi$ electron overlap and the O (hydroxyl)-C-C-C torsional angle in such a way that disfavors the interactions with the intended target (Table 1).

|  |  |  |
| :---: | :---: | :---: |
| Compounds | R | $\mathrm{IC}_{50}(\mu \mathrm{M})$ |
| 6a |  | $>10$ |
| 6b |  | >10 |
| 6c | $\underbrace{\xi}$ | 3.8 |
| 6d | ~ | >10 |
| 6 e |  | 6.6 |
| 6 f |  | >10 |

Table 1: Class I Analogues

### 3.2 Results for Class II Analogues

For analogues in class $I I, 9 \mathbf{g}^{*}$ have $\mathrm{IC}_{50}$ value of less $1 \mu \mathrm{M}$, and the other analogues (9a, 9b, 9c) have a lower $\mathrm{IC}_{50}$ value than most compounds in class I . When comparing class II and class I, the result showed that para-subtittuions exhibited better
inhibition of HIF-1 activity than those analogues with ortho-substitutions. Specifically, $\mathbf{9} \mathbf{g}^{*}$ with a Boc-protected piperidine substitution on the left side inhibited completely HIF-1 activity most effectively with an $\mathrm{IC}_{50}$ of $0.89 \mu \mathrm{M}$. (Table 2).



Table 2: Class II Analogues

### 3.3 Results for Class III Analogues

Reviewing Table 3 below and Table 2 above, the analogues of class III were seen to show better $\mathrm{IC}_{50}$ values than $\mathbf{9 b} \mathbf{~} \mathbf{9 c}$ and $\mathbf{9 d ^ { \# }}$ in class II. Although there are only minor differences in the structure, meta-substituted compounds exhibited lower $\mathrm{IC}_{50}$ values than para-substituted compounds, especially compound $\mathbf{1 7 d}$. The $\mathrm{IC}_{50}$ value of $\mathbf{1 7 d}$ is $0.5 \mu \mathrm{M}$, which is lower than the $\mathrm{IC}_{50}$ value of compound $\mathbf{2}$, but the other compounds were still three- to five-fold less potent compared to compound 2. Temporarily, class III and class II without 9g* and 17d were not considered as potent HIF inhibitors due to the low biology activities.


| Compounds | R | $\mathrm{IC}_{50}(\mu \mathrm{M})$ |
| :---: | :---: | :---: |
| 17a |  | 2.12 |
| 17b |  | >10 |
| 17c | $\sim_{2}$ | 2.5 |
| 17d |  | 0.5 |
| 17e* |  | 1.65 |

Table 3: Class III Analogues.

### 3.4 Results for Class IV Analogues

Class IV analogues shown in Table 4 can best be compared to Class I analogues except there are two major differences: 1) the use of a protected pyragollol on the right side instead of the protected catechol, and 2) a methylene that separates the oxygen atom of the ether and the phenyl ring on the left side. The $\mathrm{IC}_{50}$ values of three compounds in class IV were determined and compared, and it was found that 22c showed the best inhibitory activity with an $\mathrm{IC}_{50}$ value of $2.2 \mu \mathrm{M}$ (Table 4). In contrast, 22a* was seen to give $\mathrm{IC}_{50}$ over $10 \mu \mathrm{M}$. Silimar to class I , the resulting value of this class indicated that ortho-modications on the left might twist molecules into a less suitable conformation, unable to inhibit HIF-1 activity.


| Compounds | R | $\mathrm{IC}_{50}(\mu \mathrm{M})$ |
| :---: | :---: | :---: |
| 22a* $^{*}$ |  | $>10$ |
| 22b |  |  |
| 22c |  |  |

Table 4: Class IV Analogues

### 3.5 Results for Class V Analogues

The analogues from class V have the best $\mathrm{IC}_{50}$ values of all these five classes, listed in Table 5. Notably, 25b has better $\mathrm{IC}_{50}$ value than the lead, 2, at $0.58 \mu \mathrm{M} . \mathbf{2 5 d}$ has the same $\mathrm{IC}_{50}$ value as compound $\mathbf{2}$; thus it can be considered as the same potent compound. This class of compounds, featuring an additional methyl group on the left ring and a protected pyrogallol ion the right, yields the most active class of compounds, and are likely the best suited for interaction with the HIF-1 target. Following class V , such compounds can be considered as potent inhibitors and evaluated further (Table 5).


| Compounds | R | $\mathrm{IC}_{50}(\mu \mathrm{M})$ |
| :---: | :---: | :---: |
| 25a | 25b |  |
| 25c |  | 0.62 |
|  |  | 0.4 |
| 25d |  | 0.75 |


| 25 e | 0.7 |  |
| :--- | :--- | :--- |
| 25 m |  | 1.2 |

Table 5: Class V Analogues.

## 4 CONCLUSIONS AND FUTURE WORK

### 4.1 Structure Activity Relationship Study

In conclusion, 27 analogs were synthesized within five different classes. According to the HIF inhibition results from the luciferase assay described above, a structure-activity relationship (SAR) was developed and analyzed. For the right side, protected pyragollol or protected catechol was used on the phenyl ring, and both of them are important to the activity. The different positions substituted on left phenyl ring were compared according to results from the luciferase assay, and the additional methyl group in Class V gave the best results. Otherwise, an important factor in improving inhibitory activity is the substituted para-position of Class II. Compared with the ortho-substituted analogues were found to be the least potent inhibitors of the three different types of compounds. The results suggest that ortho-substituted compounds may twist the molecule unfavorably, resulting in a decrease of activity. However, for the compounds with a Boc-protected piperidine, they exhibited better activity than there comparable analogues, no matter the substitution (Class II and IV).

### 4.2 Future Work

For future work, more tri-substituted analogs with methyl group of Class V will be synthesized by same method. In this study, class V compounds, especially 25b were considered to be potent inhibitors. In the future, other similar compounds will be pursued, such as compounds with a methoxy group on 3'-position of the left phenyl ring.

## 5 EXPERIMENTAL

In this study, all starting materials were purchased from Sigma-Aldrich or Oakwood chemicals without further purification. All intermediates from this study were synthesized by according to literature reports. ${ }^{1} \mathrm{H}$ amd ${ }^{13} \mathrm{C}$ NMR spectra were recorded at 400 MHz and 100 MHz , respectively, on a Bruker Avance 400 NMR spectrometer. The solvent for dissolving all compounds in this study to record NMR was $\mathrm{CDCl}_{3}$. All mass spectra analyses were obtained by the mass spectratrometry facilities at Georgia State University.

General Procedure for the Synthesis of 4a to $\mathbf{4 f}$. One equivalent of $\mathbf{3}$ was dissolved in DMF. One equivalent of bromide and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$ were added. The solution was stirred overnight at $78^{\circ} \mathrm{C}$, and then ethyl acetate was added. The combined organic solution was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The crude residue was purified by column chromatography with $100: 1$ hexane-ethyl acetate as the leuent. $\mathbf{4 a}-\mathbf{4 f}$ were synthesized by according to literature reports. ${ }^{[34-38]}$

2-(Allyloxy)benzaldehyde (4a). Yield: 98\%, ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 10.56(\mathrm{~s}, 1 \mathrm{H}), 7.87(\mathrm{~d}$, $1 \mathrm{H}), 7.69(\mathrm{t}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.07-6.99(\mathrm{~m}, 2 \mathrm{H}), 6.13-6.06(\mathrm{~m}, 1 \mathrm{H}), 5.50(\mathrm{~d}, J=16 \mathrm{~Hz}, 1 \mathrm{H}), 5.39$ $(\mathrm{d}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 4.68 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 2 \mathrm{H})$.

2-Isopropoxybenzaldehyde (4b). Yield: $99 \%$, ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 10.52(\mathrm{~s}, 1 \mathrm{H}), 7.84(\mathrm{~d}$, $J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.55-7.51(\mathrm{~m}, 1 \mathrm{H}), 7.02-6.99(\mathrm{~m}, 2 \mathrm{H}), 4.71(\mathrm{t}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 1.42 \mathrm{ppm}(\mathrm{d}, J=$ $4 \mathrm{~Hz}, 6 \mathrm{H}$ ).

2-Propoxybenzaldehyde (4c). Yield: 97\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.52(\mathrm{~s}, 1 \mathrm{H}), 7.82-7.80$ $(\mathrm{m}, 1 \mathrm{H}), 7.49(\mathrm{t}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.99-6.94(\mathrm{~m}, 2 \mathrm{H}), 4.02(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 1.88-1.83(\mathrm{~m}, 2 \mathrm{H})$, 1.09-1.07 ppm (m, 3H).

2-(Hexyloxy)benzaldehyde (4d). Yield: 98\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.51(\mathrm{~d}, \mathrm{~J}=4 \mathrm{~Hz}, 1 \mathrm{H})$, 7.83-7.81 (m, 1H), 7.54-7.49 (m, 1H), 7.01-6.96 (m, 2H), 4.08-4.04 (m, 2H), 1.86-1.81 (m, 2H), $1.49(\mathrm{~s}, 2 \mathrm{H}), 1.26(\mathrm{~s}, 4 \mathrm{H}), 0.92 \mathrm{ppm}(\mathrm{t}, J=4 \mathrm{~Hz}, 3 \mathrm{H})$.
2-(Cyclohexylmethoxy)benzaldehyde (4e). Yield: 97\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.52(\mathrm{~d}, \mathrm{~J}$ $=4 \mathrm{~Hz}, 1 \mathrm{H}), 7.83-7.81(\mathrm{~m}, 1 \mathrm{H}), 7.54-7.49(\mathrm{~m}, 1 \mathrm{H}), 7.01-6.95(\mathrm{~m}, 2 \mathrm{H}), 4.08-4.04(\mathrm{~m}, 2 \mathrm{H}), 1.86-$ $1.81(\mathrm{~m}, 2 \mathrm{H}), 1.49(\mathrm{~s}, 2 \mathrm{H}), 1.35(\mathrm{~s}, 5 \mathrm{H}), 0.91 \mathrm{ppm}(\mathrm{t}, J=4 \mathrm{~Hz}, 3 \mathrm{H})$.
2-(Benzyloxy)benzaldehyde (4f). Yield: 99\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.59$ (s, 1H), 7.90$7.88(\mathrm{~m}, 1 \mathrm{H}), 7.55(\mathrm{t}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.48-7.38(\mathrm{~m}, 5 \mathrm{H}), 7.08(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 5.22 \mathrm{ppm}(\mathrm{s}, 2 \mathrm{H})$.

General Procedure for the Synthesis of 6a to $\mathbf{6 f}$. One equivalent of $\mathbf{5}$ was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, and then 1.1 equivalent of $n$-butyllithium was added. The reaction was stirred for 1.5 hour, followed by the dropwise addition of 1 equivalent of $\mathbf{4}$ in dry THF under Ar. The reaction was stirred for 2 hours at $-78{ }^{0} \mathrm{C}$. Then ethyl acetate was added followed by $\mathrm{NH}_{4} \mathrm{Cl}$. The mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-acetone as the eluent.
(2-(Allyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (6a). Yield: 56\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.29-7.26(\mathrm{~m}, 3 \mathrm{H}), 7.00(\mathrm{t}, J=20 \mathrm{~Hz}, 1 \mathrm{H}), 6.91-6.89(\mathrm{~m}, 2 \mathrm{H}), 6.83(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H})$, 6.08-5.95 (m, 2H), 5.39-5.26 (m, 2H), 4.58-4.56 (m, 2H), 3.87 (d, $J=4 \mathrm{~Hz}, 6 \mathrm{H}), 3.03 \mathrm{ppm}(\mathrm{d}, J$ $=12 \mathrm{~Hz}, 1 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 155.64,148.76,148.13,136.00,132.95,132.49,128.59$,
$127.70,121.02,118.79,117.64,111.97,110.81,110.00,72.02,68.92,55.90,55.82 \mathrm{ppm}$. HRMS (ESI) $m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{20} \mathrm{O}_{4}+\mathrm{Na}^{+}: 323.1259$, found 323.1261.
(2-Isopropoxyphenyl)(3,4-dimethoxyphenyl)methanol (6b). Yield: 62\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.28-7.23 (m, 2H), 7.02 ( $\left.\mathrm{s}, 1 \mathrm{H}\right), ~ 6.93-6.81(\mathrm{~m}, 4 \mathrm{H}), 5.97(\mathrm{~s}, 1 \mathrm{H}), 4.65-4.59(\mathrm{~m}, 1 \mathrm{H})$, $3.88(\mathrm{~d}, J=4 \mathrm{~Hz}, 6 \mathrm{H}), 3.28(\mathrm{~s}, 1 \mathrm{H}), 1.31(\mathrm{~d}, J=4 \mathrm{~Hz}, 3 \mathrm{H}), 1.27 \mathrm{ppm}(\mathrm{d}, J=4 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 154.93,148.70,148.04,136.23,132.90,128.51,128.00,120.44,118.83,112.68$, $110.74,109.99,72.65,70.01,55.92,55.81,22.16,21.98 \mathrm{ppm}$. HRMS (ESI) $m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}-\mathrm{OH}^{-}: 285.1485$, found: 285.1473 .
(2-Propoxyphenyl)(3,4-dimethoxyphenyl)methanol (6c). Yield: 69\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28-7.23(\mathrm{~m}, 2 \mathrm{H}), 6.97-6.88(\mathrm{~m}, 4 \mathrm{H}), 6.84(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 6.03(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H})$, 3.99-3.93 (m, 2H), $3.89(\mathrm{~d}, J=4 \mathrm{~Hz}, 6 \mathrm{H}), 3.14(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 1.81-1.79(\mathrm{~m}, 2 \mathrm{H}), 1.01 \mathrm{ppm}(\mathrm{t}$, $J=12 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.21,148.76,148.11,136.02,132.11,128.64,127.73$, $120.60,118.83,111.45,110.79,109.98,72.37,69.62,55.91,55.81,22.63,10.64 \mathrm{ppm}$. HRMS (ESI ${ }^{+}$) $m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}+\mathrm{Na}^{+}: 325.1416$, found 325.1426 .
(2-(Hexyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (6d). Yield: 61\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.28-7.22 (m, 3H), 6.97-6.82 (m, 4H), 4.02-3.97 (m, 2H), 3.93-3.89 (m, 6H), 3.17 $(\mathrm{d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.78-1.74(\mathrm{~m}, 2 \mathrm{H}), 1.33-1.26(\mathrm{~m}, 6 \mathrm{H}), 0.91 \mathrm{ppm}(\mathrm{t}, J=16 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.24,148.77,148.11,136.04,132.07,128.64,127.76,120.57,118.82,111.44$, $110.75,109.96,72.44,68.06,55.88,55.79,31.54,29.26,25.76,22.56,14.03 \mathrm{ppm}$. HRMS $\left(\mathrm{ESI}^{+}\right) m / z$ calculated for $\mathrm{C}_{21} \mathrm{H}_{28} \mathrm{O}_{4}+\mathrm{Na}^{+}: 367.1885$, found 367.1877.
(2-(Cyclohexylmethoxy)phenyl)(3,4-dimethoxyphenyl)methanol (6e). Yield: 55\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.28-7.19 (m, 4H), 6.99-6.79 (m, 3H), $6.00(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.87-3.85$ $(\mathrm{m}, 4 \mathrm{H}), 3.79-3.75(\mathrm{~m}, 2 \mathrm{H}), 3.14(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.75-1.71(\mathrm{~m}, 6 \mathrm{H}), 1.26 \mathrm{ppm}(\mathrm{t}, J=16 \mathrm{~Hz}$, $7 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.31,148.79,148.12,136.03,132.05,128.66,127.82,120.49$, $118.82,111.34,110.83,109.97,73.47,72.42,55.93,55.80,37.77,29.90,29.86,26.40,26.32$, 25.82 ppm . HRMS (ESI ${ }^{+}$) $\mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{22} \mathrm{H}_{28} \mathrm{O}_{4}+\mathrm{Na}^{+}: 379.1885$, found 379.1886 .
(2-(Benzyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (6f). Yield: 64\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.33-7.26 (m, 8H), 7.03-6.97 (m, 3H), 6.84-6.80 (m, 2 H$), 6.05(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H})$, $5.06(\mathrm{~s}, 2 \mathrm{H}), 3.88(\mathrm{~s}, 3 \mathrm{H}), 3.77(\mathrm{~s}, 3 \mathrm{H}), 2.96(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.59 \mathrm{ppm}(\mathrm{s}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 155.76,148.75,148.11,136.58,136.14,132.45,128.67,128.58,128.09,127.71$,
127.48, 121.09, 118.73, 111.95, 110.84, 110.05, 72.29, 70.18, 55.94, 55.71 ppm. HRMS (ESI ${ }^{+}$) $m / z$ calculated for $\mathrm{C}_{22} \mathrm{H}_{22} \mathrm{O}_{4}+\mathrm{Na}^{+}: 373.1416$, found 373.1398.

General Procedure for the Synthesis of 8a to 8c. One equivalent of $\mathbf{7}$ was dissolved in DMF. One equivalent of bromide and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$ were added. The solution was stirred overnight at $78{ }^{\circ} \mathrm{C}$, and then ethyl acetate was added. The combined organic solution was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The crude residue was purified by column chromatography with $100: 1$ hexane-ethyl acetate as the eluent. 8a-8c were synthesized by according to literature reports. ${ }^{[39-40]}$
4-Isobutoxybenzaldehyde (8a). Yield: $95 \% .{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 9.89$ (s, 1H), 7.84 (d, $J=$ $8 \mathrm{~Hz}, 2 \mathrm{H}), 7.01(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 3.82(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.17-2.10(\mathrm{~m}, 1 \mathrm{H}), 1.05 \mathrm{ppm}(\mathrm{t}, J=$ $8 \mathrm{~Hz}, 6 \mathrm{H}$ ).

4-(Hexyloxy)benzaldehyde (8b). Yield: $97 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 9.89(\mathrm{~s}, 1 \mathrm{H}), 7.84(\mathrm{~d}, J$ $=8 \mathrm{~Hz}, 2 \mathrm{H}), 7.01(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 4.06(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 1.86-1.79(\mathrm{~m}, 2 \mathrm{H}), 1.51-1.36(\mathrm{~m}, 6 \mathrm{H})$, $0.95 \mathrm{ppm}(\mathrm{t}, J=8 \mathrm{~Hz}, 3 \mathrm{H})$.
4-(CyclohexyImethoxy)benzaldehyde (8c). Yield: 97\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 9.90$ (s, $1 \mathrm{H}), 7.84(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 7.01(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 3.86(\mathrm{~d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 1.91-1.72(\mathrm{~m}, 6 \mathrm{H})$, $1.35-1.21(\mathrm{~m}, 3 \mathrm{H}), 1.13-1.07 \mathrm{ppm}(\mathrm{m}, 2 \mathrm{H})$.

General Procedure for the Synthesis of 9a to 9c. One equivalent of $\mathbf{5}$ was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, and 1.1 equivalent of $n$-butyllithium was added. The reaction was stirred for 1.5 hour, followed by the dropwise addition of 1 equivalent of $\mathbf{8}$ in dry THF under Ar. The reaction was stirred for another 2 hours at $-78{ }^{\circ} \mathrm{C}$. Then ethyl acetate was added followed by $\mathrm{NH}_{4} \mathrm{Cl}$. The mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-acetone as the eluent.
(4-Isobutoxyphenyl)(3,4-dimethoxyphenyl)methanol (9a). Yield: $66 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.27(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.94(\mathrm{~s}, 1 \mathrm{H}), 6.93-6.84(\mathrm{~m}, 4 \mathrm{H}), 5.75(\mathrm{~s}, 1 \mathrm{H}), 3.86(\mathrm{~d}, J=$ $4 \mathrm{~Hz}, 6 \mathrm{H}), 3.72(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.35(\mathrm{~s}, 1 \mathrm{H}), 2.12-2.06(\mathrm{~m}, 1 \mathrm{H}), 1.04 \mathrm{ppm}(\mathrm{d}, J=4 \mathrm{~Hz}, 6 \mathrm{H})$.
${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 158.72,148.99,148.33,136.85,136.02,127.76,118.79,114.45,110.92$, 109.70, 75.56, 74.48, 55.92, 55.85, 28.28, 19.28 ppm . HRMS (ESI ${ }^{+} \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{19} \mathrm{H}_{24} \mathrm{O}_{4}-\mathrm{OH}^{-}: 299.1642$, found 299.1644.
(4-(Hexyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (9b). Yield: 71\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.30-7.27(\mathrm{~m}, 3 \mathrm{H}), 6.95-6.82(\mathrm{~m}, 4 \mathrm{H}), 5.79(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.96(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H})$, $3.88(\mathrm{~d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}), 2.15(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.80-1.75(\mathrm{~m}, 2 \mathrm{H}), 1.49(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 1.36-1.33$ $(\mathrm{m}, 4 \mathrm{H}), 0.94-0.91 \mathrm{ppm}(\mathrm{m}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 158.63,149.01,148.36,136.79$, $135.99,127.77,118.78,114.44,110.91,109.68,75.60,68.05,55.93,55.86,31.59,29.25,25.73$, 22.61, 14.04 ppm. HRMS $\left(\mathrm{ESI}^{+}\right) \mathrm{m} / z$ calculated for $\mathrm{C}_{21} \mathrm{H}_{28} \mathrm{O}_{4}-\mathrm{OH}^{-}$: 327.1955, found 327.1941 .
(4-(Cyclohexylmethoxy)phenyl)(3,4-dimethoxyphenyl)methanol (9c). Yield: 54\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28(\mathrm{~s}, 2 \mathrm{H}), 6.95-6.83(\mathrm{~m}, 5 \mathrm{H}), 5.78(\mathrm{~s}, 1 \mathrm{H}), 3.87(\mathrm{~d}, J=4 \mathrm{~Hz}, 6 \mathrm{H}), 3.76$ $(\mathrm{d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.14(\mathrm{~s}, 1 \mathrm{H}), 1.89-1.70(\mathrm{~m}, 6 \mathrm{H}), 1.33-1.20(\mathrm{~m}, 3 \mathrm{H}), 1.10-1.01 \mathrm{ppm}(\mathrm{m}, 2 \mathrm{H})$. ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 158.77,148.99,148.33,136.85,135.97,127.76,118.78,114.43,110.92$, 109.70, 75.57, 73.55, 55.93, 55.85, 37.70, 29.93, 26.54, $25.82 \mathrm{ppm} . \mathrm{HRMS}_{\left(\mathrm{ESI}^{+}\right) \mathrm{m} / \mathrm{z}}$ calculated for $\mathrm{C}_{22} \mathrm{H}_{28} \mathrm{O}_{4}-\mathrm{OH}^{-}: 339.1955$, found 339.1938.

General Procedure for the Synthesis of 11a and 11b. One equivalent of 10 was dissolved in $\mathrm{H}_{2} \mathrm{O}$. Then 1.1 equivalents of $\mathrm{Boc}_{2} \mathrm{O}$ was added in solution. The mixture was stirred overnight at room temperature, followed by the addition of ethyl acetate. The mixture was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. 11a-11b were synthesized by according to literature reports. ${ }^{[41-42]}$
Tert-butyl 4-(hydroxymethyl)piperidine-1-carboxylate (11a). Yield: 99\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 4.15(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 3.52(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.75-2.69(\mathrm{~m}, 2 \mathrm{H}), 1.75-1.63(\mathrm{~m}$, $4 \mathrm{H}), 1.49$ ( $\mathrm{s}, 9 \mathrm{H}$ ), $1.20 \mathrm{ppm}(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H})$.
Tert-butyl 4-(2-hydroxyethyl)piperidine-1-carboxylate (11b). Yield: 96\%. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 3.89(\mathrm{~m}, 2 \mathrm{H}), 3.51-3.48(\mathrm{~m}, 2 \mathrm{H}), 3.16-3.14(\mathrm{~m}, 1 \mathrm{H}), 2.54(\mathrm{~m}, 2 \mathrm{H}), 1.54-$ $1.46(\mathrm{~m}, 3 \mathrm{H}), 1.38-1.34(\mathrm{~m}, 4 \mathrm{H}), 1.30(\mathrm{~s}, 9 \mathrm{H}) 1.00-0.94(\mathrm{~m}, 2 \mathrm{H}) \mathrm{ppm}$.

General Procedure for the Synthesis of 12*. One equivalent of $\mathbf{1 1 b}$ was dissolved in THF. Then 1.1 equivalents of TsCl and 1.1 equivalents of $\mathrm{NEt}_{3}$ were added in solution. The mixture was stirred overnight at room temperature, followed by the addition of ethyl acetate. The solution was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated.

Tert-butyl 4-(2-(tosyloxy)ethyl)piperidine-1-carboxylate (12*). Yield: $86 \%$. ${ }^{1}$ H NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.71(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 7.28(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 4.05-3.98(\mathrm{~m}, 4 \mathrm{H}), 2.54(\mathrm{~m}, 2 \mathrm{H}), 2.37$ $(\mathrm{s}, 3 \mathrm{H}), 1.51-1.44(\mathrm{~m}, 5 \mathrm{H}), 1.38(\mathrm{~s}, 9 \mathrm{H}) 0.99-0.94(\mathrm{~m}, 2 \mathrm{H}) \mathrm{ppm} .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 170.9$, $154.6,144.8,132.9,129.8,127.8,79.2,67.9,60.2,43.5,35.1,32.1,31.5,28.3,21.5,20.9 \mathrm{ppm}$.

General Procedure for the Synthesis of $14 \mathbf{a}^{\#}$ to $\mathbf{1 4 d}^{*}$. One equivalent of $\mathbf{1 3}$ was dissolved in DMF. To this solution, one equivalent of bromide or $\mathbf{1 2}$ and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$ were added. The solution was stirred 5 hours at $90{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate. The combined organic solution was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The crude residue was purified by column chromatography with 100:1 hexane-ethyl acetate as the leuent. $\mathbf{1 4 a}^{\#} \mathbf{- 1 4 d}{ }^{*}$ were synthesized by according to literature reports. ${ }^{[43-44]}$

1-(Allyloxy)-4-bromobenzene (14a ${ }^{\#}$ ). Yield: $42 \% .{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.37$ (d, $J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.80(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.08-5.98(\mathrm{~m}, 1 \mathrm{H}), 5.40(\mathrm{~d}, J=16 \mathrm{~Hz}, 2 \mathrm{H}), 5.30(\mathrm{~d}, J=$ $12 \mathrm{~Hz}, 2 \mathrm{H}), 4.51(\mathrm{~s}, 2 \mathrm{H}) \mathrm{ppm} .{ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 157.7,132.8,132.2,117.9,116.5$, 113.0, 69.0 ppm .

1-Bromo-4-isopropoxybenzene (14b). Yield: $96 \% .{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.47-7.43 (m, $2 \mathrm{H}), 6.83-6.79(\mathrm{~m}, 2 \mathrm{H}), 4.52-4.48(\mathrm{~m}, 1 \mathrm{H}), 1.33 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 6 \mathrm{H})$.
1-Bromo-4-propoxybenzene (14c). Yield: 98\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ 7.39-7.36 (m, 2H), 6.82-6.79 (m, 2H), $3.90(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 1.86-1.78(\mathrm{~m}, 2 \mathrm{H}), 1.05(\mathrm{t}, J=16 \mathrm{~Hz}, 3 \mathrm{H}) \mathrm{ppm}$.

Tert-butyl 4-(2-(4-bromophenoxy)ethyl)piperidine-1-carboxylate (14d*). Yield: 85\%. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.34(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.74(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 4.07(\mathrm{~m}, 2 \mathrm{H}), 3.95$ $(\mathrm{t}, J=7 \mathrm{~Hz}, 2 \mathrm{H}), 2.72-2.66(\mathrm{~m}, 2 \mathrm{H}), 1.70-1.67(\mathrm{~m}, 5 \mathrm{H}), 1.44(\mathrm{~s}, 9 \mathrm{H}), 1.18-1.13(\mathrm{~m}, 2 \mathrm{H}) \mathrm{ppm}$.
${ }^{13} \mathrm{C}$ NMR (100 MHz, $\mathrm{CDCl}_{3}$ ): $\delta 158.0,156.1,154.9,132.2,132.1,117.3,116.2,79.4,65.6,44.0$, 35.6, 32.9, 32.0, 28.4 ppm .

General Procedure for the Synthesis of $\mathbf{9} d^{\#}$ to $\mathbf{9} \mathbf{g}^{*}$. One equivalent of $\mathbf{1 4}$ was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, and 1.1 equivalent of $n$-butyllithium was added. The reaction was stirred 1.5 hour, followed by the dropwise addition of 1 equivalent of 3,4-dimethoxybenzaldehyde in dry THF under Ar. The reaction was stirred for another 2 hours at $-78{ }^{0} \mathrm{C}$. Then ethyl acetate was added followed by $\mathrm{NH}_{4} \mathrm{Cl}$. The mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-actone as the leuent.
(4-(Allyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (9d"). Yield: 20\%. ${ }^{1} \mathrm{H}$ NMR (400 $\left.\mathrm{MHz}, \mathrm{CDCl}_{3}\right): \delta 7.27(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.91-6.81(\mathrm{~m}, 5 \mathrm{H}), 6.09-5.99(\mathrm{~m}, 1 \mathrm{H}), 5.76(\mathrm{~s}, 1 \mathrm{H}), 5.40$ $(\mathrm{d}, J=16 \mathrm{~Hz}, 2 \mathrm{H}), 5.27(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 4.52(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 3.86(\mathrm{~s}, 3 \mathrm{H}), 3.84(\mathrm{~s}, 3 \mathrm{H}), 2.13$ ( $\mathrm{s}, 1 \mathrm{H}$ ). ${ }^{13} \mathrm{CNMR}$ and HRMS are based on MG-1-15.
(4-Isopropoxyphenyl)(3,4-dimethoxyphenyl)methanol (9e). Yield: 57\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.24(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.92(\mathrm{~s}, 1 \mathrm{H}), 6.92-6.80(\mathrm{~m}, 4 \mathrm{H}), 5.70(\mathrm{~s}, 1 \mathrm{H}), 4.56-4.51(\mathrm{~m}$, $1 \mathrm{H}), 3.84(\mathrm{~d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}), 2.61(\mathrm{~s}, 1 \mathrm{H}), 1.33 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta$ $157.25,148.93,148.25,136.86,136.10,127.84,118.79,115.73,110.87,109.70,75.47,69.89$, 55.90, 55.82, 22.06 ppm. HRMS (ESI $) ~ m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}-\mathrm{OH}^{-}: 285.1485$, found 285.1484 .
(4-Propoxyphenyl)(3,4-dimethoxyphenyl)methanol (9f). Yield: $62 \%$. ${ }^{1} \mathrm{H}$ NMR ( 400 $\left.\mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.23(\mathrm{~d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 6.91(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 6.87-6.80(\mathrm{~m}, 4 \mathrm{H}), 5.70(\mathrm{~s}, 1 \mathrm{H})$, $3.91(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 3.89(\mathrm{~d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}), 2.61(\mathrm{~s}, 1 \mathrm{H}), 1.85-1.76(\mathrm{~m}, 2 \mathrm{H}), 1.04 \mathrm{ppm}(\mathrm{t}, J=$ $16 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 158.52,148.96,148.28,136.92,136.13,127.78,118.80$, $114.39,110.91,109.73,75.47,69.53,55.90,55.82,22.59,10.54 \mathrm{ppm} . \operatorname{HRMS}\left(\mathrm{ESI}^{+}\right) \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}-\mathrm{OH}^{-}: 285.1485$, found 285.1480.

Tert-butyl 4-(2-(4-((3,4-
dimethoxyphenyl)(hydroxy)methyl)phenoxy)ethyl)piperidine-1-carboxylate (9g*).

Yield: 16\%. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 7.25(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.91(\mathrm{~s}, 1 \mathrm{H}), 6.83-6.79(\mathrm{~m}$, $4 \mathrm{H}), 5.74(\mathrm{~s}, 1 \mathrm{H}), 3.99-3.96(\mathrm{~m}, 2 \mathrm{H}), 3.84(\mathrm{~s}, 3 \mathrm{H}), 3.83(\mathrm{~s}, 3 \mathrm{H}), 2.71-2.65(\mathrm{~m}, 2 \mathrm{H}), 2.35(\mathrm{~s}, 1 \mathrm{H})$, $1.70-1.68(\mathrm{~m}, 5 \mathrm{H}), 1.44(\mathrm{~s}, 9 \mathrm{H}), 1.16-1.13(\mathrm{~m}, 2 \mathrm{H}) \mathrm{ppm} .{ }^{13} \mathrm{C}$ NMR $\left(100 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 158.3$, $154.8,149.0,148.3,136.8,136.2,127.7,118.7,114.3,110.9,109.6,79.2,76.7,75.5,65.3,60.3$, 55.9, 55.8, 35.7, 32.9, 32.0, 28.4, 21.0, 14.2 ppm. HRMS is in Dr. Jalisa Holmes Ferguson's data information

General Procedure for the Synthesis of 16a to $\mathbf{1 6 c}$. One equivalent of $\mathbf{1 5}$ was dissolved in DMF, followed by the addition of one equivalent of bromide and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$. The reaction was stirred overnight at $78{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate. The resulting mixture was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The resulting residue was purified by column chromatography with 100:1 hexane-ethyl acetate as the eluent. 16a-16c were synthesized by according to literature reports. ${ }^{[45-46]}$
3-Isobutoxybenzaldehyde (16a). Yield: 96\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 9.99(\mathrm{~s}, 1 \mathrm{H}), 7.47-7.40$ $(\mathrm{m}, 3 \mathrm{H}), 7.21-7.18(\mathrm{~m}, 1 \mathrm{H}), 3.80(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.16-2.10(\mathrm{~m}, 1 \mathrm{H}), 1.05 \mathrm{ppm}(\mathrm{t}, J=8 \mathrm{~Hz}$, $6 \mathrm{H})$.
3-Isopropoxybenzaldehyde (16b). Yield: $95 \%$. ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 9.98$ (s, 1H), 7.45$7.39(\mathrm{~m}, 3 \mathrm{H}), 7.18-7.15(\mathrm{~m}, 1 \mathrm{H}), 4.68-4.62(\mathrm{~m}, 1 \mathrm{H}), 1.38 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 6 \mathrm{H})$.
3-Propoxybenzaldehyde (16c). Yield: $92 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 9.99(\mathrm{~s}, 1 \mathrm{H}), 7.46-7.40$ $(\mathrm{m}, 3 \mathrm{H}), 7.21-7.18(\mathrm{~m}, 1 \mathrm{H}), 4.00(\mathrm{t}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 1.89-1.81(\mathrm{~m}, 2 \mathrm{H}), 1.07 \mathrm{ppm}(\mathrm{t}, J=8 \mathrm{~Hz}, 3 \mathrm{H})$.

General Procedure for the Synthesis of 17a to 17c. One equivalent of 5 was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, followed by the addition of 1.1 equivalent of $n$-butyllithium. The reaction was stirred for 1.5 hour, and then 1 equivalent of $\mathbf{1 6}$ in dry THF under Ar was added drop-wise. The reaction was stirred for another 2 hours at $-78{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate to the solution. Then $\mathrm{NH}_{4} \mathrm{Cl}$ was added to quench the reaction. The resulting reaction mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-acetone as the eluent.
(3-Isobutoxyphenyl)(3,4-dimethoxyphenyl)methanol (17a). Yield: $68 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28-7.23(\mathrm{~m}, 1 \mathrm{H}), 6.97-6.90(\mathrm{~m}, 4 \mathrm{H}), 6.85-6.81(\mathrm{~m}, 2 \mathrm{H}), 5.76(\mathrm{~s}, 1 \mathrm{H}), 3.87(\mathrm{~d}, J=$ $4 \mathrm{~Hz}, 6 \mathrm{H}), 3.72(\mathrm{~d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.33(\mathrm{~s}, 1 \mathrm{H}), 2.10-2.07(\mathrm{~m}, 1 \mathrm{H}), 1.04 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 6 \mathrm{H})$. ${ }^{13} \mathrm{C}_{\mathrm{NMR}}\left(\mathrm{CDCl}_{3}\right): \delta 159.45,149.04,148.49,145.49,136.48,129.43,118.99,118.57,113.45$, $112.66,110.95,109.80,75.94,74.39,55.92,55.87,28.32,19.30 \mathrm{ppm}$. HRMS $^{\left(\text {ESI }^{+}\right) ~ m / z}$ calculated for $\mathrm{C}_{19} \mathrm{H}_{24} \mathrm{O}_{4}-\mathrm{OH}^{-}: 299.1642$, found 299.1640.
(3-Isopropoxyphenyl)(3,4-dimethoxyphenyl)methanol (17b). Yield: 72\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.20(\mathrm{t}, J=16 \mathrm{~Hz}, 1 \mathrm{H}), 6.92-6.86(\mathrm{~m}, 4 \mathrm{H}), 6.81-6.77(\mathrm{~m}, 2 \mathrm{H}), 5.70(\mathrm{~s}, 1 \mathrm{H}), 4.57-$ $4.51(\mathrm{~m}, 1 \mathrm{H}), 3.88(\mathrm{~d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}), 2.70(\mathrm{~s}, 1 \mathrm{H}), 1.29 \mathrm{ppm}(\mathrm{d}, J=8 \mathrm{~Hz}, 6 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 157.94,148.95,148.36,145.67,136.57,129.45,118.97,118.64,114.62,114.09$, $110.90,109.75,75.80,69.76,55.89,55.81,22.06,22.04 \mathrm{ppm}$. HRMS $\left(\mathrm{ESI}^{+}\right) \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}-\mathrm{OH}^{-}: 285.1485$, found 285.1481.
(3-Propoxyphenyl)(3,4-dimethoxyphenyl)methanol (17c). Yield: 67\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28-7.22(\mathrm{~m}, 1 \mathrm{H}), 6.98-6.84(\mathrm{~m}, 4 \mathrm{H}), 6.82-6.79(\mathrm{~m}, 2 \mathrm{H}), 5.71(\mathrm{~s}, 1 \mathrm{H}), 3.90(\mathrm{t}, J=$ $8 \mathrm{~Hz}, 2 \mathrm{H}), 3.84(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.66(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.83-1.77(\mathrm{~m}, 2 \mathrm{H}), 1.04 \mathrm{ppm}(\mathrm{t}, J=$ $8 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 159.26,149.00,148.42,145.61,136.58,129.41,118.98$, $118.64,113.37,112.68,110.95,109.82,75.83,69.45,55.90,55.84,22.61,10.56$ ppm. HRMS (ESI ${ }^{+}$) $m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{4}-\mathrm{OH}^{-}: 285.1485$, found 285.1483.

General Procedure for the Synthesis of 19a and 19b. One equivalent of 18 was dissolved in DMF. To this solution, one equivalent of bromide and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$ were added. The solution was stirred 5 hours at $90{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate. The combined organic solution was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The crude residue was purified by column chromatography with 100:1 hexaneethyl acetate as the eluent. 19a-19b* were synthesized by according to literature reports. ${ }^{[47-48]}$
1-(Allyloxy)-3-bromobenzene (19a). Yield: 95\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.24-7.14(\mathrm{~m}, 3 \mathrm{H})$, 6.89-6.86 (m, 1H), 6.14-6.04 (m, 1H), 5.48-5.44 (m, 1H), 5.39-5.35 (m, 1H), 4.55-4.52 ppm (m, $2 \mathrm{H})$.

1-Bromo-3-(hexyloxy)benzene (19b)*. Yield: $65 \%$. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.13(\mathrm{t}$, $J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.07-7.06(\mathrm{~m}, 2 \mathrm{H}), 6.83(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 3.93(\mathrm{t}, J=7 \mathrm{~Hz}, 2 \mathrm{H}), 1.77(\mathrm{q}, 2 \mathrm{H})$,
1.47-1.44(m, 2H), 1.36-1.34(m, 4H), $0.93(\mathrm{t}, J=7 \mathrm{~Hz}, 3 \mathrm{H}) \mathrm{ppm} .{ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 159.9,130.4,123.5,122.8,117.7,113.5,68.2,31.5,29.1,25.7,22.6,14.0 \mathrm{ppm}$.

General Procedure for the Synthesis of 17d and 17e. One equivalent of 19 was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, and 1.1 equivalent of $n$ butyllithium was added. The reaction was stirred 1.5 hour, followed by the dropwise addition of 1 equivalent of 3,4-dimethoxybenzaldehyde in dry THF under Ar. The reaction was stirred for another 2 hours at $-78{ }^{\circ} \mathrm{C}$. Then ethyl acetate was added followed by $\mathrm{NH}_{4} \mathrm{Cl}$. The mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-actone as the eluent.
(3-(Allyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (17d). Yield: 64\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.27(\mathrm{t}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 6.99-6.89(\mathrm{~m}, 4 \mathrm{H}), 6.84(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.10-6.03(\mathrm{~m}, 1 \mathrm{H})$, $5.78(\mathrm{~s}, 1 \mathrm{H}), 5.44(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 5.39(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 4.55-4.53(\mathrm{~m}, 2 \mathrm{H}), 3.88(\mathrm{~d}, J=8 \mathrm{~Hz}$, $6 \mathrm{H}), 2.24 \mathrm{ppm}(\mathrm{s}, 1 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 158.68,148.99,148.41,145.72,136.56,133.29$, $129.42,119.00,118.98,117.66,113.56,112.90,110.96,109.82,75.73,68.74,55.90,55.83$ ppm. HRMS ( $\mathrm{ESI}^{+}$) $m / z$ calculated for $\mathrm{C}_{18} \mathrm{H}_{20} \mathrm{O}_{4}-\mathrm{OH}^{-}: 283.1329$, found 283.1326.
(3-(Hexyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (17e*). Yield: 93\%. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.22(\mathrm{t}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.93-6.86(\mathrm{~m}, 4 \mathrm{H}), 6.82-6.77(\mathrm{~m}, 2 \mathrm{H}), 5.74(\mathrm{~s}, 1 \mathrm{H})$, $3.92(\mathrm{t}, J=7 \mathrm{~Hz}, 2 \mathrm{H}), 3.85(\mathrm{~s}, 3 \mathrm{H}), 3.84(\mathrm{~s}, 3 \mathrm{H}), 2.30(\mathrm{bs}, 1 \mathrm{H}), 1.75(\mathrm{p}, J=7 \mathrm{~Hz}, 2 \mathrm{H}), 1.45-1.40$ $(\mathrm{m}, 2 \mathrm{H}), 1.34-1.30(\mathrm{~m}, 4 \mathrm{H}), 0.90(\mathrm{t}, J=7 \mathrm{~Hz}, 1 \mathrm{H}) \mathrm{ppm} .{ }^{13} \mathrm{C} \operatorname{NMR}\left(100 \mathrm{MHz}, \mathrm{CDCl}_{3}\right): \delta 159.4$, $149.1,148.5,145.6,136.5,129.5,119.0,118.6,113.5,112.7,111.0,109.8,76.0,68.0,56.0$, 55.9, 31.7, 29.3, 25.8, 22.7, 14.1 ppm. HRMS is in Dr. Jalisa Holmes Ferguson's data information.

General Procedure for the Synthesis of 21a to 21c. One equivalent of $\mathbf{2 0}$ was dissolved in DMF, followed by the addition of 1 equivalent of alcohol and 2 equivalents of NaH . The reaction was stirred overnight at $0{ }^{0} \mathrm{C}$ to room temperature. To this solution, ethyl acetate was added. The resulting mixture was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and
concentrated. The residue was purified by column chromatography with 100:1 hexane-ethyl acetate as the eluent. 21a-21c were synthesized by according to literature reports. ${ }^{[49-51]}$

1-Bromo-2-(phenoxymethyl)benzene (21a). Yield: $99 \%$. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta$ 7.69-7.65 (m, 2H), 7.43-7.39 (m, 3H), 7.26 (t, $J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.11-7.07(\mathrm{~m}, 3 \mathrm{H}), 5.23(\mathrm{~s}, 2 \mathrm{H})$ ppm. ${ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta$ 158.5, 136.4, 132.6, 129.6, 129.2, 128.9, 127.6, 122.3, 121.2, 114.9, 69.3 ppm .

Tert-butyl 4-(((2-bromobenzyl)oxy)methyl)piperidine-1-carboxylate (21b). Yield: $96 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.56-7.54(\mathrm{~m}, 1 \mathrm{H}), 7.49-7.47(\mathrm{~m}, 1 \mathrm{H}), 7.33(\mathrm{t}, J=16 \mathrm{~Hz}, 1 \mathrm{H}), 7.18$ $(\mathrm{t}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 4.57(\mathrm{~s}, 2 \mathrm{H}), 3.42(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 1.80-1.77(\mathrm{~m}, 4 \mathrm{H}), 1.48(\mathrm{~s}, 9 \mathrm{H}), 1.28-1.15$ ppm (m, 5H).

Tert-butyl 4-(2-((2-bromobenzyl)oxy)ethyl)piperidine-1-carboxylate (21c). Yield: $96 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.56(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 7.47(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 7.33(\mathrm{t}, J=16 \mathrm{~Hz}, 1 \mathrm{H})$, $7.17(\mathrm{t}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 4.57(\mathrm{~s}, 2 \mathrm{H}), 3.62(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 1.70-1.59(\mathrm{~m}, 6 \mathrm{H}), 1.47(\mathrm{t}, J=$ $16 \mathrm{~Hz}, 9 \mathrm{H}), 1.13 \mathrm{ppm}(\mathrm{d}, J=12 \mathrm{~Hz}, 3 \mathrm{H})$.

General Procedure for the Synthesis of 22a to 22c. One equivalent of 21 was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, followed by the addition of 1.1 equivalent of $n$-butyllithium. The reaction was stirred for 1.5 hour, and then 1 equivalent of 3,4,5trimethoxybenzaldehyde in dry THF under Ar was added drop-wise. The reaction was stirred for another 2 hours at $-78{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate to the solution. Then $\mathrm{NH}_{4} \mathrm{Cl}$ was added to quench the reaction. The resulting reaction mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-acetone as the leuent.
(2-(Phenoxymethyl)phenyl)(3,4,5-trimethoxyphenyl)methanol (22a*). Yield: 58\%. ${ }^{1} \mathrm{H}^{\mathrm{NMR}}\left(\mathrm{CDCl}_{3}\right) \delta 7.44(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 7.33-7.28(\mathrm{~m}, 5 \mathrm{H}), 7.00(\mathrm{t}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 6.92-6.90$ $(\mathrm{m}, 2 \mathrm{H}), 6.55(\mathrm{~s}, 2 \mathrm{H}), 6.07(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 5.02-4.94(\mathrm{dd}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 3.86(\mathrm{~d}, J=4 \mathrm{~Hz}$, $3 \mathrm{H}), 3.72(\mathrm{~d}, J=4 \mathrm{~Hz}, 6 \mathrm{H}), 2.92 \mathrm{ppm}(\mathrm{d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR and HRMS are in Dr. Jalisa Holmes Ferguson's data information.

## Tert-butyl

Yield: $55 \%{ }^{1}{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.35-7.23(\mathrm{~m}, 4 \mathrm{H}), 6.61(\mathrm{~s}, 2 \mathrm{H}), 5.95(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 4.61$ $(\mathrm{d}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 4.40(\mathrm{~d}, J=12 \mathrm{~Hz}, 1 \mathrm{H}), 4.17-4.12(\mathrm{~m}, 2 \mathrm{H}), 3.86(\mathrm{~s}, 3 \mathrm{H}), 3.81(\mathrm{~s}, 6 \mathrm{H}), 3.34(\mathrm{t}$, $J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.67(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 1.79-1.69(\mathrm{~m}, 4 \mathrm{H}), 1.46(\mathrm{~s}, 9 \mathrm{H}), 1.17-1.13 \mathrm{ppm}(\mathrm{m}, 2 \mathrm{H})$. ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 154.80,153.12,143.32,138.44,136.98,135.29,130.76,128.94,128.88$, 127.93, 103.57, 79.36, 75.51, 73.81, 72.46, 60.89, 56.09, 43.36, 36.50, 29.07, 28.45 ppm. HRMS (ESI') $m / z$ calculated for $\mathrm{C}_{28} \mathrm{H}_{39} \mathrm{NO}_{7}-\mathrm{OH}^{-}: 484.2694$, found 484.2691. $1.46(\mathrm{~s}, 9 \mathrm{H}), 1.12-1.09 \mathrm{ppm}(\mathrm{m}, 2 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 154.85,153.10,143.45,138.50$, 136.94, 135.39, 130.78, 129.10, 128.91, 127.92, 103.51, 79.26, 73.88, 72.32, 68.15, 60.90 , 56.09, 43.81, 36.13, 32.90, 32.06, 28.46 ppm . HRMS (ESI ${ }^{+} \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{29} \mathrm{H}_{41} \mathrm{NO}_{7}-$ $\mathrm{OH}^{-}: 498.2850$, found 498.2827.

General Procedure for the Synthesis of 24a to 24f. One equivalent of $\mathbf{2 3}$ was dissolved in DMF. To this solution, 1 equivalents of RBr and 2 equivalents of $\mathrm{K}_{2} \mathrm{CO}_{3}$ were added. The reaction was stirred overnight at $78{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate. The resulting mixture was washed with water and brine, dried over $\mathrm{MgSO}_{4}$, and concentrated. The resulting residue was purified by column chromatography with 100:1 hexane-ethyl acetate. 24a-24f were synthesized by according to literature reports. ${ }^{[52-56]}$
1-(Allyloxy)-4-bromo-2-methylbenzene (24a). Yield: 97\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) ~ \delta ~ 7.29-~$ $7.24(\mathrm{~m}, 2 \mathrm{H}), 6.70(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.11-6.04(\mathrm{~m}, 1 \mathrm{H}), 5.47(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 5.33(\mathrm{~d}, J=4 \mathrm{~Hz}$, $1 \mathrm{H}), 4.55-4.53$ (m, 2H), $2.26 \mathrm{ppm}(\mathrm{s}, 3 \mathrm{H})$.
4-Bromo-1-isobutoxy-2-methylbenzene (24b). Yield: 96\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.29-$ 7.24 (m, 2H), 6.70 (d, $J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.11-6.04$ (m, 1H), 5.47 (d, $J=4 \mathrm{~Hz}, 1 \mathrm{H}), 5.33$ (d, $J=4 \mathrm{~Hz}$, $1 \mathrm{H}), 4.55-4.53(\mathrm{~m}, 2 \mathrm{H}), 2.26 \mathrm{ppm}(\mathrm{s}, 3 \mathrm{H})$.

4-Bromo-1-isopropoxy-2-methylbenzene (24c). Yield: $95 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28-$ $7.25(\mathrm{~m}, 2 \mathrm{H}), 6.69(\mathrm{~d} . J=8 \mathrm{~Hz}, 1 \mathrm{H}), 3.73(\mathrm{~d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.24(\mathrm{~s}, 3 \mathrm{H}), 2.19-2.09(\mathrm{~m}, 1 \mathrm{H}), 1.08$ ppm (d, $J=4 \mathrm{~Hz}, 6 \mathrm{H})$.

4-Bromo-2-methyl-1-propoxybenzene (24d). Yield: $97 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.25$ (d, $J$ $=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.68(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.91(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.21(\mathrm{~s}, 3 \mathrm{H}), 1.90-1.80(\mathrm{~m}, 2 \mathrm{H}), 1.08$ ppm (t, $J=8 \mathrm{~Hz}, 3 \mathrm{H})$.
4-Bromo-1-(hexyloxy)-2-methylbenzene (24e). Yield: $99 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \quad \delta 7.24$ $(\mathrm{d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 6.69(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.91(\mathrm{t}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.22(\mathrm{~s}, 3 \mathrm{H}), 1.86-1.79(\mathrm{~m}, 2 \mathrm{H})$, $1.52-1.48(\mathrm{~m}, 1 \mathrm{H}), 1.40-1.33(\mathrm{~m}, 5 \mathrm{H}), 0.92 \mathrm{ppm}(\mathrm{s}, 3 \mathrm{H})$.
4-Bromo-1-(cyclohexylmethoxy)-2-methylbenzene (24f). Yield: 92\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.28-7.24(\mathrm{~m}, 2 \mathrm{H}), 6.67(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.73(\mathrm{~d}, J=4 \mathrm{~Hz}, 2 \mathrm{H}), 2.21(\mathrm{~s}, 3 \mathrm{H}), 1.89-$ $1.72(\mathrm{~m}, 5 \mathrm{H}), 1.34-1.23(\mathrm{~m}, 4 \mathrm{H}), 1.11-1.08 \mathrm{ppm}(\mathrm{m}, 2 \mathrm{H})$.

General Procedure for Synthesis of 25a to 25f. One equivalent of $\mathbf{2 4}$ was dried for half an hour under oil-vacuum pump, and dissolved in dry THF under argon gas. The solution was cooled in a dry ice-acetone bath for 30 minutes, followed by the addition of 1.1 equivalent of $n$ butyllithium. The reaction was stirred for 1.5 hour, and then 1 equivalent of 3,4,5trimethoxybenzaldehyde in dry THF under Ar was added drop-wise. The reaction was stirred for another 2 hours at $-78{ }^{\circ} \mathrm{C}$, followed by the addition of ethyl acetate to the solution. Then $\mathrm{NH}_{4} \mathrm{Cl}$ was added to quench the reaction. The resulting reaction mixture was washed with water, dried over $\mathrm{MgSO}_{4}$, and concentrated. The residue was purified by column chromatography with 100:1 hexane-acetone as the leuent.
(4-(Allyloxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25a). Yield: 60\%. ${ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 7.16-7.15(\mathrm{~m}, 2 \mathrm{H}), 6.79(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 6.63(\mathrm{~s}, 2 \mathrm{H}), 6.12-6.05(\mathrm{~m}, 1 \mathrm{H})$, $5.71(\mathrm{~s}, 1 \mathrm{H}), 5.46(\mathrm{dd}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 5.31-5.27(\mathrm{~m}, 1 \mathrm{H}), 4.56-4.54(\mathrm{~m}, 2 \mathrm{H}), 3.86-3.85(\mathrm{~m}, 9 \mathrm{H})$, $2.26(\mathrm{~s}, 3 \mathrm{H}), 2.21 \mathrm{ppm}(\mathrm{s}, 1 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.35,153.21,139.75,137.11,135.64$, $133.50,129.16,127.17,125.01,117.00,111.12,103.39,75.97,68.77,60.83,56.11,16.41 \mathrm{ppm}$. HRMS (ESI ${ }^{+}$) $m / z$ calculated for $\mathrm{C}_{20} \mathrm{H}_{24} \mathrm{O}_{5}-\mathrm{OH}^{-}: 327.1591$, found 327.1592.
(4-Isobutoxy-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25b). Yield: 59\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.15-7.13(\mathrm{~m}, 2 \mathrm{H}), 6.77(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.64(\mathrm{~s}, 2 \mathrm{H}), 5.71(\mathrm{~d}, J=4 \mathrm{~Hz}$, $1 \mathrm{H}), 3.86(\mathrm{~d}, J=4 \mathrm{~Hz}, 9 \mathrm{H}), 3.73(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.24(\mathrm{~s}, 3 \mathrm{H}), 2.15-2.11(\mathrm{~m}, 2 \mathrm{H}), 1.05 \mathrm{ppm}(\mathrm{d}$, $J=8 \mathrm{~Hz}, 6 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): ~ \delta 156.84,153.14,139.99,136.99,135.33,128.99,126.94$, $125.02,110.62,103.38,75.92,74.35,60.80,56.07,28.43,19.33,16.34 \mathrm{ppm}$. HRMS $\left(\mathrm{ESI}^{+}\right) \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{21} \mathrm{H}_{28} \mathrm{O}_{5}-\mathrm{OH}^{-}: 343.1904$, found 343.1905.
(4-Isopropoxy-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25c). Yield: $43 \%$. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.16-7.11(\mathrm{~m}, 2 \mathrm{H}), 6.81(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.64(\mathrm{~s}, 2 \mathrm{H}), 5.72(\mathrm{~s}, 1 \mathrm{H})$, $4.54-4.51(\mathrm{~m}, 1 \mathrm{H}), 3.86(\mathrm{t}, J=4 \mathrm{~Hz}, 9 \mathrm{H}), 2.21(\mathrm{~s}, 3 \mathrm{H}), 2.13(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.35 \mathrm{ppm}(\mathrm{d}, J=$ $4 \mathrm{~Hz}, 6 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 153.21,139.74,135.28,129.27,124.97,112.82,112.50$, 104.27, 103.40, 76.04, 70.28, 60.84, 56.12, 56.03, 22.25, $16.56 \mathrm{ppm} . \operatorname{HRMS}\left(\mathrm{ESI}^{+}\right) \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{20} \mathrm{H}_{26} \mathrm{O}_{5}-\mathrm{OH}^{-}: 329.1747$, found 329.1732.
(3-Methyl-4-propoxyphenyl)(3,4,5-trimethoxyphenyl)methanol (25d). Yield: 69\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.13(\mathrm{t}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 6.78(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.63(\mathrm{~s}, 2 \mathrm{H}), 5.70(\mathrm{~s}, 1 \mathrm{H}), 3.93$ (t, $J=8 \mathrm{~Hz}, 2 \mathrm{H}), 3.88(\mathrm{~s}, 9 \mathrm{H}), 2.34(\mathrm{~s}, 1 \mathrm{H}), 2.23(\mathrm{~s}, 3 \mathrm{H}), 1.86-1.81(\mathrm{~m}, 2 \mathrm{H}), 1.07 \mathrm{ppm}(\mathrm{t}, J=$ $8 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.65,153.17,139.88,137.04,135.30,129.03,127.02$, $125.03,110.75,103.38,75.97,69.54,60.82,56.10,22.71,16.33,10.66 \mathrm{ppm}$. HRMS (ESI ${ }^{+} \mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{20} \mathrm{H}_{26} \mathrm{O}_{5}-\mathrm{OH}^{-}: 329.1747$, found 329.1747.
(4-(Hexyloxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25e). Yield: 63\%. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.15-7.13(\mathrm{~m}, 2 \mathrm{H}), 6.78(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.64(\mathrm{~s}, 2 \mathrm{H}), 5.72(\mathrm{~s}, 1 \mathrm{H}), 3.97$ $(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 3.85(\mathrm{~d}, J=4 \mathrm{~Hz}, 9 \mathrm{H}), 2.23(\mathrm{~s}, 3 \mathrm{H}), 2.19(\mathrm{~s}, 1 \mathrm{H}), 1.83-1.77(\mathrm{~m}, 2 \mathrm{H}), 1.51-1.47$ $(\mathrm{m}, 2 \mathrm{H}), 1.38-1.34(\mathrm{~m}, 4 \mathrm{H}), 0.92 \mathrm{ppm}(\mathrm{t}, J=12 \mathrm{~Hz}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 156.92,153.21$, $139.79,137.09,135.21,129.02,127.06,125.03,110.72,103.37,76.04,68.05,60.84,56.12$, 31.57, 29.30, 25.82, 22.62, 16.36, 14.03 ppm. HRMS (ESI ${ }^{+}$m/z calculated for $\mathrm{C}_{23} \mathrm{H}_{32} \mathrm{O}_{5}-$ $\mathrm{OH}^{-}: 371.2217$, found 371.2219.
(4-(Cyclohexylmethoxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25f).
Yield: $62 \%{ }^{1}{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 7.15-7.13(\mathrm{~m}, 2 \mathrm{H}), 6.77(\mathrm{~d}, J=8 \mathrm{~Hz}, 1 \mathrm{H}), 6.64(\mathrm{~s}, 2 \mathrm{H}), 5.72(\mathrm{~d}$, $J=4 \mathrm{~Hz}, 1 \mathrm{H}), 3.89(\mathrm{~s}, 9 \mathrm{H}), 3.76(\mathrm{~d}, J=8 \mathrm{~Hz}, 2 \mathrm{H}), 2.23(\mathrm{~s}, 3 \mathrm{H}), 2.15(\mathrm{~d}, J=4 \mathrm{~Hz}, 1 \mathrm{H}), 1.90-1.74$
(m, 6H), 1.25-1.08 ppm (m, 5H). ${ }^{13} \mathrm{C}$ NMR (100 MHz, CDCl3): $\delta 156.91,153.14,139.96$, $136.99,135.26,128.97,126.97,125.01,110.62,103.36,75.94,73.45,60.80,56.08,37.84$, 29.97, 26.57, 25.89, 16.38 ppm . HRMS ( $\mathrm{ESI}^{+}$) $\mathrm{m} / \mathrm{z}$ calculated for $\mathrm{C}_{24} \mathrm{H}_{32} \mathrm{O}_{5}-\mathrm{OH}^{-}: 383.2217$, found 383.2213.

## REFERENCES

1. Ma, X.; Yu, H., Global Burden of Cancer. Yale J Biol Med 2006, 79(3-4), 85-9.
2. Cell, Volume 144, Issue 5, 4 March 2011, Page 646-674
3. Sonnenschein, C.; Soto, A. M., The aging of the 2000 and 2011 Hallmarks of Cancer, Review: a critique. J. Biosci 2013, 38 (3), 651-63.
4. Lazebnik, Y., What are the Hallmarks of Cancer. Nat. Rev. Cancer 2010,10 (4), 232-3.
5. Burroughs, S. K.; Kaluz, S.; Wang, D.; Wang, K.; Van Meir, E. G.; Wang, B., Hypoxia inducible factor pathway inhibitors as anticancer therapeutics. Future Med Chem 2013, 5 (5), 553-72.
6. Tan, C.; de Noronha, R. G.; Devi, N. S.; Jabbar, A. A.; Kaluz, S.; Liu, Y.; Mooring, S. R.; Nicolaou, K. C.; Wang, B.; Van Meir, E. G., Sulfonamides as a new scaffold for hypoxia inducible factor pathway inhibitors. Bioorg Med Chem Lett 2011, 21 (18), 5528-32.
7. Gleadle, J.M.; Mole, D. R.; Pugh, C. W., Hypoxia-inducible factors: where, when and why? Kidney International 2015, 69(1), 15-17.
8. Bruick. R. K., Oxygen sensing in the hypoxic response pathway: regulation of the hypoxia- inducible transcription factor. Genes \& Dev 2003, 17, 2614-2623.
9. Mooring, S. R.; Jin, H.; Devi, N. S.; Jabbar, A. A.; Kaluz, S.; Liu, Y.; Van Meir, E. G.; Wang, B., Design and synthesis of novel small-molecule inhibitors of the hypoxia inducible factor pathway. J Med Chem 2011, 54 (24), 8471-89.
10. Ke, Q.; Costa, M., Hypoxia-inducible factor-1 (HIF-1). Mol Pharmacol 2006, 70 (5), 1469-80.
11. Ziello, J.; Jovin, I.; Huang, Y., Hypoxia-Inducible Factor (HIF)-1 Regulatory Pathway and its Potential for Therapeutic Intervention in Malignancy and Ischemia. Yale J Biol Med 2007, 80(2), 51-60.
12. Vadlapatla, R. K.; Vadlapudi, A. D.; Mitra, A. K., Hypoxia-Inducible Factor-1 (HIF-1): A Potential Target for Intervention in Ocular Neovascular Diseases. Curr Drug Targets 2013, 14(8), 919-935.
13. Dengler, V. L.; Galbraith, M. D.; Espinosa, J. M., Transcriptional regulation by hypoxia inducible factors. Crit Rev Biochem Mol Biol 2014, 49 (1), 1-15.
14. Shang, F.; Taylor, A., Roles for the ubiquitin-proteasome pathway in protein quality control and signaling in the retina: implications in the pathogenesis of age-related macular degeneration. Mol Aspects Med 2012, 33 (4), 446-66.
15. Speer, R. E.; Karuppagounder, S. S.; Basso, M.; Sleiman, S. F.; Kumar, A.; Brand, D.; Smirnova, N.; Gazaryan, I.; Khim, S. J.; Ratan, R. R., Hypoxia-inducible factor prolyl hydroxylases as targets for neuroprotection by "antioxidant" metal chelators: From ferroptosis to stroke. Free Radic Biol Med 2013, 62, 26-36.
16. Rabinowitz, M. H., Inhibition of hypoxia-inducible factor prolyl hydroxylase domain oxygen sensors: tricking the body into mounting orchestrated survival and repair responses. $J$ Med Chem 2013, 56 (23), 9369-402.
17. Zheng, G.; Cox, T.; Tribbey, L.; Wang, G. Z.; Iacoban, P.; Booher, M. E.; Gabriel, G. J.; Zhou, L.; Bae, N.; Rowles, J.; He, C.; Olsen, M. J., Synthesis of a FTO inhibitor with anticonvulsant activity. ACS Chem Neurosci 2014, 5 (8), 658-65.
18. Percy, M. J.; Mooney, S. M.; McMullin, M. F.; Flores, A.; Lappin, T. R.; Lee, F. S., A common polymorphism in the oxygen-dependent degradation (ODD) domain of hypoxia
inducible factor-1alpha (HIF-1alpha) does not impair Pro-564 hydroxylation. Mol Cancer 2003, 2, 31 .
19. Poon, E.; Harris, A. L.; Ashcroft, M., Targeting the hypoxia-inducible factor (HIF) pathway in cancer. Expert Reviews in Molecular Medicine 2009, 11, e26.
20. Liu, W.; Shen, S. M.; Zhao, X. Y.; Chen, G. Q., Targeted genes and interacting proteins of hypoxia inducible factor-1. Int J Biochem Mol Biol 2012, 3(2), 165.
21. Kaur, B.; Khwaja, F. W.; Severson, E. A.; Matheny, S. L.; Brat, D. J.; Van Meir, E. G., Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. Neuro Oncol 2005, 7 (2), 134-53.
22. Huang, L. E.; Bunn, H. F., Hypoxia-inducible factor and its biomedical relevance. J Biol Chem 2003, 278 (22), 19575-8.
23. Roskoski, R., Jr., Vascular endothelial growth factor (VEGF) signaling in tumor progression. Crit Rev Oncol Hematol 2007, 62 (3), 179-213.
24. What is erythropoietin? https://www.themmrf.org/multiple-myeloma-knowledge-center/myeloma-treatments-guide/growth-factors/erythropeietin/
25. Debeljak, N.; Solar, P.; Sytkowski, A. J., Erythropoietin and cancer: the unintended consequences of anemia correction. Front Immunol 2014, 5, 563.
26. Tan, C.; de Noronha, R. G.; Roecker, A. J.; Pyrzynska, B.; Khwaja, F.; Zhang, Z.; Zhang, H.; Teng, Q.; Nicholson, A. C.; Giannakakou, P.; Zhou, W.; Olson, J. J.; Pereira, M. M.; Nicolaou, K. C.; Van Meir, E. G., Identification of a novel small-molecule inhibitor of the hypoxia-inducible factor 1 pathway. Cancer Res 2005, 65(2), 605-12.
27. Semenza, G. L., Hypoxia-inducible factors: mediators of cancer progression and targets for cancer therapy. Trends Pharmacol Sci 2012, 33 (4), 207-14.
28. Onnis, B.; Rapisarda, A.; Melillo, G., Development of HIF-1 inhibitors for cancer therapy. J Cell Mol Med 2009, 13 (9A), 2780-6.
29. Terzuoli, E.; Puppo, M.; Rapisarda, A.; Uranchimeg, B.; Cao, L.; Burger, A. M.; Ziche, M.; Melillo, G., Aminoflavone, a ligand of the aryl hydrocarbon receptor, inhibits HIF-1alpha expression in an AhR-independent fashion. Cancer Res 2010, 70 (17), 6837-48.
30. Kwon, D. Y.; Lee, H. E.; Weitzel, D. H.; Park, K.; Lee, S. H.; Lee, C. T.; Stephenson, T.
N.; Park, H.; Fitzgerald, M. C.; Chi, J. T.; Mook, R. A., Jr.; Dewhirst, M. W.; Lee, Y. M.; Hong, J., Synthesis and Biological Evaluation of Manassantin Analogues for Hypoxia-Inducible Factor 1alpha Inhibition. $J$ Med Chem 2015, 58 (19), 7659-71.
31. Hossain, C. F.; Kim, Y. P.; Baerson, S. R.; Zhang, L.; Bruick, R. K.; Mohammed, K. A.; Agarwal, A. K.; Nagle, D. G.; Zhou, Y. D., Saururus cernuus lignans--potent small molecule inhibitors of hypoxia-inducible factor-1. Biochem Biophys Res Commun 2005, 333 (3), 102633.
32. Seo, C. S.; Lee, W. H.; Chung, H. W.; Chang, E. J.; Lee, S. H.; Jahng, Y.; Hwang, B. Y.; Son, J. K.; Han, S. B.; Kim, Y., Manassantin A and B from Saururus chinensis inhibiting cellular melanin production. Phytother Res 2009, 23 (11), 1531-6.
33. Yin, S.; Kaluz, S.; Devi, N. S.; Jabbar, A. A.; de Noronha, R. G.; Mun, J.; Zhang, Z.; Boreddy, P. R.; Wang, W.; Wang, Z.; Abbruscato, T.; Chen, Z.; Olson, J. J.; Zhang, R.; Goodman, M. M.; Nicolaou, K. C.; Van Meir, E. G., Arylsulfonamide KCN1 inhibits in vivo glioma growth and interferes with HIF signaling by disrupting HIF-1alpha interaction with cofactors p300/CBP. Clin Cancer Res 2012, 18 (24), 6623-33.
34. Liao, J.; Fan, L.; Guo, W.; Zhang, Z.; Li, J.; Zhu, C.; Ren, Y.; Wu, W.; Jiang, H., Palladium-Catalyzed Fluoroalkylative Cyclization of Olefins. Org Lett 2017, 19 (5), 1008-1011.
35. Chen, Y. H.; Chang, C. Y.; Chang, C. F.; Chen, P. C.; Lee, Y. T.; Chern, C. Y.; Tsai, J. N., Pro-Angiogenic Effects of Chalcone Derivatives in Zebrafish Embryos in Vivo. Molecules 2015, 20 (7), 12512-24.
36. Liu, W.; Wang, Y.; Sun, M.; Zhang, D.; Zheng, M.; Yang, W., Alkoxy-position effects on piezofluorochromism and aggregation-induced emission of 9,10-
bis(alkoxystyryl)anthracenes. Chem Commun (Camb) 2013, 49 (54), 6042-4.
37. Deng, H.; Su, Y.; Hu, M.; Jin, X.; He, L.; Pang, Y.; Dong, R.; Zhu, X., Multicolor Fluorescent Polymers Inspired from Green Fluorescent Protein. Macromolecules 2015, 48 (16), 5969-5979.
38. Wang, D.; Wang, L.; Wu, Y.; Song, S.; Feng, J.; Zhang, X., Natural $\alpha$-methylenelactam analogues: Design, synthesis and evaluation of $\alpha$-alkenyl- $\gamma$ and $\delta$-lactams as potential antifungal agents against Colletotrichum orbiculare. Eur J Med Chem 2017, 130, 286-304.
39. Xiong, Y.; Yan, X.; Ma, Y.; Li, Y.; Yin, G.; Chen, L., Regulating the piezofluorochromism of 9,10-bis(butoxystyryl)anthracenes by isomerization of butyl groups. Chem. Comтип 2015, 51, 3403-3406.
40. Li, J.; He, J., Synthesis of Sequence-Regulated Polymers: Alternating Polyacetylene through Regioselective Anionic Polymerization of Butadiene Derivatives. ACS Macro Letters 2015, 4 (4), 372-376.
41. Li, Z.; Yazaki, R.; Ohshima, T., Chemo- and Regioselective Direct Functional Group Installation through Catalytic Hydroxy Group Selective Conjugate Addition of Amino Alcohols to alpha,beta-Unsaturated Sulfonyl Compounds. Org Lett 2016, 18 (14), 3350-3.
42. Wei, H.; Li, D.; Yang, X.; Shang, H.; Fan, S.; Li, Y.; Song, D., Design and Synthesis of Vandetanib Derivatives Containing Nitroimidazole Groups as Tyrosine Kinase Inhibitors in Normoxia and Hypoxia. Molecules 2016, 21 (12).
43. Arnatt, CK.; Adams, JL.; Zhang, Z.; Haney, KM.; Li, G.; Zhang, Y., Design, syntheses, and characterization of piperazine based chemokine receptor CCR5 antagonists as anti prostate cancer agents. Bioorg Med Chem Lett 2014, 24(10), 2319-23.
44. Bang, JS.; Kim, YJ.; Song, J.; Yoo, JS.; Lee, S.; Lee, MJ.; Min, H.; Hwang, KW.; Min, KH., Small molecules that regulate zymosan phagocytosis of macrophage through deactivation of Rho GTPases. Bioorg Med Chem 2012, 20(17), 5262-8.
45. Brodney, M. A.; Barreiro, G.; Oglivie, K.; Hajos-Korcsok, E.; Murray, J.; Vajdos, F.; Ambroise, C.; Christoffersen, C.; Fisher, K.; Lanyon, L.; Liu, J.; Nolan, C. E.; Withka, J. M.; Borzilleri, K. A.; Efremov, I.; Oborski, C. E.; Varghese, A.; O’Neill, B. T., Spirocyclic Sulfamides as $\beta$-Secretase 1 (BACE-1) Inhibitors for the Treatment of Alzheimer's Disease: Utilization of Structure Based Drug Design, WaterMap, and CNS Penetration Studies To Identify Centrally Efficacious Inhibitors. J Med Chem 2012, 55(21), 9224-39.
46. Liu, W.; Wang, Y.; Sun, M.; Zhang, D.; Zheng, M.; Yang, W., Alkoxy-position effects on piezofluorochromism and aggregation-induced emission of 9,10bis(alkoxystyryl)anthracenes. Chem Coтmии (Camb) 2013, 49 (54), 6042-4.
47. Minutolo, F.; Bellini, R.; Bertini, S.; Carboni, I.; Lapucci, A.; Pistolesi, L.; Prota, G.; Rapposelli, S.; Solati, F.; Tuccinardi, T.; Martinelli, A.; Stossi, F.; Carlson, K. E.;
Katzenellenbogen, B. S.; Katzenellenbogen, J. A.; Macchia, M., Monoaryl-Substituted Salicylaldoximes as Ligands for Estrogen Receptor $\beta$. J. Med. Chem. 2008, 51, 1344-51.
48. Liu, H. Y.; Wu, P. J.; Kuo, S. Y.; Chen, C. P.; Chang, E. H.; Wu, C. Y.; Chan, Y. H., Quinoxaline-Based Polymer Dots with Ultrabright Red to Near-Infrared Fluorescence for In Vivo Biological Imaging. J Am Chem Soc 2015, 137 (32), 10420-9.
49. Corrie, T. J.; Ball, L. T.; Russell, C. A.; Lloyd-Jones, G. C., Au-Catalyzed Biaryl Coupling To Generate 5- to 9-Membered Rings: Turnover-Limiting Reductive Elimination versus pi-Complexation. J Am Chem Soc 2017, 139 (1), 245-254.
50. Budzik, B.; Garzya, V.; Shi, D.; Walker, G. Lauchart, Y.; Lucas, A. J.; Rivero, R. A.; Langmead C. J.; Wastson, J.; Wu, Z.; Forbes, I. T.; Jin, J., 2’ biaryl amides as novel and subtype selective M1 agonists. Part II: Further optimization and profiling, Bioorg Med Chem Lett 2010, 20(12), 3545-9.
51. Bissantz, Caterina et al, Preparation of 8-hydroxyquinazolin-4(3H)-one derivatives as catechol-O-methyltransferase (COMT) inhibitors, PCT Int. Appl. Patent 2014102233, Jul 03, 2014.
52. Shiraishi, M.; Baba, M.; Seto, M.; Nanzaki, N.; Nishimura, O., Preparation of benzothiepinecarboxanilides and related compounds as CCR-5 antagonists. PCT Int. Appl Patent, 2000037455, Jun 29, 2000.
53. Andreini, M.; Gabellieri, E.; Guba, W.; Marconi, G.; Narquizian, R.; Power, E.; Travagli, M.; Woltering, T.; Wostl, W., Preparation of 4, 4-diphenyl-4, 5-dihydro-oxazol-2ylamin derivatives as $\beta$-secretase inhibitors. U. S. Patent 2009-0209529, Feb 12, 2009.
54. Richards, S. J.; von Geldern, T. W.; Jacobson, P.; Wilcox, D.; Nguyen, P.; Ohman, L.; Osterlund, M.; Gelius, B.; Grynfarb, M.; Goos-Nilsson, A.; Wang, J.; Fung, S.; Kalmanovich, M., Synthesis and activity of novel bile-acid conjugated glucocorticoid receptor antagonists, Bioorg Med Chem Lett 2006, 16(23), 6086-90.
55. Kobayashi, Takeshi et al, Smectiv liquid-crystalline materials, liquid-crystal displays, and spatial optical modulators, Jpn. Kokai Tokkyo Koho, Patent 2011088987, May 06, 2011. 56. Baloglu, Erkan et al, Substituted 2, 3-dihydrobenzofuranyl compounds as cytotoxic agents and their preparation, PCT Int. Appl., Patent 2014085607, Jun 05, 2014.

## APPENDICES

## Spectra

## Class I: Ortho-phenolic ether benzhydrol analogues

2-(allyloxy)benzaldehyde (4a)



## 2-isopropoxybenzaldehyde (4b)




## 2-propoxybenzaldehyde (4c)




2-(hexyloxy)benzaldehyde (4d).



## 2-(cyclohexylmethoxy)benzaldehyde (4e)




## 2-(benzyloxy)benzaldehyde (4f)




## (2-(allyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (6a)




(3,4-dimethoxyphenyl)(2-isopropoxyphenyl)methanol (6b)




LY I 6 ESIPOS BWANG 012017 \#145-158 RT: 2.04-2.22 AV: 14 4E7
T: FTMS + p ESIFull ms [100.00-1000.00]
4 E7


## (3,4-dimethoxyphenyl)(2-propoxyphenyl)methanol (6c)






## (3,4-dimethoxyphenyl)(2-(hexyloxy)phenyl)methanol (6d)






## (2-(cyclohexylmethoxy)phenyl)(3,4-dimethoxyphenyl)methanol (6e)






## (2-(benzyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (6f)






## Class II: Para-phenolic ether benzhydrol analogues

## 4-isobutoxybenzaldehyde (8a).




## 4-(hexyloxy)benzaldehyde (8b)




## 4-(cyclohexylmethoxy)benzaldehyde (8c)




## 1-bromo-4-isopropoxybenzene (14b)




## 1-bromo-4-propoxybenzene (14c)




Tert-butyl 4-(2-(4-bromophenoxy)ethyl)piperidine-1-carboxylate (14d)* Spectra will be in Jalisa Ferguson's information.

## (3,4-dimethoxyphenyl)(4-isobutoxyphenyl)methanol (9a)





| 170 | 160 | 150 | 140 | 130 | 120 | 110 | 100 |  | 80 | 70 | 60 | 50 | 40 | 30 | 20 | 10 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  | 1 (ppm) |  |  |  |  |  |  |  |  |



## (3,4-dimethoxyphenyl)(4-(hexyloxy)phenyl)methanol (9b)






## (4-(cyclohexylmethoxy)phenyl)(3,4-dimethoxyphenyl)methanol (9c)






## (3,4-dimethoxyphenyl)(4-isopropoxyphenyl)methanol (9e)





## (3,4-dimethoxyphenyl)(4-propoxyphenyl)methanol (9f)




tert-butyl 4-(2-(4-((3,4-dimethoxyphenyl)(hydroxy)methyl)phenoxy)ethyl)piperidine-1carboxylate (9g)* Spectra will be in Jalisa Ferguson's information.

## Class III: Meta-phenolic ether benzhydrol analogues

3-isobutoxybenzaldehyde (16a)



## 3-isopropoxybenzaldehyde (16b)




## 3-propoxybenzaldehyde (16c)




## 1-(allyloxy)-3-bromobenzene (19a).




## 1-bromo-3-(hexyloxy)benzene (19b)*

## (3,4-dimethoxyphenyl)(3-isobutoxyphenyl)methanol (17a)






## (3,4-dimethoxyphenyl)(3-isopropoxyphenyl)methanol (17b)





## (3,4-dimethoxyphenyl)(3-propoxyphenyl)methanol (17c)






## (3-(allyloxy)phenyl)(3,4-dimethoxyphenyl)methanol (17d)





(3,4-dimethoxyphenyl)(3-(hexyloxy)phenyl)methanol (17e)* Spectra will be in Jalisa Ferguson's information.

## Class IV : Ortho-benzyl ether benzhydrol analogues.

1-bromo-2-(phenoxymethyl)benzene (21a)



## tert-butyl 4-(((2-bromobenzyl)oxy)methyl)piperidine-1-carboxylate (21b)




## tert-butyl 4-(2-((2-bromobenzyl)oxy)ethyl)piperidine-1-carboxylate (21c)




## (2-(phenoxymethyl)phenyl)(3,4,5-trimethoxyphenyl)methanol (22a)*




CNMR was obtained from Jalisa Holmes.
tert-butyl 4-(((2-(hydroxy(3,4,5-trimethoxyphenyl)methyl)benzyl)oxy)methyl)piperidine-1carboxylate (22b)




tert-butyl 4-(2-((2-(hydroxy(3,4,5-trimethoxyphenyl)methyl)benzyl)oxy)ethyl)piperidine-1carboxylate (22c)




## Class V: Tri-substituted phenolic ether benzhydrol analogues

## 1-(allyloxy)-4-bromo-2-methylbenzene (24a)




## 4-bromo-1-isobutoxy-2-methylbenzene (24b)




## 4-bromo-1-isopropoxy-2-methylbenzene (24c)




## 4-bromo-2-methyl-1-propoxybenzene (24d)




## 4-bromo-1-(hexyloxy)-2-methylbenzene (24e)




## 4-bromo-1-(cyclohexylmethoxy)-2-methylbenzene (24f)




## (4-(allyloxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25a)






## (4-isobutoxy-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25b)






## (4-isopropoxy-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25c)






## (3-methyl-4-propoxyphenyl)(3,4,5-trimethoxyphenyl)methanol (25d)






## (4-(hexyloxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25e)






## (4-(cyclohexylmethoxy)-3-methylphenyl)(3,4,5-trimethoxyphenyl)methanol (25f)






