Technical University of Denmark



Visualizing infections and immune mechanisms in zebrafish

Jørgensen, Louise ; Korbut, Rozalia; Mehrdana, Foojan; Kania, Per Walter; Schmidt, Jacob Günther; Otani, Maki; Buchmann, Kurt

Publication date: 2017

Document Version Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA):

Jørgensen, L. V. G., Korbut, R., Mehrdana, F., Kania, P. W., Schmidt, J. G., Otani, M., & Buchmann, K. (2017). Visualizing infections and immune mechanisms in zebrafish. Abstract from Bioimaging workshop, Copenhagen, Denmark.

DTU Library Technical Information Center of Denmark

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from the public portal for the purpose of private study or research.

- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Visualizing infections and immune mechanisms in zebrafish

Louise von Gersdorff Jørgensen *, Rozalia Korbut, Foojan Mehrdana, Per Walter Kania, Jacob Günther Schmidt, Maki Ohtani and Kurt Buchmann

Laboratory of Aquatic Pathobiology, Department of Veterinary and Animal Sciences, University of Copenhagen, Denmark

The zebrafish has become an important model to study a wide spectrum of vertebrate biological processes. It is used within all major biological sciences and has been especially valuable within developmental biology and genetics. Within the last 20 years the amount of papers describing zebrafish as a model has increased dramatically. It is a valuable model because of several characteristics: it is a vertebrate, it has external embryological development and is fully transparent for the first 24 h post hatching, development is fast; the vertebrate organs can be identified within 48 hours (ears, eyes, brain, internal organs), embryos are simple to genetically manipulate, the genome has been mapped, thousands of transgenic lines exist and a pair of breeders are able to produce up to 300 eggs per week making it a high-throughput but low-cost model organism. Unique tools for non-invasive in vivo imaging has been developed and high quality intravital microscopy can be conducted without comparison. A transparent line has also been developed where transparency is maintained throughout the lifespan of the fish facilitating in vivo visualization of e.g. the spread of fluorescence-tagged cancer cells, immunological reactions during e.g. transplant rejections or the spread and pathogenicity of pathogens. We have, in our laboratory, used the zebrafish as a model for aquacultured fish species and their pathogens. We have 1) visualized antigen uptake in vivo following a bath in a soup containing fluorescent inactivated bacteria, 2) visualized the spread of a bacterium after either a bath or an injection in the peritoneal cavity, and 3) visualized (images + videos) the behaviour of certain immune cells, called neutrophils during a parasite disease. Using adult fish of a transgenic line with GFP-tagged neutrophils, we got an unprecedented view into the interactions between the parasites and the neutrophils.