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Impact of environmental conditions on biomass yield, quality, and bio-mitigation capacity of *Saccharina latissima*

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ABSTRACT: Seaweeds are attractive as a sustainable aquaculture crop for food, feed, bioenergy and biomolecules. Further, the non-value ecosystem services of seaweed cultivation (i.e. nutrient recapture) are gaining interest as an instrument towards sustainable aquaculture and for fulfilling the aims of the EU Marine Strategy Framework Directive. Environmental factors determine the yield and quality of the cultivated seaweed biomass and, in return, the seaweed aquaculture affects the marine environment by nutrient assimilation. Consequently, site selection is critical for obtaining optimal biomass yield and quality and for successful bio-mitigation. In this study, 5 sites for cultivation of *Saccharina latissima* were selected within a eutrophic water body to guide site selection for future kelp cultivation activities. Results were coupled to marine monitoring data to explore the relationship between environmental conditions and cultivation success. The biomass yields fluctuated 10-fold between sites due to local variations in light and nutrient availability. Yields were generally low, i.e. up to 510 g fresh weight (FW) per meter seeded line; however, the dry matter contents of protein and high-value pigments were high (up to 17 % protein and 0.1 % fucoxanthin). Growth performance, biomass quality and bio-mitigation potential was restricted by low availability of light and bioavailable phosphorus, and biofouling through juvenile suspension feeders was a critical factor at all cultivation sites. At specific sites, the tissue metal contents (Pb and Hg) exceeded the limit values for feed or food. Our results emphasize the importance of careful site selection before establishing large-scale cultivation, and stress the challenges and benefits of kelp cultivation in eutrophic waters.

KEY WORDS: Eutrophication · Limfjorden · Seaweed farming · Metals · Nitrogen · Phosphorus · Site quality · Ecosystem service

INTRODUCTION

Cultivation of macroalgae is a rapidly growing industry in a global perspective (FAO 2016). The main driver is the establishment of a production of marine-

based biomass for food, energy, protein and biomolecules (Bruton et al. 2009, Kraan 2013, Wei et al. 2013), but also exploitation of the bio-mitigation capacity of the algae is in focus (Troell et al. 1999, Castine et al. 2013, Marinho et al. 2015a). The non-use value eco-

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system service (Daly 1998) provided by cultivated algae in terms of recapturing nutrients in coastal areas is of commercial and societal interest—as compensation for increased aquaculture activities (Sanderson et al. 2012, Handå et al. 2013, Smale et al. 2013, Holdt & Edwards 2014, Marinho et al. 2015a) or as a potential instrument for circular nutrient management, improving the ecological status of eutrophic marine areas (Seghetta et al. 2016) in line with the EU Marine Strategy Framework Directive (EU 2008a, 2014).

In Europe, the effort concerning cultivation of large brown algae (Laminariales), in particular, is increasing. The most commonly cultivated brown algae species in Europe, *Saccharina latissima* ((Linnaeus) C.E. Lane, C. Mayes, Druehl & G.W. Saunders), has been cultivated on a smaller or larger scale in Ireland, Scotland, Germany, Holland, Spain, Norway, Faroe Islands and Denmark (i.e. Buck & Buchholz 2004, 2005, Buck et al. 2008, Werner et al. 2009, Wegeberg 2010, Edwards & Watson 2011, Forbord et al. 2012, Handå et al. 2013, Peteiro & Freire 2013b, Wegeberg et al. 2013, Marinho et al. 2015a). The achieved biomass yields and the biochemical composition of the biomass vary considerably seasonally and spatially, primarily because of different environmental conditions (Edwards & Watson 2011, Handå et al. 2013, Peteiro & Freire 2013a,b, Marinho et al. 2015a). In return, the algae production also exerts an impact on the environmental conditions through the removal of nutrients (Troell et al. 1999, Stephens et al. 2014, Marinho et al. 2015a). Thus, algae cultivation sites should be carefully selected for optimizing biomass production, biomass quality as well as the non-value ecosystem services (Kerrison et al. 2015). Further, the seasonal timing of the deployment and harvest of the algae needs to be optimized according to local environmental conditions. The focus of the optimization, i.e. high protein yield or high carbohydrate yield, will depend on the final application of the biomass. Sporophytes of *Laminaria* species store nutrients for length growth during periods when environmental nutrients concentrations are high (Bartsch et al. 2008). Thus, high environmental nutrient concentrations favour high tissue nitrogen (N) concentrations: in wild *S. latissima* up to 3.5% N of dry matter (DM) (Gevaert et al. 2001, Nielsen et al. 2014), up to 5.0% N of DM when cultivated in close proximity to fish aquaculture (Handå et al. 2013, Marinho et al. 2015a) and even up to 6.7% N of DM when cultivated under highly eutrophic conditions (Nielsen 2015). The molar N:phosphorus (P) ratio is commonly in the range of 9–25:1 (Atkinson & Smith 1983), and P con-

centrations of up to 0.8% of DM are reported in nutrient-rich waters (Marinho et al. 2015a). High tissue N concentrations reflect a correspondingly high content of proteins (Manns et al. 2014, Marinho et al. 2015b, Angell et al. 2016). Consequently, both the biomass quality and the bio-mitigation capacity of the produced algae increase in nutrient-rich waters, increasing the value of the biomass, as well as improving the environmental condition of the water body through harvest, and thus removal of nutrients.

Of the 21 Danish water bodies, Limfjorden receives the highest annual net supply of nutrients (8.2 t N and 0.30 t P km⁻² y⁻¹; Seghetta et al. 2016). These high nutrient loadings have caused a regime shift in the fjord from benthic to pelagic primary production (Krause-Jensen et al. 2012). The high pelagic primary production supports a substantial stock of benthic suspension feeders, including blue mussels *Mytilus edulis* L., supporting a local mussel fishery (Maar et al. 2010, Timmermann et al. 2014). Mussel farming has been successfully tested as an instrument to recapture nutrients and improve the ecological status of Limfjorden (Petersen et al. 2014), and farming of long-line blue mussels is an emerging business in Limfjorden. Along with the development of the mussel farming industry, interest in macroalgae cultivation is increasing, partly because the 2 crops may be cultivated using the same structures (Nielsen 2015). Due to the high environmental nutrient concentrations, cultivation of large brown algae in a water body like Limfjorden would theoretically hold a potential for the production of a *Saccharina* biomass with high protein content, representing a higher value for the food or feed market. At the same time the potential of seaweed cultivation as an instrument for circular nutrient management would be maximized. Despite the relatively small size of Limfjorden (1500 km²), local environmental conditions differ considerably between the different basins (Maar et al. 2010). Cultivation of *S. latissima* has to date been documented only once at 1 site in Limfjorden, indicating a potential for cultivation of *S. latissima*. This study, however, also demonstrates the need for investigating optimal timing of cultivation and harvest in order to maximize biomass yield and avoid biofouling (Wegeberg 2010).

Testing and evaluating the interactions between local environmental conditions and biomass yield, quality and potential for bio-mitigation through nutrient recapture of cultivated kelps in coastal waters is needed before implementing cultivation on a larger scale. This applies not only to Limfjorden, but to any water body where macroalgae cultivation is intended.

The aim of this study was to compare the biomass yield, bio-mitigation capacity and nutritional quality for food and feed of *S. latissima* cultivated at 5 sites in Limfjorden as well as to explore the influence of local environmental conditions on these parameters with the purpose of guiding site selection and timing of harvest. The 5 selected cultivation sites each represented their basin in Limfjorden, with the basins characterized by different environmental conditions regarding salinity, turbidity, nutrient availability and sediment metal concentrations.

MATERIALS AND METHODS

Study area and cultivation sites

Limfjorden is a shallow, semi-enclosed estuary located between the North Sea and the Kattegat (Fig. 1). The total surface area of the fjord is ~1500 km² and the average depth is 4.6 m. The total catchment area is 7587 km² and is predominantly agricultural land. Despite a small tidal amplitude, tidal forces and wind are the drivers of the annual net flow of 6.8 km³ of water from the North Sea via the Thyborøn channel in the west through Limfjorden to the Kattegat. Limfjorden consists of several relatively shallow water basins connected by narrow and deep sounds. The big broads have water depths of 5–8 m, whereas the sounds have depths of 18–22 m, the deepest point being Oddesund (28 m). The average salinity varies from 32–34 in the western part to 19–25 in the central and eastern part (Lyngby et al. 1999, Markager et al. 2006, Krause-Jensen et al. 2012, Timmermann et al. 2014).

Five existing mussel farms were selected as experimental cultivation sites (Fig. 1, Table 1) for the

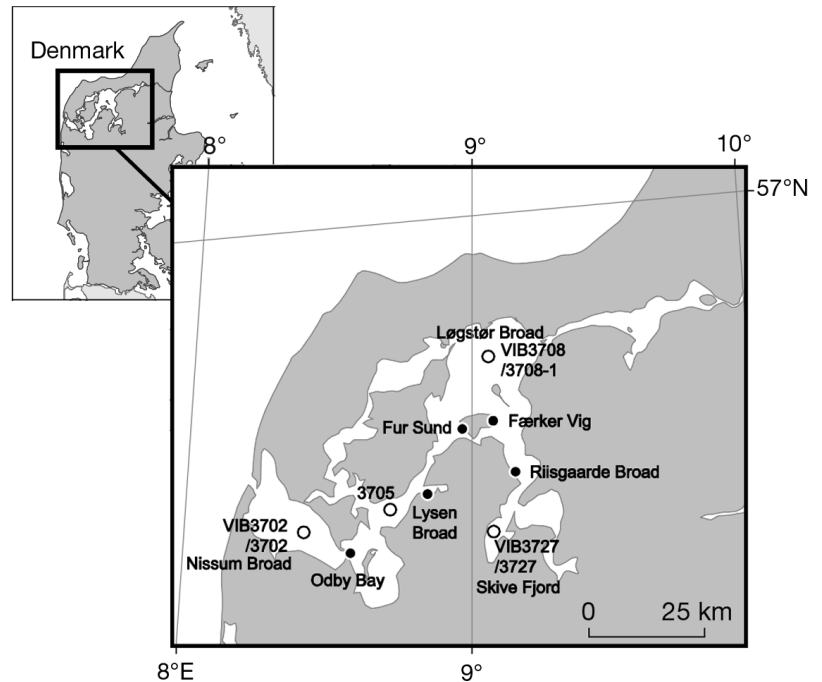


Fig. 1. Location of Limfjorden in Denmark and the 5 *Saccharina latissima* cultivation sites (filled circles) and 4 environmental monitoring stations (open circles). Stns VIB3702, VIB3708 and VIB 3727 are pelagic stations for monitoring water quality. Stns 3702, 3705, 3708-1 and 3727 are stations for monitoring benthic metal concentrations

following reasons: (1) they were each located in a distinct basin of Limfjorden; (2) aquaculture licenses were already active; (3) the mussel cultivation structures could be used for the seaweed cultivation; and (4) the 5 basins were covered by the Danish National Monitoring and Assessment Program for the Aquatic and Terrestrial Environment (NOVANA).

Environmental data

For each of the 5 cultivation sites, the data on biomass yield and quality were coupled to environmental data from an environmental monitoring station located centrally within each basin (Fig. 1, Table 2).

Table 1. Location, size and characteristics of the 5 *Saccharina latissima* cultivation sites in Limfjorden, Denmark. N: north, E: east, S: south, W: west

Site	Position		Basin	Size (m × m)	Depth (m)	Sea bed classification	Degree of exposure
	Latitude (°N)	Longitude (°E)					
Odby Bay	56.577	8.570	Nissum/Kaas Broad	300 × 300	4	Soft mud	Exposed to winds from E and S
Lysen Broad	56.692	8.841	Sallingsund	250 × 500	2–5	Fine sand with clay	Protected
Fur Sund	56.816	8.968	Fur Sund	250 × 750	5	Rocky/sandy	Exposed to strong currents
Færker Vig	56.834	9.073	Løgstør Broad	300 × 300	4	Hard sand/stone	Protected
Riisgaarde Broad	56.736	9.151	Riisgaarde Broad/ Skive Fjord	250 × 500	10	Soft mud	Exposed to winds from N, E, S

Table 2. Deployment and sampling dates at the 5 *Saccharina latissima* cultivation sites in Limfjorden, batches of seeded lines, as well as identification numbers of and distance to the environmental monitoring stations (pelagic and sediment stations, see Fig. 1) from which data were used for analyses

Site	Deployment date	Sampling 1 date	Sampling 2 date	Batch	Pelagic station	Sediment station	Distance (km)
Odby Bay	Dec 6, 2011	Apr 11, 2012	Jun 12, 2012	1	VIB3702	3702	14
Lysen Broad	Dec 6, 2011	Apr 11, 2012	Jun 12, 2012	1	– ^a	3705	8.5
Fur Sund	Dec 6, 2011	Apr 11, 2012	Jun 12, 2012	1	VIB3708	3708-1	20
Færker Vig	Oct 28, 2011	Apr 11, 2012	May 25, 2012	2	VIB3708	3708-1	14
Riisgarde Broad	Dec 6, 2011	Apr 11, 2012	Jun 12, 2012	1	VIB3727	3727	14

^aNo pelagic monitoring station was within proximity to the cultivation site at Lysen Broad

Data from the monitoring stations were retrieved from NOVANA through the National Database for Marine Data (ODAM) (Fig. 1, Table 1).

Data on water temperature, salinity, turbidity and concentrations of oxygen, chlorophyll *a* (chl *a*), inorganic nutrients (dissolved inorganic N [DIN = NO₂⁻-N, NO₃⁻-N, NH₄⁺-N], dissolved inorganic bioavailable P [ortho-P]) and sediment metals were collected and analysed using standard methods according to the current national Technical Instructions for Marine Monitoring (Markager 2004, Pedersen et al. 2004, Larsen 2013, Markager & Fossing 2013, Vang 2013, Vang & Hansen 2013). Sampling was performed on average every 2–3 wk. Sampling of sediment was performed every 1–5 yr. By trapezoidal integration, all pelagic environmental data were calculated into weighted averages over 2 periods up to the time point of each biomass sampling—early spring: the period of detectable growth from 1 February 2012 to Sampling 1, 11 April 2012; and late spring: the last part of the grow-out period from Sampling 1 (11 April 2012) to Sampling 2 (25 May or 12 June 2012) (see next section and Table 2).

Data regarding temperature, salinity and turbidity were differentiated according to the actual cultivation depths (1.5 and 2.5 m, respectively). Data regarding nutrients, oxygen and chl *a* were only available from 1 m of depth, but no significant stratification prevailed during the cultivation period. Sediment metal concentration data were averaged for each station over a period covering the preceding 10 yr (2003–2012). Data on local incoming light was supplied from the Danish Meteorological Institute.

Ideally, cultivation sites and monitoring stations could have been geographically closer. However, the data from the environmental monitoring stations was considered as being representative for the cultivation sites despite the distances of 8–20 km between monitoring station and cultivation site for a number of

reasons: (1) other studies correlating monitoring data and macrovegetation performance in Limfjorden generally achieve good correlations (e.g. Krause-Jensen et al. 2012), (2) the experimental period from winter to early summer is a period of maximal wind-driven circulation (Wiles et al. 2006), and absence of vertical stratification (Christiansen et al. 2006), (3) mixing was confirmed as no stratification was observed during the experimental period, and (4) sites and stations were located in the more open parts of the basins in proximity to point sources of run-off from land. Coupling of biomass yield and quality to environmental data for the cultivation site at Lysen Broad was not possible, as only data on sediment chemistry was available from the environmental monitoring station in this basin.

Cultivation and sampling of *Saccharina latissima*

Two batches of *S. latissima* seeded lines were used in the cultivation experiment (Table 2). Batch 1 consisted of 500 m of ready-made seeded line (diameter: 6 mm) produced by direct sporulation (Wegeberg 2010) at Blue Food A/S, Denmark. This batch was delivered to the Danish Shellfish Centre on 5 December 2011, kept in running seawater overnight and deployed the following day at 4 sites: Odby Bay, Lysen Broad, Fur Sund and Riisgarde Broad. Batch 2 was deployed at Færker Vig and consisted of 125 m of seeded line (diameter: 6 mm), also produced through direct sporulation, but at the Danish Shellfish Centre during August 2011. Both batches were produced from fertile material from a *S. latissima* population in the Danish Belt Sea, and visual inspection of the lines upon deployment did not reveal any difference between the 2 batches in quality, density or size of the juvenile sporophytes. Length of the seedlings at deployment was ~1 mm. All lines were

deployed as vertical droppers, each 2.5 m long, attached to a horizontal long-line with a 50 cm tethering line. The droppers were interspaced by 40 cm along the horizontal long-line. During the grow-out period, the horizontal long-lines were kept 50 cm below the water surface to avoid disturbance by floating ice and heavy storms. Consequently, the seeded lines were positioned between 1 and 3.5 m depth.

Sampling of biomass was performed twice by random selection of 3 droppers from each site: Sampling 1 on 11 April 2012 and Sampling 2 on either 25 May or 12 June 2012 (Table 2). The lines were brought to the laboratory where the upper 2 m of each line was divided into 2 sections: the upper section represented the seeded line hanging in 1–2 m depth (average 1.5 m), and the lower section representing the seeded line hanging at a depth of 2–3 m (average 2.5 m). The remaining 50 cm of each seeded line with the attached bottom weight was discarded due to lack of biomass. The following parameters were recorded for both sections of the lines: total weight of sample (seeded line + algae + epiphytes), weight of seeded line, weight of algae, weight and taxonomy of dominating biofouling epiphytic organisms, and finally, average sporophyte frond length, based on 15 randomly selected sporophytes. After sampling, tissue samples were stored at -20°C until biochemical analyses were performed. Due to increasingly heavy biofouling by epiphytic organisms over time, algae material harvested from late May and onwards (Sampling 2) was fully covered with epiphytic organisms such as ascidians and juvenile mussels, and thus considered unsuitable for food or feed applications. Therefore only algae material sampled in April (Sampling 1) was used in the biochemical analyses. Due to very limited biomass harvested from Fur Sund at 2.5 m in April, only pigment analyses were performed on this biomass.

Calculations of growth rates and biomass yields

Specific growth rates (SGRs) were calculated from measurements of the fresh weight (FW) per running meter of seeded line as:

$$\text{SGR}(\%) = 100 \times \frac{\ln\left(\frac{\text{FW}_t}{\text{FW}_0}\right)}{t} \quad (1)$$

where FW_0 and FW_t corresponded to the fresh weight of *S. latissima* per m of seeded line at time 0 and after t days of cultivation, respectively. Biomass yields were reported as g FW per m of seeded line (g

FW m^{-1}). The average frond length of the *S. latissima* sporophytes was calculated as an average length \pm SE of the 15 randomly selected sporophytes from each sample.

Saccharina latissima tissue biochemistry

DM, ash, carbon (C), N and P. Algae samples were freeze-dried at -40°C and homogenized by dry milling. DM content was calculated as percentage of FW. A known amount of dry algae was combusted at 550°C for 2 h, and the ash fraction was calculated as percentage of DM. Concentrations of C and N in the freeze-dried algae tissue were analysed by Pregl-Dumas ignition in pure oxygen atmosphere followed by chromatographic separation of C and N with detection of the individual elements by thermal conductivity (Culmo 2010). Total P content of the algae biomass was as analysed spectrophotometrically according to standard methods (Grasshoff et al. 1983). Prior to analysis, the dried and homogenized samples were heated at 550°C for 2 h, autoclaved with 2 M hydrogen chloride (HCl) (20 mg DM for 7 ml acid), and finally filtered through GFF filters (Whatman).

Metals. Metal concentrations (As, Cd, Hg, Pb) were determined by inductively coupled plasma-mass Spectrometry (ICP-MS). In short, a 0.2 g dry subsample was digested in a closed vessel microwave oven using 5 ml of nitric acid (7 M) and 1 ml of hydrogen peroxide, then diluted to 50 ml with milliQ water, followed by ICP-MS determination using internal standards of Rh, Ir and Ge to correct for drift (see Nielsen et al. 2012). Certified reference material of macroalgae from IAEA-140 (Coquery et al. 2000) was used for quality assurance.

Pigments. Pigment concentrations (chl *a*, fucoxanthin, violaxanthin and β -carotene) were determined using acetone extraction and quantification by HPLC as described in Boderskov et al. (2016). Pigment standards were obtained from DHI Laboratory Products.

Crude protein and amino acids (AAs). Crude protein and AA composition were analysed only for samples from Færker Vig. Total organic bound crude protein was determined by the Kjeldahl principle according to Nordic Committee on Food Analysis (2005). Protein content was calculated by multiplying the amount of N by a factor of 5 and expressed as percent of DM (Angell et al. 2016). The determination of AAs was done by HPLC according to EU 152/2009 (A) and ISO 13903:2005. AA contents were expressed as percentage of DM.

Data analysis

For comparing growth performance and biomass quality between sites and depths, 2-way ANOVA (using Tukey's post hoc analysis) and linear regression analyses were performed using JMP 10.0 (SAS Institute). Explorative data analysis was performed to identify significant correlation patterns between macroalgae growth and environmental parameters. Data were log transformed in order to obtain normal distribution and homogeneity of variance for the residuals of the models. Multivariate data analysis (MVDA) was performed to guide model selection of variables to be tested using general linear models (GLM). Partial least square regression (PLS-R) was used as explorative technique for pattern recognition using the Unscrambler v.10.2 (CAMO Software). Biomass yield and biofouling in early and late spring as well as bio-mitigation capacity, i.e. N and P content in the harvested seaweed biomass, were selected as Y-variables in the PLS-R models and modelled using environmental parameters, characterising the marine growth environment surrounding the individual cultivation sites, as original explanatory variables (data not shown). GLMs were used to assess the effect of light, salinity, availability of ortho-P, temperature and environmental N:P ratio (NP_E) on growth performance, biofouling, and biomass quality. The environmental parameters were selected as independent variables based on the indicative impact on the dependent variable (biomass growth parameters and quality), as observed from MVDA (data not shown). As several of the independent variables showed strong correlations (Pearson, Table S1 in the Supplement at www.int-res.com/articles/suppl/q008p619_supp.pdf), the independent variables were split into 2 models to avoid issues with collinearity—Model 1: light, salinity, and ortho-P; Model 2: temperature, salinity and NP_E. These analyses were performed in SAS 9.3 (SAS Institute) using the Proc mixed function with cultivation site as a random factor. The level of significance applied was 0.05, unless mentioned otherwise.

RESULTS

Environmental conditions

The environmental conditions differed among the basins of Limfjorden (Fig. 2, Table 1). Differences were most pronounced with regard to salinity, light, and concentrations of inorganic nutrients and chl *a*.

Salinity

The salinity in the different basins decreased with increasing distance from the North Sea: Nissum Broad, 29.0–31.9; Løgstør Broad, 26.0–28.9; and Skive Fjord, 23.5–26.5. In Nissum Broad, the salinity increased slightly over the grow-out period, whereas in Løgstør Broad and Skive Fjord the salinity decreased over the period, reflecting a stronger influence of run-off from land (Fig. 2A). No pronounced stratification of the water column was observed from the monitoring data during the grow-out period at any of the stations (data not shown).

Temperature

Generally, the differences in temperature among stations were minor ($<1^{\circ}\text{C}$), and even less between the 2 cultivation depths at any station. The temperatures experienced during the full grow-out period ranged from minimum temperatures in all basins measured on 1 February (between -0.2 and 1.5°C) to maximum temperatures in June (13.8 – 14.1°C) (Fig. 2B).

Inorganic nutrients

The average concentrations of DIN from deployment to April ranged between 20 and 40 μM ; however, with concentrations up to 58 μM in Skive Fjord in winter and early spring (Fig. 2C). In late spring, between April and June, the DIN concentrations decreased <2 μM in Nissum Broad, but remained high between 10 and 20 μM in the other basins. In all periods, the highest DIN concentrations were measured near Skive Fjord and the lowest in Nissum Broad. Concentrations of ortho-P were high during the winter period (0.4 – 0.9 μM), but decreased below 0.1 μM during the spring bloom from February to April (Fig. 2D).

Pelagic chl *a*

In early spring, February and March, the phytoplankton concentrations peaked with 12 and 16 μg chl *a* l^{-1} in Løgstør and Nissum Broad, respectively (Fig. 2E). In Skive Fjord, the highest chl *a* concentrations were measured in early June (14 μg chl *a* l^{-1}).

Light

The photon flux density generally decreased by $\sim 50\%$ from 1.5 to 2.5 m, emphasizing the high turbidity of Limfjorden (Fig. 2F). The algae at 1.5 m experienced an average of 400–700 μmol photons m^{-2} s^{-1} in late

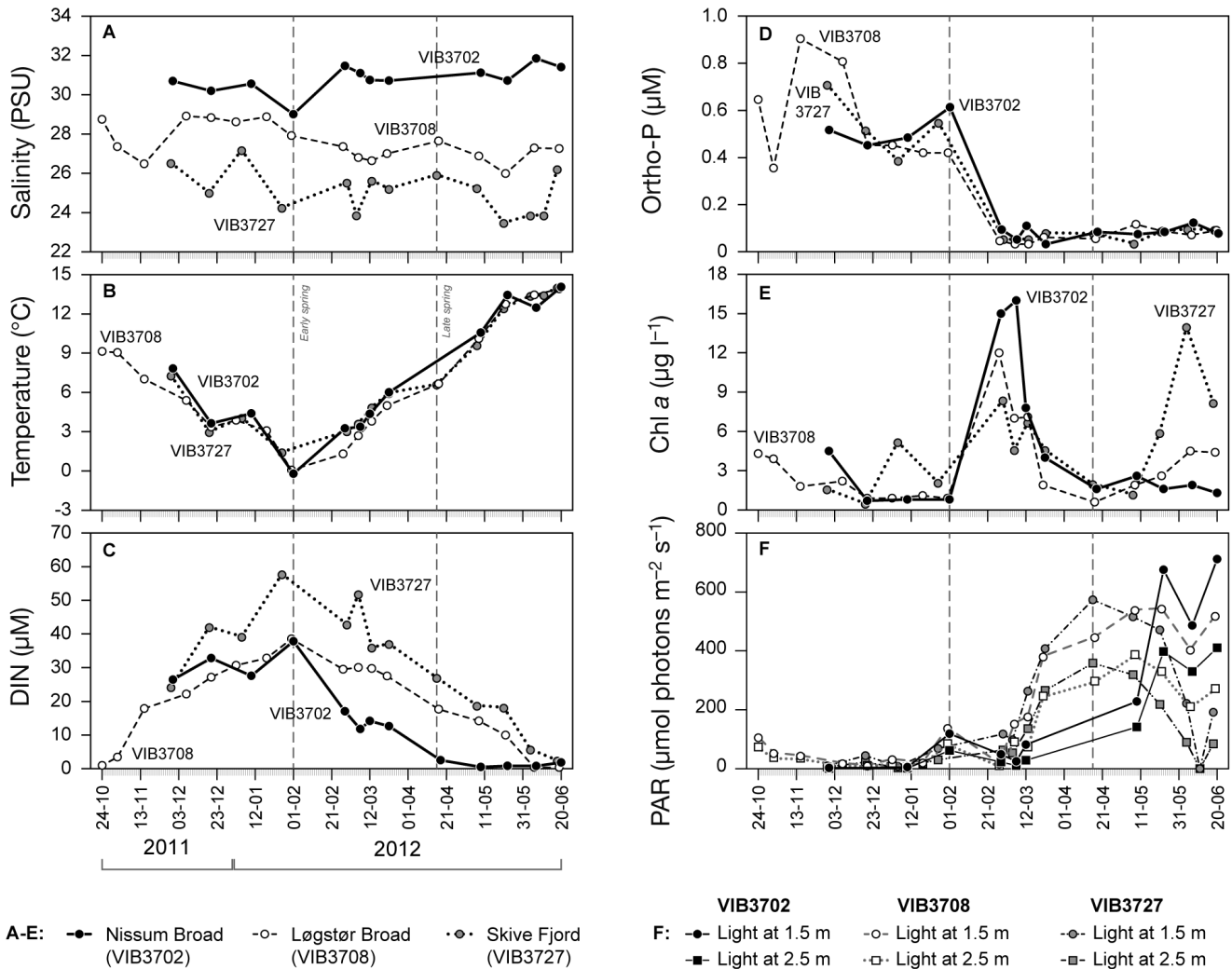


Fig. 2. Seasonal pattern of selected environmental parameters at the 3 pelagic stations (see Table 2) during the grow-out period of *Saccharina latissima* (October–December 2011 to June 2012): (A) salinity, (B) temperature, (C) dissolved inorganic nitrogen (DIN), (D) ortho-phosphate, (E) chlorophyll *a* and (F) photosynthetically active radiation (PAR) estimated at the cultivation depths (1.5 and 2.5 m). Data represent the actual measured values

spring, whereas the algae at 2.5 m only experienced up to 400 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in the same period.

Overall, a high degree of inter-correlation between the key environmental parameters was observed (Table S1 in the Supplement). In early spring (February to April) the concentration of phytoplankton biomass (chl *a*) correlated strongly to the concentrations of dissolved inorganic nutrients. During early spring, the concentrations of pelagic chl *a* correlated positively to ortho-P, and negatively to DIN concentrations, whereas light availability correlated negatively to ortho-P concentrations. The DIN concentrations were negatively correlated to salinity. In late spring, the pelagic phytoplankton biomass was negatively correlated to salinity and positively to temperature.

Saccharina latissima growth performance

Biomass yield, frond length, SGR and biofouling

At all cultivation sites, the biomass yields and frond lengths were higher at 1.5 m than at 2.5 m depth (Fig. 3A,B, Table 3). The highest biomass yield in April (mean \pm SE: $510 \pm 66 \text{ g FW m}^{-1}$) as well as the longest fronds in April and June ($40.9 \pm 3.7 \text{ cm}$ in April and $33.7 \pm 9.0 \text{ cm}$ in June) were achieved at Færker Vig at 1.5 m (Fig. 3A,B, Table 4). In June, the biomass yield in Odby Bay and Færker Vig at 1.5 m was significantly higher than at the remaining 3 sites (Table 4). At 2.5 m, the highest biomass yield in June was obtained in Odby

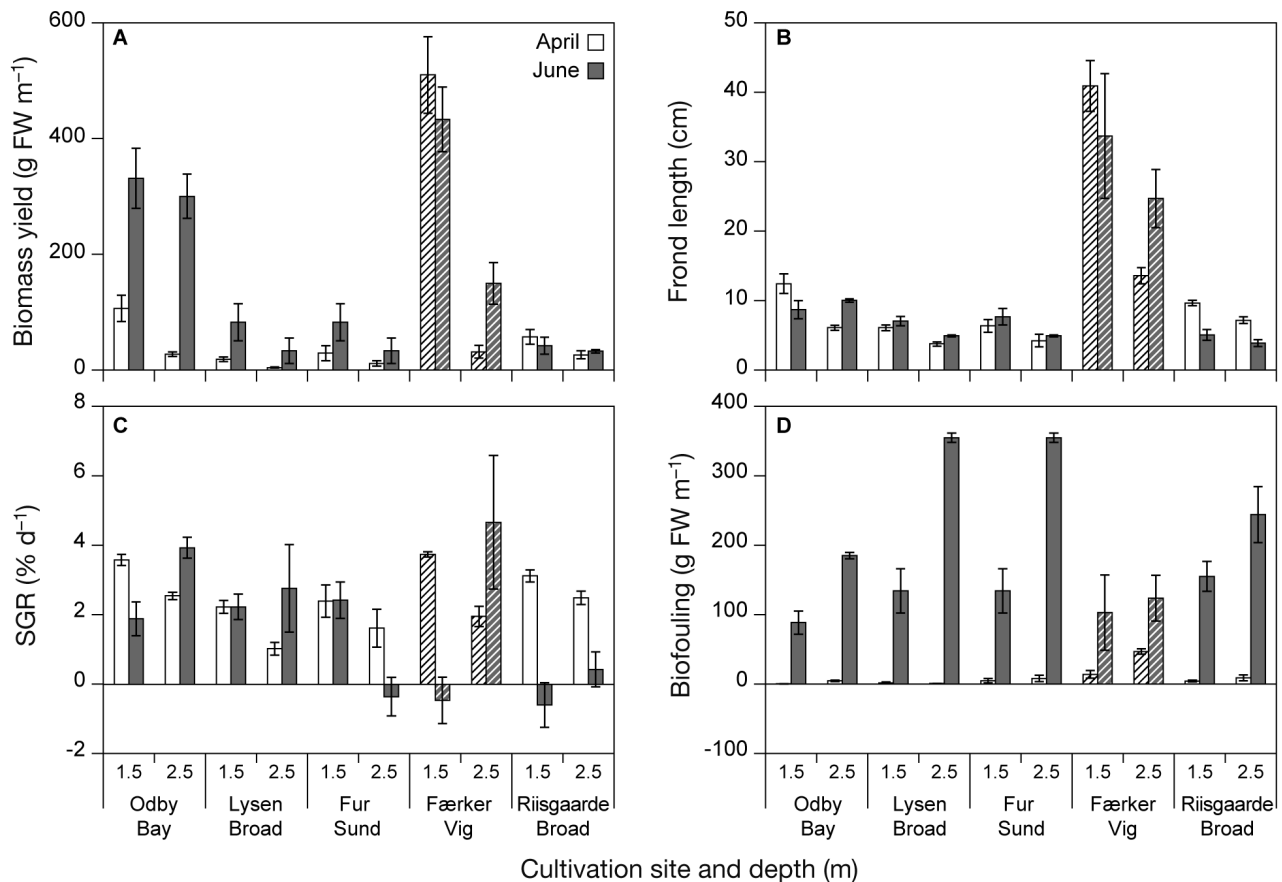


Fig. 3. Growth performance and biofouling of *Saccharina latissima* at the 5 cultivation sites (see Fig. 1): (A) biomass yields, (B) average frond lengths ($n = 15$), (C) specific growth rates (SGR), and (D) biofouling of *S. latissima* at 1.5 and 2.5 m depths at the 5 cultivation sites sampled in April (white bars) and June (grey bars), respectively. Solid bars represent batch 1 of seeded lines, and crossed bars represent batch 2. Data represent means \pm SE, $n = 3$

Bay, whereas the longest fronds were found in Færker Vig (Fig. 3A,B, Table 4).

The SGR (in the period from deployment to April) reflected the same pattern as the biomass yield at 1.5 m depth: Færker Vig ($3.8\% \text{ d}^{-1}$) > Odby Bay ($3.7\% \text{ d}^{-1}$) > Riisgaarde Broad ($3.1\% \text{ d}^{-1}$) > Fur Sund ($2.4\% \text{ d}^{-1}$) > Lysen Broad ($2.2\% \text{ d}^{-1}$), and all with significantly higher SGRs at 1.5 m as compared to 2.5 m (Fig. 3C, Table 3). However, from April to June, the SGR decreased for the algae nearest to the surface (1.5 m) at Odby Bay, Færker Vig and Riisgaarde Broad and at the 2 latter sites to negative values. At Lysen Broad and Fur Sund, the SGR of the algae near the surface was constant throughout the full growth-out period. Regarding the algae growing at 2.5 m from April to June, diverging trends were observed: at 3 cultivation sites (Odby Bay, Lysen Broad and Færker Vig) the SGRs exceeded the SGRs at 1.5 m in the early growth period, whereas at the other 2 sites (Fur Sund and Riisgaarde Broad), the SGRs decreased to around or below zero.

The degree of biofouling increased dramatically at all sites from April to June, and was in June significantly higher at 2.5 m than at 1.5 m depth, with the one exception of Færker Vig (Fig. 3D). In June, the biomass yield of biofouling organisms (predominantly hydroids, juvenile *M. edulis* and ascidians) exceeded the biomass yields of *S. latissima* at 3 sites (Lysen Broad, Fur Sund and Riisgaarde Broad) at both depths (Fig. 3A,D).

In early spring, the growth performance (biomass increase [Fig. 4A], length growth and SGR) was positively correlated to the light availability, with also salinity and ortho-P availability being positively correlated to length growth and SGR (Fig. 4B, statistics are provided in Table S2 in the Supplement), respectively. The total biomass yield in June was negatively correlated to the degree of biofouling in late spring (linear regression, $p = 0.003$, $R^2 = 0.28$). (Fig. 4C). The biofouling in late spring was positively correlated to the sea temperature at the cultivation depth between April and June (Fig. 4D, Table S2).

***Saccharina latissima* biomass quality**

DM, tissue N and P

The DM content of the algae varied between 6.3 and 16.8% of fresh weight (Fig. 5A). The C content generally ranged between 26.8 and 33.4% of DM, except at Fur Sund, where the C content was significantly lower (15.3–20.5% of DM). At Odby Bay and Fur Sund, the tissue C concentrations were significantly higher in the biomass closest to the surface. This was not the case at the other sites (Fig. 5B, Table 3).

The tissue N concentration in April was significantly higher in biomass from Odby Bay (4.5% of DM) than from any of the other cultivation sites (3.5–4.0% of DM) (Fig. 5C, Table 4). Only at Riisgaarde Broad was there a significantly higher N concentration in the algae cultivated at 2.5 m than at 1.5 m depth. The tissue P content was significantly higher in the algae cultivated at 2.5 m than at 1.5 m in Odby Bay and Riisgaarde Broad, where the P content in the algae from 2.5 m was up to 0.28% of DM compared to 0.11% of DM at 1.5 m (Fig. 5D).

The bio-mitigation capacity of N and P varied between sites and depths from (mean \pm SE) 0.02 ± 0.01 to 1.84 ± 0.24 g N m⁻¹ and 0.001 ± 0.0004 to 0.05 ± 0.01 g P m⁻¹, respectively (Fig. 5E,F), reflecting predominantly the large fluctuations in biomass yields (Fig. 3A).

The environmental concentration of ortho-P was positively related to the tissue DM and N contents (Fig. 4B), while not related to the tissue P content

Table 3. Dependency of *Saccharina latissima* growth performance and biomass quality on cultivation site and depth, as well as the interaction between the two. p-values from 2-way ANOVA, data were log transformed prior to analysis. Statistical significance ($p > 0.05$) is indicated in **bold**. A: April, J: June. (–) designates a negative correlation, otherwise the correlation is positive

Parameter	Cultivation site \times Depth	Cultivation site	Depth
Growth performance			
Yield (g m ⁻¹) (A)	0.064	<0.001	<0.001(–)
Yield (g m ⁻¹) (J)	0.336	<0.001	0.004(–)
Length (cm) (A)	0.014	<0.001	<0.001(–)
Length (cm) (J)	0.398	<0.001	0.036(–)
SGR (% d ⁻¹) (A)	0.535	0.003	<0.001
SGR (% d ⁻¹) (J)	0.057	0.025	0.118
Biofouling (g m ⁻¹) (A)	0.417	<0.001	0.097
Biofouling (g m ⁻¹) (J)	0.937	0.058	0.003
Biomass quality (A)			
DM (% FW)	0.021	<0.001	0.643
C (% DM)	0.030	<0.001	<0.001(–)
N (% DM)	0.001	<0.001	0.959
P (% DM)	0.006	0.008	<0.001
Chl a (mg g DM ⁻¹)	0.029	0.005	<0.001
Fucoxanthin (mg g DM ⁻¹)	0.433	0.006	<0.001
Violaxanthin (mg g DM ⁻¹)	0.302	0.003	0.004
Beta-carotene (mg g DM ⁻¹)	0.316	0.026	0.005
As (mg kg DM ⁻¹)	0.249	<0.001	0.003
Hg (mg kg DM ⁻¹)	0.057	0.050	0.761
Pb (mg kg DM ⁻¹)	0.003	<0.001	<0.001
Cd (mg kg DM ⁻¹)	0.926	<0.001	0.831

(Table S2). Temperature was positively correlated to the tissue DM, N and P contents (Table S2). Light availability correlated positively to the tissue C content, but negatively to P content (Table S2).

Table 4. Tukey post-hoc pairwise comparisons of *Saccharina latissima* growth performance in April and June, and tissue biochemistry in April among cultivation sites at 1.5 and 2.5 m depth. Different letters are assigned to significantly different results. Fuco: fucoxanthin, Viola: violaxanthin, β -car: β -carotene

Site	Yield (Apr)	Yield (Jun)	FronD length (Apr)	FronD length (Jun)	Biofou-ling (Apr)	Biofou-ling (Jun)	DM	N	P	C	Chl a	Fuco	Viola	β -car
1.5 m depth														
FærkerVig	A	A	A	A	A	A	B	B	A	B	AB	B	AB	A
Odby Bay	B	A	B	B	B	A	AB	A	A	A	A	A	A	A
Riisgaarde Broad	B	B	B	B	AB	A	A	B	A	A	A	B	B	A
Fur Sund	B	B	B	B	AB	A	C	B	A	C	B	B	–	A
Lysen Broad	B	B	B	B	AB	A	A	B	A	A	A	AB	AB	A
2.5 m depth														
Færker Vig	A	B	A	A	A	C	B	B	B	B	BC	ABC	AB	A
Odby Bay	A	A	BC	B	B	BC	A	A	A	AB	A	A	A	A
Riisgaarde Broad	A	B	B	B	B	B	A	A	AB	A	AB	AB	AB	A
Fur Sund	A	B	BC	B	B	A	B	C	B	C	C	C	–	A
Lysen Broad	A	B	C	B	B	A	–	–	–	–	BC	BC	B	A

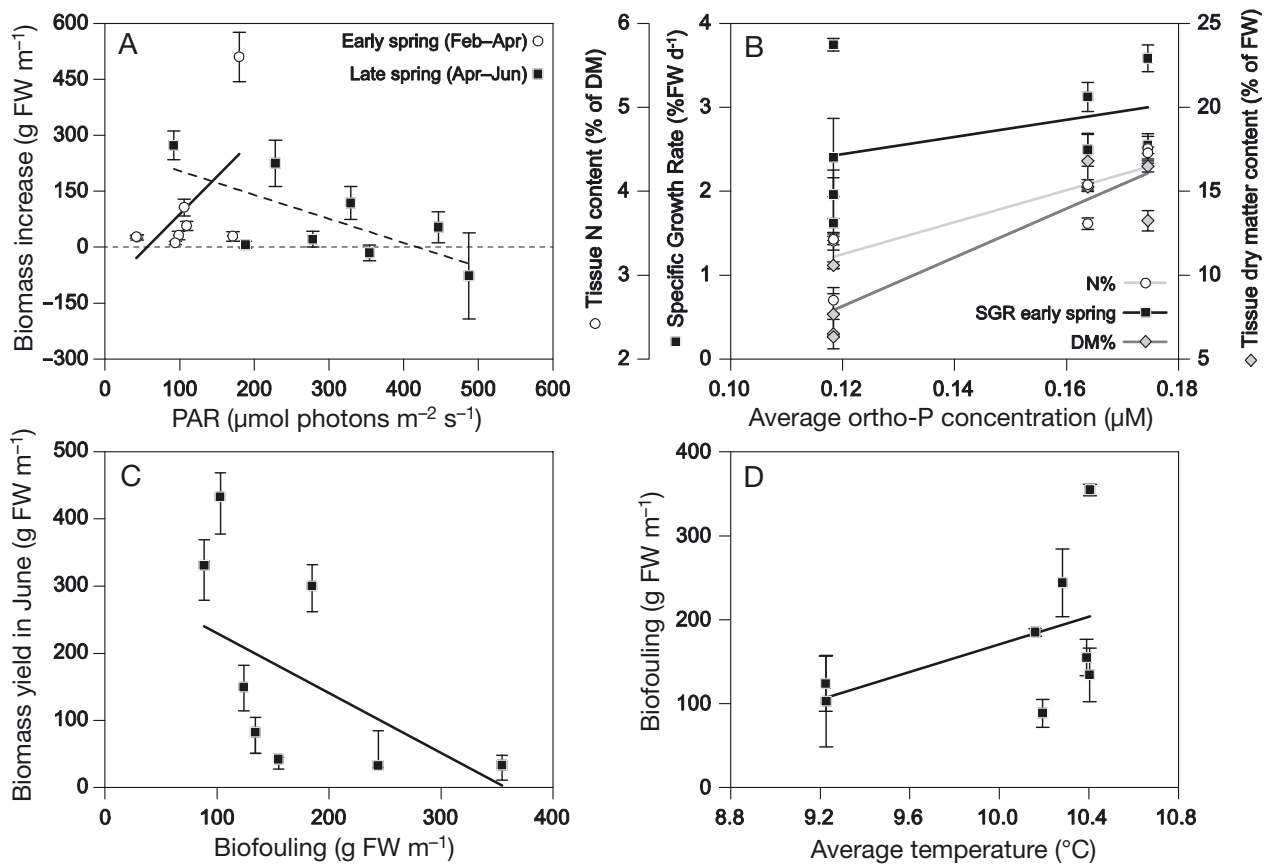


Fig. 4. Correlations between environmental parameters and *Saccharina latissima* biomass yields and quality. (A) Biomass increase or decrease in each of the 2 periods (early and late spring) as a function of light availability; (B) specific growth rate, tissue N and dry matter (DM) as a function of P availability in early spring; (C) biomass yield in June as a function of the biofouling in June (linear regression, $p = 0.003$, $R^2 = 0.28$); (D) biofouling in June as a function of average water temperature. Data represent means \pm SE, $n = 3$. Datapoints represent values for 4 stations (2 depths each). Significant correlations are indicated by solid lines, non-significant relations by dotted lines. Statistics for correlations in (A,B,D) are given in Table S2 in the Supplement at www.int-res.com/articles/suppl/q008p619_supp.pdf

The average molar tissue ratios of C:N and N:P were (mean \pm SE) 9.0 ± 0.3 and 69.7 ± 4.8 , respectively, indicating strong P-limitation already in early spring (data not shown).

Protein and AAs

The content of crude protein in the *S. latissima* biomass from Færker Vig was $17.0 \pm 0.2\%$ and $16.0 \pm 0.1\%$ of DM in the biomass at 1.5 m and 2.5 m, respectively. The essential AAs (EAAs) constituted 23.8 ± 0.2 and $27 \pm 1.9\%$ of the total AAs (TAAs), at 1.5 m and 2.5 m depth, respectively. The specific EAA, methionine, constituted $1.25 \pm 0.04\%$ (1.5 m) and $1.37 \pm 0.10\%$ (2.5 m) of the TAAs, whereas another EAA, lysine, constituted $3.25 \pm 0.07\%$ (1.5 m) and $4.06 \pm 0.51\%$ of TAAs (2.5 m).

Pigments

The tissue pigment contents ranged from 1.19–2.49 mg chl *a* g DM⁻¹, 0.62–1.09 mg fucoxanthin g DM⁻¹, 0.01–0.04 mg violaxanthin g DM⁻¹ and 0.01–0.03 mg β -carotene g DM⁻¹ (Fig. 6). Higher contents of fucoxanthin, violaxanthin and β -carotene were found in algae cultivated at 2.5 m depth, than at 1.5 m. Also, there was a significant difference in the content of the 3 pigments among sites (Table 3), with algae cultivated at Odby Bay yielding the highest concentrations. Regarding chl *a*, there was a significant interaction effect between site and depth, with higher concentrations of chl *a* at 2.5 m depth as compared to 1.5 m (Table 3), except at Lysen Broad, where no significant difference in the chl *a* content between cultivation depths was observed (Fig. 6A, Table 3). The

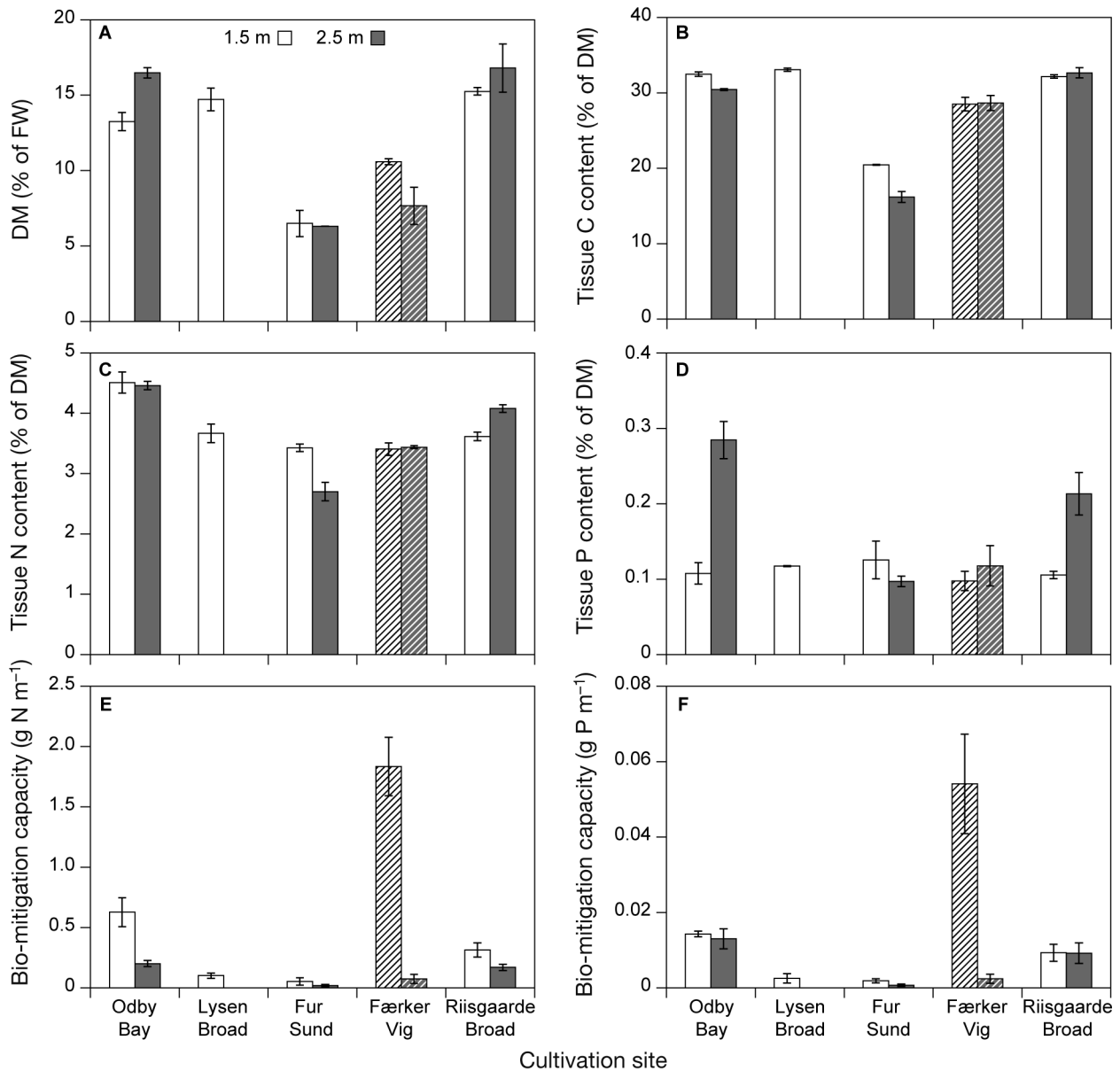


Fig. 5. Tissue concentrations of (A) dry matter (DM; % of fresh weight), (B) carbon (C), (C) nitrogen (N), (D) phosphorus (P), as well as bio-mitigation capacity of (E) N and (F) P of cultivation of *Saccharina latissima* harvested in April at the 5 cultivation sites at 1.5 (white bars) and 2.5 m depth (grey bars). Solid bars represent batch 1 of seeded lines, and crossed bars represent batch 2. Data represent means \pm SE, $n = 3$

tissue concentrations of all pigments were negatively related to light availability (Table S2).

Harmful metals

The tissue concentrations of the harmful metals As, Pb and Cd showed significant differences between sites and/or cultivation depths (Tables 3 & 5), with higher concentration of Pb and lower concentrations of As at Fur Sund as compared to the other sites. No

significant differences were observed in tissue Hg concentrations between sites and cultivation depths (Tables 3 & 5).

Metal concentrations ranged between (mean \pm SE) 9.90 ± 0.93 and 31.67 ± 1.07 mg As kg DM⁻¹, 0.91 ± 0.13 and 1.72 ± 0.08 mg Cd kg DM⁻¹, 1.11 ± 0.20 and 17.60 ± 3.33 mg Pb kg DM⁻¹, and between 0.18 ± 0.01 and 1.03 ± 0.40 mg Hg kg DM⁻¹ (Table 5). The tissue concentrations of As were positively correlated to SGR (linear regression, $p = 0.004$, $R^2 = 0.285$, $F = 10.556$, $df = 23$, $slope = 6.207$), whereas the tissue Cd

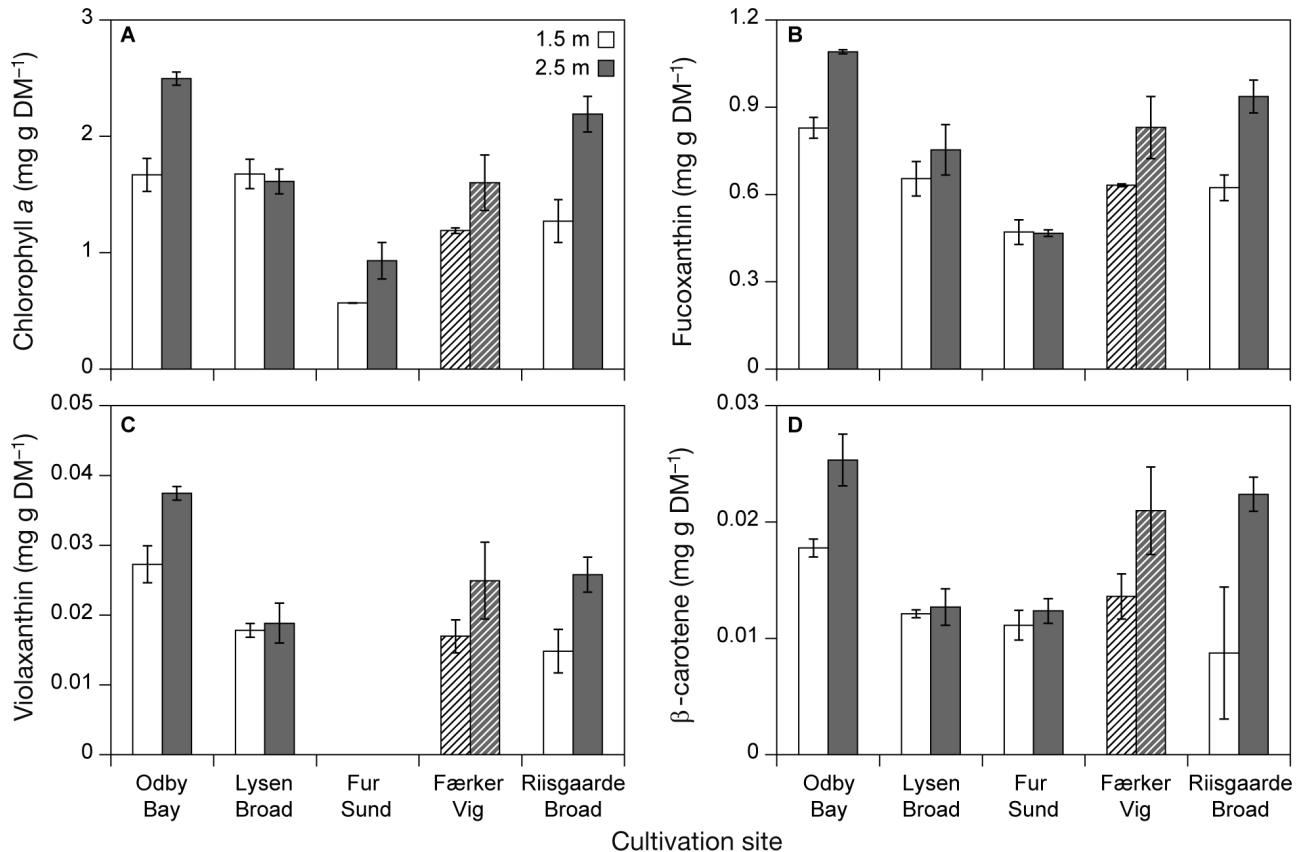


Fig. 6. Tissue concentrations of the pigments (A) chlorophyll *a*, (B) fucoxanthin, (C) violaxanthin and (D) β -carotene in *Saccharina latissima* harvested in April, at 1.5 (white bars) and 2.5 m depths (grey bars). Solid bars represent batch 1 of seeded lines, and crossed bars represent batch 2. Data represent means \pm SE, $n = 3$

concentrations were positively correlated to sediment concentrations of Cd (linear regression, $p < 0.0001$, $R^2 = 0.679$, $F = 51.878$, $df = 23$, slope = 2.327).

DISCUSSION

Pelagic environment

The lack of correlation between the availability of DIN and ortho-P indicated different origin of the 2 nutrients. The availability of DIN was negatively correlated to salinity, indicating input with freshwater run-off from the surrounding agricultural areas. The effect of freshwater run-off was also observed in Løgstør Broad and Skive Fjord as a decrease in salinity over the cultivation period.

The positive correlation between ortho-P and chl *a* indicated that the availability of P was controlling the primary production in Limfjorden in early spring, with DIN concentrations being too high to be limiting. P-limitation has previously been observed in

eutrophic coastal regions, including parts of Limfjorden (Lyngby 1990, Holmboe et al. 1999, Lyngby et al. 1999, Pedersen et al. 2010), as a consequence of a more efficient sewage treatment reducing the emissions of P as compared to N to the marine environment (Conley et al. 2000, Kronvang et al. 2005). In late spring the lack of correlation between nutrients and pelagic phytoplankton biomass indicated that other factors came into play controlling phytoplankton biomass, potentially grazing (Maar et al. 2010), as also indicated by the increasing density of biofouling organisms (filter-feeders).

The general inverse reflection of the pelagic phytoplankton biomass (chl *a*) by the photon flux density at cultivation depth indicated a close coupling between pelagic phytoplankton density and turbidity, as is common for Limfjorden (Krause-Jensen et al. 2012). However, impaired light conditions were also observed in winter in particular in Nissum Broad and Skive Fjord, most likely as a consequence of high wind speeds causing resuspension (Nissum) and/or soft sediment that is easily resuspended (Skive).

Table 5. Tissue metal concentrations of the biomass harvested in April, expressed as ppm of fresh biomass (fresh weight) (mg kg FW⁻¹) and as ppm of dry matter (mg kg DM⁻¹). The concentrations are compared to the limit values of fresh biomass for food and food supplement according to the EU food legislation (EU 2008b), to the limit values of dry biomass for use in feed according to the EU feed legislation (EU 2013), as well as to the limit values according to the Danish regulations on sludge used as fertilizer (Danish Ministry of Environment 2006). Data are mean ± SE, n = 3. nd: no data. Underscored numbers indicate tissue concentrations exceeding limit values for food supplement. Numbers in **bold** indicate tissue concentrations exceeding limit values for feed, and numbers in *italics* indicate tissue concentrations exceeding limit values for use as fertilizer

Cultivation site	Depth (m)	As	Cd	Pb	Hg
Concentration in fresh biomass (mg kg FW⁻¹)					
Odby Bay	1.5	4.02 ± 0.18	0.16 ± 0.02	0.15 ± 0.03	0.02 ± 0.00
	2.5	3.31 ± 0.21	0.19 ± 0.00	1.09 ± 0.03	0.08 ± 0.01
Lysen Broad	1.5	2.34 ± 1.17	0.16 ± 0.08	0.31 ± 0.18	0.04 ± 0.02
	2.5	nd	nd	nd	nd
Fur Sund	1.5	0.67 ± 0.02	0.06 ± 0.00	1.11 ± 0.15	nd
	2.5	0.42 ± 0.21	0.04 ± 0.02	0.68 ± 0.34	nd
Færker Vig	1.5	3.35 ± 0.07	0.11 ± 0.01	0.27 ± 0.08	0.04 ± 0.01
	2.5	2.08 ± 0.60	0.08 ± 0.02	0.26 ± 0.04	0.02 ± 0.01
Riisgaarde Broad	1.5	3.38 ± 0.18	0.26 ± 0.01	0.20 ± 0.02	<u>0.16 ± 0.06</u>
	2.5	3.18 ± 0.51	0.27 ± 0.03	0.43 ± 0.02	0.08 ± 0.01
Limit values, fresh biomass					
Mussels		–	1	1.5	0.5
Fish meat		–	0.05–0.3	0.3	0.05–1
Food supplement		–	1–3 ^a	3	0.1
Concentration in dry biomass (mg kg DM⁻¹)					
Odby Bay	1.5	30.37 ± 0.38	1.19 ± 0.06	1.11 ± 0.20	0.18 ± 0.01
	2.5	20.06 ± 0.87	1.18 ± 0.02	6.65 ± 0.28	0.51 ± 0.06
Lysen Broad	1.5	24.65 ± 1.09	1.42 ± 0.06	3.60 ± 0.70	0.32 ± 0.08
	2.5	nd	nd	nd	nd
Fur Sund	1.5	10.61 ± 1.38	<i>0.91 ± 0.13</i>	17.60 ± 3.33	nd
	2.5	9.90 ± 0.93	<i>0.94 ± 0.01</i>	16.20 ± 0.16	nd
Færker Vig	1.5	31.67 ± 1.07	1.08 ± 0.11	2.55 ± 0.83	0.39 ± 0.10
	2.5	26.11 ± 3.40	1.04 ± 0.12	3.66 ± 0.91	0.35 ± 0.15
Riisgaarde Broad	1.5	22.15 ± 0.83	1.72 ± 0.08	1.34 ± 0.10	<i>1.03 ± 0.40</i>
	2.5	18.65 ± 1.42	1.63 ± 0.03	2.63 ± 0.31	0.51 ± 0.07
Limit values, dry biomass					
Feed	40 (10 ^b)	1–2 ^b	5 ^b –10		
Sludge	25 ^c	0.8	120	0.8	

^aFood supplement derived from seaweed; ^bcomplete feed (As, Cd for pet animals based on seaweed); ^climit value only for use as fertilizer in private gardens

Biomass yield and growth performance

The *Saccharina latissima* biomass yields and sporophyte lengths obtained at the 5 sites in Limfjorden were generally low compared to values reported from other cultivation trials in Europe (Peteiro & Freire 2009, 2013b, Edwards & Watson 2011, Forbord et al. 2012, Handå et al. 2013), and in particular in Denmark (Marinho et al. 2015a, Nielsen 2015). The frond lengths were comparable to trials from Norway

with a shorter grow-out period (84 vs. 166 d at Færker Vig) (Forbord et al. 2012), and the sporophyte lengths obtained at Lysen Broad were fully comparable to earlier trials at the same site (Wegeberg 2010). Only sporophytes obtained in the German Baltic Sea were smaller, with maximum lengths of 20 cm obtained in a 1 yr grow-out period (Rössner & Krost 2012). The low biomass yields were explained by several factors: (1) light limitation; (2) P-limitation reducing the SGR, and contents of DM and N in the biomass, and (3) the high degree of biofouling forcing an early harvest.

The higher yield observed at Færker Vig was most likely a consequence of the combination of less turbid waters at this site during winter and early spring as well as the earlier deployment (October instead of December), which may have given the juvenile sporophytes there a head start in growth, as has been documented from trials in Spain and Ireland (Peteiro & Freire 2009, Edwards & Watson 2011). The fact that the sporophytes were derived from a different batch of seeded lines and potentially could have been of superior quality was not supported by visual inspection at deployment. Comparing only sporophytes from Batch 1, the growth performance was best at Odby Bay, where light conditions improved markedly from April to June.

The results in general support light as a main controlling factor for growth of cultivated *S. latissima* in Limfjorden in spring. In early spring, light was positively correlated to growth and the average PAR (100–400 μmol photons m⁻² s⁻¹) reflected a photon flux density within the range reported to saturate photosynthesis of *S. latissima* (E_{sat} 20–500 μmol photons m⁻² s⁻¹; Bartsch et al. 2008). In contrast, in late spring, the available PAR exceeded E_{sat}, and in this period the higher average PAR appeared to have a negative effect on growth since mainly the sporophytes from the deepest cultivation depth showed increased growth rates between April and June. A positive correlation between frond length and salinity has previously

been reported (Wegeberg 2010). The low biomass yields were explained by several factors: (1) light limitation; (2) P-limitation reducing the SGR, and contents of DM and N in the biomass, and (3) the high degree of biofouling forcing an early harvest.

been suggested (Nielsen et al. 2014), and reduced frond length at lower salinities may be a consequence of increased allocation of energy to osmoregulation at the expense of growth.

The high turbidity of the waters in Limfjorden generally limited the extent of the vertical production potential. The turbidity in Limfjorden is primarily a consequence of high nutrient loadings supporting a high pelagic primary production (Krause-Jensen et al. 2012). Historically, the primary production in marine waters is considered to be controlled by N availability (Howarth 1988). Limfjorden, however, is an estuary with strong influence of freshwater run-off from agricultural land, and in this study, DIN was available in the water column until late spring/early summer. The limiting nutrient appeared to be P, since the bioavailable P disappeared with the onset of the phytoplankton spring bloom in early spring, and subsequently, P availability appeared to control the primary production. That P rather than N availability controlled the growth performance of *S. latissima* in this study was supported by several observations: tissue N:P ratios were, already in early spring, almost 3 times as high as other reports of kelp N:P ratios (9–25:1) (Atkinson & Smith 1983); the tissue P contents were generally below the P concentration defined as being critical for growth (0.22% P of DM, 69.4 $\mu\text{mol P g}^{-1}$ DM) as suggested by Pedersen et al. (2010), whereas the tissue N concentrations were not below the concentrations critical for growth (N_C) of 1.71, as suggested for brown algae (Pedersen & Borum 1997), and 1.88 specifically for *S. latissima* (Chapman et al. 1978), and finally, SGRs in early spring corresponded positively to the ortho-P concentrations. Phosphorus limitation of macroalgae growth has previously been observed (Pedersen et al. 2010).

Biofouling of the biomass precluded a late summer harvest of the sporophytes and thus, a biomass build-up over summer. Devastation of biomass by biofouling has been reported from cultivation trials in Norway (Handå et al. 2013), Spain (Peteiro & Freire 2013a) as well as from other trials in Denmark (Wegeberg 2010, Marinho et al. 2015a, Nielsen 2015), and the phenomenon appears to be coupled to relatively sheltered locations with established natural or cultured populations of suspension-feeders. Temperature generally exerts positive control on the growth and development of juvenile filter-feeders (Widdows 1991, Nasrolahi et al. 2013), and in this study biofouling was positively correlated to temperature, even within a very narrow range of temperature differences. In the eutrophic environment in

Limfjorden, food (phytoplankton) is not a limiting factor for the juvenile filter-feeders, whereas suitable substrate for settling might be. Thus, any substrate introduced in the water column, including macroalgae sporophytes, has the risk of becoming fully overgrown. In this study, the degree of biofouling was most pronounced at the deeper cultivation depths, but did not correspond to the estimated degree of exposure at the individual cultivation sites. The negative correlation between biofouling and length growth may indicate that heavily bio-fouled fronds did not grow well, or that once the fronds were sufficiently long in early spring, they were able to avoid the biofouling, the latter partly being supported by recent findings showing that dense natural kelp canopies tend to be less prone to settling of epiphytic organisms (Bennett et al. 2015).

Biomass quality

If harvested before the onset of biofouling, *S. latissima* cultivated in Færker Vig, Limfjorden, provided a rich source of protein, essential AAs and pigments with bioactive properties suitable for food or feed purposes. Availability of ortho-P influenced the quality of the biomass, significantly increasing tissue DM and N content.

As for growth performance, the biochemical composition of *S. latissima* biomass showed large differences among cultivations sites. Tissue P concentrations were generally in the same range as reported from cultivation trials in Kattegat, Denmark (Marinho et al. 2015a). A doubling of tissue P concentrations in macroalgae cultivated at 2.5 m depth at 2 sites (Odbay Bay and Riisgaarde Broad), where the seabed was characterized by soft mud, indicated local differences in resuspension events as also indicated by the poorer light conditions at these sites during winter and early spring. The N content of 3–4.5% of DM in April was high for this time of the year compared to natural populations and cultivated biomass from other locations in Denmark (Nielsen et al. 2014, 2016, Marinho et al. 2015a) and was more comparable to N contents obtained in close proximity to fish farms or in late autumn/winter months where environmental N concentrations are naturally higher (Gevaert et al. 2001, Handå et al. 2013).

The high tissue N concentrations were indicative of high tissue protein concentration in the range of 16.0–17.0%. Compared to other cultivation trials in Denmark, this protein content was high for April (Marinho et al. 2015b), but comparable to what has

been reported elsewhere (Black 1950). The ratio of EAAs, and the content of methionine and lysine in the biomass in Færker Vig were higher than described from *S. latissima* biomass cultivated in proximity to fish cages, and thus the *S. latissima* biomass from Limfjorden represented a biomass with an attractive profile for applications within food or feed (Marinho et al. 2015b).

Light availability influenced biomass quality, correlating positively to tissue C content, but negatively to the tissue concentrations of P and all pigments. The pigment contents in *S. latissima* from the 5 sites varied by a factor of 2–5 and were generally high due to the turbid conditions, in particular in the deeper cultivated biomass. The tissue contents of chl *a* and fucoxanthin in the biomass were up to 8 and 5 times higher, respectively, than the tissue contents in *S. latissima* fronds cultivated in autumn under low light conditions in tanks (Boderskov et al. 2016). The antioxidant and other bioactive properties of fucoxanthin have recently drawn attention as being active against obesity and diabetes (Miyashita et al. 2011, D’Orazio et al. 2012). Thus, high contents of this pigment in kelp biomass are attractive for applications in (functional) food and feed.

The positive effects of temperature on DM, N and P tissue contents may in part be explained by increased activity of enzymes involved in nutrient assimilation over the range of temperatures experienced during early spring (Davison & Davison 1987).

Only extreme levels of pollution are considered to cause significant reduction in production of marine plants (Sharp et al. 1988); however, tissue concentrations of specific metals (i.e. As, Cd, Hg and Pb) may prevent the use of the produced biomass for food, food supplement, feed or fertilizer (Miljøstyrelsen 2006, EU 2008b, 2013). In this study, we only had access to sediment concentrations of selected metals from the national environmental monitoring program, as water concentrations are not monitored. For this reason, we had no basis for estimating the environmental metal concentrations experienced by the algae, and the potential direct consequential physiological impacts. However, through the sediment concentrations, we may get an indication of the local level of environmental pollution and an indication of whether this may be a predictive tool in future site selection. The tissue concentrations of As, Pb and Hg fluctuated by a factor of 3–5 between the 5 cultivation sites, whereas the tissue concentrations of Cd were relatively constant. The tissue metal concentrations in this study did not exceed the limit values set for human consumption, and only at one site (Riis-

gaard Broad) would the tissue Hg concentrations prevent the use of the biomass for food supplements. For use in animal feed, the Pb concentrations in the biomass cultivated at Fur Sund and Odby Bay (2.5 m) exceeded limit values, whereas the Cd concentrations would prevent the use for fertilizer of the biomass cultivated at any of the sites (limit value = 0.8 mg kg DM⁻¹, Danish Ministry of Environment 2006). The As concentrations found in this study did not exceed limit values for use in food or feed, and they were generally lower than what has been found in natural populations in more open Danish waters (Nielsen et al. 2016). Since tissue As concentrations were positively correlated to growth, bioaccumulation may explain the higher As concentrations found in older individuals in natural populations, as compared to the 1-yr-old cultivated individuals in this study. The linear correlation between tissue and sediment Cd concentrations indicated that elevated sediment concentrations of Cd may cause increased availability and hence uptake into the seaweed tissue. At the 2 stations with the highest sediment Cd concentrations (Riisgaard: 0.44 ppm and Lysen: 0.25 ppm), the seabed sediment and depth, as well as the degree of exposure, differed. At Riisgaard the sediment was soft and muddy, and a high degree of exposure increased the risk of resuspension of the sediment into the water column, potentially increasing the availability of Cd to the seaweed. Below the cultivation structures at the more sheltered and shallow site in Lysen Broad, the seabed consisted of fine sand and clay, and there the depth was lower. Thus, despite different conditions regarding sediment, depth and exposure, the sediment concentration of Cd demonstrated a potential value as an instrument in site selection.

Bio-mitigation

The bio-mitigation capacity of *S. latissima* in this study proved to be relatively poor in comparison with other studies of Laminariales in Denmark (Marinho et al. 2015a) and Scotland (Sanderson et al. 2012), where up to 4 and 1.4 times more N was removed per metre of seeded line, respectively. The low bio-mitigation capacity was primarily a consequence of the low biomass yields obtained due to turbid waters, P-limitation and biofouling. Thus, in highly eutrophic waters such as Limfjorden, the pelagic primary productivity limits the efficiency of kelp cultivation as a tool for bio-mitigation of N. Consequently, care should be taken when extrapolating the bio-mitigation ca-

capacities described in the literature to any cultivation site assuming high areal productivity (i.e. Holdt & Edwards 2014). This study highlights the limitations and challenges of kelp production for bio-mitigation purposes in eutrophic waters, where bio-mitigation is needed the most.

Site selection

Even within the relatively homogenous eutrophic Limfjorden, production yields varied by a factor of 10 between different basins. Environmental monitoring data proved useful as predictive instruments for site selection. Regarding the pelagic parameters, generally, highly N-enriched sites with low light availability, high pelagic N:P ratios and high chl *a* concentrations should be avoided, as they supported a lower biomass production and, in conjunction with marginally higher temperatures during spring, also presented a higher risk of biofouling.

Regarding sediment characteristics, 2 recommendations for site selection are suggested: (1) kelp cultivation should be reconsidered in shallow areas dominated by soft muddy seabed, as resuspension events tend to increase turbidity; and (2) sediment Cd concentrations could be investigated as a part of site selection. High sediment Cd concentrations were reflected as high Cd concentrations in seaweed biomass, and depending on the post-harvest use of the biomass, high tissue Cd concentrations may have a strong negative impact on biomass value.

CONCLUSIONS

Basin-scale differences in light and nutrient availability, seabed properties and sediment metal concentrations cause pronounced local differences in the suitability of an area for cultivation of *Saccharina latissima* in terms of biomass yield and quality as well as bio-mitigation, and hence, impact the profitability of potential seaweed production. When selecting sites for cultivation of *S. latissima*, highly N-enriched sites with low light availability, high pelagic N:P ratios and chl *a* concentrations, and high sediment Cd concentration should be avoided. The highly N-enriched waters of Limfjorden appeared less suitable for efficient biomass production of *S. latissima* due to reduced light conditions and P-limitation in early spring, and a high risk of devastating biofouling impairing growth performance, bio-mitigation capacity as well as biomass quality. However, *S. latissima* bio-

mass harvested in spring in Limfjorden had a high content of pigments and protein with a beneficial amino acid composition, and proved highly suitable for food or feed purposes.

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