1	Isotopic variation of dissolved and colloidal iron and copper in a carbonat													
2	floodplain soil after experimental flooding													
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15 Abstract

Many floodplain soils worldwide are contaminated by present and past industrial and mining activities. During flooding redox potential decreases, triggering the release of dissolved and colloidal metals. We used an anaerobic microcosm incubation to simulate flooding of a carbonate-rich floodplain soil for 40 days. Soil solution samples were extracted to determine the release of dissolved ($<0.02 \mu m$) and colloidal fractions (0.02 - 10 μm). We analyzed stable isotope ratios of colloidal and dissolved Fe and Cu representing two groups of metals with different release behavior; release of Fe was steadily increasing, while Cu peaked sharply after flooding and decreased afterwards. The temporal trend of δ^{56} Fe values of total Fe in

solution indicated dissimilatory iron reduction. The apparent isotopic fractionation between 24 dissolved and colloidal Fe (Δ^{56} Fe_{dissolved-colloidal} = δ^{56} Fe_{dissolved} - δ^{56} Fe_{colloidal}) varied between 25 $0.31\pm0.04\%$ and $-1.86\pm0.26\%$. Low δ^{56} Fe_{colloidal} (-1.16\pm0.04\%) values on day 4 of the 26 experiment suggested colloid formation by precipitation of dissolved Fe, while the strong 27 temporal variation in Δ^{56} Fe_{dissolved-colloidal} indicated subsequent changes in colloid mineralogy, 28 sorption to soil components and/or electron transfer-atom exchange. The variations in δ^{65} Cu 29 values (Δ^{65} Cu_{dissolved-colloidal} from 0.81±0.03‰ to 1.58±0.09‰) are probably linked to 30 changing oxidation state of colloidal Cu. While at the beginning of the experiment colloidal 31 32 Cu and solid soil Cu exchange, these systems decouple after the onset of sulfate reduction in the second half of the experiment. The experimental results fit well to findings from 33 redoximorphic soils described in the literature and highlight the importance of colloids for 34 metal release and the isotopic pattern in carbonatic soils. 35

36 Keywords: Copper, Iron, Soil, Redox Processes, Colloids, Stable Metal Isotopes

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38 Highlights

39	•	Colloidal Fe precipitates from dissolved Fe in soil solution and does not originate from
40		the mobilization of solid Fe minerals.

- The δ⁵⁶Fe value of colloidal Fe changes with time indicating changes in mineralogy or
 electron transfer-atom exchange.
- Colloidal and solid soil Cu exchange occurs until sulfate reduction starts, afterwards
 colloidal Cu decouples from solid soil and colloids aggregate.
- The Δ^{65} Cu_{dissolved-colloidal} values are probably driven by the redox state of colloidal Cu.
- Experimental findings agree well with field observations.



Present and past industrial and mining activities release contaminants into rivers, which 53 continuously accumulate in floodplain soils (Burton et al., 2008; Frohne et al., 2011; 54 55 Hindersmann and Mansfeldt, 2014). Depending on the frequency of flooding, metals might be remobilized and contaminate ground and surface waters (Du Laing et al., 2007; Hindersmann 56 et al., 2014; Mansfeldt and Overesch, 2013). The mobility of metals mainly depends on 57 organic matter concentrations and composition, soil mineral composition, redox conditions, 58 and carbonate concentrations (Du Laing et al., 2009). When soils are flooded, the redox 59 60 potential (E_h) decreases and Fe and Mn (oxyhydr)oxides are dissolved in many soils releasing associated trace elements (e.g., As, Ba, Co, Cr, V; Abgottspon et al., 2015; Hindersmann and 61 Mansfeldt, 2014; Weber et al., 2009b). In some soils with permanent water saturation, the 62 63 pool of reducible Fe and Mn is depleted and no further mobilization occurs under anoxic conditions. The release kinetics depends strongly on crystallinity of Fe and Mn minerals and 64 microbial community composition in the soil (Lovley, 1991; Schuth et al., 2015). In 65 66 carbonatic soils, the pH is buffered by the carbonic acid-hydrogen carbonate buffer system and proton release during formation of minerals like siderite and rhodochrosite (Abgottspon et 67 al., 2015; Ponnamperuma, 1972). Furthermore, changes to anoxic conditions may cause 68 microbial formation of reduced metal colloids (e.g., Cu⁰, Weber et al., 2009b). When E_h 69 decreases, microbial sulfate reduction may be initialized and mobility of metals can be limited 70 71 by the formation of or co-precipitation with sulfides (Borch et al., 2010; Weber et al., 2009a). However, sulfate reduction may also favor the release of metal-sulfide colloids of some 72 elements (e.g., Ag, Cd, Cu, Hg, Pb) resulting in enhanced metal mobility during several days 73

74 after flooding (Abgottspon et al., 2015; Hofacker et al., 2013; Weber et al., 2009b). The 75 formation of sulfides is limited by the available sulfate content and controlled by the 76 solubility of the corresponding metal sulfides (Weber et al., 2009a). Beside the formation of

organic, biomineralized and sulfide colloids, also colloids of Fe and Mn minerals may favor 77 78 trace element transport (Hassellov and von der Kammer, 2008). The colloids may form because of precipitation of siderite and rhodochrosite in carbonatic soils or the precipitation of 79 mixed valence (oxyhydr)oxides (e.g., magnetite or green rust; Ratering and Schnell, 2000; 80 Tadanier et al., 2005; Taylor, 1980). Another colloid formation process is the partial reduction 81 82 of ferrihydrite by microorganisms which may initialize deflocculation and release of the 83 remaining ferrihydrite as colloids along with associated non-Fe elements (Tadanier et al., 2005). When soil flooding ends and redox conditions change back from anoxic to oxic, 84 sulfides will be oxidized, releasing metals like Cd, Cu and Zn to pore waters, which may 85 86 (co)precipitate with (oxyhydr)oxides (Contin et al., 2007; Du Laing et al., 2007; Frohne et al., 2011). 87

Abgottspon et al. (2015) differentiated trace metals into two groups with different 88 89 release behavior in water-saturated soils. When E_h decreased (from 300 mV to 0 mV), Group I metals (Ba, Co, Cr, Ni, and V) were constantly released during 40 days, predominantly in 90 91 dissolved form (<50 % as colloids) closely correlated with the release of Fe and Mn by 92 reductive dissolution of (oxyhydr)oxides. The increase in colloidal concentrations of Group I metals is probably attributable to coprecipitation of dissolved metals with and sorption to 93 94 colloidal Fe and Mn minerals. Group II metals (Ag, Cd, Cu and Pb) were rapidly released after 2-6 days of flooding, strongly dominated by the colloidal fraction (>80%; 0.02-10 µm). 95 In this study, we focus on the stable isotope ratios of Fe and Cu as representatives of Group I 96 97 and Group II metals, respectively, to further explore the mechanisms underlying the observed differences in the release behavior of the two metal groups. 98

99 The stable isotope ratios of Fe and Cu bear a high potential to investigate long-term 100 biogeochemical processes in soils, especially redox processes (Babcsanyi et al., 2014; Bigalke 101 et al., 2013; Bigalke et al., 2010b; Mansfeldt et al., 2012; Wiederhold et al., 2007a). Iron is an 102 important driver of biogeochemical processes and trace metal behavior in soils. Changing the

oxidation state of Fe results in marked Fe isotope fractionation causing variations in Fe 103 isotopic composition among Fe pools in anoxic soils with δ^{56} Fe values ranging from -5% to 104 +1‰ (Johnson et al., 2008). Under anoxic conditions, dissimilatory iron reduction (DIR) by 105 bacteria initializes three main Fe reaction pools, i.e. $Fe^{2+}_{(aq)}$, $Fe^{2+}_{(sorb)}$ and $Fe^{3+}_{(reac)}$. Some of 106 the dissolved $Fe^{2+}_{(aq)}$ may sorb to the oxide surface as $Fe^{2+}_{(sorb)}$. Electron transfer between 107 $Fe^{2+}(sorb)$ and Fe^{3+} creates a reactive layer of Fe^{3+} at the oxide surface ($Fe^{3+}(reac)$) and is 108 associated with ⁵⁶Fe enrichment in the solid phase. Subsequently, DIR releases isotopically 109 light Fe into solution while heavy Fe isotopes are preferentially adsorbed on Fe oxide surfaces 110 $(\Delta^{56}$ Fe_{mineral-solution} up to 4‰; Frierdich et al., 2014; Icopini et al., 2004; Mikutta et al., 2009; 111 Teutsch et al., 2005). The apparent isotope fractionation between $Fe^{2+}_{(aq)}$ and $Fe^{3+}_{(reac)}$ is 112 identical regardless of bacteria species or type of oxides yielding Δ^{56} Fe_{Fe(II)-Fe(III)reac} values of 113 ~ -3‰ (Crosby et al., 2007). When the reduction continues, increased Fe^{2+} concentrations in 114 solution result in a shift to a heavier Fe isotopic composition of dissolved Fe because of 115 116 isotopic pool-size effects. This fractionation depends on the relative proportion of the three main reactive Fe pools (Crosby et al., 2007). Schuth et al. (2015) conducted incubation 117 experiments under controlled E_h conditions and found strong variations in bulk δ^{56} Fe values 118 in soil solutions of a surface soil horizon, while a permanently anoxic lower soil horizon 119 showed no Fe release and no response of δ^{56} Fe values to flooding, probably because of a more 120 stable crystalline Fe oxide mineralogy or an absence of Fe-reducing bacteria. However, at 121 constant E_h a steady state of δ^{56} Fe values and dissolved total Fe was observed by Schuth et al. 122 (2015) for the topsoil because of the balance of Fe release and removal from solution. Garnier 123 at al. (2017) and Thompson et al. (2007) found Rayleigh-type fractionation during DIR in 124 redox-influenced soils but smaller fractionation compared to the experimentally determined 125 fractionation factors, because of additional processes occurring in soil (e.g. organic 126 complexation). 127

Ilina et al. (2013a) studied iron isotope variations in river, lake and mire waters and 128 soil solutions of different filtration sizes reaching from 100 μ m to <1 kDa. The δ^{57} Fe values 129 of mire waters (0.12 to 0.66 %) and soil solutions (-0.20 to -0.42 %) showed no systematic 130 differences except for the <1 kDa fraction, which was the lightest (-0.83 to -0.91 ‰). River 131 waters became constantly heavier (up to 4.2 %) with decreasing pore size, which was 132 explained by stronger Fe binding of low molecular weight organic compounds compared to 133 bigger particles like Fe (oxyhydr)oxides (Ilina et al., 2013a). A similar observation was made 134 by Schuth and Mansfeldt (2016) for stagnant water, where smaller particles indicated a slight 135 136 enrichment in ⁵⁶Fe. Furthermore, other mechanisms can induce Fe isotope fractionation. Heavy Fe isotopes are preferentially adsorbed or precipitated on the bacterial cell surface 137 $(\Delta^{56}\text{Fe}_{cell-solution} \text{ up to } 1.7\%)$ and complexed by organic ligands $(\Delta^{56}\text{Fe}_{complex-solution} \text{ of } 0.6\%)$; 138 Brantley et al., 2001; Dideriksen et al., 2008; Morgan et al., 2010; Mulholland et al., 2015). 139 Precipitation of siderite caused a lighter Fe isotope ratio in the siderite (Δ^{56} Fe_{dissolved-siderite} = 140 0.0-0.9%; Johnson et al., 2005; Wiesli et al., 2004). While kinetic FeS precipitation caused an 141 enrichment of light Fe in the precipitate (Δ^{56} Fe_{Fe(II)ac-FeS} = 0.85±0.30‰; Butler et al., 2005), 142 143 precipitation of mackinawite at equilibrium conditions caused oppositional fractionation of Δ^{56} Fe_{Fe(II)ad-mackinawite} = -0.32±0.29‰ to -0.64±0.36‰ (Guilbaud et al., 2011; Wu et al., 2011). 144 The precipitation of magnetite similarly causes an enrichment of heavy isotopes in the 145 precipitates (Δ^{56} Fe_{Fe(II)aq-magnetite} = -1.34±0.11‰ to -1.56±0.20‰; Frierdich et al., 2014; 146 Johnson et al., 2005). 147

148Sorption of Cu to Al and Fe (oxyhydr)oxides causes an enrichment of heavy Cu on the

surface of these minerals with Δ^{65} Cu_{solid-solution} values of up to 1‰ (Balistrieri et al., 2008;

- 150 Pokrovsky et al., 2008). Sorption to organic ligands shows varying fractionation with
- 151 Δ^{65} Cu_{solid-solution} values between -0.3 and 0.8‰ depending on type of organic ligand and pH
- (Bigalke et al., 2010a; Navarette et al., 2011; Ryan et al., 2014). Lighter Cu isotopes are
- preferentially adsorbed on clay mineral surfaces with Δ^{65} Cu_{adsorbed-solution} values of -0.3‰ (Li

et al., 2015). Copper uptake into bacteria also causes various Cu isotope fractionations 154 depending on organism and pH. Mainly the lighter Cu isotope is preferentially incorporated 155 into the live bacteria cell (Δ^{65} Cu_{bacteria-solution} = 0 to -4.4 ‰; Navarette et al., 2011; Pokrovsky 156 et al., 2008; Zhu et al., 2002). In redox reactions, the reduced Cu species is enriched in lighter 157 Cu isotopes (e.g., Δ^{65} Cu_{solution-covellite} = 3.06‰; Ehrlich et al., 2004 and Δ^{65} Cu_{Cu2+-CuI} = 4‰; 158 Zhu et al., 2002). Bigalke et al. (2010b, 2011, 2013) found episodically water-saturated soil to 159 be enriched in heavy Cu isotopes, which they attributed to the loss of light Cu by leaching of 160 reduced colloidal Cu forms. Ilina et al. (2013b) found no significant variations (δ^{65} Cu values 161 of 0.36 to 0.46 ‰) between differently filtered fractions (100µm - <1kDa) of river water. 162 163 which they attributed to similar bonding strength of Cu fulvic complexes of different mass. Vance et al. (2008) reported lighter δ^{65} Cu values (-0.24 to -1.02‰) in the particulate fraction 164 of rivers compared to the dissolved phase (0.42 to 0.94‰) and explained this with strong 165 bonding of dissolved Cu to organic complexes. 166 167 In this study, we investigate the isotope geochemistry of dissolved and colloidal Fe and Cu in a water-saturated carbonatic soil. Our goals are to better understand how a) Fe (oxyhydr)oxide 168 dissolution and Fe colloid formation and b) Cu release and colloid formation in soils affect 169

results to field observations previously reported in the literature.

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2. Materials and methods

A Calcaric Fluvisol (IUSS, 2014), with high calcium carbonate content (420 g kg⁻¹) was sampled near the city of Aesch in Switzerland (47°28'39" N / 7°36'21" E). The soil had a neutral pH (7.4), an organic carbon concentration of 21.7 g kg⁻¹, a total Fe concentration of 12.6 g kg⁻¹ and a dithioniteextractable Fe concentration of 7.87 g kg⁻¹ (Abgottspon et al., 2015). More details about the soil are given in Table S1. The soil was located next to the river Birs on a floodplain with temporary flooding (Kayser et al., 2006). The sampling site was moderately contaminated with several trace metals (e.g.,

isotopic composition of soils and soil solutions. We furthermore will link our experimental

Cd, Cr, Cu, Pb, Table S1) by an adjacent non-ferrous metal smelter, causing elevated Cu 180 concentrations of 122 mg kg⁻¹ (Kusonwiriyawong et al., 2016). About 10 kg of topsoil were sampled 181 182 from the Ah horizon (0-0.15 m depth) in March 2011. The soil was air-dried and sieved to <2 mm. 183 The setup of the incubation experiment was similar to that of Weber et al. (2009b) and identical to that 184 described in Abgottspon et al. (2015). In fact, samples were taken from exactly the same experiment 185 described in Abgottspon et al. (2015). In summary, 1.7 L of artificial river water were filled in 3-L PE 186 microcosms. The artificial river water (prepared from ultrapure water and clean salts) had a similar ionic strength and ionic composition as the river Birs next to the sampling site (0.92 mmol L⁻¹ CaCl₂, 187 0.98 mmol L⁻¹ NaNO₃, 0.84 mmol L⁻¹ MgSO₄), but did not contain DOC, trace metals, or suspended 188 189 matter. Then, 1.7 kg of air-dry soil was slowly added to the microcosm and stirred to ensure homogenization and removal of air bubbles, resulting in complete water saturation. After 190 191 homogenization, the soil material was allowed to settle and around 2 cm of stagnant water covered the 192 soil (Figure S1). Three replicate microcosms and one blank (i.e. only artificial river water) were set up 193 in a glovebox (GS Glovebox Plexiglas, GS Glovebox Systemtechnik, Malsch, Germany) that was 194 purged with nitrogen (Figure S2). The soil solution was extracted with PE suction cups with a 10-µm 195 nominal pore size (ecoTech GmbH, Bonn, Germany) manually located in the lower half of the 196 microcosms with a syringe. On each sampling date, 50 mL of solution was extracted. Over the whole 197 experiment, we extracted 700 mL of a total solution volume of 1700 mL. Nevertheless, in the lower 198 part of the microcosm the soil was always water-saturated (and the suction cup at this depth always 199 flooded). Our extraction of solution might have simulated slow drainage as is also observed in natural 200 soils. We realized mass balance calculations to check how repeated sampling of the microcosms may affect Fe and Cu concentrations and δ^{56} Fe and δ^{65} Cu values. Over the whole experiment we extracted 201 202 0.06% and 0.18% of the total Fe and Cu in the system, respectively. Taking the isotopic composition 203 into account, the isotopic change of the total pool related to the sampling is insignificant (-0.001‰ and 204 0.002‰) for δ^{56} Fe and δ^{65} Cu, respectively. These effects are much smaller than our analytical 205 precision. Therefore, we conclude that the sampling will not significantly affect the isotopic 206 composition of the total metal pools. The Eh and pH values were analysed directly after sampling in 207 the glovebox. To rule out that the initial mobilization of colloids is an artifact caused by manually

208 sampling of the soil solution, Al concentrations were analyzed and found to be consistently low (on 209 average 0.5 µmol L⁻¹). As Al is a major constituent of the soil (2.6 wt. %), low Al concentrations 210 render the artificial release of soil colloids (e.g., clay minerals, Al hydroxides) unlikely. The solution is supposed to contain the total (colloidal + dissolved) element concentration (<10 µm). To determine 211 dissolved element concentrations, samples were manually filtered to <0.02 µm using syringe filters 212 (Anotop 25 Plus, Whatman, Bottingen, Switzerland) in the glove box directly after sampling. The Fe 213 214 and Cu concentrations were analyzed by ICP-MS (7700x, Agilent, Santa Clara, California) using In 215 and Rh as internal standards. The difference between total and dissolved elemental concentrations was 216 assumed to be the colloidal fraction $(0.02-10 \,\mu\text{m})$. All unfiltered and filtered solutions were acidified with suprapur HNO₃ (ROTH, Karlsruhe, Germany) and stored at -20°C prior to isotope analysis. The 217 218 Fe²⁺ concentration in the unfiltered solution was determined photometrically (Spectroquant Pharo 100, 219 MERCK). Each 5-mL aliquot of unfiltered soil solution was treated with 50 µL of H₂SO₄ (25%, p.a.), 0.5 mL of ammonium-acetate-acetic acid (400 g L⁻¹ ammonium acetate, 50% acetic acid) and 0.2 mL 220 of phenanthrolin solution (5 g L⁻¹ 1,10-phenanthroliumchlorid) and made up to 10 mL with Millipore 221 222 water in the glovebox. Samples were measured after 15 minutes at the wavelength of 510 nm. A detailed evolution of E_{h} , pH, SO₄²⁻, S²⁻, Fe²⁺ C and metal concentrations in 2-4 day resolution is given 223 224 in Tables S2 & S3 and Abgottspon et al. (2015).

For total digestion of the bulk soil sample, approximately 70 mg of soil was digested 225 in HNO₃, HF and H₂O₂ (3:2:1) in PFA beakers (Savillex, MN, USA) at least 24 h on a hot 226 plate at 120 °C. The digested samples were evaporated until dryness at 70 °C. Soil solution 227 samples were evaporated in PFA vials (Savillex, MN, USA). The dried residues were digested 228 using a mixture of concentrated HNO₃ and H₂O₂ (ratio 1:1) at 120°C for at least 3-4 hours and 229 evaporated to dryness. The dried soil and solution samples were subsequently re-fluxed in 230 HCl and HNO₃ (3:1) at least 3 h at 120 °C and finally dissolved in 7 mol L^{-1} HCl + 0.001% 231 H_2O_2 for ion exchange separation. The total (<10µm) and dissolved (<0.02µm) fractions were 232 analyzed in samples from Days 0, 2, 4, 6, 8, 11, 13, 16, 19, 22, 26, 30, 34 and 39. After 8 days 233 of incubation, the dissolved Cu concentrations (<0.02µm) were too low to get the required 234

300 ng of Cu for the isotope analysis. Thus, samples from the dissolved fraction of days 8, 10, 235 236 13, and 16 were combined to a single composite sample and another composite sample was prepared from days 19, 22, 26, 30, 34 and 39. In these composite samples of the dissolved 237 fraction, we also measured stable Fe isotope ratios after purification (see below). In addition, 238 for Fe isotope measurement samples from the three replicates microcosms were combined to a 239 composite sample of days 4 and 6, respectively, for both total (<10µm) and dissolved 240 $(<0.02\mu m)$ fractions because of too low mass (< 7µg Fe) of Fe for isotopic analysis at the 241 beginning of the experiment. Copper and Fe fractions were purified as described in Bigalke et 242 al. (2013). For solution samples, a single-step ion-exchange approach was sufficient to 243 244 separate Cu and Fe from other matrix elements, while bulk soil samples were purified twice for Cu isotope analysis. All Fe and Cu fractions yielded an average recovery of 100.6±1.3% 245 (mean±SD) and 101.1±2.1% (mean±SD), respectively. All samples which did not yield a 246 247 recovery of within $100 \pm 6\%$ were discarded, and the purification process was repeated. The acids (HNO₃ and HCl) used in this study were purified by sub-boiling distillation in quartz 248 249 stills or were purchased in suprapur quality. Other reagents were of suprapur quality. All 250 solutions and dilute acids were prepared using 18 MΩ grade water (EMD Millipore, MA, USA). All sample preparations were performed in the clean chemistry laboratory at the 251 Institute of Geology, University of Bern. The procedural blank was 3.1±1.8 ng (n=3) for Fe 252 253 and 2.3±0.2 ng (n=3) for Cu, compared to at least 7000 and 300 ng of Fe and Cu, respectively, in the samples. 254

Iron and Cu isotope measurements were performed on a Thermo-Scientific Neptune Plus MC-ICP-MS at the Leibniz University Hannover, Germany. The Fe isotopic compositions were analyzed in the high mass resolution mode to resolve isobaric interferences of 40 Ar 14 N ${}^{+}$, 40 Ar 16 O ${}^{+}$, and 40 Ar 16 OH ${}^{+}$ on 54 Fe, 56 Fe, and 57 Fe (Weyer and Schwieters, 2003). Chromium (53 Cr) was monitored to correct isobaric interferences of 54 Cr on 54 Fe. The standard-sample bracketing method was applied in combination with external element doping for mass bias correction. For the latter, the 5 mg L⁻¹ Ni standard NIST 986 (National Institute of Standards and Technology, Gaithersburg, MA, USA) was doped to all Fe standards (IRMM-014, Institute for Reference Materials and Measurements, Geel, Belgium) and samples for mass-bias correction (Oeser et al., 2014). The Fe isotope standard (IRMM-014) and the samples were diluted to yield a concentration of 7 mg L⁻¹ Fe with 2% HNO₃. Every sample was at least analyzed twice and the mean δ^{56} Fe value is given. The δ^{56} Fe values are reported relative to IRMM-014.

In a three-isotope plot, δ^{56} Fe and δ^{57} Fe values define a linear correlation 268 (slope=1.4664, R^2 = 0.99, Figure S3) following the theoretical mass-dependent fractionation 269 line (δ^{57} Fe ~1.5 x δ^{56} Fe) which indicates the absence of isobaric interferences (Malinovsky et 270 al., 2003). The accuracy and precision of the Fe isotope analysis was monitored using an Fe-271 free matrix sample (BCR-2, Basalt Columbia River 2, USGS, Reston, VA, USA) that was 272 273 doped with Fe of known isotopic composition, and an in-house standard (an Fe salt from ETH Zurich, Switzerland). The Fe-free matrix samples were prepared from the matrix fraction 274 275 derived from the purification of the original samples which were doped with the Fe standard (Merck Certipur[®], Darmstadt, Germany) and processed in the same way as the samples. The 276 average δ^{56} Fe value of the matrix fraction was 0.19±0.03‰ (mean ±2SD, n=5) which is 277 identical to the pure Merck Fe value (0.19±0.02‰, n=3). The USGS basalt BCR-2 yielded a 278 279 mean δ^{56} Fe value of 0.07±0.02‰ (2SD, n=4) which agrees well with previously published values ranging from 0.05±0.02‰ to 0.11±0.03‰ (Craddock and Dauphas, 2011; Liu et al., 280 2014a; Schuth et al., 2015; Weyer et al., 2005). The average δ^{56} Fe value of our in-house 281 standard was $-0.69\pm0.02\%$ (mean ± 2 SD, n=6) and again in good agreement with previously 282 published values varying from -0.71±0.18‰ to -0.73±0.10‰ (Fehr et al., 2008; Kiczka et al., 283 2011; Mansfeldt et al., 2012; Schuth et al., 2015). 284

For Cu isotope analysis, standard-sample bracketing was conducted in combination with external element doping for mass-bias correction. Again the Ni standard NIST 986 was

used for the mass-bias correction by analyzing ⁶²Ni/⁶⁰Ni (for details see Lazarov and Horn, 287 2015). The samples were diluted to yield a final concentration of 300 μ g Cu L⁻¹ and 1000 μ g 288 Ni L⁻¹ and were measured with MC-ICP-MS in low resolution mode. Every sample was at 289 least analyzed twice and the mean was reported. The δ^{65} Cu values are reported relative to the 290 Cu standard NIST SRM 976 after external mass-bias correction using the exponential law. 291 The success of the Cu purification and the accuracy and precision of the Cu isotope analysis 292 was verified by using Cu-free matrix samples that were subsequently doped with Cu of 293 294 known isotopic composition, and a basalt reference material (BCR-2). The Cu-free matrix samples were prepared from the matrix fraction derived from the purification of the original 295 samples and doped with the ERM®-AE633 Cu isotope standards (Institute for Reference 296 Materials and Measurements, Geel, Belgium), which is isotopically indistinguishable from 297 NIST 976 (Moeller et al., 2012). The doped matrices were treated and purified in the same 298 manner as the original samples. The average δ^{65} Cu value of the matrix samples was -299 $0.02\pm0.01\%$ (mean \pm 2SD, n=3) which is identical to the value of ERM[®]-AE633 (-300 0.01±0.05‰; Moeller et al., 2012). The BCR-2 reference material yielded a mean of 301 δ^{65} Cu_{NIST976} = 0.18±0.01‰ (2SD, n=3), which is in good agreement with previously 302 published data ranging from 0.14±0.05‰ to 0.22±0.06‰ (Moeller et al., 2012). The external 303 reproducibility was evaluated with the in-house Cu standard (NBS C 125-2) yielding an 304 average δ^{65} Cu value of 0.37±0.06‰ (mean ±2SD, n=10). 305

306 Most isotopic compositions from our experiments are reported as means of the 307 triplicate microcosm incubations (Table 1). Therefore, error bars in the figures do not reflect 308 measurement uncertainties but variations among the triplicate microcosms reflecting the 309 natural soil heterogeneity. The isotopic composition of the metal colloidal fraction ($\delta_{coll} =$ 310 δ^{56} Fe and δ^{65} Cu values) was calculated from the difference in the isotopic composition of 311 total and dissolved metal concentrations in solution based on isotopic mass balance, as show 312 in Equation 1. To calculate colloidal concentrations and δ values of the composite samples, we averaged the values of the colloidal concentrations and the δ values based on mass balance approaches, as shown in Equation 2 with δ_{ss} and C_{ss} being the delta value and the concentration of the respective subsample. In Equation 1-5 δ_{tot} , δ_{diss} and δ_{coll} are the δ^{56} Fe or δ^{65} Cu and c_{tot} , c_{diss} and c_{coll} the concentrations of the total, dissolved and colloidal fraction, while the prefix 2SD indicate the respective 2SD uncertainty.

318
$$\delta_{coll} = \frac{(\delta_{tot} \times [\sigma_{tot}]) - (\delta_{diss} \times [\sigma_{diss}])}{[\sigma_{tot}] - [\sigma_{diss}]} \quad (1)$$

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$$\delta_{bulk} = \frac{\sum_{ss=1}^{s} \delta_{ss} \times [c_{ss}]}{\sum_{ss=1}^{s} [c_{ss}]} \quad (2)$$

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320 The error of the δ value of the colloidal fraction calculated according to the general formula
321 of error propagation (Taylor, 1997) as shown in Equation 3.

$$2SD\delta_{coll} = \sqrt{\left(\frac{\partial\delta_{coll}}{\partial\delta_{tot}}2SD\delta_{tot}\right)^2 + \left(\frac{\partial\delta_{coll}}{\partialc_{tot}}2SDc_{tot}\right)^2 + \left(\frac{\partial\delta_{coll}}{\partial\delta_{diss}}2SD\delta_{diss}\right)^2 + \left(\frac{\partial\delta_{coll}}{\partial c_{diss}}2SDc_{diss}\right)^2}$$

$$323 \qquad (3)$$

324 If we apply Equation 3 to Equation 1, we get Equation 4, which allows calculating the

uncertainty of the δ values of the colloidal fraction as displayed in Table 1.

$$= \sqrt{\left(\frac{c_{tot}}{c_{tot} - c_{diss}}2SD\delta_{tot}\right)^2 + \left(-\frac{c_{diss}(\delta_{tot} - \delta_{diss})}{(c_{tot} - c_{diss})^2}2SDc_{tot}\right)^2 + \left(-\frac{c_{diss}}{c_{tot} - c_{diss}}2SD\delta_{diss}\right)^2 + \left(\frac{(\delta_{tot} - \delta_{diss})c_{tot}}{(c_{diss} - c_{tot})^2}2SDc_{diss}\right)^2}$$

$$326$$

$$327$$

$$(4)$$

Finally, the error of the
$$\Delta_{dissolved-colloidal}$$
 values was calculated according to equation 5

329
$$2SD\Delta_{diss-coll} = \sqrt{(2SD\delta_{diss})^2 + (2SD\delta_{coll})^2}$$
(5)

Element	Bulk soil /	Total				Dissolved					Colloidal				
	day after flooding	Conc. (µmol L ⁻¹)	SD	δ value (‰)	2SD	n ^a	Conc. (µmol L ⁻¹)	SD	δ value (‰)	2SD	nª	Conc. ^h (µmol L ⁻¹)	SD ^e	δ value ^f (‰)	2SD ^g
	Soil	226000 ^b	22920 ^b	0.04	0.01	3	-	-	-	-	-	-	-	-	-
	4	41.4	2.86	-1.09	0.01	1	9.89	3.95	-0.85	0.01	1	31.6	4.88	-1.16	0.04
	6	116	0.24	-0.77	0.01	1	10.99	2.70	-1.56	0.01	1	105	2.71	-0.68	0.03
Fe	8	181	7.36	-0.80	0.05	3	1100	19.8	-1.86 ^b	0.03	1	197°	29.4	0.03	0.26
	16	452	25.6	-0.64	0.04	3	119								
	39	673	9.17	-0.44	0.03	3	394 ^d	24.3	-0.98 ^c	0.01	1	203 ^d	25.9	0.32	0.14
	Soil	1920 ^b	70 ^b	-0.08	0.04	3	-	-	-	_	-	-	_	-	-
	0	5.09	0.12	0.12	0.11	3	4.60	0.29	0.10	0.11	3	0.49	0.31	0.36	1.55
	2	23.0	1.59	-1.01	0.04	3	5.04	4.15	-0.37	0.04	3	18.0	4.45	-1.19	0.20
_	4	20.4	2.30	-0.73	0.10	3	3.19	0.29	0.28	0.03	3	17.2	2.32	-0.92	0.12
Cu	6	21.5	1.60	-0.50	0.13	3	0.53	0.23	1.04	0.03	1	21.0	1.62	-0.54	0.13
	8	14.4	1.46	-0.30	0.07	3	0.070	0.03	0.68 ^c	0.04	1	10.1	1.37	-0.23	0.07
	16	5.80	1.28	-0.14	0.08	3	0.07								
	39	2.64	0.82	-0.41	0.07	3	0.07 ^d	0.02	0.38 ^d	0.03	1	2.59	0.82	-0.43	0.07

Table 1 Concentration and isotopic compositions of Fe and Cu in bulk soil prior to flooding, total ($<10\mu$ m), dissolved ($<0.02\mu$ m) and (calculated) colloidal (0.02-10 µm) fractions of soil solution during 40 days of experimental flooding.

^aNumber of replicate preparations and analysis, ^bConcentrations in the solid soil are in µmol kg⁻¹, ^cBulked sample combined from days 8 to 16,

^dBulked sample combined from days 19 to 39, ^eCalculated by error propagation of standard deviation in total and dissolved concentration,

^fCalculated by the difference between the δ^{65} Cu values of total and dissolved Cu (Eq. 1), ^gCalculated by error propagation according to Eq. S1-S4,

³³⁵ ^hCalculated by substracting the dissolved from the total concentrations.

336 3. Results

Iron was first mainly released in colloidal form, which changed after 19 days to 337 predominantly dissolved Fe (Figure 1 and Table S3, Abgottspon et al., 2015). On day 39, only 338 22% of the total Fe was still in colloidal form. The sum of total and dissolved Fe 339 concentrations increased continuously until the end of the incubation, while the colloidal 340 concentrations increased until day 16 and decreased afterwards (Figure 1). From day 2 on, the 341 total Fe in solution was dominated by Fe^{2+} , but the concentration of Fe^{2+} started to deviate 342 substantially from that of total Fe from day 13 on. After day 13, the Fe²⁺ concentrations 343 showed only a slight increase until day 22 and a decrease afterwards. From day 22 on, the 344 dissolved Fe concentration was even higher than that of total Fe²⁺ indicating that Fe³⁺ was 345 also present in the dissolved phase. On day 29, only 33% of the total Fe in solution was still 346 Fe²⁺. 347

The Fe isotope composition of total Fe in solution started at negative δ^{56} Fe values on day 348 4 and afterwards increased slightly (Figure 2b). The δ^{56} Fe value of total Fe in solution was 349 correlated with the total Fe concentrations in solution (Figure 3). The δ^{56} Fe value of the 350 colloidal Fe sharply increased on day 6 and then gradually continued to increase until the end 351 of incubation, and was parallel to the evolution of the δ^{56} Fe values of the total Fe in solution 352 (Figure 2b). The δ^{56} Fe values of the dissolved fraction were higher than the colloidal fraction 353 on day 4. Then, δ^{56} Fe values of the dissolved fraction continuously decreased from day 4 to 354 day 8-16. After 19-39 days, the δ^{56} Fe values of dissolved Fe shifted to heavier values (Figure 355 2b). The Δ^{56} Fe_{dissolved-colloidal} value decreased from 0.31±0.04‰ on day 4 to -1.86±0.26‰ on 356 day 12 and thereafter increased slightly (Figure 4a). The δ^{56} Fe value of the bulk soil was 357 0.04±0.01‰ (n=3). 358

Copper was also mainly released in colloidal form but in contrast to Fe, Cu release peaked around day 6 and decreased thereafter (Figure 2c, Abgottspon et al., 2015). The δ^{65} Cu values of the total, dissolved and colloidal Cu fractions did not differ from each other and

were close to that of the bulk soil at the start of the experiment (day 0; Figure 2d). With 362 strongly increasing Cu concentrations in solution on day 2, the δ^{65} Cu value of total Cu in 363 solution shifted to lower values and later on back towards initial values (Figure 2d). The 364 δ^{65} Cu value in both colloidal and dissolved Cu decreased markedly on day 2. After day 2, the 365 δ^{65} Cu value of the colloidal Cu increased until day 16; thereafter the δ^{65} Cu value decreased 366 slightly until the end of the experiment. The δ^{65} Cu value of the dissolved fraction increased as 367 well until day 6, but then, decreased towards the end of the experiment. The Δ^{65} Cu_{dissolved}-368 colloidal value changed from a maximum of 1.58±0.13‰ on day 6 back to 0.81±0.03‰ at the 369 end of the experiment (Figure 4b). The Δ^{65} Cu_{dissolved-colloidal} from day 0 of the experiment could 370 not be used because of the high uncertainty of the δ^{65} Cu_{coll} value at this day (Tab. 1). The 371 δ^{65} Cu value of the bulk soil was -0.08±0.04‰ (n=3). 372

373



374

Figure 1 Fe concentrations in total, dissolved and colloidal Fe during the experiment. Total, dissolved and colloidal Fe concentrations were taken from Abgottspon et al. (2015). The Fe²⁺ concentrations were specifically measured for this study in unfiltered samples and therefore represent total Fe²⁺ in solution including dissolved an colloidal fractions.

379



Figure 2 Temporal variation in a) Fe concentration, b) Fe isotope compositions, c) Cu concentration, and d) Cu isotope compositions during 40 days of flooding.
 Connection lines of the single data points are displayed to guide the eye. Plateau-like connection lines for Fe and Cu are displayed because of the pooling of the

samples from different days. So every step corresponds to one sample. In Panels b and d, the dashed horizontal lines show the δ^{56} Fe (0.04±0.01‰) and δ^{65} Cu (-

384 0.08±0.04‰) values of the bulk soil, respectively. Vertical bars in Panels a and c indicate SD of concentrations from triplicate microcosm experiments. Vertical

bars in Panels b and d indicate 2SD for isotope values. Error bars are smaller than the symbols unless visible. Please note the different scales.

398

4. Discussion

387 4.1. Fe isotopes

After the flooding of soils, oxygen is consumed and electrons are released by 388 microorganisms gaining energy from oxidation of organic matter. When the dissolved oxygen is 389 used up, the electrons are transferred to alternative electron acceptors (Lovley, 1991). Iron 390 391 (oxyhydr)oxides become the dominant alternative electron acceptors in flooded soils at sufficiently low E_h (< 150 mV). In our experiment, the E_h value decreased quickly, reached 90 392 mV four days after flooding and stabilized around 0 mV afterwards (Abgottspon et al., 2015). 393 394 This decrease potentially triggers DIR, which is reflected in the temporal course of the total Fe concentration and δ^{56} Fe value in solution in our experiment (Figure 3) and agrees with findings 395 of Crosby et al. (2007), Johnson et al. (2005), but also with abiotic reduction reactions 396 (Wiederhold et al., 2006). 397



Figure 3 δ^{56} Fe values of total Fe in solution as a function of Fe concentrations during 40 days of flooding. Bars indicate SD of Fe concentrations from triplicate microcosm experiments and 2SD of Fe isotope 400 401 compositions. Error bars are smaller than the symbols unless visible. The dotted line represents a logarithmic trend line.

402

From days 2-10, nearly all Fe in solution was Fe^{2+} , while the contribution of Fe^{2+} to total 403 Fe in solution decreased after day 10 (Fig. 1), which is similar to the increase in Fe^{3+} 404 concentrations in solution reported by Schuth et al. (2015). After day 19, total dissolved Fe 405 showed higher concentrations than Fe^{2+} , indicating the presence of Fe^{3+} even in the dissolved 406 phase. Because the solubility of Fe^{3+} is very low, this finding may only be explained by small 407 Fe^{3+} colloids, which passed through the 0.02 µm filter (and are therefore operationally defined as 408 dissolved species), or by the complexation of Fe^{3+} by organic substances, forming soluble 409 complexes. Another possible explanation might be an analytical complication in the 410 determination of the Fe²⁺ concentrations. If the Fe colloids at the end of the experiment did not 411 fully dissolve when the sulphuric acid was added, the Fe^{2+} concentration might have been 412 underestimated. 413

The pooling of the samples from different days, which was necessary because of the low 414 Fe concentrations at the beginning of the experiment and low dissolved Cu concentrations at the 415 416 end of the experiment, limit the temporal information of the Fe data. Therefore, we focus the 417 further discussion on the Fe colloid release under anoxic conditions in soils, a process for which no Fe isotope data have yet been reported. The colloidal release plays an important role in our 418 419 carbonatic floodplain soil. Up to 90% of the Fe was released in colloidal form on day 6 (Table S3; Abgottspon et al., 2015). Even in the later phase of the experiment (after day 10), colloidal 420 Fe still accounted for 20-50% of total Fe in solution (Figure 2a). The lack of colloidal organic 421 carbon and Al in solution (Abgottspon et al., 2015) indicate no co-mobilisation with OM or 422 colloidal soil constituents like clay minerals, but rather Fe to be the main component of the 423

colloids (Table S2, S3). At the beginning of the experiment, total Fe and Fe^{2+} concentrations in 424 solution were identical, highlighting that colloids consisted of reduced Fe and are not Fe 425 (oxyhydr)oxides released into soil solution (Figure 1). The colloidal Fe pool had low δ^{56} Fe 426 values at the beginning, hence supporting the idea of precipitation from reduced light Fe^{2+}_{aq} 427 428 rather than mobilization by deflocculation of ferrihydrite, which should have a more positive value (span of the amorphous Fe oxide fraction in hydromorphic and oxic soils approx. -0.7 to 429 430 0.4 ‰; Guelke et al., 2010; Schuth et al., 2015; Wiederhold et al., 2007a; Wiederhold et al., 431 2007b) compared to the colloidal fraction on day 4 of our experiment $(-1.16\pm0.04\%)$. Indeed, the initial Δ^{56} Fe_{dissolved-colloidal} value was positive and fitted the values reported for siderite 432 precipitation in laboratory experiments (Δ^{56} Fe_{dissolved-siderite} values between 0.0 and 0.9‰; 433 434 Johnson et al., 2005; Wiesli et al., 2004) and similar to that of lake sediments with supposed 435 siderite precipitation (Figure 4a, Teutsch et al., 2009). However, when looking at the proposed Δ^{56} Fe_{dissolved-colloidal} value, it has to be kept in mind that isotopic fractionation of Fe in solution can 436 also be influenced by interactions with the solid Fe pool in soil. Dissolved Fe²⁺ released by DIR 437 438 might be sorbed to Fe (oxyhydr)oxide surfaces in soil and undergo electron transfer-atom exchange reactions, a process which influences the δ^{56} Fe value in solution (Crosby et al., 2005; 439 2007) and might also affect the Δ^{56} Fe_{dissolved-colloidal} values. 440







443 Figure 4 Temporal variations in apparent isotopic fractionation between dissolved and colloidal a) Fe and

b) Cu. Horizontal error bars represent the time interval of the pooled samples. Vertical error bars

represent the 2SD calculated according to Equations 3-5.

446

447 Abgottspon et al. (2015), suggested siderite precipitation from day 8 on in the same 448 experiment. However, there is a time gap between the positive Δ^{56} Fe_{dissolved-colloidal} on day 4 and 449 the proposed siderite precipitation. Furthermore, a confirmation of siderite formation or the 450 precipitation of other minerals by XRD analysis was not possible because of the low mass of 451 colloidal material on the filters and interference of the filter material (an alumina-based 452 membrane).

Siderite can be formed by the reaction between Fe²⁺ produced by DIR and HCO₃⁻ produced by 453 454 microbial oxidation of organic matter (Mansfeldt et al., 2012; Wiesli et al., 2004). In our soil, high concentrations of inorganic carbon in solution (10 mmol L⁻¹ on day 4, Abgottspon et al., 455 2015) might cause a precipitation of FeCO₃ directly from dissolved Fe^{2+} and HCO_3^{-} in solution 456 457 at the beginning of the experiment. This is in agreement with the presence of colloidal inorganic 458 carbon between days 2-19 of the experiment (Abgottspon et al., 2015), a dominance of colloidal Fe and the fact that total Fe in solution was Fe²⁺ until day 10 of the experiment. Therefore, even 459 460 if we have no spectroscopic proof for siderite formation, we consider siderite to be the most 461 likely Fe mineral in colloidal Fe based on solution chemistry, isotope results and previous findings (Abgottspon et al., 2015; Du Laing et al., 2009; Mansfeldt et al., 2012; Wiesli et al., 462 2004). However, already on day 6, the Δ^{56} Fedissolved-colloidal changed, which may be explained by 463 the following scenarios: 464

1. Changing colloid mineralogy. Mineralogy of the colloids may change with time, depending on changing solution chemistry and microbial community (Fulda et al., 2013; Roh et al., 2003). As an example, siderite and magnetite are formed in similar environments depending on the composition of the surrounding air and dissolved gases, the presence or absence of the carbonic acid-hydrogencarbonate buffer, pH, incubation temperature, type of

substrate, incubation time, and bacteria species (Roh et al., 2003). Sulfate reduction might 470 induce formation of Fe sulfides (Borch et al., 2010). The change from Fe^{2+} to Fe^{3+} in the soil 471 solution and thus changing proportions of Fe^{2+} and Fe^{3+} in the dissolved and colloidal phases 472 might explain the variations in the Δ^{56} Fe_{dissolved-colloidal} values. Furthermore, the big gap 473 between total Fe and total Fe^{2+} indicate that colloids change from a Fe^{2+} dominated mineral to 474 a mineral containing also Fe³⁺. Observed Δ^{56} Fe_{dissolved-colloidal} values are consistent with those 475 expected for a number of Fe minerals like e.g., mackinawite or magnetite (Frierdich et al., 476 2014; Guilbaud et al., 2011; Johnson et al., 2005; Wu et al., 2011). 477

Sorption and electron transfer-atom exchange reactions. The shift to lower 478 2. Δ^{56} Fe_{dissolved-colloidal} values can also be caused by sorption and possibly electron transfer-atom 479 exchange reactions at the surface of the colloids or soil components (Crosby et al., 2005; 480 2007, Liu et al., 2015). The divalent dissolved Fe can be sorbed on these surfaces and electron 481 transfer-atom exchange reactions may occur, which change isotopic signatures of Fe in 482 solution (e.g., Frierdich et al., 2014). Sorption and electron transfer-atom exchange reactions 483 on colloid surfaces would increase δ^{56} Fe values in colloids and decrease δ^{56} Fe values in 484 485 solution, which is consistent with our results until day 16. However, also sorption to Fe oxy(hydr)oxide surfaces in the soil and electron transfer-atom exchange reactions, might 486 explain a significant part of the observed δ^{56} Fe variation in the solution (Reddy et al., 2015). 487 After 16 days of flooding, the shift of dissolved Fe to higher δ^{56} Fe values can be attributed to 488 an isotopic pool size effect caused by proceeding DIR and the growth of the dissolved relative 489 to the colloidal Fe fraction (Figure 2b). 490

491 However, the changing δ^{56} Fe values together with the increasing or constant colloid 492 concentration, indicated that new Fe colloids were formed throughout, while the concentrations 493 of Cu colloids steadily decreased probably because of aggregation and sedimentation.

494 In a recent experiment, the same soil was incubated under the same experimental conditions 495 and changes in the partitioning of Fe into five operatively defined fractions (F1-F5; NH₄NO₃extractable, NaOAc-extractable, NH₄Ox-extractable, hot H₂O₂/NH₄OAc-extractable and residual 496 497 fractions, respectively) were investigated (Abgottspon et al., 2015). At the beginning of the 498 experiment 81 % of the Fe in the soil was in the residual fraction which is supposed to consist 499 mainly of silicate-bound Fe and crystalline Fe (oxyhydr)oxides, the second important fraction 500 was the oxalate-extractable fraction (15 %, amorphous Fe oxides), while the other fractions had 501 minor importance (< 3 %). Over the time of the incubation, Fe concentrations in F1-F4 502 increased, indicating a substantial redistribution of Fe, which is only partly mirrored in the solution chemistry. 503

A variation in solid phase δ^{56} Fe values of up to 0.93‰ was reported for soils which are 504 505 seasonally or permanently water-saturated (Fekiacova et al., 2013; Liu et al., 2014b; Mansfeldt et al., 2012; Schuth et al., 2015; Wiederhold et al., 2007a). This can be attributed to the 506 mobilization of reduced isotopically light Fe, which might be leached, leaving the soil enriched 507 508 in heavier Fe isotopes (Schuth and Mansfeldt, 2016). The fractionation follows an apparent 509 Rayleigh-type behavior with smaller fractionation factors in soils compared to experimentally 510 found values, because of additional processes (e.g., organic complexation and reprecipitation) occurring in the complex soil system (Garnier et al. 2017, Thompson et al., 2007). In contrast, 511 most soils that developed under oxic conditions only display a small variation in bulk solid phase 512 δ^{56} Fe values of 0.08‰ (Podzols show variations in δ^{56} Fe values of up to 0.6‰; Poitrasson et al., 513

514 2008; Wiederhold et al., 2007b). In agreement with these findings, we observed a release of light 515 Fe into the soil solution ranging from -1.1‰ to -0.4‰ (Figure 2b). The evolution of the δ^{56} Fe 516 values in total Fe during our experiment showed a time dependency with low δ^{56} Fe values 517 shortly after flooding which successively approached bulk soil values with increasing time of 518 flooding.

These findings agree with the interpretation of the variation in δ^{56} Fe values of total Fe in the 519 water of the Rio Negro River, where a substantial temporal variation was observed (dos Santos 520 Pinheiro et al., 2014). In Rio Negro River water, the lowest δ^{56} Fe values were coupled to strong 521 rain events, which cause a reductive mobilization of Fe from the soils in the river catchment 522 (Bergquist and Boyle, 2006). Hence, colloidal Fe release in certain anoxic soils significantly 523 524 affects the temporal isotopic variability of the colloidal fraction. For a detailed understanding of 525 the processes causing the isotopic changes in the colloid fraction, further investigation of the 526 associated processes and mineralogy will be needed which is beyond the scope of this study.

Overall, the agreement of the results from our microcosm experiment and field observations 527 528 confirms that such experiments can be used for future research into Fe isotopic behavior in 529 anoxic soils. A big advantage of the microcosms is that soil solution samples can be extracted under controlled oxygen-free conditions and allow e.g., for the separation of colloids from 530 dissolved Fe in anoxic samples or photometric Fe²⁺ analysis. This separation is hardly possible 531 under oxic field conditions as reduced Fe^{2+} will almost immediately precipitate as Fe 532 (oxyhydr)oxides when coming in contact with the atmosphere. In spite of the limitations of our 533 study because we had to pool bulk samples of several dates and the lack of direct 534 characterization of the mineral phases, our results indicate that the Fe release in colloidal form 535

536 contributes substantially to total Fe concentrations and δ^{56} Fe values in solution and thus to Fe 537 mobility in our study soil.

538

539 **4.2. Cu isotopes**

540 The initial apparent isotopic fractionation between Cu in solution (total, dissolved and colloidal) and in bulk soil was small (Figure 2d). With increasing time after flooding, the 541 542 fractionation between bulk soil and solution and between dissolved and colloidal Cu species in 543 solution increased, with total and colloidal fraction becoming lighter and the dissolved fraction 544 becoming heavier. Relating the Cu isotope ratios to a similar experiment in which Cu speciation was investigated by XAS on a polluted soil from the River Mulde in Germany, offers a possible 545 546 explanation for most of the observed isotope fractionations. The Cu concentration peak on day 2 and the strong shift to negative δ^{65} Cu values of total Cu in solution could be caused by Cu-547 carrying bacteria cells which were dominating the colloidal phase in studies with other soils 548 549 (Hofacker et al., 2013; Weber et al., 2009a). The simultaneously observed heavy Cu isotope ratio 550 of the remaining Cu in solution would then be the consequence of previous preferential uptake of ⁶³Cu by the bacteria (Navarette et al., 2011; Pokrovsky et al., 2008), Cu reduction in the cell and 551 the change to trigonal coordination (Weber et al., 2009a). The shift in δ^{65} Cu values between 552 553 dissolved and colloidal Cu was similar to previously reported Cu isotope fractionation that was linked to the uptake of Cu into bacteria with Δ^{65} Cu_(solution-bacteria) values between +1.0‰ and 554 +4.4‰ (Navarette et al., 2011). The change of δ^{65} Cu values in solution between days 2 and 6, 555 while total Cu concentrations remained stable, can only be explained by the exchange of solution 556 Cu with solid soil Cu pools. The shift to higher δ^{65} Cu values from -1.01±0.04‰ to -0.50±0.13‰ 557 in total Cu in solution requires the removal of isotopically light Cu from solution and/or release 558

559 of heavy Cu into solution. In the experiments of Hofacker et al. (2013) and Weber et al. (2009a), bacteria released Cu⁺, which disproportioned into Cu²⁺, released into solution, and zero-valent 560 Cu (Cu⁰) colloids, in this phase of the incubation. In solution, Cu²⁺ released from bacteria was 561 partly reduced and precipitated together with the already present Cu in soil solution and 562 exchangeable Cu from the solid soil as Cu_xS (Weber et al., 2009b). If this process also occurred 563 in our experiment the Cu_xS would be a mixture of soil Cu and solution Cu and would probably 564 carry a heavier isotopic signal than that of the Cu in the bacteria. After day 6 with the onset of 565 sulfate reduction (Table S2, Abgottspon et al., 2015), Cu isotope composition and apparent 566 fractionation between dissolved and colloidal Cu stabilized (average values of δ^{65} Cu = -567 $0.29\pm0.15\%$ and Δ^{65} Cu_{dissolved-colloidal} = $0.87\pm0.09\%$) despite decreasing Cu concentrations in 568 solution. These findings indicate that the further decrease of colloidal Cu was caused by a 569 570 progressive aggregation and sedimentation of colloids, which does not cause a pronounced isotopic fractionation (Figure 4b). Even if the results from the soil investigated by Hofacker et al. 571 (2013) and Weber et al. (2009a) cannot be directly transferred to our soil because of the acid soil 572 used by these authors and our slightly alkaline soil, Abgottspon et al. (2015) showed that in our 573 soil decreasing metal mobilization paralleled increasing sulfate reduction. However, the fact that 574 575 we did not observe colloidal organic carbon at the beginning of the experiment (Table S2) disagrees with the findings of bacterial release causing the increase of colloidal Cu at the 576 beginning of the experiment (Hofacker et al., 2013; Weber et al. 2009a). Assuming a C/Cu (w/w) 577 ratio of about 4000 as described for methanogenic bacteria (Scherer et al., 1983), a bacterial 578 release causing 23 µmol Cu L⁻¹ on day 2 should be clearly visible also in colloidal C (approx. 579 500 mmol L⁻¹). Even if the strong variation of Cu concentrations in bacteria is taken into 580

account, the colloidal C concentration should be clearly higher than the measured 0.2 mmol L⁻¹
C_{coll} (Table S2), if bacteria caused the colloidal Cu concentration on day 2.

One might hypothesize that the reported changes of Cu redox states (Weber et al., 2009a) 583 explain much of the observed Δ^{65} Cu_{dissolved-colloidal} values in soil solution, which are 0.82±0.20‰ 584 on day 2, when Cu is supposed to occur as Cu⁺ and reach a maximum value of 1.58±0.13‰ on 585 day 6 as a result of the presumed dominance of Cu⁰. After sulfate reduction on days 6-8 (Table 586 S1), Δ^{65} Cu_{dissolved-colloidal} values changed back to 0.81±0.03‰, in line with the assumption that 587 588 Cu⁺ was the dominant Cu species (Weber et al., 2009a; Figure 4b). However, it is not clear, if the same redox transformations as described by Weber et al. (2009a) occur in the soil used in our 589 experiment. Generally, the isotopic fractionation associated with abiotic oxidative leaching of 590 reduced Cu minerals (chalcocite, chalcopyrite, enargite) is in the range of Δ^{65} Cu_{aq-mineral} = 0 to 591 2.7‰ in laboratory experiments and field surveys (Fernandez and Borrok, 2009; Kimball et al., 592 2008; Mathur et al., 2005) and agree well with theoretical estimations stating that Cu(I) 593 complexes preferentially enrich ⁶³Cu and their precipitation results in the enrichment of ⁶⁵Cu in 594 595 the fluid (Fuji et al., 2013). This pattern supports the interpretation that the observed Δ^{65} Cu_{dissolved-colloidal} values in our experiment are caused by precipitation of reduced colloidal Cu 596 species. However, fractionation has been reported to be irrelevant or even in the opposite 597 direction in the presence of bacteria (Kimball et al., 2008, Mathur et al., 2005). The latter has 598 599 been explained with the accumulation of heavy isotopes on the bacteria and preferential precipitation of heavy Cu on bacteria surfaces. The biotic induced isotope fractionation is 600 counteracting the isotope fractionation introduced by abitic redox processes. Thus in a system 601 with reducing conditions and bacteria the δ^{65} Cu values in solution are determined by the balance 602 of Cu released abiotically and Cu that interacts with the cells and biotic precipitates (Mathur et 603

al., 2005, Kimball et al., 2008). If we transfer the latter findings to our Δ^{65} Cu_{dissolved-colloidal} values, this might indicate that abiotic reduction rather that bacterial Cu precipitation is governing colloid formation. However, the influence of microorganisms as well as other soil process (e.g. organic complexation or reprecipitation as described by Thompson et al., 2007 for Fe), might decrease the apparent fractionation in our soil, compared to experimentally determined fractionation factors in less complex systems (e.g., Fernandez and Borrok, 2009; Zhu et al., 2002).

611 In a recent experiment (Kusonwiriyawong et al., 2016), we determined the changes in Cu partitioning and the δ^{65} Cu values in five fractions of a sequential extraction (F1–F5; NH₄NO₃-612 extractable, NaOAc-extractable, NH₄Ox-extractable, hot H₂O₂/NH₄OAc-extractable and residual 613 fractions, respectively) in the same soils that we used in this study. We found that in the dry soil, 614 δ^{65} Cu values in F1–F4 followed the estimated bonding strengths of Cu in the respective 615 fractions, indicating equilibrium distribution of Cu at the beginning of the experiment. After 616 flooding, Cu concentrations decreased in F1-F3 and increased in F4-F5. Overall, 73% of the 617 total Cu was redistributed among the five studied fractions. The strongest variations in δ^{65} Cu 618 values occurred in F3 (0.09±0.07‰ to 1.43±0.13‰) and F4 (-0.24±0.07‰ to 0.55±0.07‰), 619 while flooding had no or small effects on the δ^{65} Cu values of F1, F2 and F5. So even if F1 620 621 should be the Cu pool, which interacts most strongly with the solution, it did not mirror the shifts of the dissolved fractions in this experiment. The results from Kusonwiriyawong et al. (2016) 622 suggest a direct transfer of Cu from F3 to F4 because both concentration changes and changes in 623 δ^{65} Cu values were balanced between the two fractions. The responses of Cu partitioning and 624 δ^{65} Cu values to flooding are in line with the formation of CuxS or other reduced Cu species and 625 626 the reduction of Cu associated with Fe (oxyhydr)oxides.

627 One factor which can only be traced with the used isotopic approach is the exchange of Cu in solution with solid soil Cu, which is most pronounced between days 2-6. On day 8, the 628 total δ^{65} Cu value of the soil solution showed good agreement with that of Fraction 4 of the 629 sequential extraction of the solid soil by Kusonwiriyawong et al. (2016) on day 7, which is 630 supposed to represent reduced Cu phases. The agreement in these two δ^{65} Cu values supports the 631 assumption, that at this time colloidal Cu in solution and solid soil Cu are coupled. However, the 632 Cu isotope signatures in Fraction 4 in the solid soil changed to positive values $(0.55\pm0.07\%)$ on 633 day 35, while dissolved Cu still showed negative δ^{65} Cu values and did not change significantly 634 between days 8 and 39. This suggests that after the initial peak Cu colloid concentration, colloid 635 formation and exchange with the soil stopped, while aggregation in solution decreased the Cu 636 colloid concentration. 637

Babcsanyi et al. (2014) and Bigalke et al. (2010b, 2011, 2013) investigated different oxic 638 and anoxic soils and wetland systems and found that Cu isotope signatures were isotopically 639 heavier in temporally flooded soils than in soils, which developed in an oxic environment. They 640 concluded that isotopically light Cu was released from flooded wetlands. Furthermore, these 641 642 authors suggested that leaching of reduced Cu colloids was responsible for the observed apparent 643 fractionation. This assumption is confirmed by our findings, which demonstrate a strong release of light colloidal Cu shortly after flooding. Leaching of this Cu from the soil would cause the 644 described shift to heavier δ^{65} Cu values. The extent of the fractionation in real soils would depend 645 on the progress of leaching. In our study soil, the lowest δ^{65} Cu value for total Cu in solution 646 occurred on day 2. Assuming a similar δ^{65} Cu value (-1.0‰) of the total Cu released into solution 647 648 and leached over several redox cycles and a leaching of e.g., 10% of the initial Cu in bulk soil, the total Cu in bulk soil should be isotopically shifted from initially 0 to +0.1%. 649

This mobilization of Cu has to be considered in the management of polluted sites, by preventing changes of redox conditions. Especially, floodplain soils, which show high metal concentrations in many cases, might directly leach the Cu to the associated river, and thus negatively affect the river ecosystem. Because the colloids are carrying up to 100% of Cu and associated trace metals in soil solution and will strongly affect transport behavior and toxicity (Griffitt et al., 2007; Ju-Nam and Lead, 2008), the extent and stability of colloids formed in carbonatic soils need to be considered in risk assessments.

657

658 **5.** Conclusions

Total δ^{56} Fe values in solution followed the pattern of DIR. The Δ^{56} Fe_{dissolved-colloidal} values 659 660 indicated that dissolved Fe precipitated directly to form colloids, which accounted for 90% of total Fe at the beginning to 20% of total Fe at the end of the experiment. While 661 siderite precipitation might be one reason for colloidal Fe formation, a change of 662 Δ^{56} Fe_{dissolved-colloidal} values with time indicates either changes in the colloid mineralogy, or 663 sorption and possible electron transfer-atom exchange reactions with colloidal Fe or Fe 664 minerals in the soil solid phase. The results match previous findings from lake, river and 665 soil environments and underline the significance of colloid formation from dissolved Fe 666 in some anoxic soil environments. The strong changes in Δ^{56} Fe_{dissolved-colloidal} values raise 667 668 new questions about formation and changes of the Fe colloid composition under anoxic conditions. 669

The Δ⁶⁵Cu_{dissolved-colloidal} seemed to be mainly driven by Cu redox state. The δ⁶⁵Cu values
 indicated a strong exchange of colloidal Cu with solid soil Cu and a decoupling of these
 two pools with beginning sulfate reduction and decreasing Cu concentrations in solution.

The comparison of the experimental results with previous findings from hydromorphic soils and wetlands indicates that Cu colloid formation might be an important factor driving Cu leaching in these environments. Therefore, Cu colloid formation should be accounted for in risk assessment of e.g., polluted floodplain soils and similar environments.

In this study, the first results of Fe and Cu isotopic composition of colloids forming in anoxic soil environments are presented. Both elements are redox sensitive and probably undergo reduction, but show distinctly different temporal concentration and isotopic patterns. Both elements have a considerable colloidal component in common. Colloidal Fe and Cu might cause colloidal co-mobilization of a line of other potential toxic elements in soils. Furthermore, colloidal properties need to be considered in toxicity and mobility assessments.

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693 Supporting Material

Supporting material for this manuscript is available, including soil characterization, solution
chemistry data and figures illustrating the experimental setup and a three isotope plot for the Fe
isotope analysis.

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