

## **Biodiversity impacts of bioenergy production: Microalgae vs. first generation biofuels**

**Diego F. Correa<sup>1, 2\*</sup>, Hawthorne L. Beyer<sup>2</sup>, Hugh P. Possingham<sup>2</sup>, Skye R. Thomas-Hall<sup>1</sup>, Peer M. Schenk<sup>1</sup>**

<sup>1</sup>Algae Biotechnology Laboratory, School of Agriculture and Food Sciences, The University of Queensland, Brisbane, Queensland 4072, Australia

<sup>2</sup>ARC Centre of Excellence for Environmental Decisions, The University of Queensland, Brisbane, Queensland 4072, Australia

E-mail addresses: [h.beyer@uq.edu.au](mailto:h.beyer@uq.edu.au) (H. L. Beyer)

[h.possingham@uq.edu.au](mailto:h.possingham@uq.edu.au) (H.P. Possingham)

[s.thomashall@uq.edu.au](mailto:s.thomashall@uq.edu.au) (S.R. Thomas-Hall)

[p.schenk@uq.edu.au](mailto:p.schenk@uq.edu.au) (P.M. Schenk)

\*Corresponding author.

E-mail address: [diegofelipecorrea@gmail.com](mailto:diegofelipecorrea@gmail.com) (D.F. Correa)

## **ABSTRACT**

Energy and fuel demands, which are currently met primarily using fossil fuels, are expected to increase substantially in the coming decades. Burning fossil fuels results in the increase of net atmospheric CO<sub>2</sub> and climate change, hence there is widespread interest in identifying sustainable alternative fuel sources. Biofuels are one such alternative involving the production of different fuels which include biodiesel and bioethanol from plants. However, the environmental impacts of biofuels are not well understood. First generation biofuels (i.e. biofuels derived from edible biomass including crops such as maize and sugarcane) require extensive agricultural areas to produce sufficient quantities to replace fossil fuels, resulting in competition with food production, increased land clearing and pollution associated with agricultural production and harvesting. Microalgal production systems are a promising alternative that suffer from fewer environmental impacts. Here, we evaluate the potential impacts of microalgal production systems on biodiversity compared to first generation biofuels, through a review of studies and a comparison of environmental pressures that directly or indirectly impact biodiversity. We also compare the area of cultivation land required to meet gasoline and distillate fuel oil demands globally, accounting for spatial variation in productivity and energy consumption. We conclude that microalgal systems exert fewer pressures on biodiversity per unit of fuel generated compared to first generation biofuels, mainly because of reductions in direct and indirect land-use change, water consumption if water is recycled, and no application of pesticides. Further improvements of technologies and production methods, including optimization of productivities per unit area, colocation with wastewater systems and industrial CO<sub>2</sub> sources, nutrient and water recycling and use of coproducts for internal energy generation, would

further increase CO<sub>2</sub> savings. Overall pollution reductions can be achieved through increased energy efficiencies, along with nutrient and water recycling. Microalgal systems provide strong potential for meeting global energy demand sustainably.

**Keywords:** biofuel crops, ecological footprint, land-use change, life cycle assessment, tropic, vertebrate

## 1. INTRODUCTION

Future energy demands are expected to increase as a result of ongoing population and economic growth. According to the IEA [1], energy consumption could increase between 17% and 50% by 2040 relative to 2012, reaching around 15,629 and 20,039 million tons of oil equivalent (Mtoe) respectively. Meeting these demands under current levels of fossil fuels exploitation—with coal, oil and gas accounting for 82% of total primary energy consumption in 2012—is likely to drive increases in global atmospheric temperature above 3.6°C by 2100 in comparison to pre-industrial levels [1], leading to widespread changes in ecological communities and increases in extinction risks for species [2, 3]

Although a system that combines energy derived from the wind, water and sunlight has been proposed for supplying global energy demands [4], fuels with their high energy density will still be a major component in the future to power large machinery, planes, and ships. Biofuels, defined as high-density energy carriers derived from biomass transformation, could be a sustainable alternative to replace fossil fuels [5-7], especially for the transport sector [1, 8], which in 2012 accounted for around 23% of total CO<sub>2</sub> emissions [1]. Burning biofuel releases carbon that has been already fixed by plants through

photosynthesis and thus, in theory, should not increase the net atmospheric CO<sub>2</sub> content [9, 10]. However, there are concerns about the environmental impacts that a widespread adoption of biofuels could exert at a global scale, which could lead to further environmental degradation depending on the production system and initial land-use [11, 12]. Furthermore, environmental impacts are a function of differences in energy demands per country and regional variation in biofuels' productivities.

Currently, biofuels are primarily produced in the forms of bioethanol and biodiesel derived from food crops (i.e. first generation biofuels). It is estimated that between 2013 and 2015 around 77% of produced bioethanol was based on the processing of maize and sugarcane; while around 81% of biodiesel was produced from vegetable oils [13]. Because first generation biofuels compete with agricultural lands, environmental degradation—including biodiversity losses due to land clearing of biodiverse systems—has been associated with biofuels' expansion [12, 14-19]. Furthermore, biofuel production can increase the magnitude of other pressures that directly or indirectly affect biodiversity, including CO<sub>2</sub> emissions from land-use change [20-22] and production systems [23-25], emission of pollutants [7, 17] and depletion of water [26-28].

Microalgal production systems, which include open ponds and closed photobioreactors [29-33] could overcome several drawbacks of first generation biofuels, because they offer higher biomass yields than terrestrial crops per unit area, can be grown on non-arable lands, can make use of brackish or seawater, and can be coupled with wastewater systems and industrial CO<sub>2</sub> sources, helping in water remediation and in CO<sub>2</sub> emission reductions [29, 30, 32, 34-36].

Previous work on microalgal production systems has addressed several environmental impacts of microalgal biofuel production, including resource consumption and pollution [37-42], water consumption [43] and potential impacts of genetically modified strains [44]. However, no study has focused on biodiversity or compared the potential impacts of microalgal systems on biodiversity in relation to first generation biofuels.

Here, we review the potential impacts of microalgal systems for biofuel production on biodiversity in contrast to first generation biofuels, focusing particularly on vertebrates in tropical and subtropical biodiverse regions of the world [45, 46] where the potential for agricultural expansion, including first generation biofuels, is greatest [47, 48]. We classify the different factors that affect biodiversity as due biofuel production, using the DPSIR framework which, based on Driving forces, Pressures, States, Impacts and Responses, is useful for describing the interactions between society and the environment [49, 50]. Then, we identify and compare the different pressures—defined as anthropogenic factors that induce environmental impacts [51]—that directly and indirectly impact biodiversity, when using microalgal systems or first generation biofuels. Accounting for spatial variation in productivity and energy consumption, we estimate the area of cultivation land required to meet gasoline and distillate fuel oil for each country using either microalgal systems or first generation biofuels, to investigate the relative feasibility of adopting biofuels as a substitute energy source.

## **2. MATERIALS AND METHODS**

Relevant literature was identified in April 2016 using the Science Citation Index Expanded (SCI-EXPANDED) and the Emerging Sources Citation Index (*ESCI*) in Web of Science, with the following combinations of keywords: (biofuel OR bioenergy) AND (biodiversity OR wildlife), (biofuel OR bioenergy) AND (fish\* OR bird\* OR avian OR mammal\* OR reptil\* OR amphibian\*). A citation report was made using Web of Science in order to show the progress in the field. Papers were screened to identify those that relate first generation biofuels or microalgae with impacts in tropical and subtropical areas of the world (i.e. between parallels 38°N and 38°S). We used these studies to identify the impacts that biofuel production has on biodiversity, the anthropogenic factors that induce impacts on biodiversity (i.e. pressures), as well as the mechanisms and processes by which those impacts occur.

Further comparisons between microalgal systems and first generation biofuels were based on pressures that directly or indirectly have shown to impact biodiversity. Environmental pressures were schematized based on the DPSIR causal framework [49, 50]. The DPSIR framework has been adopted by the European Environmental Agency [50] and has been widely applied for understanding relationships between factors that drive impacts on the environment and society responses [12], for allowing communication between scientists [52] as well as a tool for decision making [53]. For this comparison, life cycle assessments for microalgal production systems were reviewed.

An estimate of the area of cultivation land required by microalgal systems and first generation biofuels and microalgal systems to meet each country's 2010 gasoline and distillate fuel oil demands [54] was developed. The average yield of crops that could be

used for ethanol and biodiesel production between 2005 and 2014 was calculated using the “FAOSTAT” database [55] for each country. Average ethanol yields were then estimated using conversion efficiencies from feedstocks [56-59] and average biodiesel yields were estimated using reported lipid contents and oil-specific densities per crop [56, 60], assuming lipid extraction efficiencies of 90% and lipid conversion efficiencies of 90%. For microalgal systems, lipid yields were obtained using the global map developed by Moody, McGinty [61]. The most frequent value of lipid yield per country was obtained based on an area weighted average. The total area of cultivation land required to meet each country’s gasoline and distillate fuel oil needs was then calculated by dividing their annual consumption in 2010 ( $\text{GJ year}^{-1}$ ) by the average biofuel yield per country ( $\text{GJ ha}^{-1} \text{ year}^{-1}$ ) (see Appendix A for details about calculations).

### **3. RESULTS AND DISCUSSION**

We identified 898 papers addressing the impacts of biofuels on biodiversity, 101 of which related first generation biofuels or microalgal systems to biodiversity in tropical and subtropical regions of the world. From this only three studies focused specifically on microalgal systems [62-64] (Tables A1 and A2, Appendix A). A citation report generated in Web of Science shows the increasing trend in number of citations for recent years, from five citations in 1993 to 5036 citations in 2015 and 4243 citations in 2016 (Fig. 1).

Increases in population growth, energy and food demands, and replacement of fossil fuels were identified as the main drivers for biodiversity changes arising from biofuel expansion.

A wide range of pressures that affect biodiversity were identified (Fig. 2). Because first generation biofuels make use of food crops, the pressures that impact biodiversity are closely related to those found for agricultural systems [65, 66]. These pressures corresponded to changes in land-use, overexploitation of resources, pollution and changes in environmental conditions that directly or indirectly impact biodiversity: land-use change (direct, indirect) and land-use intensification, increases in greenhouse gas emissions (leading to global warming), pesticide and fertilizer pollution, water depletion, overexploitation of soils (including soil erosion), increases in invasive species and genetic pollution, emissions of air pollutants and changes in environmental conditions that affect regional climate.

These pressures alter the state of ecosystems, resulting in a series of impacts on biodiversity (Fig. 2). Responses of society to these impacts may increase or decrease their magnitude. For instance, adaptation measures to climate change may drive further environmental degradation without an adequate planning for biodiversity conservation [67, 68], which outlines the importance of defining priorities that satisfy societal needs at the minimum costs for biodiversity [69].

These pressures can directly or indirectly impact biodiversity through several mechanisms. For instance, land-use change directly decreases available habitat, but can also lead to fragmentation that further increases potential extinction risks in the remaining habitat patches [70, 71]. Furthermore, the magnitude of biodiversity impacts resulting from biofuel crop expansion was found to be a function of initial land-use, type of biofuel system and its associated management practices and production technologies, and landscape

configurations between biofuel crops and native ecosystems [12, 72]. We examine each category of pressure in detail in the following sections.

### **3.1 Direct land-use change and land-use intensification**

Land-use change and land-use intensification were reported as the main pressures negatively impacting biodiversity due to the expansion of first generation biofuel systems [12, 73]. Replacement of native ecosystems and cropping intensification has been linked to habitat loss and degradation, decreases in richness and abundance of native vertebrates, affecting species of high conservation concern [12, 18, 74-78]. Furthermore, species that make use of biofuel plantations are mostly considered generalists and of low conservation value [15, 18, 72, 79-84].

Impacts on biodiversity depended upon the initial land-use, the type of biofuel production system and the landscape configuration. Reductions of species diversity are larger when transforming very biodiverse ecosystems [19] such as tropical forests [47, 75] and savannas [22, 77] and when using biofuel production systems that require a larger area per unit of energy produced [85]. In some circumstances, where biofuel crops recreate ecological conditions needed for the survival of native species, vertebrate diversity could increase. For instance, it has been proposed that the replacement of degraded lands by several biofuel crops could increase biodiversity values. In the Indonesian tropics, if degraded *Imperata* grasslands are replaced with oil palm plantations, which are structurally and functionally more complex than pastures, diversity of forest dependent vertebrates is expected to increase [84]—though mostly for low conservation value species—and lead to less pressure

on forests [86]. In the USA, large patches of perennial crops (e.g. switchgrass, *Miscanthus*, mixed-grass prairies) are expected to be better than annual crops (e.g. maize) for maintaining populations of grassland specialists including endangered vertebrates (e.g. the Henslow's sparrow), provided that management practices (e.g. application of pesticides and harvesting) do not negatively affect the fitness of species [14, 18, 87].

If large patches of forests remain near to biofuel plantations, several forest species can use oil palm plantations, even endangered vertebrates. For instance, it has been shown that chimpanzees can make use of oil palm plantations, eating young leaves, flowers and fruits when other sources of food are scarce [88]. Populations of large and medium sized felids can make use of oil palm plantations if native forest tracts remain [82]. However, the benefits may be diminished by negative interactions between humans and wildlife where species are perceived as pests, or where they are systematically hunted [89-91].

Microalgal cultivation systems need less land than first generation biofuels in order to produce the same amount of energy, and thus it is expected that their widespread adoption would lead to less direct land-use changes and lower relative habitat losses for native species. However, estimates for lipid productivities are very wide, ranging between 2.3 and 136.9 kl ha<sup>-1</sup>year<sup>-1</sup> [92]. Thus, we compare potential land savings based on a more conservative worldwide lipid estimation developed by Moody, McGinty [61], which closely resembles calculated productivities in experimental outdoor raceway ponds [93].

Our calculations show that microalgal cultivation systems consistently need less land than first generation biofuels (Fig. 3). For instance, in order to meet the USA gasoline and distillate fuel oil demands, microalgal systems would need 23.7% the area needed by olives

and 40.8% the area needed by sugar beets cropped within the country (Tables A.5 and A.6, Appendix A). This is an optimistic scenario for first generation biofuels because yields are based on areas where crops grow well, and it is assumed that these crops can be readily used for biofuel production. For microalgal systems, conservative yields are assumed, based on the area-weighted average of average lipid yields within each country based on the Moody, McGinty [61] global estimates.

Furthermore, microalgae can be grown in areas not suitable for other crops (i.e. in poor soils and in regions with low precipitation values) [30, 32]. If microalgal production proves to be feasible in these areas, less land-use change and intensification in highly biodiverse regions is expected, although marginal or degraded lands can still retain considerable biodiversity values [74, 94], and construction of microalgal facilities will inevitably decrease available habitat for native species [63].

### **3.2 Indirect land-use change**

Leakage effects result when economic activities are displaced into different regions where biofuels are grown [95]. Indirect land-use change occurs when agricultural lands are displaced into regions previously occupied by native ecosystems or non-intensive production systems including extensive pastures and agroforestry systems [17, 96-99]. For example, the European Union biofuel policies are expected to lead to increased land-use changes outside Europe and transfer environmental impacts to more biodiverse regions [100-102]. Biofuel cropping has also been related to indirect land-use change as a result of

complex interactions between economic factors, including increases in food prices and economic incentives for biofuel production [103-105].

Infrastructure development associated with agricultural expansion can further drive land-use changes, as shown in tropical remote regions, where deforestation increases due to higher accessibility and cropping profitability when roads are constructed or paved [48, 106]. In fact, oil palm and soybean expansion are related to road expansion and further deforestation in Southeast Asia and South America [84, 107]. For instance, in the Brazilian Cerrado, increased accessibility to forests around soybean plantations has led to further deforestation for fueling the steel industry, which not only decreases the area of remaining forests but also generates profits for further soybean expansion [107, 108].

Microalgal systems are not considered to drive indirect land-use change [109]. This is because if they are produced in degraded, dry or marginal lands that are less suitable for food production, less competition with agricultural lands would occur, which is expected to lead to fewer leakage effects, land clearing and transformation of biodiverse systems. However, this assumption is contingent upon the feasibility of microalgal biofuel production in areas not suitable for agriculture production.

### **3.3 Increases in greenhouse gas emissions**

Biofuel expansion affects the emission of greenhouse gasses via land-use change and energy intensive production systems [110], while coproducts can help in decreasing greenhouse gas emissions. These emissions of greenhouse gasses have been linked to local extinction and habitat shifts for native species through global warming [2, 3].

### **3.3.1 Greenhouse gas emissions as a result of land-use change**

Clearing of rich carbon systems releases CO<sub>2</sub> when plant biomass is burnt and soil organic carbon is lost [111-113]. Fargione, Hill [20] estimated that 17 years would be needed by sugarcane ethanol production systems to recapture the CO<sub>2</sub> emitted after replacing Cerrado grasslands in Brazil. Oil palm production systems replacing peatland rainforests in Indonesia would need 423 years to recapture the emitted CO<sub>2</sub> [20]. [15] estimated that in Southeast Asia the replacement of native forests into oil palm can emit between 163 and 1,550 tons ha<sup>-1</sup> of stored carbon. If peatland forests are transformed into oil palm crops, it could take up to 692 years by oil palm production systems to recapture this released carbon. Based on satellite images, Koh, Miettinen [16] estimated that between 2000 and 2010 conversion of forests into oil palm plantations in Malaysia, Borneo and Sumatra led to the loss of around 140 million Mg of aboveground biomass carbon. Even in tropical grasslands, significant carbon emissions are expected to occur if replaced by biofuel production systems [22]. Furthermore, first generation biofuel production can lead to indirect land-use changes, which would further drive clearing of native ecosystems for crop production, and thus increases greenhouse emissions [21].

Initial land-use is expected to alter the magnitude of CO<sub>2</sub> emissions under the construction of microalgal farms [114]. Because microalgal production systems need less land for producing the same amount of energy than terrestrial crops, and their production can be achieved in places with naturally lower carbon stocks (i.e. degraded and dry areas), it would be expected that much less CO<sub>2</sub> would be released following direct land-use changes

when using microalgal systems compared to first generation biofuels. If degraded areas, dry areas and marginal lands are used for microalgal biofuel production, or even for the production of microalgal animal feed, less competition with crops is expected to occur, leading to less indirect land-use changes and lower CO<sub>2</sub> emissions. In fact, it has been estimated that the global expansion of microalgae as a feedstock for animal feed, in areas not suitable for agricultural production, could free almost 2 billion hectares of pastures and feed crops, where forest plantations can be established for bioenergy production and habitat restoration, leading to net atmospheric CO<sub>2</sub> reductions [115]. However, if rich carbon systems are used for microalgal production, CO<sub>2</sub> emissions may become substantial. For instance, Quiroz-Arita, Yilmaz [116] estimate that within the USA the CO<sub>2</sub> savings of microalgal systems may decrease between 3% and 85% as a result of losses in aboveground biomass and soil carbon associated to land-use changes.

### **3.3.2 Production technologies and greenhouse gas emissions**

Biofuel production systems and their associated cropping management practices and conversion technologies affect the balance of greenhouse gas emissions [110]. In agriculture, greenhouse gas emissions come from energy consumed along the production chain (CO<sub>2</sub> emissions), fertilizer use (liberation of N<sub>2</sub>O and CO<sub>2</sub>), cultivation in flooding conditions (CH<sub>4</sub> emissions) and several soil management practices including tillage, addition of lime and irrigation frequency [117].

As a result, crops with lower fertilizer requirements, coupled with management practices that optimize nutrient uptake and soil carbon storage, and less energy-intensive production

technologies, would lead to lower greenhouse gas emissions. For instance, in the USA biodiesel production from soybeans captures more greenhouse gasses than bioethanol production from maize (41% vs. 12% respectively; taking into account energy used for crop cultivation, biofuel production and transport), mainly because of lower agricultural inputs and less intensive processes for biofuel production [7].

In comparison to terrestrial crops, microalgal systems can offer higher CO<sub>2</sub> savings when using efficient technologies under optimal production conditions [118]. However, a consensus in an optimal production technology—that maximizes both cost-effectiveness and reductions in CO<sub>2</sub> emissions—has not been reached. This is because large commercial microalgal farms for biofuel production have not been deployed, and because of difficulties in comparing studies that have different system boundaries, sources of electrical energy, functional units, influence of coproducts and model parameters [92, 118, 119] (Table 1).

Open raceway ponds are estimated to be energetically more efficient than photobioreactors [42, 120], leading to higher CO<sub>2</sub> savings [121]. Using open ponds, carbon savings can increase due to higher productivities per unit area [40, 114, 122, 123], colocation of microalgal systems with CO<sub>2</sub> sources (e.g. use of flue gas) [38-40, 43, 124, 125] or wastewater systems [38, 39], use of technologies that allow nutrient recycling (e.g. water recycling) [43, 121, 124] and production of energy (e.g. anaerobic digestion for producing methane which can be used for electricity generation) [41, 92, 120, 122, 126-128]. However, Clarens, Nassau [38] suggest that anaerobic digestion for nutrient recycling and energy production is not the best approach for reducing greenhouse gas emissions compared to direct combustion of algal biomass, although increases in digestibility,

methane production and nutrient recovery could increase the environmental benefits of this technology.

Increasing the energy efficiency of production methods is also important for reducing emissions, such as through improved water pumping methods and more efficient lipid extraction processes [42, 123]. In fact, wet extraction routes have potential for decreasing energy inputs and increase CO<sub>2</sub> savings [37, 42, 122], especially through hydrothermal liquefaction [126, 129]. Overall, increasing low-carbon energy sources for microalgal production systems, including heat, electricity grid, fertilizers, transport and building materials not derived from fossil fuels, would lead to further carbon savings [125].

Thus, substantial increased carbon savings in comparison to first generation biofuels are feasible. For instance, Lardon, Helias [37] estimated that assuming biomass productivities between 20 and 30 g m<sup>-2</sup> d<sup>-1</sup> for *Chlorella vulgaris* grown in open raceway ponds under Mediterranean conditions, and using wet extraction lipid routes, microalgal production could lead to less global warming potential when compared to soybean and conventional diesel, but not to oil palm or rapeseed. However, this study did not take into account nutrient recycling through anaerobic digestion or culture medium recycling. Stephenson, Kazamia [114] estimated that the production of *C. vulgaris* in open raceway ponds under U.K. conditions could lead to higher carbon savings than biofuel obtained from soybean, sunflower and rapeseed grown in South Africa or from oil palm in Malaysia; assuming higher lipid productivities, production in degraded lands, use of flue gas from power stations, nutrient recycling and energy production through anaerobic digestion, and lower velocities for microalgal cultivation media. Clarens, Nassau [38] found that, assuming biomass yields of 91.1 Mg ha<sup>-1</sup> year<sup>-1</sup> for brackish water species grown in Southwestern

USA conditions and lipid contents of 19.6%, greenhouse gas emissions per kilometer travelled would be lower compared to rapeseed.

### **3.3.3 Influence of coproducts in greenhouse gas emissions**

Coproducts of biofuel production help in increasing CO<sub>2</sub> savings. These include dried distillers grains, feed products, CO<sub>2</sub>, starch, syrup and oils (e.g. corn oil) in the case of bioethanol production from sugar and starch crops, as well as protein meal and glycerol from biodiesel production [9]. Microalgal systems can be designed to produce not only biodiesel or bioethanol as main biofuel products but also a wide arrange of coproducts that can be used for energy production, food and animal feed [35, 130]. For instance, using wet conversion routes it is possible to produce biodiesel, carbon monoxide, hydrogen, methane, ethane and propane, while through dry conversion, biodiesel, glycerol, pyrolysis oil and biogas can be produced [131].

Coproducts are considered fundamental for increasing the cost-effectiveness and sustainability of microalgal biofuel production systems [118, 130]. In particular, methane production has been identified as a key coproduct that increases carbon savings when it is combusted for replacing external energy requirements [41, 92, 120, 122, 128, 132].

### **3.4 Pesticide and fertilizer pollution**

Pesticides and fertilizers can impact vertebrate biodiversity in biofuel crops and non-target areas, negatively affecting native ecosystems. Pesticides can directly and indirectly lead to

the collapse of vertebrate populations as a result of several mechanisms, including direct poisoning, reduced amounts of prey, increases in occurrence of diseases [133-136] and decreases in fruit productivities when pollinator biodiversity is negatively affected [137].

Overuse of fertilizers can pollute soils with heavy metals that bioaccumulate in vertebrates [138] and indirectly alter biodiversity through increases in greenhouse gas emissions [117]. Eutrophication of aquatic systems as a consequence of runoff can lead to oxygen depletion and bioaccumulation of toxins produced by toxic algae blooms [139] and occurrence of diseases (e.g. nitrate accumulation in vertebrates) [140]. Besides the global warming potential of NO<sub>x</sub>, increases in nitrogen compounds in the atmosphere have been suggested to reduce plant diversity and alter ecosystem functioning [141, 142].

The release of pollutants depends upon the type of biofuel production system and its associated management practices. For example, soybean cropping in the USA uses lower amounts of fertilizers and pesticides when compared to maize, leading to the release of 1% of the N, 8.3% of the P, and 13% of the pesticides, per net energy gain, used for maize ethanol production [7].

Unlike first generation biofuels, microalgal cultivation does not require the use of pesticides [35, 37, 143]. When grown in photobioreactors, contamination of cultures by pathogens and algae grazers does not often occur [32, 144]. In open ponds, methods other than pesticide addition help to decrease the incidence of undesired organisms, such as increases in pH and free ammonia concentrations [64, 93, 145, 146].

Microalgal systems make use of fertilizers mainly in the forms of nitrates, ammonium and phosphate [42]. It has been estimated that the production of 1 kg of biodiesel from *C.*

*vulgaris* grown in open raceway ponds under California conditions, needs 0.33, 0.71, 0.58, 0.27, and 0.15 kg of nitrogen, phosphorous, potassium, magnesium, and sulfur respectively [147]. At Pinjarra Hills (Brisbane, Australia) the production of 1 kg of biodiesel from *Scenedesmus dimorphus* requires 0.04 kg of monoammonium phosphate, 0.02 kg of magnesium sulfate, 0.2 kg of ammonium sulfate, plus 0.004 kg of micronutrients [93]. However, microalgal systems have lower eutrophication potential than first generation biofuels [37, 39, 41, 127], primarily because runoff can be controlled in contrast to terrestrial crops [39]. In fact, if cultivation wastewater is recycled, fertilizers would not reach aquatic systems, eliminating gray water footprints [131], and reducing nutrient requirements [43, 147]. For instance Yang, Xu [147] estimate that water recycling in open ponds using *C. vulgaris* could reduce fertilizers use by around 55%; and if using seawater or wastewater the use of nitrogen would decrease by 94%. Using sea/waste-water for algal culture can reduce nitrogen usage by 94% and eliminate the need for potassium, magnesium, and sulfur. However, if wastewater reaches aquatic systems negative impacts on biodiversity are expected due to eutrophication [42, 63, 64, 131].

### **3.5 Water depletion**

Water depletion can affect biodiversity associated with water systems, because of direct withdrawals and changes in water quality, including increases in salinity and concentrations of minerals [148]. The water footprint (WF) can be divided into green WF (volume of rainwater consumed), blue WF (volume of surface and groundwater consumed) and gray WF (volume of polluted water) [149]. Microalgal systems have a green and blue WF as a

result of evaporative losses in raceway open ponds, evaporative cooling in photobioreactors and evaporation from dry biodiesel conversion routes, while if wastewater is recycled or treated the gray WF should be zero [131]. As a consequence, for open ponds in California, the water footprint is expected to be reduced by around 84% if water is recycled, and by around 90% if seawater or wastewater are used [147].

Green and blue WFs using wet conversion routes and recycling water are between 2.7 and 32.6 kL per GJ of produced green diesel [131] (Table A.12, Appendix A). These values are lower than those obtained for terrestrial biofuel crops such as soybean, sugarcane, maize, rapeseed and sugar beet (Fig. 4). The variation in water requirements is a function of lipid productivity, local weather conditions and the architecture of the microalgal production system (photobioreactors or open ponds), being highest when using open ponds in places with high evaporation rates [43] and low lipid productivities [131]. Other factors that affect water consumption are the medium preference of microalgal strains (fresh, brackish or saline water) and the conversion technologies for biodiesel production (thermal drying and pyrolysis in dry conversion route vs. water reuse in wet conversion route), being higher when using freshwater species and when using dry conversion routes [131]. However, water use would be higher if it is not recycled. For instance, Clarens, Resurreccion [39] show that open raceway ponds in Virginia, Iowa, and California conditions, would need more water than rapeseed and maize cropped in the same locations, provided that there is not water recycling.

### **3.6 Overexploitation of soils**

Soils are considered a renewable resource when managed in a sustainable way, by avoiding erosion and maintaining or increasing fertility and soil biodiversity [150]. Fertile soils increase food security, decrease desertification, help in climate change mitigation and increase biodiversity [151-153]. Biofuel production systems may negatively affect in-situ soil productivity, when using management practices that increase soil erosion and affect physical, chemical and biological properties in soils (e.g. indiscriminate tillage) [154, 155]. Additionally, soil erosion can negatively affect aquatic biodiversity due to eutrophication, sedimentation and the alteration of physical and chemical properties in aquatic systems [152].

Soils are not used directly for microalgal production systems. However, construction of open ponds could increase soil erosion, soil compaction and alter soil properties including texture and fertility [62, 63], if soil conservation practices are not implemented. The construction of elevated ponds (e.g. using bricks) could decrease soil removal (which could be around 225,000 and 450,000 tons ha<sup>-1</sup>, assuming pond depths between 15 and 30 cm and soil bulk densities at 1.5 g/cm<sup>3</sup>), although at higher economic costs. After ponds are constructed, soil erosion is expected to be lower than in agricultural production systems, which have reported erosion rates between 0.5 and 400 tons ha<sup>-1</sup> year<sup>-1</sup> [151, 156].

### **3.7 Increases in invasive species and genetic pollution**

Invasive species are a major threat to biodiversity [157, 158]. Biofuel crops can increase the occurrence of invasive species within and outside plantations, creating more favorable environmental conditions for the arrival and persistence of invasive organisms [159].

Furthermore, some species may become invasive as a result of their increased propagule production, dispersal and/or persistence abilities [160, 161]. Crops like sugarcane, soybean, sugar beet and maize are not considered invasive, while others have traits that increase their invasive potential (e.g. rapeseed produces large seed quantities that can be dispersed by a wide range of agents, and can hybridize with wild native varieties) [161, 162] (Table 2). As a consequence, it has been estimated that terrestrial plants suitable for biofuel production have two to four times higher potential than other crops to become naturalized or become invasive [161, 163].

In relation to microalgal production systems, the potential invasion of water systems could happen if leakage of growth medium, which may include genetically engineered species, occurs [14, 42, 62]. This is because the same traits that allow them to grow in a wide range of environmental conditions predispose them for invasiveness potential [164]. If toxic species are released (e.g. *Anabaena circinalis*, *Oscillatoria agardhii*, *Cylindrospermopsis raciborskii*) unexpected changes in ecosystem function could occur under favorable environmental conditions (e.g. expansion of toxic algae blooms in eutrophic aquatic systems) [161, 165]. However, if native or local microalgal strains are used for biofuel production, or if water is recycled, invasion potential is expected to decrease.

### **3.8 Emissions of air pollutants and changes in factors that affect regional climate**

In addition to greenhouse gases (section 3.2), the production and use of biofuels generate toxic substances that are released into the air, and that can negatively impact ecosystem functions and biodiversity. These pollutants include nitrogen oxides (NO<sub>x</sub>), ammonia

(NH<sub>3</sub>), carbon monoxide (CO), volatile organic compounds (VOC), particulate matter (PM), oxides of sulfur (SO<sub>x</sub>) [166], methyl bromide (CH<sub>3</sub>Br) [167] and nitrous oxide (N<sub>2</sub>O) [168]. These pollutants can be produced during cropping (including fuel combustion for machinery operation during cropping practices, chemical applications and soil disturbance), biofuel production and combustion [166], and during the construction of facilities, extraction and shipping of resources [41]. They lead to increases in acidification (i.e. acid rain), ozone layer depletion, and photochemical oxidation, among other environmental impacts [169]. Their effects include changes in the structure and function of both terrestrial and aquatic ecosystems, including alterations in species composition [170-172].

Pollutant release differs among biofuel production systems. For instance, taking into account total life-cycle emissions, it has been shown that soybean biodiesel produced in the USA releases less air pollutants when compared to corn ethanol per net energy gain [7], while corn grain ethanol blended with gasoline (E-85) increases the amount of emitted pollutants (CO, VOC, PM<sub>10</sub>, NO<sub>x</sub>, SO<sub>x</sub>) in comparison to gasoline per unit of energy released upon combustion [173]. After accounting for cultivation and harvesting, it is estimated that in the USA corn grain ethanol would emit more pollutants per produced gallon than ethanol from switchgrass, corn stover, wheat straw and forest residues [166].

Compared to first generation biofuels, emission of air pollutants can be lower for microalgal systems [41, 127]. Collet, Helias [41] estimated that biodiesel produced from *C. vulgaris* grown in open raceway ponds in Mediterranean conditions coupled with anaerobic digestion and assuming low-energy cultivation systems, led to lower potential acidification, ozone layer depletion and photochemical oxidation per MJ of combusted fuel than first generation biofuels after accounting for extraction and shipping of resources,

cultivation and biofuel production, and construction and dismantling of facilities. Using the same species and open raceway ponds in Mediterranean conditions, Lardon, Helias [37] found lower acidification potential in comparison to rapeseed, but higher ozone layer depletion and photochemical oxidation when compared to first generation biofuels. However, they did not account for nutrient recycling (e.g. using anaerobic digestion), which would lead to lower energetic burdens and decrease air pollutants.

Air pollution may also impact biodiversity via changes in atmospheric temperature and weather patterns: the release of substances that increase tropospheric ozone (CO, NO<sub>x</sub>, VOC, CH<sub>4</sub>) exacerbates global warming potential, while the release of aerosol particles (including sulfate, organic carbon, black carbon, biomass burning, nitrate and mineral dust aerosols) increase albedo and thus exert an atmospheric cooling effect [174]. Furthermore, it has been shown that aerosols affect not only cloud albedo but also the size and number of droplets in clouds, which can alter precipitation regimes worldwide depending on meteorological conditions [175-177]. Changes in surface albedo (that result from land-use change), coupled with increases in tropospheric ozone and aerosols, can alter atmospheric temperature and precipitation patterns, with potential impacts on ecosystems. While deforestation for biofuel production would decrease regional humidity and increase atmospheric temperature, evaporation from microalgal ponds could have the opposite effect [64], with potential increases in regional precipitation and additional cooling effects as water evaporates [177].

### **3.9 Considerations for the adoption of sustainable biofuel production systems**

Transforming biodiverse landscapes into biofuel cropping systems is a detrimental practice for the short and long-term conservation of biodiversity. Biofuel production should only be promoted where few direct and indirect impacts on biodiverse systems are expected; implying that crops with low biofuel yields or crops that compete with available lands for agriculture or for the conservation of biodiversity should be avoided. Currently, biofuel is primarily produced from suboptimal crops that do not have the highest biofuel yields (Figs. A.1 to A.5, Tables A.7 to A.11, Appendix A) and that compete with agricultural lands or highly biodiverse landscapes. Thus, biofuel production systems, management practices and production technologies that have lower environmental footprints should be encouraged. This means that only systems with low potential to cause direct and indirect land-use change of agricultural lands and biodiverse regions and that offer higher carbon savings should be deployed, and also those systems with high freshwater consumption, high potential for pollution, soil degradation, and high invasiveness should be avoided.

#### **4. CONCLUSIONS**

The main pressures negatively impacting biodiversity due to biofuel production are direct and indirect land-use change, particularly when ecosystems with high biodiversity values (e.g. tropical and subtropical forests, native grasslands) are transformed into biofuel crops. Several other pressures that negatively impact biodiversity include greenhouse gas emissions, pesticide and fertilizer pollution, water depletion, overexploitation of soils, invasive species and genetic pollution, emissions of air pollutants and changes in factors

that affect regional climate (e.g. alterations in albedo and evapotranspiration patterns), which directly or indirectly impact biodiversity.

Biofuel production systems and their associated management practices influence the magnitude of the impacts on biodiversity. In relation to land-use change, systems with higher productivity per unit area are expected to lead to less direct and indirect land-use changes, especially if their cultivation does not occupy fertile agricultural lands and does not compete with areas of high biodiversity value. Higher greenhouse gas savings would be achieved both when transforming low carbon systems (e.g. eroded lands) into biofuel crops and when using biofuel systems with lower energy intensive processes. Pollution would be reduced using systems with lower fertilizer and pesticide inputs, combined with less energy intensive processes that are currently powered by fossil fuels. Furthermore, biofuel and their associated management practices can be designed to achieve better water efficiency, less soil degradation (e.g. low soil erosion), and reduced invasive species and genes potential.

We estimated that microalgal production systems would need substantially less cultivation land compared to first generation biofuels per unit of produced energy, making them the most feasible option worldwide in term of reduced land needs, especially within tropical and subtropical regions of the world where they achieve higher productivities. Open ponds are the preferred option for increasing carbon savings, because of their lower energy-intensive production processes compared to photobioreactors. Increased carbon savings in microalgal systems can be achieved with the optimization of productivities per unit area, colocation with industrial CO<sub>2</sub> sources or wastewater systems, nutrient recycling and energy production (e.g. using anaerobic digestion and recycling water), and use of

coproducts for reducing external energy sources (e.g. combustion of methane for internal electricity generation). Increasing energy efficiencies (e.g. using wet conversion routes for biodiesel production) and replacement of external fossil energy sources is expected to reduce greenhouse gas emissions. Increased energy efficiencies and nutrient recycling are expected to decrease emissions of air pollutants (NO<sub>x</sub>, NH<sub>3</sub>, CO, VOC, PM, SO<sub>x</sub>, N<sub>2</sub>O). Water recycling is also essential in order to reduce the gray water footprint, avoid pollution derived from the release of growth medium and decrease the chances of spreading invasive and potentially harmful microalgal strains.

Finally, we call for a better inclusion of biodiversity in future studies on environmental impacts of biofuel production systems as it is currently underrepresented, particularly in life cycle assessments [178-180].

## **ACKNOWLEDGEMENTS**

D.F. Correa acknowledges financial support for Ph.D. studies by COLCIENCIAS (Convocatoria 529 para estudios de Doctorado en el exterior) and the University of Queensland (APA scholarship). We thank Dr. Jason Quinn at the Colorado State University, USA, for providing databases about estimates on lipid productivities for microalgal systems at a global scale.

Table 1. Comparison between several life cycle assessments developed to date, in relation to system boundaries, main processing technologies, measured environmental impacts and main results. Open raceway pond (OP), photobioreactor (PB), open raceway pond integrated with photobioreactor (OP-PB), Not Stated (N.S.)

Species	Growing	System boundaries	Main processing technologies	First gen. biofuels	Measured environmental impacts	Main results	Notes	Ref.
<i>Chlorella vulgaris</i>	OP	Cradle-to-combustion analysis for the fuel, cradle-to-grave analysis for the facility. Includes extraction and production of raw materials, facility construction and dismantling, biofuel production and use in the engine.	1) Advanced drying followed by hexane extraction. 2) Direct extraction from the wet algal paste.	Rapeseed, soybean, oil palm	Abiotic depletion, potential acidification, eutrophication, global warming potential, ozone layer depletion, marine toxicity, human toxicity, land competition, emission of ionizing radiation, photochemical oxidation.	Lower land competition and eutrophication compared to first generation biofuels. Lower acidification potential in comparison to rapeseed and lower human toxicity in comparison to rapeseed and oil palm. Lower global warming potential in comparison to soybean. Higher abiotic depletion, ozone layer depletion, marine toxicity, ionizing radiation and photochemical oxidation compared to first generation biofuels.	Assumed biomass productivities at 20-30 g m <sup>-2</sup> day <sup>-1</sup> in Mediterranean conditions. Functional unit as the combustion of 1 MJ of fuel in a diesel engine.	[37]
<i>Chlorella vulgaris</i>	OP	Microalgal cultivation to downstream fuel production. Includes cultivation, harvesting, dewatering, oil extraction, oil upgrading and nutrient recycling.	Harvesting by flocculation and centrifugation, followed by dry conversion routes for lipids (transesterification) or wet conversion routes lipids (hydrogenation).	Several vegetable oils and sugar crops	Land use	Large positive energy balance in comparison to first generation biofuels can be achieved. Potential to increase productivity and decrease nutrient usage by nitrogen deprivation during growing. Larger land savings when increasing productivity per unit area.	Assumed lipid contents between 19.7-43% and 15% of nutrient recycling for wet processing route.	[123]
<i>Chlorella vulgaris</i>	OP	Production, harvesting and concentration of algae, methane extraction and combustion, facility construction and dismantling, extraction and shipping of resources.	Harvesting by settling and centrifugation followed by injection in anaerobic digesters, biogas burning and production, CO <sub>2</sub> reinjection into cultures.	Rapeseed, oil palm	Abiotic depletion, potential acidification, eutrophication, global warming potential, ozone layer depletion, human toxicity, land competition, emission of ionizing radiation, photochemical oxidation.	Lower impacts compared to first generation biofuels for acidification, eutrophication, ozone layer depletion and photochemical oxidation, when assuming low energy consumption by paddlewheels and pumping water [39]. Global warming potential decreases when assuming low energy consumption.	Assumed biomass productivities of 25 g m <sup>-2</sup> day <sup>-1</sup> in Mediterranean conditions (Narbonne, France). Low energy consumption of paddlewheels and pumping water is assumed based on Clarens, Resurreccion [39]. Functional unit as the combustion of 1 MJ of fuel in an internal combustion engine.	[41]
<i>Chlorella vulgaris</i>	OP	Cultivation, harvesting, lipid extraction, fuel distribution and combustion by end users.	Harvesting by flocculation, drying and algae oil extraction.	N.S.	Greenhouse gas emissions.	Higher CO <sub>2</sub> emissions compared to conventional diesel for most scenarios.	Assumed biomass productivities of 75 tons ha <sup>-1</sup> year <sup>-1</sup> and average algae oil content of 30-70%. Explicit analyses in U.K., France, Brazil, China, Nigeria and Saudi Arabia. Assumes use of CO <sub>2</sub> from nearby power plants (12.5%). Includes three options for coproduct use: co-firing of biomass residues, direct combustion in a biomass/heating system or a biomass combined heat and power unit. Functional unit set as 1 MJ of biodiesel produced from algae oil.	[125]

<i>Chlorella vulgaris</i>	OP	Cradle-to-gate, including processes upstream of dried biomass production.	Harvesting by centrifugation or chamber filter press followed by two algal drying options (natural gas based drying or waste heat drying).	N.S.	Greenhouse gas emissions, direct water demands.	Greenhouse gas savings for 5 out of 8 scenarios analyzed. Water demands were related to geographic locations and their local evaporation rates.	Assumed algae composition of 20% lipids, 25% carbohydrates and 50% protein at 21 geographic locations in the contiguous USA. Includes colocation with natural gas power plant and water recycling. Functional unit as 1 MJ of dried algal biomass.	[43]
<i>Chlorella vulgaris</i>	OP	Culture, harvest, drying, extraction and esterification.	Drying and lipid extraction.	Maize, potato, sugarcane, sugar beet, sorghum, soybean	Water footprint, nutrient depletion.	The water footprint is in general lower compared to first generation biofuels, and lowest if recycling water or using wastewater/seawater. Nutrient usage is lower when recycling water and when using wastewater or seawater.	Assumed use of freshwater, seawater, and wastewater in California conditions.	[147]
<i>Chlorella vulgaris</i>	OP, PB	Cultivation, harvesting and lipid extraction, anaerobic digestion, oil extraction, esterification, transport of oil and final combustion in vehicles.	Harvesting by flocculation, followed by centrifugation (for open raceway ponds), cell disruption by homogenization, hexane lipid extraction, anaerobic digestion for onsite electricity use.	Rapeseed, sunflower, soybean, oil palm	Global warming potential (CO <sub>2</sub> , NO <sub>2</sub> , CH <sub>4</sub> ), water depletion.	Lower global warming potential for open raceway ponds and compared to rapeseed, sunflower and soybean biodiesel grown in arable lands in South Africa and compared to oil palm grown in Malaysia. Higher water requirements for photobioreactors under U.K conditions.	Assumed oil productivities at 40 tons ha <sup>-1</sup> year <sup>-1</sup> and production in degraded lands in U.K. Assumes nitrogen deprivation, coproduct allocation, use of flue gas from power stations (12.5% CO <sub>2</sub> ). Functional unit as the combustion of 1 ton of biodiesel in a car engine filled at a U.K. station.	[114]
N.S.	N.S.	Well-to-wheel. Includes cultivation, processing and biofuel production, transport and final use of biodiesel.	Harvesting, and extraction followed by transesterification and excess methanol recycling.	Soybean	Abiotic depletion potential, global warming potential, ozone depletion potential, photochemical oxidation potential, acidification potential, eutrophication potential, human toxicity potential, freshwater aquatic ecotoxicity potential, marine aquatic ecotoxicity potential, terrestrial ecotoxicity potential.	Lower impacts in comparison to first generation biofuels for most assessed impacts.	Assumed biomass productivities between 5-50 g m <sup>-2</sup> day <sup>-1</sup> and lipid contents between 15-80% in China conditions. Includes coproduct allocation and analyses for water recycling. Functional unit as 1 MJ of energy from biodiesel well-to-wheel.	[127]
N.S.	OP	Cradle-to-gate, including the processes upstream of dry biomass production.	Harvesting through flocculation and centrifugation.	Rapeseed, maize	Water use, greenhouse gas emissions, eutrophication potential, land use.	Higher impacts than first generation biofuels in terms of energy use, greenhouse gas emissions and water use, mainly driven by demand for CO <sub>2</sub> and fertilizer. Lower impacts for land use and eutrophication potential compared to first generation biofuels. Using wastewater leads to CO <sub>2</sub> savings and decreases water footprint.	This model was run for Virginia, Iowa, and California, USA. Included scenarios for colocation with wastewater and industrial CO <sub>2</sub> sources. Functional unit as 317 GJ of biomass-derived energy.	[39]
N.S.	OP-PB	Well-to-pump. From cultivation to biofuel final use at refueling stations.	1) Filtration through chamber filter press followed by drying and hexane extraction. 2) Centrifugation followed by drying and hexane extraction.	Soybean	CO <sub>2</sub> emissions, emissions of air pollutants (VOC, CO, NO <sub>x</sub> , particulate matter, SO <sub>x</sub> , CH <sub>4</sub> ).	Higher CO <sub>2</sub> savings in comparison to soybeans when using filter press and coproduct allocation. High energy consumption for thermal algae dewatering.	Assumed 30% lipids, 31% carbohydrates and 37.5% proteins. Includes recycling of water and addition of external CO <sub>2</sub> sources. Includes coproduct allocations. Functional unit as 1,000 MJ of energy at a refueling station.	[124]

N.S.	OP-PB	Well-to-wheel. Includes cultivation, harvesting and dewatering, lipid extraction, lipid conversion to a liquid transportation fuel, and coproducts from defatted algae.	1) Harvesting by best filter press followed by wet lipid extraction and hydrothermal liquefaction, hydrotreatment for lipid conversion and use of anaerobic digestion or animal feed. 2) Harvesting by centrifugation followed by hexane lipid extraction, lipid conversion by transesterification and use of anaerobic digestion or animal feed.	N.S.	Global warming potential.	Lower global warming potential for wet lipid extraction routes compared to dry extraction and for high productivity scenarios.	Estimated ranges of expected values of life cycle assessment metrics based on Monte Carlo simulations. Assumed 1210-ha microalgal facility using seawater and three ranges in biomass productivities: low (2.4–16 g m <sup>-2</sup> day <sup>-1</sup> ), base (17–33 g m <sup>-2</sup> day <sup>-1</sup> ), and high (34–50 g m <sup>-2</sup> day <sup>-1</sup> ). Functional unit defined as 1 MJ of liquid biofuel (biodiesel or “green” diesel).	[122]
N.S.	OP, PB	Cradle-to-wheel. From upstream of the delivered energy product to consumer use (passenger automobile).	Harvesting by auto-flocculation followed by thickening, homogenization, lipid extraction, solvent recovery and anaerobic digestion, belt-filter pressing and transesterification for biodiesel production.	N.S.	Climate change (global warming potential from greenhouse gas emissions), net water use, net eutrophication potential.	Open ponds that use brackish water are the preferred option for decreasing global warming potential.	Assumed biomass yields between 41.6-95.7 Mg ha <sup>-1</sup> year <sup>-1</sup> , and lipid contents between 13.4-32.4% using freshwater and brackish water species. Assumes use of virgin CO <sub>2</sub> from commercial sources. Includes production of biodiesel and methane-derived bioelectricity.	[120]
N.S.	OP, PB	Several system boundaries based on reviewed studies.	Several processing technologies based on reviewed studies.	Sugarcane	CO <sub>2</sub> emissions.	Higher variability in CO <sub>2</sub> emissions in comparison to sugarcane. Emissions decrease when using open raceway ponds and when recycling water.	Reviews different studies that relate CO <sub>2</sub> emissions with production technologies.	[121]
N.S.	OP	Cradle-to-grave, excluding the production facilities and its construction	Addition of flocculants for algae concentration followed by heating, centrifugation and lipid extraction using methanol and a catalyst. Anaerobic digestion for electricity generation.	Rapeseed	Greenhouse gas emissions (CO <sub>2</sub> , CH <sub>4</sub> , NO <sub>2</sub> ).	Higher CO <sub>2</sub> savings in comparison to rapeseed, highest when assuming high algae productivities and when using CO <sub>2</sub> from an ammonia plant.	Assumed biomass productivities at 15-30 g m <sup>-2</sup> day <sup>-1</sup> and use of salt water in Australian conditions. Includes three options for CO <sub>2</sub> feeding: in pure form from an ammonia plant, from flue gas (15% concentration) or delivered by truck in liquefied form. Functional unit as combustion of enough fuel in an articulated truck diesel engine to transport one ton of freight one kilometer.	[40]

<i>Nannochloropsis salina</i>	OP-PB	Well-to-pump, including microalgal cultivation through the delivery of fuel to the filling station. Well-to-wheel for comparison with conventional diesel.	Harvesting by settling, dissolved air flotation and centrifugation, followed by pressure homogenization, hexane extraction and nutrient recovery through anaerobic digestion.	N.S.	Greenhouse gas emissions (CO <sub>2</sub> , CH <sub>4</sub> , NO <sub>2</sub> ).	Lower CO <sub>2</sub> emissions for the scenario that includes improved algae productivity and anaerobic digestion.	Assumed biomass productivities at 25 g m <sup>-2</sup> day <sup>-1</sup> and lipid concentrations between 25-50%. Four scenarios were taken into account: baseline, improved algal productivity, supercritical CO <sub>2</sub> extraction, no nutrient recycling (lipid extracted biomass used as cattle feed). Functional unit as 1MJ of biofuel produced.	[128]
<i>Nannochloropsis</i> sp.	OP-PB	Cradle-to-gate, including microalgal cultivation through biodiesel production.	Dewatering and drying through the use of flocculants and centrifugation, followed hexane extraction and transesterification.	N.S.	CO <sub>2</sub> emissions.	High energy consumption for lipid extraction and biodiesel production. CO <sub>2</sub> savings were not found.	Assumed biomass productivities of 25 g m <sup>-2</sup> day <sup>-1</sup> in Singapore conditions and using seawater. Lipid contents between 25-45%. Functional unit as 1 MJ biofuel.	[181]
<i>Phaeodactylum</i> sp., <i>Tetraselmis</i> sp.	OP	Well-to-wheel. Includes extraction of raw materials, cultivation and lipid extraction, conversion and use of biofuel in vehicles.	Cultivation followed by harvesting through auto-flocculation, thickening and homogenization. Several scenarios for biomass processing: 1) Anaerobic digestion of bulk algae biomass for production of electricity from methane. 2) Production of biodiesel from algae lipids coupled with anaerobic digestion for producing electricity. 3) Production of biodiesel from lipids and direct combustion for electricity production from residual algae biomass. 4) Direct combustion of algae biomass for producing electricity.	Rapeseed, maize	Net energy use, water use, and greenhouse gas emissions.	Highest energy efficiencies when using direct combustion of algae biomass for producing electricity, and lowest when producing biodiesel from algae lipids coupled with anaerobic digestion for producing electricity. Use of wastewater and flue gas increases energy efficiencies. Algae systems are better than rapeseed and maize in relation to vehicle kilometers traveled per ha. Greenhouse gas emissions and water used per kilometer traveled are lower compared to rapeseed	Assumed biomass yields of 91.1 Mg ha <sup>-1</sup> year <sup>-1</sup> and lipid contents at 19.6% using brackish water species in Southwestern USA conditions and in marginal lands. Includes scenarios for CO <sub>2</sub> sources: virgin CO <sub>2</sub> , carbon capture from coal-fired using chemical sorption, use of flue gas 12.5% CO <sub>2</sub> power plant. Includes one scenario for wastewater use. Makes use of stochastic inputs to capture uncertainty in processes. Functional unit as usable energy production per unit land area (vehicle kilometers traveled per ha) and environmental burdens (net energy use, water use, and greenhouse gas emissions per vehicle kilometers traveled).	[38]
<i>Scenedesmus dimorphus</i>	OP	Well-to-pump. Cultivation, dewatering, thermochemical bio-oil recovery, bio-oil stabilization, conversion to renewable diesel, and transport to the pump.	Harvesting by membrane filtration and centrifugation, followed by thermochemical conversion (hydrothermal liquefaction vs. pyrolysis)	Maize, soybean	Net energy ratio, greenhouse gas emissions.	Hydrothermal liquefaction leads to carbon savings in contrast to pyrolysis. Carbon savings are higher in comparison to maize bioethanol.	Biomass productivities at 6.5 g m <sup>-2</sup> day <sup>-1</sup> in Arizona conditions.	[129]

Several species	OP	Upstream resources, cultivation, conversion into biodiesel followed by anaerobic digestion.	Normalization of studies based on cultivation in open ponds, conversion into biodiesel and use of anaerobic digestion.	Maize, soybean	Greenhouse gas emissions	Energy consumption and greenhouse gas emissions would be similar to those obtained for terrestrial alternatives.	Meta-analysis based on six life cycle assessments for microalgal biofuel production. Functional unit set as 1,000 L biodiesel.	[118]
Several species	OP	Several system boundaries based on reviewed studies.	Several technologies based on the reviewed studies.	Rapeseed	Greenhouse gas emissions. Overview for water use, land use, nutrient and fertilizer use, carbon fertilization, fossil fuel inputs, eutrophication, genetically modified algae, algal toxicity.	Decreases in CO <sub>2</sub> for raceway ponds compared to photobioreactors, reaching similar values to those obtained for rapeseed. Major energy inputs are associated with pumping and mixing during cultivation and to the provision of heat for algae drying.	Review of seven life cycle assessments.	[42]
Several species	OP, PB	Several system boundaries based on reviewed studies.	Several biomass processing methods including solvent extraction, hydrothermal liquefaction, secretion, pyrolysis, supercritical water, in-situ transesterification.	N.S.	Global warming potential.	Global warming potential varies between production technologies and system boundaries. Thermochemical conversion and anaerobic digestion seem promising alternatives that reduce energy inputs.	Review that includes information about global warming potential for a set of microalgal production technologies.	[92]
<i>Tetraselmis chui</i>	PB	Cradle-to-grave, including cultivation, harvesting, processing and products (utilization and consumption).	Harvesting through primary to tertiary dewatering and spray drying, followed by slow pyrolysis, oil extraction by solvent and production of biogas, bio-oil, biodiesel and biochar.	Rapeseed, soybean	Global warming, abiotic resource depletion (excluding water), land transformation and use, water resource depletion, eutrophication, acidification, eco-toxicity, human toxicity, photochemical smog, ozone depletion, ionizing radiation, respiratory effects.	Lower global warming and land use in comparison to first generation biofuels. Higher eutrophication, water use, ecotoxicity, acidification, photochemical smog and respiratory effects in comparison to first generation biofuels. Improvements are expected if using hydrothermal liquefaction.	The system was modeled in Queensland conditions, Australia. Includes coproduct allocation, CO <sub>2</sub> addition from power plant station (13%), water use for evaporative cooling and water recycling. Functional units defined as 1 MJ of pyrolysis biogas combusted for electricity and 1 MJ of pyrolysis bio-oil combusted for electricity or extracted lipid refined for transport fuel.	[126]

Table 2. Comparison of widely used first generation biofuel crops in relation to their potential for genetic pollution and invasiveness.

<b>Biofuel crop</b>	<b>Center of origin</b>	<b>Dispersal units</b>	<b>Non-human effective dispersal vectors</b>	<b>Reported genetic pollution</b>	<b>Reported invasiveness</b>
Oil palm ( <i>Elaeis guineensis</i> )	Tropical Africa [182]	Seeds	Animals	No	Yes [183, 184]
Maize ( <i>Zea mays</i> )	Americas	Seeds	N.A.	Yes [185, 186]	No
Rapeseed ( <i>Brassica napus</i> )	Mediterranean region [187]	Seeds	Autochory, wind, water, animals [188]	Yes [189, 190]	Yes [191, 192]
Sugarcane ( <i>Saccharum</i> sp.)	Tropical region [193]	Cuts, seeds (low viability of seeds)	N.A.	No	No
Soybeans ( <i>Glycine max</i> )	China	Seeds	Autochory [194]	Yes [195, 196]	No

Figure 1. Citation report using the Science Citation Index Expanded (SCI-EXPANDED) and the Emerging Sources Citation Index (*ESCI*) in Web of Science and the following combination of keywords: (biofuel OR bioenergy) AND (biodiversity OR wildlife), (biofuel OR bioenergy) AND (fish\* OR bird\* OR avian OR mammal\* OR reptil\* OR amphibian\*).

Figure 2. Drivers, Pressures, States, Impacts and Responses of biofuel production on biodiversity, based on the reviewed literature and following the DPSIR framework.

Figure 3. Superimposed circles showing the area of cultivation land (km<sup>2</sup>) required to meet gasoline and distillate fuel oil demands for each country in 2010, when comparing microalgal systems with the most productive biodiesel and bioethanol crop per country. For first generation biofuels, yields are based on areas where crops possibly grow best (average yields between 2005 and 2014) [55], while the most frequent value (area-weighted average) of average lipid yield within countries is used for microalgal systems. Microalgal lipid estimations are based on Moody, McGinty [61] .

Figure 4. Water consumption per unit of produced energy (GJ) derived from biodiesel (soybean, oil palm, microalgae) and bioethanol (maize, sugarcane). Based on calculations by Gerbens-Leenes, Xu [131] for wet conversion of microalgal biodiesel and assuming water recycling. Available water footprints for first generation biofuels were obtained from Mekonnen and Hoekstra [149]. Microalgal systems in New Mexico and Perth consist of open ponds using salty water (OPS), in Hawaii correspond to a combination of open ponds and photobioreactors using fresh water (OPF-PBF), in Italy consist of photobioreactors

using salty water (PBS), and in the Netherlands, France and Algeria consist of photobioreactors using fresh water (PBF).

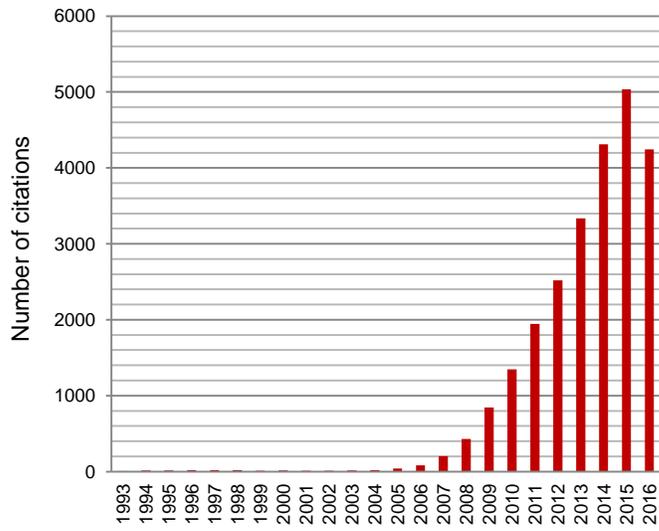


Figure 1.

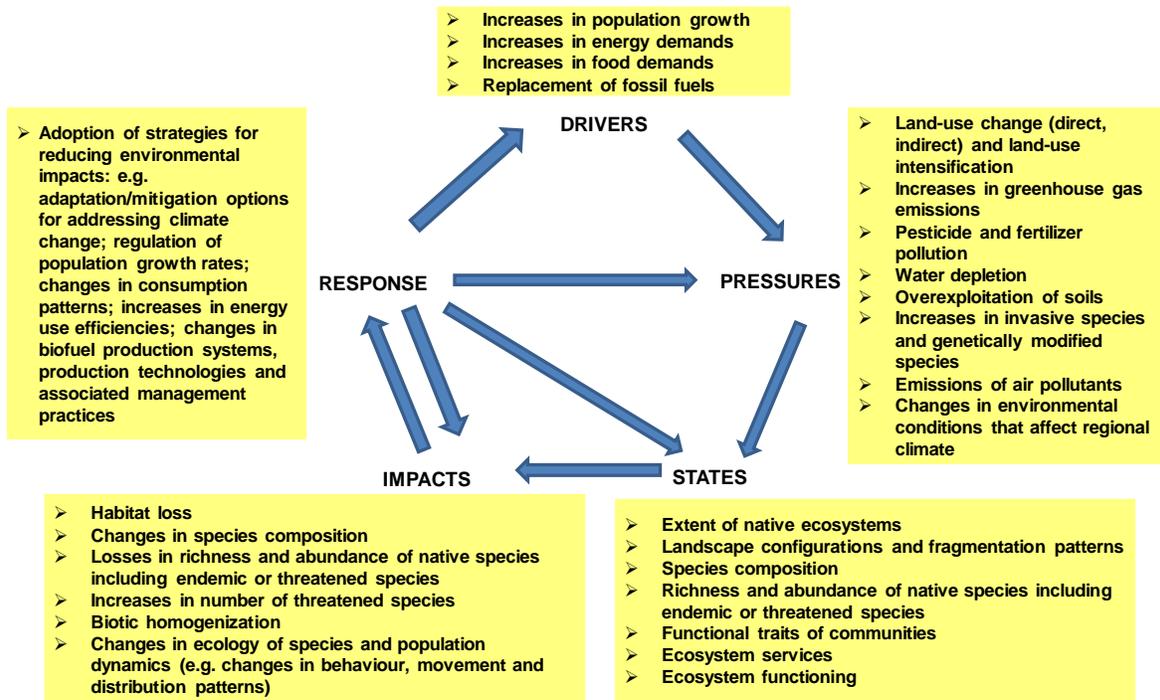


Figure 2.

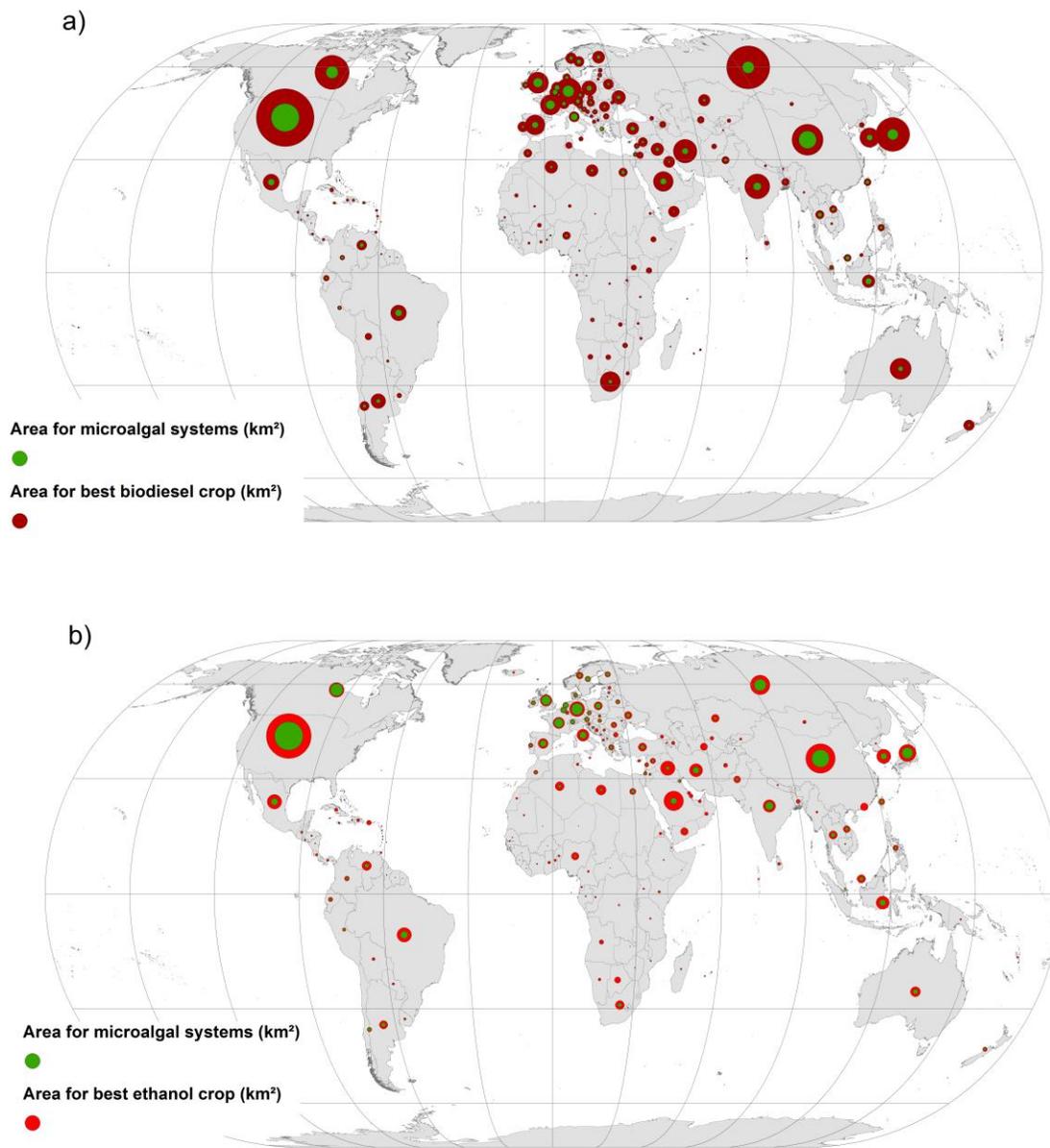


Figure 3.

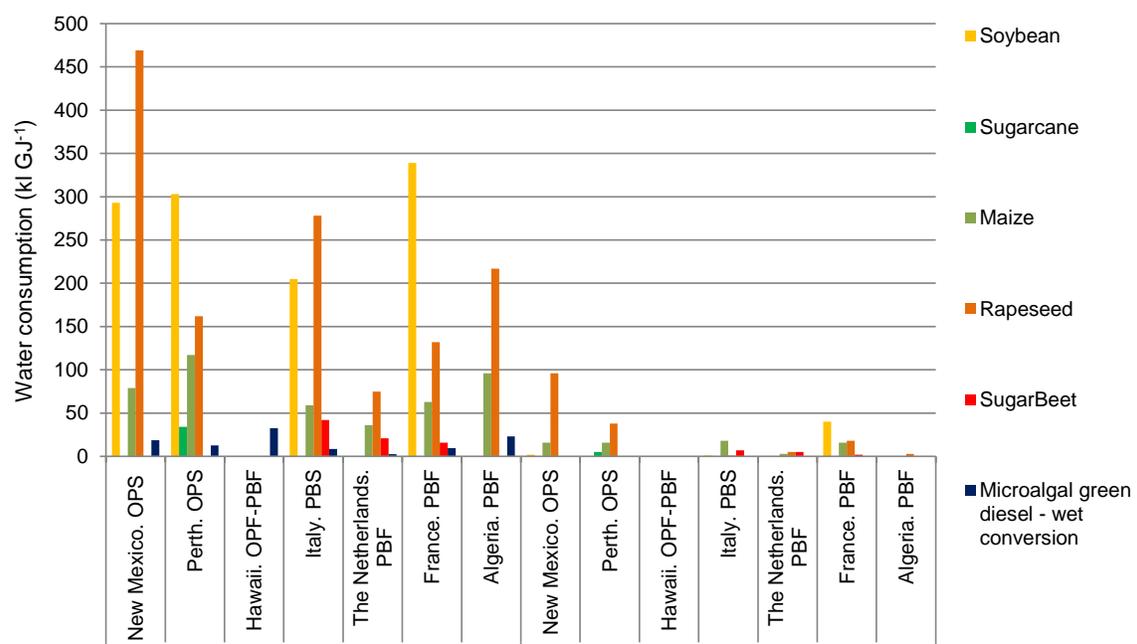


Figure 4.

## LITERATURE CITED

- [1] IEA. World Energy Outlook 2014. Paris, France: IEA Publications; 2014.
- [2] Bellard C, Bertelsmeier C, Leadley P, Thuiller W, Courchamp F. Impacts of climate change on the future of biodiversity. *Ecology letters*. 2012;15:365-77.
- [3] Thomas CD, Cameron A, Green RE, Bakkenes M, Beaumont LJ, Collingham YC, et al. Extinction risk from climate change. *Nature*. 2004;427:145-8.
- [4] Jacobson MZ, Delucchi MA. Providing all global energy with wind, water, and solar power, Part I: Technologies, energy resources, quantities and areas of infrastructure, and materials. *Energy Policy*. 2011;39:1154-69.
- [5] Goldemberg J. The promise of clean energy. *Energy Policy*. 2006;34:2185-90.
- [6] Panwar NL, Kaushik SC, Kothari S. Role of renewable energy sources in environmental protection: A review. *Renewable and Sustainable Energy Reviews*. 2011;15:1513-24.
- [7] Hill J, Nelson E, Tilman D, Polasky S, Tiffany D. Environmental, economic, and energetic costs and benefits of biodiesel and ethanol biofuels. *Proceedings of the National Academy of Sciences of the United States of America*. 2006;103:11206-10.
- [8] Williams CL, Dahiya A, Porter P. Introduction to Bioenergy. In: Dahiya A, editor. *Bioenergy Biomass to biofuels*. London: Academic Press; 2015. p. 5-36.
- [9] Naik SN, Goud VV, Rout PK, Dalai AK. Production of first and second generation biofuels: A comprehensive review. *Renewable and Sustainable Energy Reviews*. 2010;14:578-97.
- [10] McKendry P. Energy production from biomass (part 1): overview of biomass. *Bioresource technology*. 2002;83:37-46.
- [11] Tilman D, Socolow R, Foley JA, Hill J, Larson E, Lynd L, et al. Beneficial biofuels—the food, energy, and environment trilemma. *Science*. 2009;325:270-1.
- [12] Immerzeel DJ, Verweij P, Hilst F, Faaij AP. Biodiversity impacts of bioenergy crop production: a state-of-the-art review. *GCB Bioenergy*. 2014;6:183-209.
- [13] OECD/FAO. *OECD-FAO Agricultural Outlook 2016-2025*. Paris: OECD Publishing; 2016.
- [14] Fargione JE, Cooper TR, Flaspohler DJ, Hill J, Lehman C, Tilman D, et al. Bioenergy and Wildlife: Threats and Opportunities for Grassland Conservation. *BioScience*. 2009;59:767-77.
- [15] Danielsen F, Beukema H, Burgess ND, Parish F, Bruhl CA, Donald PF, et al. Biofuel plantations on forested lands: double jeopardy for biodiversity and climate. *Conservation biology : the journal of the Society for Conservation Biology*. 2009;23:348-58.

- [16] Koh LP, Miettinen J, Liew SC, Ghazoul J. Remotely sensed evidence of tropical peatland conversion to oil palm. *Proceedings of the National Academy of Sciences of the United States of America*. 2011;108:5127-32.
- [17] Fargione JE, Plevin RJ, Hill JD. The Ecological Impact of Biofuels. *Annual Review of Ecology, Evolution, and Systematics*. 2010;41:351-77.
- [18] Fletcher RJ, Robertson BA, Evans J, Doran PJ, Alavalapati JRR, Schemske DW. Biodiversity conservation in the era of biofuels: risks and opportunities. *Frontiers in Ecology and the Environment*. 2011;9:161-8.
- [19] Duke C, Pouyat R, Robertson G, Parton W. Ecological dimensions of biofuels. *Issues in Ecology*. 2013;17:1-17.
- [20] Fargione J, Hill J, Tilman D, Polasky S, Hawthorne P. Land clearing and the biofuel carbon debt. *Science*. 2008;319:1235-8.
- [21] Searchinger T, Heimlich R, Houghton RA, Dong F, Elobeid A, Fabiosa J, et al. Use of US croplands for biofuels increases greenhouse gases through emissions from land-use change. *Science*. 2008;319:1238-40.
- [22] Searchinger TD, Estes L, Thornton PK, Beringer T, Notenbaert A, Rubenstein D, et al. High carbon and biodiversity costs from converting Africa's wet savannahs to cropland. *Nature Climate Change*. 2015;5:481-6.
- [23] Crutzen PJ, Mosier AR, Smith KA, Winiwarter W. N<sub>2</sub>O release from agro-biofuel production negates global warming reduction by replacing fossil fuels. *Atmospheric chemistry and physics*. 2008;8:389-95.
- [24] Larson ED. A review of life-cycle analysis studies on liquid biofuel systems for the transport sector. *Energy for sustainable development*. 2006;10:109-26.
- [25] Cherubini F, Bird ND, Cowie A, Jungmeier G, Schlamadinger B, Woess-Gallasch S. Energy-and greenhouse gas-based LCA of biofuel and bioenergy systems: Key issues, ranges and recommendations. *Resources, conservation and recycling*. 2009;53:434-47.
- [26] Dominguez-Faus R, Powers SE, Burken JG, Alvarez PJ. The water footprint of biofuels: A drink or drive issue? *Environmental science & technology*. 2009;43:3005-10.
- [27] Gerbens-Leenes PW, Lienden ARv, Hoekstra AY, van der Meer TH. Biofuel scenarios in a water perspective: The global blue and green water footprint of road transport in 2030. *Global Environmental Change*. 2012;22:764-75.
- [28] Gerbens-Leenes W, Hoekstra AY, van der Meer TH. The water footprint of bioenergy. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;106:10219-23.
- [29] Chisti Y. Biodiesel from microalgae. *Biotechnology advances*. 2007;25:294-306.

- [30] Schenk PM, Thomas-Hall SR, Stephens E, Marx UC, Mussgnug JH, Posten C, et al. Second Generation Biofuels: High-Efficiency Microalgae for Biodiesel Production. *BioEnergy Research*. 2008;1:20-43.
- [31] Lundquist TJ, Woertz IC, Quinn NWT, Benemann JR. A Realistic Technology and Engineering Assessment of Algae Biofuel Production. In: Energy Biosciences Institute UoC, editor. Berkeley, California 2010.
- [32] Mata TM, Martins AA, Caetano NS. Microalgae for biodiesel production and other applications: A review. *Renewable and Sustainable Energy Reviews*. 2010;14:217-32.
- [33] Wijffels RH, Barbosa MJ. An outlook on microalgal biofuels. *Science*. 2010;329:796-9.
- [34] Chisti Y. Biodiesel from microalgae beats bioethanol. *Trends in biotechnology*. 2008;26:126-31.
- [35] Brennan L, Owende P. Biofuels from microalgae—A review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and Sustainable Energy Reviews*. 2010;14:557-77.
- [36] Sayre R. Microalgae: the potential for carbon capture. *Bioscience*. 2010;60:722-7.
- [37] Lardon L, Helias A, Sialve B, Steyer J-P, Bernard O. Life-cycle assessment of biodiesel production from microalgae. *Environmental science & technology*. 2009;43:6475-81.
- [38] Clarens AF, Nassau H, Resurreccion EP, White MA, Colosi LM. Environmental impacts of algae-derived biodiesel and bioelectricity for transportation. *Environmental science & technology*. 2011;45:7554-60.
- [39] Clarens AF, Resurreccion EP, White MA, Colosi LM. Environmental life cycle comparison of algae to other bioenergy feedstocks. *Environmental science & technology*. 2010;44:1813-9.
- [40] Campbell PK, Beer T, Batten D. Life cycle assessment of biodiesel production from microalgae in ponds. *Bioresource technology*. 2011;102:50-6.
- [41] Collet P, Helias A, Lardon L, Ras M, Goy RA, Steyer JP. Life-cycle assessment of microalgae culture coupled to biogas production. *Bioresource technology*. 2011;102:207-14.
- [42] Slade R, Bauen A. Micro-algae cultivation for biofuels: Cost, energy balance, environmental impacts and future prospects. *Biomass and Bioenergy*. 2013;53:29-38.
- [43] Zaimes GG, Khanna V. Microalgal biomass production pathways: evaluation of life cycle environmental impacts. *Biotechnology for biofuels*. 2013;6:1.
- [44] Menetrez MY. An overview of algae biofuel production and potential environmental impact. *Environmental science & technology*. 2012;46:7073-85.
- [45] Dirzo R, Raven PH. Global state of biodiversity and loss. *Annual Review of Environment and Resources*. 2003;28:137-67.

- [46] Kier G, Mutke J, Dinerstein E, Ricketts TH, Küper W, Kreft H, et al. Global patterns of plant diversity and floristic knowledge. *Journal of Biogeography*. 2005;32:1107-16.
- [47] Laurance WF. Emerging threats to tropical forests. *Annals of the Missouri Botanical Garden*. 2015;100:159-69.
- [48] Laurance WF, Sayer J, Cassman KG. Agricultural expansion and its impacts on tropical nature. *Trends in ecology & evolution*. 2014;29:107-16.
- [49] Kristensen P. The DPSIR framework. National Environmental Research Institute, Denmark. 2004;10.
- [50] Smeets E, Weterings R. Environmental indicators: Typology and overview: European Environment Agency Copenhagen; 1999.
- [51] Gabrielsen P, Bosch P. Environmental indicators: typology and use in reporting. EEA, Copenhagen. 2003.
- [52] Maxim L, Spangenberg JH, O'Connor M. An analysis of risks for biodiversity under the DPSIR framework. *Ecological Economics*. 2009;69:12-23.
- [53] Atkins JP, Burdon D, Elliott M, Gregory AJ. Management of the marine environment: integrating ecosystem services and societal benefits with the DPSIR framework in a systems approach. *Marine pollution bulletin*. 2011;62:215-26.
- [54] U.S. Energy Information Administration. U.S. Energy Information Administration; 2016.
- [55] FAO. Food and Agriculture Organization of the United Nations, Statistics Division; 2016.
- [56] El Bassam N. Handbook of bioenergy crops: a complete reference to species, development and applications. New York: Routledge; 2010.
- [57] Rajagopal D, Sexton SE, Roland-Holst D, Zilberman D. Challenge of biofuel: filling the tank without emptying the stomach? *Environmental Research Letters*. 2007;2:044004.
- [58] Wang S, Thomas K, Ingledew W, Sosulski K, Sosulski F. Rye and triticale as feedstock for fuel ethanol production. *Cereal Chemistry*. 1997;74:621-5.
- [59] de Vries SC, van de Ven GWJ, van Ittersum MK, Giller KE. Resource use efficiency and environmental performance of nine major biofuel crops, processed by first-generation conversion techniques. *Biomass and Bioenergy*. 2010;34:588-601.
- [60] Firestone D. Physical and chemical characteristics of oils, fats, and waxes (3rd Edition). Boulder, Urbana: AOCS press; 2013.
- [61] Moody JW, McGinty CM, Quinn JC. Global evaluation of biofuel potential from microalgae. *Proceedings of the National Academy of Sciences of the United States of America*. 2014;111:8691-6.

- [62] Zhu LD, Ketola T. Microalgae production as a biofuel feedstock: risks and challenges. *International Journal of Sustainable Development and World Ecology*. 2012;19:268-74.
- [63] Zhu L, Huo S, Qin L. A Microalgae-Based Biodiesel Refinery: Sustainability Concerns and Challenges. *International Journal of Green Energy*. 2015;12:595-602.
- [64] Usher PK, Ross AB, Camargo-Valero MA, Tomlin AS, Gale WF. An overview of the potential environmental impacts of large-scale microalgae cultivation. *Biofuels*. 2014;5:331-49.
- [65] McLaughlin A, Mineau P. The impact of agricultural practices on biodiversity. *Agriculture, Ecosystems & Environment*. 1995;55:201-12.
- [66] Donald PF. Biodiversity impacts of some agricultural commodity production systems. *Conservation biology*. 2004;18:17-38.
- [67] Maxwell SL, Venter O, Jones KR, Watson JE. Integrating human responses to climate change into conservation vulnerability assessments and adaptation planning. *Annals of the New York Academy of Sciences*. 2015;1355:98-116.
- [68] Watson JEM. Human Responses to Climate Change will Seriously Impact Biodiversity Conservation: It's Time We Start Planning for Them. *Conservation Letters*. 2014;7:1-2.
- [69] Balvanera P, Daily GC, Ehrlich PR, Ricketts TH, Bailey S-A, Kark S, et al. Conserving Biodiversity and Ecosystem Services. *Science*. 2001;291:2047.
- [70] Fahrig L. Effects of habitat fragmentation on biodiversity. *Annual review of ecology, evolution, and systematics*. 2003:487-515.
- [71] Krauss J, Bommarco R, Guardiola M, Heikkinen RK, Helm A, Kuussaari M, et al. Habitat fragmentation causes immediate and time-delayed biodiversity loss at different trophic levels. *Ecology letters*. 2010;13:597-605.
- [72] Azhar B, Lindenmayer DB, Wood J, Fischer J, Manning A, McElhinny C, et al. The conservation value of oil palm plantation estates, smallholdings and logged peat swamp forest for birds. *Forest Ecology and Management*. 2011;262:2306-15.
- [73] Savilaakso S, Garcia C, Garcia-Ulloa J, Ghazoul J, Groom M, Guariguata MR, et al. Systematic review of effects on biodiversity from oil palm production. *Environmental Evidence*. 2014;3:4.
- [74] Meehan TD, Hurlbert AH, Gratton C. Bird communities in future bioenergy landscapes of the Upper Midwest. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;107:18533-8.
- [75] Koh LP, Wilcove DS. Is oil palm agriculture really destroying tropical biodiversity? *Conservation Letters*. 2008;1:60-4.
- [76] Carrete M, Tella JL, Blanco G, Bertellotti M. Effects of habitat degradation on the abundance, richness and diversity of raptors across Neotropical biomes. *Biological Conservation*. 2009;142:2002-11.

- [77] Alkemade R, van Oorschot M, Miles L, Nellemann C, Bakkenes M, ten Brink B. GLOBI03: A Framework to Investigate Options for Reducing Global Terrestrial Biodiversity Loss. *Ecosystems*. 2009;12:374-90.
- [78] Buchanan GM, Butchart SHM, Dutson G, Pilgrim JD, Steininger MK, Bishop KD, et al. Using remote sensing to inform conservation status assessment: Estimates of recent deforestation rates on New Britain and the impacts upon endemic birds. *Biological Conservation*. 2008;141:56-66.
- [79] Codesido M, González-Fischer C, Bilenca D. Distributional changes of landbird species in agroecosystems of central Argentina. *The Condor*. 2011;113:266-73.
- [80] Savilaakso S, Garcia C, Garcia-Ulloa J, Ghazoul J, Groom M, Guariguata MR, et al. Systematic review of effects on biodiversity from oil palm production. *Environmental Evidence*. 2014;3:(25 February 2014)-(25 February ).
- [81] Mahood SP, Lees AC, Peres CA. Amazonian countryside habitats provide limited avian conservation value. *Biodiversity and Conservation*. 2012;21:385-405.
- [82] Vargas LEP, Laurance WF, Clements GR, Edwards W. The impacts of oil palm agriculture on Colombia's biodiversity: what we know and still need to know. *Tropical Conservation Science*. 2015;8:828-45.
- [83] Rajaratnam R, Sunquist M, Rajaratnam L, Ambu L. Diet and habitat selection of the leopard cat (*Prionailurus bengalensis borneoensis*) in an agricultural landscape in Sabah, Malaysian Borneo. *Journal of Tropical Ecology*. 2007;23:209-17.
- [84] Fitzherbert EB, Struebig MJ, Morel A, Danielsen F, Bruehl CA, Donald PF, et al. How will oil palm expansion affect biodiversity? *Trends in ecology & evolution*. 2008;23:538-45.
- [85] Geyer R, Stoms DM, Lindner JP, Davis FW, Wittstock B. Coupling GIS and LCA for biodiversity assessments of land use. *International Journal of Life Cycle Assessment*. 2010;15:454-67.
- [86] Nantha HS, Tisdell C. The orangutan-oil palm conflict: economic constraints and opportunities for conservation. *Biodiversity and Conservation*. 2009;18:487-502.
- [87] Robertson BA, Doran PJ, Loomis LR, Robertson JR, Schemske DW. Perennial biomass feedstocks enhance avian diversity. *GCB Bioenergy*. 2011;3:235-46.
- [88] Wich SA, Garcia-Ulloa J, Kuehl HS, Humle T, Lee JSH, Koh LP. Will Oil Palm's Homecoming Spell Doom for Africa's Great Apes? *Current Biology*. 2014;24:1659-63.
- [89] Treves A, Wallace RB, Naughton-Treves L, Morales A. Co-Managing Human–Wildlife Conflicts: A Review. *Human Dimensions of Wildlife*. 2006;11:383-96.
- [90] Michalski F, Boulhosa RLP, Faria A, Peres CA. Human-wildlife conflicts in a fragmented Amazonian forest landscape: determinants of large felid depredation on livestock. *Animal Conservation*. 2006;9:179-88.

- [91] Peres CA. Effects of Subsistence Hunting on Vertebrate Community Structure in Amazonian Forests. *Conservation Biology*. 2000;14:240-53.
- [92] Quinn JC, Davis R. The potentials and challenges of algae based biofuels: A review of the techno-economic, life cycle, and resource assessment modeling. *Bioresource technology*. 2015;184:444-52.
- [93] Schenk PM. Final Report: On-farm algal ponds to provide protein for northern cattle. North Sydney NSW 2059: Meat and Livestock Australia Limited; 2016. p. 46.
- [94] Plieninger T, Gaertner M. Harnessing degraded lands for biodiversity conservation. *Journal for Nature Conservation*. 2011;19:18-23.
- [95] Cottier T, Nartova O, Bigdeli SZ. International trade regulation and the mitigation of climate change: World Trade Forum. Cambridge: cambridge university Press; 2009.
- [96] Edwards R, Mulligan D, Marelli L. Indirect Land Use Change From Increased Biofuels Demand - Comparison of Models and Results for Marginal Biofuels Production from Different Feedstocks. EC Joint Research Centre, Ispra; 2010.
- [97] Lapola DM, Schaldach R, Alcamo J, Bondeau A, Koch J, Koelking C, et al. Indirect land-use changes can overcome carbon savings from biofuels in Brazil. *Proceedings of the national Academy of Sciences*. 2010;107:3388-93.
- [98] Alexandratos N, Bruinsma J. World agriculture towards 2030/2050: the 2012 revision. ESA Work Pap. 2012;3.
- [99] Castanheira EG, Grisoli R, Freire F, Pecora V, Coelho ST. Environmental sustainability of biodiesel in Brazil. *Energy Policy*. 2014;65:680-91.
- [100] Frank S, Bottcher H, Havlik P, Valin H, Mosnier A, Obersteiner M, et al. How effective are the sustainability criteria accompanying the European Union 2020 biofuel targets? *Global Change Biology Bioenergy*. 2013;5:306-14.
- [101] Pelikan J, Britz W, Hertel TW. Green Light for Green Agricultural Policies? An Analysis at Regional and Global Scales. *Journal of Agricultural Economics*. 2015;66:1-19.
- [102] Schleupner C, Schneider UA. Effects of bioenergy policies and targets on European wetland restoration options. *Environmental Science & Policy*. 2010;13:721-32.
- [103] Zilberman D, Hochman G, Rajagopal D, Sexton S, Timilsina G. The Impact of Biofuels on Commodity Food Prices: Assessment of Findings. *American Journal of Agricultural Economics*. 2013;95:275-81.
- [104] Mitchell D. A note on rising food prices. World Bank Policy Research Working Paper 4682: The World Bank, Development Prospects Group 2008.

- [105] Lambin EF, Meyfroidt P. Global land use change, economic globalization, and the looming land scarcity. *Proceedings of the National Academy of Sciences of the United States of America*. 2011;108:3465-72.
- [106] Soares-Filho BS, Nepstad DC, Curran LM, Cerqueira GC, Garcia RA, Ramos CA, et al. Modelling conservation in the Amazon basin. *Nature*. 2006;440:520-3.
- [107] Lee JSH, Garcia-Ulloa J, Koh LP. Impacts of Biofuel Expansion in Biodiversity Hotspots. In: Zachos FE, Habel JC, editors. *Biodiversity Hotspots: Distribution and Protection of Conservation Priority Areas*. Berlin, Heidelberg: Springer Berlin Heidelberg; 2011. p. 277-93.
- [108] Casson A. Oil palm, soybeans & critical habitat loss. World Wide Fund for Nature, Gland, Switzerland. 2003.
- [109] Fritsche UR, Sims REH, Monti A. Direct and indirect land-use competition issues for energy crops and their sustainable production - an overview. *Biofuels, Bioproducts and Biorefining*. 2010;4:692-704.
- [110] Creutzig F, Ravindranath N, Berndes G, Bolwig S, Bright R, Cherubini F, et al. Bioenergy and climate change mitigation: an assessment. *GCB Bioenergy*. 2015;7:916-44.
- [111] Fearnside PM. Global warming and tropical land-use change: greenhouse gas emissions from biomass burning, decomposition and soils in forest conversion, shifting cultivation and secondary vegetation. *Climatic change*. 2000;46:115-58.
- [112] Guo LB, Gifford R. Soil carbon stocks and land use change: a meta analysis. *Global change biology*. 2002;8:345-60.
- [113] Don A, Osborne B, Hastings A, Skiba U, Carter MS, Drewer J, et al. Land-use change to bioenergy production in Europe: implications for the greenhouse gas balance and soil carbon. *Gcb Bioenergy*. 2012;4:372-91.
- [114] Stephenson AL, Kazamia E, Dennis JS, Howe CJ, Scott SA, Smith AG. Life-cycle assessment of potential algal biodiesel production in the United Kingdom: a comparison of raceways and air-lift tubular bioreactors. *Energy & Fuels*. 2010;24:4062-77.
- [115] Walsh BJ, Rydzak F, Palazzo A, Kraxner F, Herrero M, Schenk PM, et al. New feed sources key to ambitious climate targets. *Carbon balance and management*. 2015;10:1-8.
- [116] Quiroz-Arita C, Yilmaz Ö, Barlak S, Catton KB, Quinn JC, Bradley TH. A geographical assessment of vegetation carbon stocks and greenhouse gas emissions on potential microalgae-based biofuel facilities in the United States. *Bioresource technology*. 2016;221:270-5.
- [117] Snyder C, Bruulsema T, Jensen T, Fixen P. Review of greenhouse gas emissions from crop production systems and fertilizer management effects. *Agriculture, Ecosystems & Environment*. 2009;133:247-66.
- [118] Liu X, Clarens AF, Colosi LM. Algae biodiesel has potential despite inconclusive results to date. *Bioresource technology*. 2012;104:803-6.

- [119] Handler RM, Canter CE, Kalnes TN, Lupton FS, Kholiqov O, Shonnard DR, et al. Evaluation of environmental impacts from microalgae cultivation in open-air raceway ponds: Analysis of the prior literature and investigation of wide variance in predicted impacts. *Algal Research*. 2012;1:83-92.
- [120] Resurreccion EP, Colosi LM, White MA, Clarens AF. Comparison of algae cultivation methods for bioenergy production using a combined life cycle assessment and life cycle costing approach. *Bioresource technology*. 2012;126:298-306.
- [121] Kendall A, Yuan J. Comparing life cycle assessments of different biofuel options. *Current opinion in chemical biology*. 2013;17:439-43.
- [122] Sills DL, Paramita V, Franke MJ, Johnson MC, Akabas TM, Greene CH, et al. Quantitative uncertainty analysis of life cycle assessment for algal biofuel production. *Environmental science & technology*. 2012;47:687-94.
- [123] Xu L, Wim Brilman DW, Withag JA, Brem G, Kersten S. Assessment of a dry and a wet route for the production of biofuels from microalgae: energy balance analysis. *Bioresource technology*. 2011;102:5113-22.
- [124] Sander K, Murthy GS. Life cycle analysis of algae biodiesel. *The International Journal of Life Cycle Assessment*. 2010;15:704-14.
- [125] Shirvani T, Yan X, Inderwildi OR, Edwards PP, King DA. Life cycle energy and greenhouse gas analysis for algae-derived biodiesel. *Energy & Environmental Science*. 2011;4:3773-8.
- [126] Grierson S, Strezov V, Bengtsson J. Life cycle assessment of a microalgae biomass cultivation, bio-oil extraction and pyrolysis processing regime. *Algal Research*. 2013;2:299-311.
- [127] Hou J, Zhang P, Yuan X, Zheng Y. Life cycle assessment of biodiesel from soybean, jatropha and microalgae in China conditions. *Renewable and Sustainable Energy Reviews*. 2011;15:5081-91.
- [128] Quinn JC, Smith TG, Downes CM, Quinn C. Microalgae to biofuels lifecycle assessment—multiple pathway evaluation. *Algal Research*. 2014;4:116-22.
- [129] Bennion EP, Ginosar DM, Moses J, Agblevor F, Quinn JC. Lifecycle assessment of microalgae to biofuel: Comparison of thermochemical processing pathways. *Applied Energy*. 2015;154:1062-71.
- [130] Wijffels RH, Barbosa MJ, Eppink MH. Microalgae for the production of bulk chemicals and biofuels. *Biofuels, Bioproducts and Biorefining*. 2010;4:287-95.
- [131] Gerbens-Leenes P, Xu L, Vries dG, Hoekstra A. The blue water footprint and land use of biofuels from algae. *Water resources research*. 2014;50:8549-63.
- [132] Sialve B, Bernet N, Bernard O. Anaerobic digestion of microalgae as a necessary step to make microalgal biodiesel sustainable. *Biotechnology advances*. 2009;27:409-16.

- [133] Gibbons D, Morrissey C, Mineau P. A review of the direct and indirect effects of neonicotinoids and fipronil on vertebrate wildlife. *Environmental Science and Pollution Research*. 2015;22:103-18.
- [134] Parsons KC, Mineau P, Renfrew RB. Effects of pesticide use in rice fields on birds. *Waterbirds*. 2010;33:193-218.
- [135] Köhler H-R, Triebkorn R. Wildlife ecotoxicology of pesticides: can we track effects to the population level and beyond? *Science*. 2013;341:759-65.
- [136] Hayes TB, Case P, Chui S, Chung D, Haeffele C, Haston K, et al. Pesticide mixtures, endocrine disruption, and amphibian declines: are we underestimating the impact? *Environmental Health Perspectives*. 2006;114:40.
- [137] Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. Global pollinator declines: trends, impacts and drivers. *Trends in ecology & evolution*. 2010;25:345-53.
- [138] Atafar Z, Mesdaghinia A, Nouri J, Homae M, Yunesian M, Ahmadimoghaddam M, et al. Effect of fertilizer application on soil heavy metal concentration. *Environmental monitoring and assessment*. 2010;160:83-9.
- [139] Anderson DM, Glibert PM, Burkholder JM. Harmful algal blooms and eutrophication: nutrient sources, composition, and consequences. *Estuaries*. 2002;25:704-26.
- [140] Guillette LJ, Edwards TM. Is nitrate an ecologically relevant endocrine disruptor in vertebrates? *Integrative and Comparative Biology*. 2005;45:19-27.
- [141] Holland EA, Dentener FJ, Braswell BH, Sulzman JM. Contemporary and pre-industrial global reactive nitrogen budgets. *Biogeochemistry*. 1999;46:7-43.
- [142] Phoenix GK, Hicks WK, Cinderby S, Kuylenstierna JC, Stock WD, Dentener FJ, et al. Atmospheric nitrogen deposition in world biodiversity hotspots: the need for a greater global perspective in assessing N deposition impacts. *Global change biology*. 2006;12:470-6.
- [143] Rodolfi L, Chini Zittelli G, Bassi N, Padovani G, Biondi N, Bonini G, et al. Microalgae for oil: strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. *Biotechnology and bioengineering*. 2009;102:100-12.
- [144] Chen CY, Yeh KL, Aisyah R, Lee DJ, Chang JS. Cultivation, photobioreactor design and harvesting of microalgae for biodiesel production: a critical review. *Bioresource technology*. 2011;102:71-81.
- [145] Park J, Craggs R, Shilton A. Wastewater treatment high rate algal ponds for biofuel production. *Bioresource technology*. 2011;102:35-42.
- [146] Schlüter M, Groeneweg J. Mass production of freshwater rotifers on liquid wastes: I. The influence of some environmental factors on population growth of *Brachionus rubens* Ehrenberg 1838. *Aquaculture*. 1981;25:17-24.

- [147] Yang J, Xu M, Zhang X, Hu Q, Sommerfeld M, Chen Y. Life-cycle analysis on biodiesel production from microalgae: water footprint and nutrients balance. *Bioresource technology*. 2011;102:159-65.
- [148] Matson PA, Parton WJ, Power A, Swift M. Agricultural intensification and ecosystem properties. *Science*. 1997;277:504-9.
- [149] Mekonnen MM, Hoekstra AY. The green, blue and grey water footprint of crops and derived crop products. *Hydrology and Earth System Sciences*. 2011;15:1577-600.
- [150] Doran JW, Zeiss MR. Soil health and sustainability: managing the biotic component of soil quality. *Applied soil ecology*. 2000;15:3-11.
- [151] Pimentel D, Harvey C, Resosudarmo P, Sinclair K, Kurz D, McNair M, et al. Environmental and economic costs of soil erosion and conservation benefits. *Science-AAAS-Weekly Paper Edition*. 1995;267:1117-22.
- [152] Pimentel D, Kounang N. Ecology of soil erosion in ecosystems. *Ecosystems*. 1998;1:416-26.
- [153] Lal R. Soils and Sustainable Agriculture: A Review. In: Lichtfouse E, Navarrete M, Debaeke P, Véronique S, Alberola C, editors. *Sustainable Agriculture*. Dordrecht: Springer Netherlands; 2009. p. 15-23.
- [154] Anderson-Teixeira KJ, Davis SC, Masters MD, Delucia EH. Changes in soil organic carbon under biofuel crops. *Gcb Bioenergy*. 2009;1:75-96.
- [155] Kim S, Dale BE. Life cycle assessment of various cropping systems utilized for producing biofuels: Bioethanol and biodiesel. *Biomass and Bioenergy*. 2005;29:426-39.
- [156] Pimentel D. Soil Erosion: A Food and Environmental Threat. *Environment, Development and Sustainability*. 2006;8:119-37.
- [157] Wilcove DS, Rothstein D, Dubow J, Phillips A, Losos E. Quantifying Threats to Imperiled Species in the United States. Assessing the relative importance of habitat destruction, alien species, pollution, overexploitation, and disease. *BioScience*. 1998;48:607-15.
- [158] Crooks JA. Characterizing ecosystem-level consequences of biological invasions: the role of ecosystem engineers. *Oikos*. 2002;97:153-66.
- [159] Richardson DM, Rejmánek M. Trees and shrubs as invasive alien species - a global review. *Diversity and Distributions*. 2011;17:788-809.
- [160] Raghu S, Anderson RC, Daehler CC, Davis AS, Wiedenmann RN, Simberloff D, et al. Adding Biofuels to the Invasive Species Fire? *Science*. 2006;313:1742.
- [161] Chimera CG, Buddenhagen CE, Clifford PM. Biofuels: the risks and dangers of introducing invasive species. *Biofuels*. 2010;1:785-96.
- [162] Davis AS, Cousens RD, Hill J, Mack RN, Simberloff D, Raghu S. Screening bioenergy feedstock crops to mitigate invasion risk. *Frontiers in Ecology and the Environment*. 2010;8:533-9.

- [163] Buddenhagen CE, Chimera C, Clifford P. Assessing biofuel crop invasiveness: a case study. *PLoS one*. 2009;4:e5261.
- [164] Phalan B. The social and environmental impacts of biofuels in Asia: An overview. *Applied Energy*. 2009;86:S21-S9.
- [165] Ditomaso JM, Reaser JK, Dionigi CP, Doering OC, Chilton E, Schardt JD, et al. Biofuel vs bioinvasion: seeding policy priorities. *Environmental science & technology*. 2010;44:6906-10.
- [166] Zhang Y, Heath G, Carpenter A, Fisher N. Air pollutant emissions inventory of large-scale production of selected biofuels feedstocks in 2022. *Biofuels, Bioproducts and Biorefining*. 2016;10:56-69.
- [167] Ristaino JB, Thomas W. Agriculture, methyl bromide, and the ozone hole: can we fill the gaps? *Plant Disease*. 1997;81:964-77.
- [168] Ravishankara A, Daniel JS, Portmann RW. Nitrous oxide (N<sub>2</sub>O): the dominant ozone-depleting substance emitted in the 21st century. *Science*. 2009;326:123-5.
- [169] Heijungs R, Guinée JB, Huppes G, Lankreijer RM, Udo de Haes HA, Wegener Sleeswijk A, et al. Environmental life cycle assessment of products: guide and backgrounds (part 1). Leiden: Centre of Environmental Science; 1992.
- [170] Barker JR, Tingey DT. Air pollution effects on biodiversity. New York: Springer Science & Business Media; 1992.
- [171] Lovett GM, Tear TH, Evers DC, Findlay SE, Cosby BJ, Dunscomb JK, et al. Effects of air pollution on ecosystems and biological diversity in the eastern United States. *Annals of the New York Academy of Sciences*. 2009;1162:99-135.
- [172] Sala OE, Chapin FS, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, et al. Global biodiversity scenarios for the year 2100. *Science*. 2000;287:1770-4.
- [173] Wang M, Weber T, Darlington T. Well-to-wheels analysis of advanced fuel/vehicle systems—a North American study of energy use, greenhouse gas emissions, and criteria pollutant emissions. *Well-to-Wheels Analysis of Advanced Fuel/Vehicle Systems: A North American Study of Energy Use, Greenhouse Gas Emissions, and Criteria Pollutant Emissions*. 2005.
- [174] Forster P, Ramaswamy V, Artaxo P, Bernsten T, Betts R, Fahey DW, et al. Changes in atmospheric constituents and in radiative forcing. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, et al., editors. Cambridge, United Kingdom: Cambridge University Press; 2007. p. 129-234.
- [175] Li Z, Niu F, Fan J, Liu Y, Rosenfeld D, Ding Y. Long-term impacts of aerosols on the vertical development of clouds and precipitation. *Nature Geoscience*. 2011;4:888-94.
- [176] Rosenfeld D, Lohmann U, Raga GB, O'Dowd CD, Kulmala M, Fuzzi S, et al. Flood or drought: how do aerosols affect precipitation? *Science*. 2008;321:1309-13.

- [177] Forster P, Ramaswamy V, Artaxo P, Berntsen T, Betts R, Fahey DW, et al. Changes in atmospheric constituents and in radiative forcing. Chapter 2. *Climate Change 2007 The Physical Science Basis* 2007.
- [178] Wiloso EI, Heijungs R, de Snoo GR. LCA of second generation bioethanol: A review and some issues to be resolved for good LCA practice. *Renewable and Sustainable Energy Reviews*. 2012;16:5295-308.
- [179] Guinée JB, Heijungs R, Huppes G, Zamagni A, Masoni P, Buonamici R, et al. Life cycle assessment: past, present, and future†. *Environmental science & technology*. 2010;45:90-6.
- [180] Cherubini F, Strømman AH. Life cycle assessment of bioenergy systems: state of the art and future challenges. *Bioresource technology*. 2011;102:437-51.
- [181] Khoo HH, Sharratt PN, Das P, Balasubramanian RK, Naraharisetti PK, Shaik S. Life cycle energy and CO<sub>2</sub> analysis of microalgae-to-biodiesel: preliminary results and comparisons. *Bioresource technology*. 2011;102:5800-7.
- [182] Corley RHV, Tinker PB. *The Origin and Development of the Oil Palm Industry*. The Oil Palm: John Wiley & Sons, Ltd; 2015. p. 1-29.
- [183] Gordon D, Tancig K, Onderdonk D, Gantz C. Assessing the invasive potential of biofuel species proposed for Florida and the United States using the Australian Weed Risk Assessment. *Biomass and Bioenergy*. 2011;35:74-9.
- [184] Meyer J-Y. Preliminary review of the invasive plants in the Pacific islands (SPREP Member Countries). *Invasive species in the Pacific: A technical review and draft regional strategy*. 2000:85-115.
- [185] Chaparro-Giraldo A, López-Pazos SA. Evidence of gene flow between transgenic and non-transgenic maize in Colombia. *Agronomía Colombiana*. 2015;33:297.
- [186] Viljoen C, Chetty L. A case study of GM maize gene flow in South Africa. *Environmental Sciences Europe*. 2011;23:1.
- [187] Rakow G. Species Origin and Economic Importance of Brassica. In: Pua E-C, Douglas CJ, editors. *Brassica*. Berlin, Heidelberg: Springer Berlin Heidelberg; 2004. p. 3-11.
- [188] Australian Government. *The biology of Brassica napus L. (canola)*. Version 2.0 ed: Department of Health and Ageing Office of the Gene Technology Regulator; 2008. p. 59.
- [189] Rieger MA, Lamond M, Preston C, Powles SB, Roush RT. Pollen-mediated movement of herbicide resistance between commercial canola fields. *Science*. 2002;296:2386-8.
- [190] Knispel AL, McLachlan SM. Landscape-scale distribution and persistence of genetically modified oilseed rape (*Brassica napus*) in Manitoba, Canada. *Environmental Science and Pollution Research*. 2010;17:13-25.

- [191] Pessel D, Lecomte J, Emeriau V, Krouti M, Messean A, Gouyon P. Persistence of oilseed rape (*Brassica napus* L.) outside of cultivated fields. *Theoretical and Applied Genetics*. 2001;102:841-6.
- [192] Kawata M, Murakami K, Ishikawa T. Dispersal and persistence of genetically modified oilseed rape around Japanese harbors. *Environmental Science and Pollution Research*. 2009;16:120-6.
- [193] Moore PH, Paterson AH, Tew T. Sugarcane: the crop, the plant, and domestication. *Sugarcane: Physiology, Biochemistry, and Functional Biology*. 2013:1-17.
- [194] Yoshimura Y, Mizuguti A, Matsuo K. Analysis of the seed dispersal patterns of wild soybean as a reference for vegetation management around genetically modified soybean fields. *Weed Biology and Management*. 2011;11:210-6.
- [195] Mallory-Smith C, Zapiola M. Gene flow from glyphosate-resistant crops. *Pest management science*. 2008;64:428-40.
- [196] Kuroda Y, Kaga A, Tomooka N, Vaughan D. Population genetic structure of Japanese wild soybean (*Glycine soja*) based on microsatellite variation. *Molecular Ecology*. 2006;15:959-74.