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SYNTHESIS, KINETIC AND CATALYTIC STUDIES OF MANGANESE COMPLEXES WITH CORROLE AND PORPHYRIN LIGANDS

A Thesis Presented to The Faculty of the Department of Chemistry Western Kentucky University Bowling Green, Kentucky

In Partial Fulfillment Of the Requirements for the Degree Master of Science

> By Haleh Jeddi

May 2017

SYNTHESIS, KINETIC AND CATALYTIC STUDIES OF MANGANESE COMPLEXES WITH CORROLE AND PORPHYRIN LIGANDS

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SYNTHESIS, KINETIC AND CATALYTIC STUDIES OF MANGANESE COMPLEXES WITH CORROLE AND PORPHYRIN LIGANDS

Haleh Jeddi	May 2017	68 Pages
Directed by: Dr. Rui Zhang	g, Dr. Kevin Williams, and Dr. Mat	thew Nee

Department of Chemistry

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High-valent transition metal-oxo intermediates play a significant role in the catalytic cycle of the ubiquitous cytochrome P450 enzymes and in biomimetic catalytic systems. In this work, manganese(III) porphyrin and corrole systems (2) were synthesized and characterized by UV-vis absorbance and ¹H-NMR, matching literature-reported spectroscopic data. Manganese(V)-oxo corroles (3) and a manganese(IV)-oxo porphyrin (4) were successfully generated by chemical oxidation using *m*-chloroperoxybenzoic acid (*m*-CPBA), and their oxidation reactions with organic reductants were comparatively investigated. Results from single-turnover kinetic studies indicate that in the tris(pentafluorophenyl)corrole system (3a), the active oxidizing intermediate differs in different solvents. The active oxidizing intermediate in acetonitrile is likely the manganese(V)-oxo species 3a. However, in dichloromethane, the active oxidant is suspected to be a putative manganese(VI)-oxo species generated by disproportionation of the manganese(V)-oxo species.

Tris(pentafluorophenyl)corrolato manganese(III) (2a) was shown to selectively catalyze sulfoxidation and epoxidation with iodobenzene diacetate [PhI(OAc)₂] as a mild oxygen source. 2a exhibited higher conversions than triphenylcorrolato manganese(III) (2b), most likely because of the higher stability of 2a compared to 2b. In

contrast, tetramesitylporphyrinato manganese(III) (2c) was more efficient in catalytic oxidations than 2a, resulting in much higher conversions, but much less selectivity. Other reported metalloporphyrin and metallocorrole systems show an accelerating effect upon addition of small amounts of water; however, neither corrole systems exhibited a positive water effect. This is attributed to the strong coordination between the manganese center and water, preventing the oxygen source from coordination.

1. INTRODUCTION

1.1 Cytochrome P450 Enzymes

The oxidation of organic substrates is monumentally important in the production of high-value chemicals.¹ The ability to perform selective oxidations in a synthetic lab also aids in the understanding of fundamental oxidative biological transformations.² However, most industrial-scale oxidations are nonselective, performed with strong, stoichiometric, environmentally hazardous oxidants, often under high heat and producing significant amounts of waste. As a result, the development of catalytic, environmentally friendly and selective oxidation methods has become one of the most important goals in oxidation chemistry.^{1,3-4}

In an effort to develop such catalysts, chemists look to natural processes. Cytochrome P450 enzymes (CYP450s) are particularly relevant. CYP450s are named for their characteristic Soret band at 450 nm in the reduced state when complexed with CO.⁵ These ubiquitous oxidative enzymes are found in most biological systems and some viruses.⁶ In the human body, these enzymes perform biosynthesis of hormones and drug metabolism.⁷ In general, CYP450s use molecular oxygen for their oxidations, transferring one oxygen to the substrate and reducing the second to water with biological reducing agents such as nicotinamide adenine dinucleotide (NADH).⁸ The oxidations performed by CYP450s are characteristically chemo-, regio- and stereoselective, so the ability to replicate the reactivity of CYP450s is a valuable synthetic tool.⁹⁻¹⁰

The classical example of a stereospecific, chemospecific and regiospecific enzyme-catalyzed oxidation is that of camphor by $CYP450_{cam}$, whose crystal structure is shown in Figure 1.¹¹ CYP450_{cam} is the most well-characterized CYP450 enzyme, named

after its substrate, camphor. CYP450_{cam} is found in *Pseudomonas putida*, a species of bacterium known for its ability to metabolize unusual substrates including camphor, caffeine and benzene. The ability of *P. putida* to metabolize these compounds is typically attributed to its powerful CYP450 monooxygenases.¹² In the case of camphor, the substrate is specifically hydroxylated to 5-*exo*-hydroxycamphor,¹¹ which is then metabolized to isobutyrate and acetate.¹³ Benzene and toluene are oxidized to benzoate and toluate, which are further metabolized into Krebs cycle intermediates as well.¹² This reaction is chemospecific as only hydroxylation takes place, regiospecific as only C5 is oxidized and stereospecific as only the *exo* product is formed. This oxidation is outlined in Scheme 1.

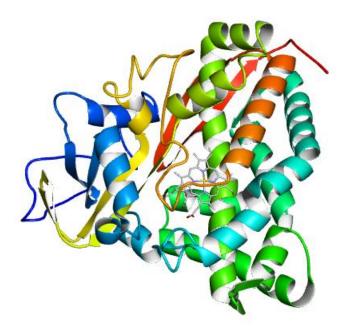
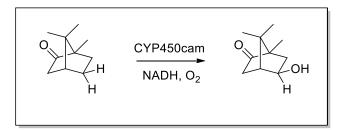


Figure 1. The crystal structure of CYP450_{cam}.¹¹



Scheme 1. The specific hydroxylation of camphor on C5, producing only the *exo* product.

CYP450s contain a heme *b* prosthetic group that acts as the active site, shown in Figure 2. A proximal cysteine ligand binds the octahedral iron(III) center, while a water molecule occupies the sixth position.¹³ The water molecule is displaced by the substrate, followed by the binding of molecular oxygen and subsequent reduction of one oxygen to water. This results in an iron(IV)-oxo radical cation, termed Compound I.¹⁴ Scheme 2 shows the catalytic cycle of CYP450s. Considering the catalytic abilities of these enzymes and the relative simplicity of the active site, the use of similar structures as biomimetic catalysts has garnered significant attention.^{15–16} The active oxidant in the catalytic cycle of CYP450s is generally suggested to be Compound I.^{14, 17} As such, highvalent metal-oxo species are particularly important intermediates in the catalytic cycles of biomimetic catalysts for a variety of oxygen atom transfer (OAT) reactions in past decades.^{18–19} Scheme 3 illustrates typical oxidation reactions catalyzed by metalloporphyrins.^{15-16, 20-21}

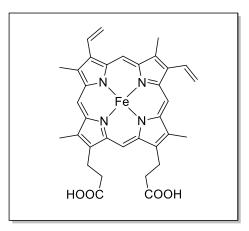
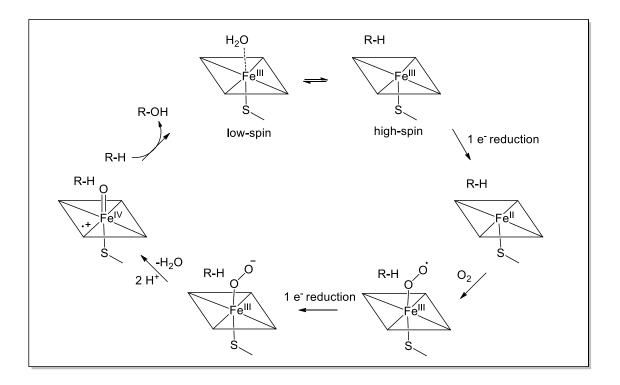
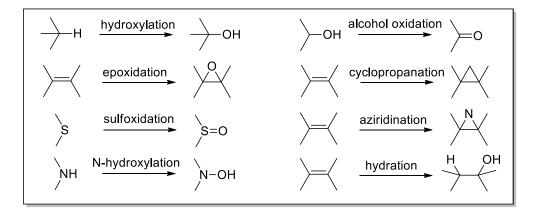


Figure 2. The structure of heme *b*, also called protoporphyrin IX.



Scheme 2. The catalytic cycle of CYP450 enzyme-catalyzed hydroxylation.



Scheme 3. Typical oxidations catalyzed by metalloporphyrins.

1.2 High-Valent Transition Metal-Oxo Species in Catalytic Oxidations

Because of their role in the catalytic cycle of CYP450s, high-valent transition metal-oxo species are of significant interest in the study of catalytic oxidations. High-valent metal-oxo species are typically considered the active oxidizing species in metalloporphyrin-mediated catalysis.²² The first reported high-valent oxo-metalloporphyrin was an iron(IV) radical cation analogous to Compound I, later implicated in the catalytic cycle of CYP450s.^{13, 17, 23} The detection of this complex was integral to the understanding of the catalytic cycle of CYP450s.¹⁴ Since the detection of this first Compound I analogue in synthetic catalysts, the iron(IV) radical cation has been implicated as the major oxidizing intermediate in a variety of iron porphyrin-catalyzed oxidations.^{13, 16}

The first high-valent manganese-oxo species was reported in 1980 by Groves et al.²⁴ Since the first report of a manganese(V)-oxo porphyrin, the development of metalloporphyrin catalysts with metals other than iron has increased. The study of high-valent manganese-oxo species is particularly interesting because iron(V)-oxo species have not been isolated in porphyrin systems, despite their proposed role in the catalytic

cycle of the enzymes.¹³ A manganese(IV)-oxo porphyrin was also reported as an active oxidizing intermediate in the catalytic cycle of an electron-deficient porphyrin for the first time in 1987.²⁵ Since then, manganese(IV) and manganese(V)-oxo porphyrins have been implicated as the active oxidants in the catalytic cycles of a variety of porphyrin catalysts.^{18, 26} Manganese(IV)-oxo radical cations have not been reported as active oxidants in the catalytic cycle of manganese porphyrins or corroles, although recently a manganese(IV)-hydroxy radical cation was characterized as a tautomer of a manganese(IV)-oxo corrole formed by protonation of the oxo ligand.²⁷ Similar manganese(IV) radical cations have been characterized by addition of Lewis acids to the oxo ligand.²⁸⁻³⁰

1.3 Metalloporphyrins as Biomimetic Catalysts

The use of metalloporphyrins as oxidation catalysts began in 1979 when Groves et al. developed a tetraphenylporphyrinato iron(III) chloride complex and used it to catalyze the epoxidation of styrene and hydroxylation of cyclohexane.³¹ An oxygen source is required to oxidize the original metalloporphyrin complex to its active high-valent metal-oxo intermediate. Groves et al. originally used iodosylbenzene (PhIO) as an oxygen source, but hydrogen peroxide (H₂O₂), iodobenzene diacetate [PhI(OAc)₂] and even molecular oxygen have been used as oxygen sources in later reports.^{4, 18, 32} The choice of oxygen source is significant in that it can affect the reactivity and stability of the catalyst. Stronger oxidants, like iodosylbenzene and hydrogen peroxide, often result in faster formation of the high-valent metal-oxo species, but catalyst bleaching is increased as a result.³³ More recently, mild oxygen sources like iodobenzene diacetate have been used with metalloporphyrins to improve catalyst stability.^{18, 34} Additionally,

environmental safety concerns have brought about the use of greener oxygen sources like hydrogen peroxide and molecular oxygen, whose only byproduct is water.^{32, 35} Figure 3 shows the structure of common metalloporphyrin catalysts.³⁶⁻³⁸ Manganese and ruthenium porphyrins are often studied because of their electronic similarity to iron coupled with the wide variety of available oxidation states in both metals. Manganese porphyrins in particular have recently been shown to efficiently and selectively catalyze epoxidations and sulfoxidations.^{18, 26}

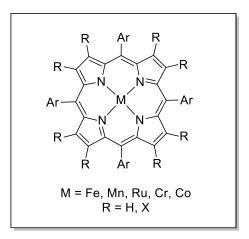


Figure 3. General structure of metalloporphyrin catalysts.

1.4 Manganese Corroles as Biomimetic Catalysts

Corroles are 19-carbon aromatic structures analogous to porphyrins (Figure 4). The backbone is similar to that of corrin, the core structure of vitamin B12; the structures differ in that corrole is fully conjugated and aromatic.³⁹ The stability of corroles differs from that of porphyrins as corroles are prone to thermal degradation, contrasting the extremely stable porphyrin ligands. However, their high-valent metal complexes are significantly more stable than the analogous porphyrin metal complexes because of the trianionic nature of the corrole ligand.⁴⁰

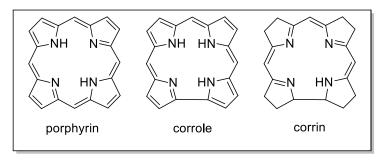
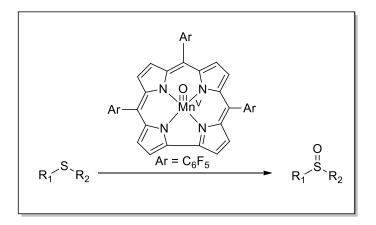


Figure 4. The structures of the porphyrin, corrole and corrin ligands.

The first corrole ligand was synthesized in 1965 by Johnson et al.⁴¹ This difficult synthesis meant that the development of corrole catalysts was impractical for many years. In 1999, Gross et al. reported a facile, solvent-free synthesis of electron-poor corroles, and since then, the development of electron-poor metallocorrole catalysts has been an active field.⁴² Metallocorroles have also been used in biochemical applications, as anticancer agents,⁴³⁻⁴⁸ in antioxidant therapy⁴⁹⁻⁵⁰ and in diabetic therapy.⁵¹ The first report of metallocorrole catalysts appeared in 2000, when an iron(III) corrole was reported to be an effective catalyst towards epoxidation and cyclopropanation using iodosylbenzene as an oxygen source.⁵² More recently, an oxo-manganese(V) complex of tris(pentafluorophenyl)corrole [Mn^V(TPFC)O] was isolated as a relatively stable solid and used for stoichiometric sulfoxidations (Scheme 4).⁵³ This work aims to illustrate the efficacy of this corrole complex towards sulfoxidations and epoxidations under both catalytic and stoichiometric conditions.



Scheme 4. The oxidation of sulfides to sulfoxides with isolated Mn^VTPFC(O).

1.5 Significance of Sulfoxidation Reactions

The selective oxidation of sulfides to sulfoxides without overoxidation to sulfones is of importance in the pharmaceutical industry.⁵⁴ Sulfoxides are commonly used in the production of antibacterial and antifungal compounds.⁵⁵ The problem of selective production of sulfoxides is most relevant in the synthesis of proton pump inhibitors, used to treat ulcers and heartburn.⁵⁶ Conventional methods for these oxidations involve the use of strong, stoichiometric oxidants, usually peroxyacids, or toxic heavy metal catalysts.^{57, 58} Thus, environmentally-friendly sulfoxidation catalysis is a growing field.⁵⁹

Manganese porphyrins and corroles have been shown to efficiently catalyze sulfoxidations.⁵³ Porphyrin catalysts typically exhibit high conversions, but the selectivities of the catalysts could be improved.¹⁸ Corrole catalysts can exhibit greater selectivities, but their catalytic conversions suffer as a result.⁶⁰ In both cases, there are significant improvements to be made for more efficient catalytic methods. These difficulties extend to epoxidations by similar complexes.⁵² There exists a demand for a

versatile catalyst that can selectively perform both sulfoxidations and hydroxylations under mild conditions with high product selectivities and turnover numbers.

2. EXPERIMENTAL SECTION

2.1 Materials

Solvents used for synthesis and column chromatography were used as received from Aldrich Chemical Co. without further purification, including acetone, dichloromethane, chloroform, acetonitrile, methanol, ethanol, diethyl ether, hexane and N,N-dimethylformamide (DMF). When used for kinetic and catalytic studies, these solvents were passed through a dry column of activated alumina (Grade I) to remove any water and impurities. Reactive substrates for kinetic and catalytic studies were also purchased from Aldrich Chemical Co. and purified before use via dry colmns of activated alumina to remove impurities. These substrates include cyclohexene, ciscyclooctene, ethylbenzene, styrene, p-chlorostyrene, p-fluorostyrene, p-methylstyrene, *p*-vinylanisole, *p*-chlorothioanisole, thioanisole, *p*-fluorothioanisole, pmethoxythioanisole and methyl *p*-tolyl sulfide. Pyrrole was freshly distilled immediately before use. Other reagents, including mesitaldehyde, pentafluorobenzaldehyde, benzaldehyde, boron trifluoride diethyl etherate (BF₃·OEt₂), 2,3-dichloro-5,6-dicyano-pbenzoquinone (DDQ), triethylamine, iodobenzene diacetate [PhI(OAc)₂], hydrochloric acid (HCl), manganese(II) acetate tetrahydrate, chloroform-d, silver nitrate, tetrabutylammonium perchlorate (TBAP) and ferrocene, were used as received without further purification.

2.2 General Procedures

2.2.1 Physical Measurements

UV-Visible absorption spectroscopy was performed on an Agilent 8453 diode array spectrophotometer. IR spectra were recorded using a Spectrum One FT-IR. ¹H-

NMR was performed on a JEOL ECA-500 MHz instrument with tetramethylsilane (TMS) as an internal standard. All chemical shifts are reported relative to TMS. Gas chromatography/mass spectrometry analyses were conducted with an Agilent GC 6890/MS 5973 equipped with a flame ionization detector (FID) and an autosampler. An Agilent J&W Cyclodex-B chiral capillary column was used for product analysis with 1,2,4-trichlorobenzene as an internal standard. Cyclic voltammetry was performed on a PARSTAT 2263 with a glassy carbon working electrode, a platinum wire counter electrode and an Ag/AgNO₃ reference electrode. Solutions of TBAP (0.1 M) and metal complex (1 mM) in acetonitrile were used for analysis.

2.2.2 Catalytic Oxidations

Unless otherwise specified, all catalytic reactions occurred in 0.5 mL of an anaerobic solvent, either dichloromethane, methanol or acetonitrile, with 1 μ mol catalyst, 0.2 mmol substrate and 0.3 mmol iodobenzene diacetate (1.5 equiv.) as an oxygen source. Catalysis with manganese corroles occurred in solutions in the absence of water, while catalysis with manganese porphyrin catalysts occurred in the presence of a small amount of water. Reactions were carried out at 23 ± 2°C. Aliquots of the reaction mixtures were analyzed by GC/MS to determine the progress of the reaction as well as the ratios of products. Reactions were run at least twice; all data reported represents the average result of these runs.

2.3 Synthesis and Spectroscopic Characterization

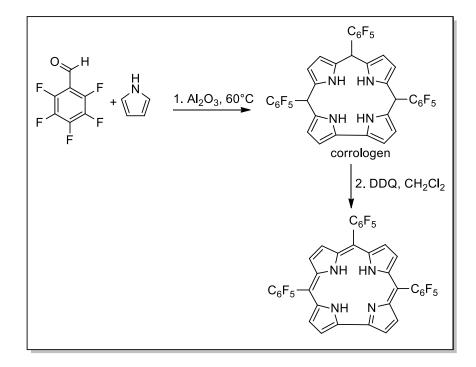
2.3.1 Tris(pentafluorophenyl)corrole (H₃TPFC, **1a**)

The synthesis of H₃TPFC was performed according to the method described by Gross et al. in 1999,⁴² shown in Scheme 5. Pentafluorobenzaldehyde (2.94 g, 15 mmol)

and distilled pyrrole (1.04 mL, 15 mmol) were dissolved in a small amount of CH_2Cl_2 (ca. 2 mL). The mixture was added to a round-bottomed flask containing alumina (3 g) as a solid support. This mixture was heated to 60°C. After the CH_2Cl_2 had completely evaporated, the mixture was allowed to react for 4 h, changing from colorless to dark brown. This first solvent-free condensation step forms the corrologen. Next, CH_2Cl_2 (50 mL) was added to the tarry mixture, and the solution was stirred for about 10 min to dissolve. This solution was filtered under vacuum. The filtrate was collected and further reacted with 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ, 1.7 g, 7.5 mmol, 2 equiv.) in a one-electron oxidation for 1 h that completed the aromatic system. Reaction progress was monitored using thin layer chromatography (TLC). A purple band was seen with strong fluorescence under UV light. If left at ambient temperature for several hours, this band turned green. The band represented the desired corrole product.

The product was isolated on a series of normal-phase columns with silica gel (SiO_2) . First, the tarry portion of the product mixture was separated from the rest of the products using a fairly short (4-5 inches) column with a mixture of dichloromethane and hexane (1:3 v/v) as the eluent. All products were collected from this column, while the tarry, insoluble impurities were removed. The desired product was isolated in an additional column. This column was much longer (8-9 inches) with dichloromethane and hexane (1:6 v/v) as the eluent. A purple band that strongly fluoresced under UV light was collected. The product and its purity were analyzed using ¹H-NMR, and a third column was performed as needed for further purification. The final product was characterized by ¹H-NMR (Figure 5) and UV-vis (Figure 6) spectroscopies, matching literature reports.^{42, 61}

H₃TPFC (**1a**) Yield% = 5% (145 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -2.25 (s, 3H, N-H), 8.65 (4H), 8.75 (2H), 9.12 (2H). UV-Vis (CH₂Cl₂) λ_{max} (nm): 408, 560, 600.



Scheme 5. The two-step synthesis of H₃TPFC (1a).

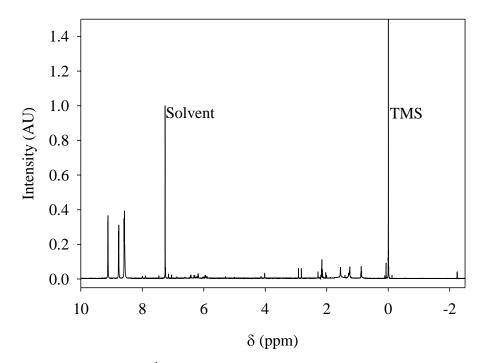


Figure 5. The ¹H-NMR spectrum of H₃TPFC (1a) in CDCl₃.

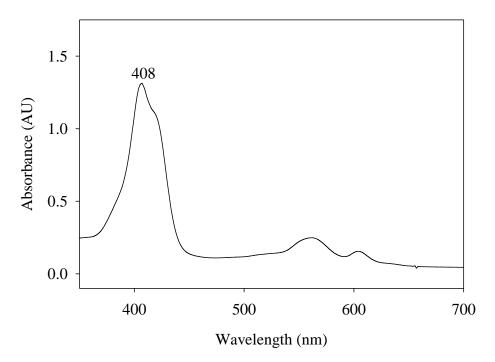
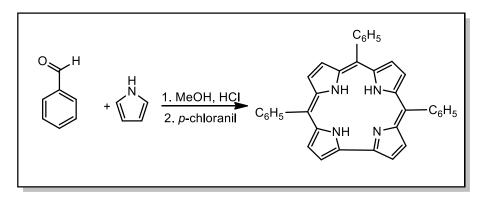


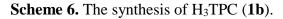
Figure 6. The UV-visible absorption spectrum of H₃TPFC (1a) in CH₂Cl₂.
2.3.2 Triphenylcorrole (H₃TPC, 1b)

This corrole was prepared based on the report of Koszarna and Gryko in 2006,⁶² as shown in Scheme 6. Benzaldehyde (510 μ L, 5 mmol), freshly distilled pyrrole (700 μ L, 10 mmol) and 500 mL of methanol were mixed in a 1 L round-bottomed flask. The solution was degassed with argon for 5 min. To start the reaction, concentrated hydrochloric acid was added as the catalyst; the mixture was stirred for 3 h and monitored by TLC. The resulting mixture was extracted with CHCl₃ (100 mL) three times, and the remaining organic layer was then washed with DI water to remove residual acid. The resulting solution was then dried with sodium sulfate. *p*-Chloranil (1.23 g, 5 mmol) was added and the mixture was refluxed for 1 h, monitored by TLC. The resulting mixture was purified on a column of silica gel and eluted with dichloromethane to yield a brown-green solid product. This product was characterized

with ¹H-NMR and UV-vis, shown in Figures 7 and 8, respectively, matching literaturereported spectra.⁶²

H₃TPC (**1b**) Yield% = 7% (80 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -2.90 (3H, N-H), 7.73-7.83 (9H), 8.17 (2H), 8.38 (4H), 8.55 (2H), 8.60 (2H), 8.87 (2H), 8.95 (2H). UV-Vis (CH₂Cl₂) λ_{max} (nm): 415, 568, 615.





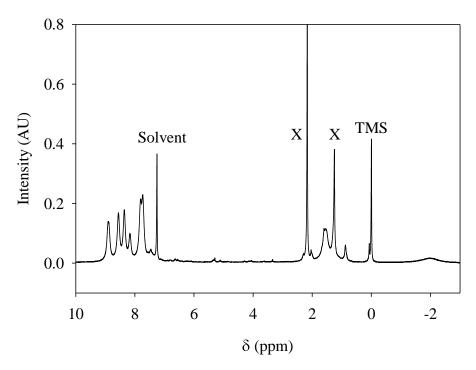


Figure 7. The ¹H-NMR spectrum of H₃TPC (1b).

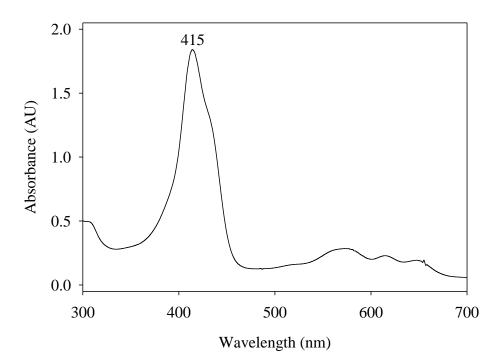


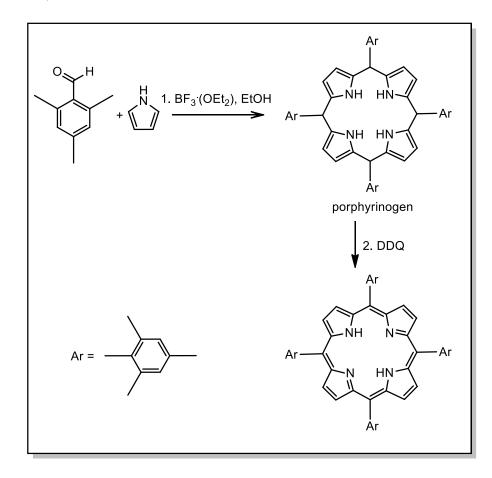
Figure 8. The UV-visible absorption spectrum of H₃TPC (1b).
2.3.3 *meso*-Tetramesitylporphyrin (H₂TMP, 1c)

This free porphyrin ligand was synthesized according to the method reported by Lindsey et al.,⁶³ shown in Scheme 7. Mesitaldehyde (736 μ L, 5 mmol), distilled pyrrole (347 μ L, 5 mmol) and ethanol (3.47 mL, 0.5% v/v) were dissolved in chloroform (500 mL) in a 1L round-bottomed flask, and the mixture was degassed under argon for 5 min. Boron trifluoride diethyl etherate (BF₃·OEt₂, 660 μ L, 1.65 mmol) was added in a dropwise manner, and the solution was stirred at room temperature for 1 h. The BF₃·OEt₂ and ethanol act as co-catalysts for the condensation step, forming the porphyrinogen. This step of the synthesis was monitored using UV-visible spectroscopy, as the intermediate product shows a distinctive band at 480 nm.

To complete the formation of the porphyrin, the porphyrinogen solution was refluxed with an excess of DDQ (1.36 g, 6 mmol) for 1 h. This one-electron oxidation process was again monitored using UV-visible spectroscopy. Triethylamine (920 μ L, 6.6

mmol) was added after the reflux to quench the highly acidic solution. The solvent was evaporated, and the crude product was washed with methanol under vacuum until the filtrate became clear. The bright purple product was further purified using a column of silica gel with dichloromethane as the eluent. The product was characterized with UV-visible spectroscopy and ¹H-NMR, shown in Figures 9 and 10, respectively, matching the literature.⁶³

H₂TMP (**1c**) Yield = 18% (185 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -2.50 (2H, N-H), 1.81 (24H, *o*-CH₃), 2.62 (12H, *p*-CH₃), 7.25 (8H, *m*-H), 8.61 (8H, pyrrolic). UV-Vis (CH₂Cl₂) λ_{max} (nm): 416, 440, 516.



Scheme 7. The synthesis of H₂TMP (1c).

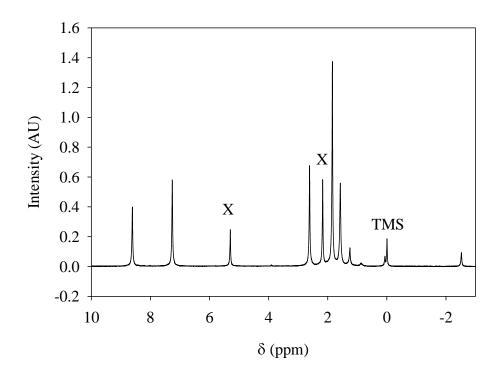


Figure 9. The ¹H-NMR spectrum of H_2 TMP (1c).

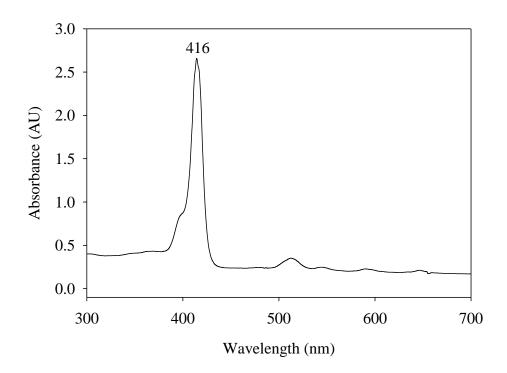


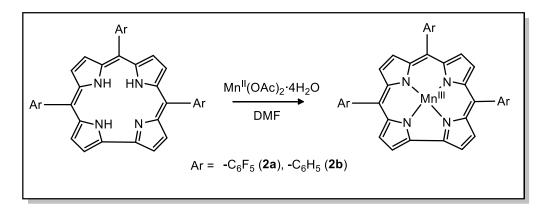
Figure 10. The UV-visible absorption spectrum of H₂TMP (1c).

2.3.4 Manganese(III) Corroles [Mn^{III}Cor·(OEt₂)₂, **2**]

The insertion of manganese into the corrole ligands was achieved by a reflux of the free ligand with manganese(II) acetate tetrahydrate in DMF, shown in Scheme 8. Free-base corrole (100 mg) and a large excess of Mn^{II}(OAc)₂·4H₂O (200 mg) were dissolved in DMF (50 mL) in a round-bottomed flask fitted with a reflux condenser. The solution was refluxed for 1 h, and reaction progress was monitored by TLC.

After reflux, DMF was removed by rotary evaporation. The crude, solid product was purified by a column of silica gel with diethyl ether as the eluent. The pure product was isolated as a dark green band. Both products were characterized by UV-vis and ¹H-NMR spectroscopies with **2a** spectra shown in Figures 11 and 12. **2b** spectra are shown in Figures 13 and 14. As expected for paramagnetic complexes, the ¹H-NMR spectra of **2a** and **2b** are characterized by an unusually large chemical shift range and exceptionally broad peaks.

Mn^{III}TPFC·(OEt₂)₂ (**2a**) Yield% = 88% (97 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -23.82, -4.16, 1.04, 3.17, 8.34, 10.80. UV-Vis (CH₂Cl₂) λ_{max} (nm): 418, 484, 602. Mn^{III}TPC·(OEt₂)₂ (**2b**) Yield% = 90% (102 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -29.93, -1.79, 1.43, 7.68, 14.28, 15.27–17.29. UV-Vis (CH₂Cl₂) λ_{max} (nm): 313, 430.



Scheme 8. Synthesis of manganese(III) corrole complexes.

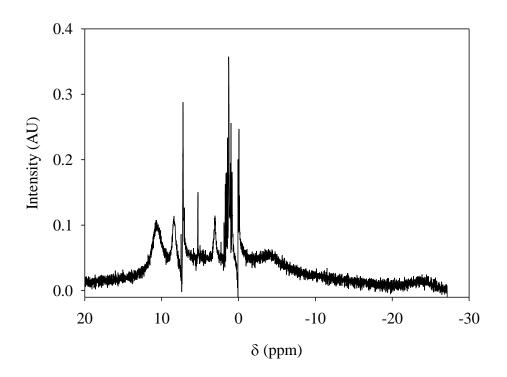


Figure 11. The ¹H-NMR spectrum of **2a** in CDCl₃.

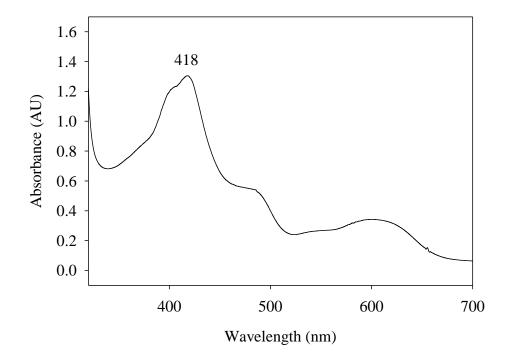


Figure 12. The UV-visible absorption spectrum of 2a in CH₂Cl₂.

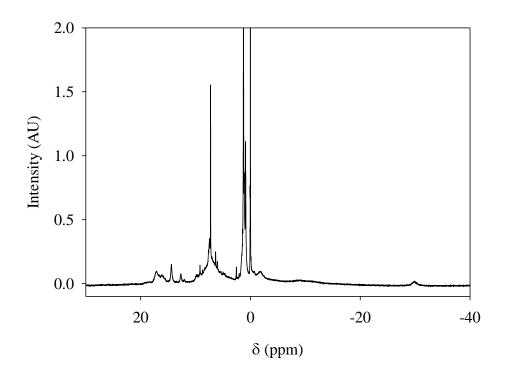


Figure 13. The ¹H-NMR spectrum of 2b in CDCl₃.

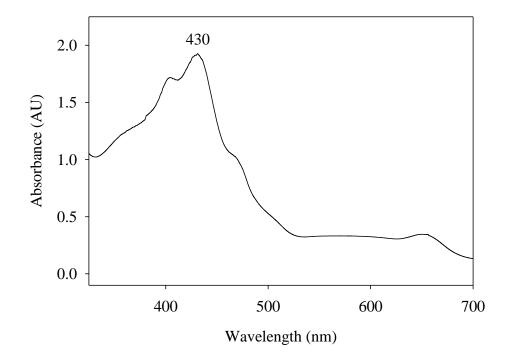


Figure 14. The UV-visible absorption spectrum of 2b in CH₂Cl₂.

2.3.5 Manganese(III) Porphyrin Chloride [Mn^{III}(TMP)Cl, 2c]

The synthesis of the manganese(III) porphyrin is similar to that of the manganese(III) corroles. The free ligand (100 mg) and a large excess of $Mn^{II}(OAc)_2 \cdot 4H_2O$ (300 mg) were dissolved in DMF (50 mL) in a 100 mL round-bottomed flask fitted with a reflux condenser. This solution was degassed for 5 min under argon, then refluxed for 1 h. Reaction progress was monitored by TLC.

After reflux, the DMF was removed by rotary evaporation, and the crude product was dissolved in dichloromethane (50 mL). This solution was mixed with hydrochloric acid (6 M, 50 mL) in order to exchange the axial ligand from hydroxy to chloride. The solution was extracted with dichloromethane (50 mL) three times. The remaining organic layer was then washed with of DI water (50 mL) three times and dried with sodium sulfate. The solvent was evaporated and the desired product was purified in a column of silica gel and eluted with a mixture of dichloromethane and hexane (1:1 v/v). The pure product was isolated as a bright green band and characterized by ¹H-NMR and UV-vis in Figure 15 and Figure 16, respectively. The manganese(III) porphyrin complex is paramagnetic, consistent with its broad ¹H-NMR spectrum.

Mn^{III}(TMP)Cl (**2c**) Yield%= 92% (105 mg). ¹H-NMR (500 MHz, CDCl₃) δ (ppm): -21.2, 1.31, 2.63, 7.57–8.94. UV-Vis (CH₂Cl₂) λ_{max} (nm): 375, 480, 585, 623.

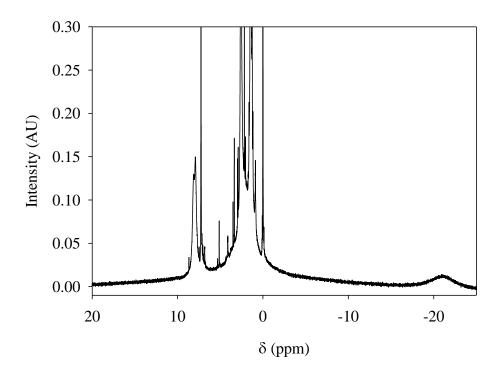


Figure 15. The ¹H-NMR spectrum of 2c in CDCl₃.

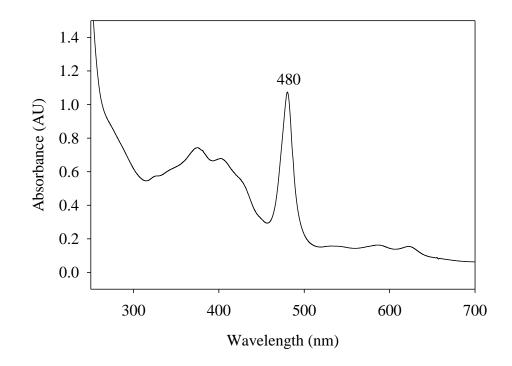


Figure 16. The UV-visible absorption spectrum of 2c in CH₂Cl₂.

3. KINETIC STUDIES OF STOICHIOMETRIC OXIDATIONS BY Mn^V(TPFC)O AND Mn^{IV}(TMP)O

3.1 Introduction

Manganese porphyrins and corroles are among the most widely used oxygen atom transfer (OAT) catalysts because of the wide variety of available oxidation states that are accessible to manganese complexes. High-valent metal-oxo species are fundamentally important due to the roles they play in catalytic OAT mechanisms. Identification of the active metal-oxo complex is decidedly complicated, as disproportionation and comproportionation mechanisms can allow for the active species to be generated in nearly undetectable amounts. This holds true for manganese(V)-oxo corroles, whose isolated forms are reported to be inactive towards olefins, despite their known catalytic activity in solution. Manganese corroles have been shown to efficiently perform stoichiometric sulfoxidations, with evidence of a multiple-pathway mechanism of oxidation that depends on the electron-rich or poor nature of the corrole ligand. This work reports that a single corrole system can access multiple pathways of the mechanism depending on the solvent used.

3.2. Results and Discussion

3.2.1 Electrochemical Studies

Cyclic voltammetry was used to characterize and compare the oxidation potentials of **2a** and **2b**. These voltammograms are shown in Figures 17 and 18, respectively. The oxidation of ferrocene under the same conditions gave $E_{1/2} = 85 \text{ mV}$. Notably, the potential at the cathodic and anodic peaks differs by less than 80 mV for both **2a** and **2b**, clearly indicating a one-electron oxidation from manganese(III) to manganese(IV) in both systems. Figure 19 shows a linear Cottrell plot⁶⁴ of peak current versus the square root of the scan rate for 2a, verifying that the process observed is diffusion-controlled.

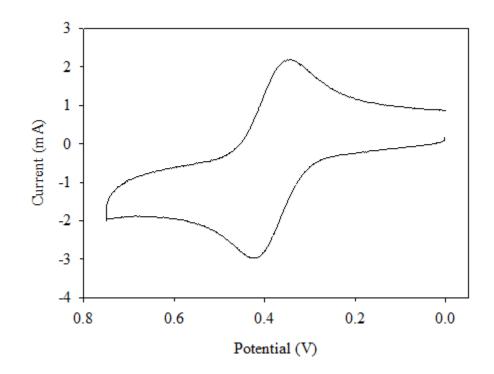


Figure 17. A cyclic voltammogram of **2a** (1 mM) in a 0.1 M TBAP/CH₃CN solution at scan rate 100 mV/s and with a three-electrode system: a glassy carbon working electrode, a platinum wire counter electrode and an Ag/AgNO₃ reference electrode. The $E_{1/2}$ of the system under these conditions is 385 mV.

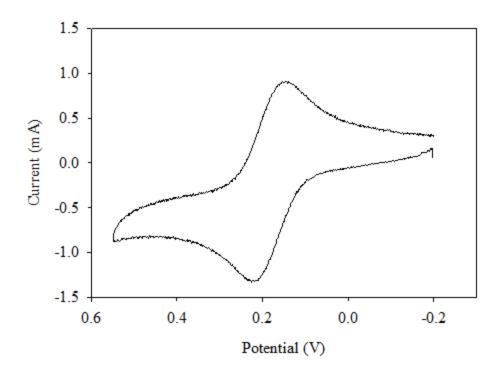


Figure 18. A cyclic voltammogram of **2b** (1 mM) in a 0.1 M TBAP/CH₃CN solution at scan rate 100 mV/s with a three-electrode system: a glassy carbon working electrode, a platinum wire counter electrode and a Ag/AgNO₃ reference electrode. The $E_{1/2}$ of the system under these conditions is 186 mV.

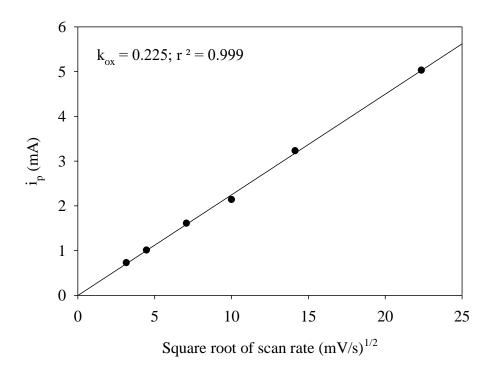


Figure 19. A Cottrell plot of the peak current of the oxidation of **2a** at varied scan rates. This plot is linear for diffusion-controlled electrochemical systems, and k_{ox} is a collection of constants specific to the system.

The relative oxidation potentials of **2a** and **2b** can be explained by the electrondemanding nature of the TPFC ligand. Upon oxidation of **2a**, the resulting manganese(IV) species is more destabilized, so it becomes much more difficult to perform this oxidation. This also implies that the oxidizing power of a high-valent metal-oxo species with a more electron-demanding corrole ligand is stronger than that of a non-electron deficient ligand in view of the electrophilic nature of the high-valent manganese-oxo species.

3.2.2 Solvent Effect

In previous work by Kumar et al., the reactivity of isolated $Mn^{V}(TPFC)O$ (**3a**) towards OAT in ethyl acetate was explored.⁵³ This work, however, aims to explore the

effects of differing solvents on the reactivity and the reaction pathways of 3a. To determine the appropriate solvent for our experiments, the stability of 3a was first examined in each of the chosen solvents: dichloromethane, acetonitrile, methanol and chloroform. **3a** was generated *in situ* in 2 mL of the chosen solvent from the corresponding precursor 2a (2.5×10^{-5} M) using *m*-CPBA (3 equiv.) as a sacrificial oxidant. *m*-CPBA was chosen because of its oxidizing power with the metallocorrole. The generation of **3a** was verified using time-resolved UV-vis spectra over 40 s, as the spectrum for **3a** is well-characterized in the literature.⁵³ For comparison, the stability of Mn^{IV}(TMP)O (4c) in various solvents was also studied. 4c was generated from its precursor 2c (1.8 \times 10⁻⁵ M) in 2.0 mL solvent with PhI(OAc)₂ (10 equiv.) as the sacrificial oxidant. $PhI(OAc)_2$ was chosen instead of *m*-CPBA because of its mild oxidizing ability, which allows for the formation of a manganese(IV)-oxo instead of a manganese(V)-oxo porphyrin. This is attributed to the comproportionation of the manganese(III) precursor and the manganese(V)-oxo species into a manganese(IV)-oxo species. Figure 20 shows the UV-vis spectra of **3a** and **4c** in CH₃CN.

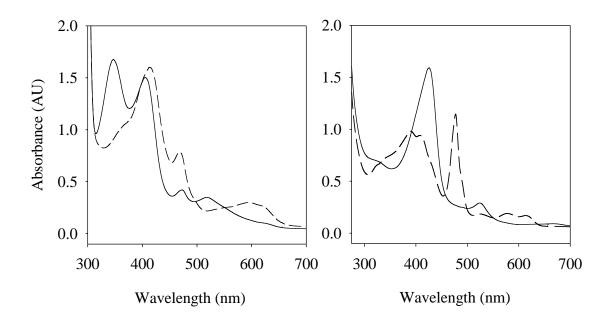


Figure 20. The UV-visible absorption spectra of 3a (left, solid line) generated by oxidation of 2a (left, dashed line) with *m*-CPBA in CH₃CN and 4c (right, solid line) generated by oxidation of 2c (right, dashed line).

3a was initially expected to regenerate **2a** over time, as is expected under catalytic conditions to allow regeneration of the catalyst. In CH₃CN, that is the case; Figure 21 shows time-resolved spectra of **3a** decaying into **2a** over the course of 3 h in CH₃CN, with an observed pseudo-first-order rate constant of $k_{obs} = 4.2 \times 10^{-5} \text{ s}^{-1}$. This decay is signaled by a decreased intensity at 349 nm as well as a distinctive peak forming at 465 nm. This is similar to previously-reported results with ethyl acetate as the solvent.⁵³ However, in both CH₂Cl₂ and CHCl₃, a suspected Mn^{IV}(TPFC)X species was observed. The identification of this species as a manganese(IV) is based on reported manganese(IV) corroles generated by Laser Flash Photolysis (LFP) studies.⁶⁵ Figure 22 shows the decay of **3a** to Mn^{IV}(TPFC)X in CH₂Cl₂ over 5 h, with $k_{obs} = 2.0 \times 10^{-5} \text{ s}^{-1}$, monitored at 349 nm. To fully investigate the kinetics of OAT by **3a**, reactions were

studied in both CH_2Cl_2 and CH_3CN . CH_2Cl_2 was chosen because of the longer lifetime of **3a** compared to that in CHCl₃.

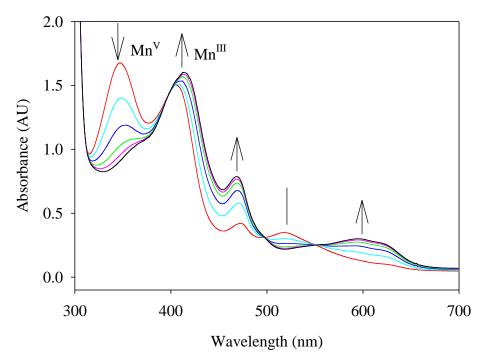


Figure 21. Time-resolved UV-vis spectra of the decay of **3a** into **2a** in CH₃CN in the presence or absence of substrate over 3 h.

The solvent effect of **3a** is markedly different from that of metalloporphyrins and other reported metallocorrole systems. Kwong et al. reported that this unique solvent effect is essentially nonexistent with **3b**, which is comparatively not electron deficient, and a manganese(IV) species is formed in all solvents.³⁵ Figure 23 shows the decay of **4c** into **2c**. Interestingly, **4c** decayed to **2c** consistently in CH₃CN, CH₂Cl₂ and CHCl₃. CH₃CN was chosen for further testing of **4c** because of its stability in the solvent.

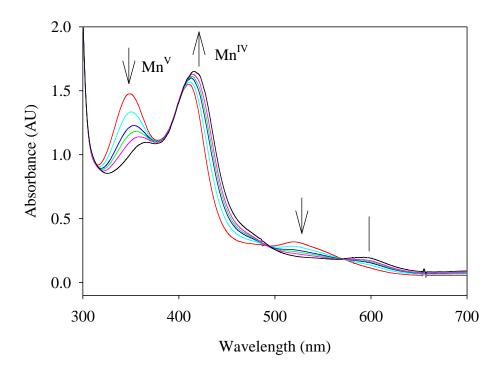


Figure 22. Time-resolved spectra of the decay of 3a in the presence or absence of substrate into a suspected Mn^{IV}(TPFC)X over 5 h.

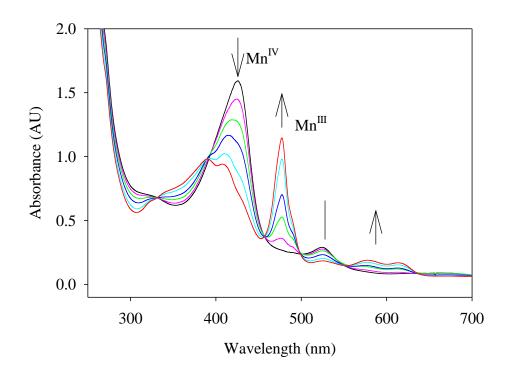


Figure 23. Time-resolved spectra of the decay of 4c into 2c in CH₃CN in the presence or absence of substrate over 20 min.

3.2.3 Kinetic Studies of Oxidations by 3a

The reactivity of **3a** towards single-turnover OAT to varied substrates was investigated under pseudo-first-order conditions. At minimum, 1000 equiv. of substrate was reacted with a freshly prepared solution of **3a** in the chosen solvent, essentially keeping the concentration of the substrate constant and allowing for an approximated first-order rate constant to be determined from the rate of decay of the 349 nm Soret band. When plotted versus [Sub], the pseudo-first-order rate constants generate a second-order rate constant that more accurately describes the reaction with a particular substrate in a specific solvent. An example kinetic trace and a representative kinetic plot are shown in Figures 24 and 25 for thioanisole. The linearity of the kinetic plot verifies that the concentration of the substrate remains essentially constant, and the R^2 values for these plots range from 0.999–0.983. Table 1 shows second-order rate constants for all substrates in each solvent.

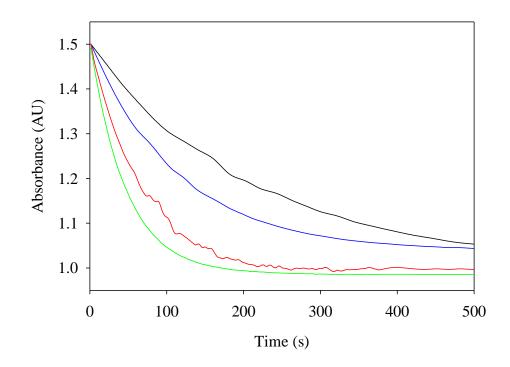


Figure 24. Kinetic traces monitored at $\lambda_{max} = 349$ nm, showing the decay of 3a (2.5 × 10⁻⁵ M) into 2a in CH₂Cl₂ with varied concentrations of thioanisole: 10 mM (black), 20 mM (blue), 30 mM (red) and 40 mM (green).

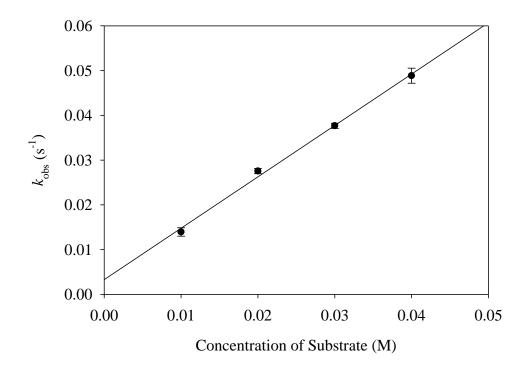


Figure 25. A plot of the observed rate constants versus the concentration of thioanisole in CH₃CN. Error bars are reported as 1σ .

Entry	Substrate	Solvent	$k_2 (\times 10^3 \mathrm{M}^{-1}\mathrm{s}^{-1})$
1	cyclohexene	CH_2Cl_2	16 ± 0.2
2		CH ₃ CN	3.9 ± 0.04
3	cis-cyclooctene	CH_2Cl_2	11 ± 0.8
4		CH ₃ CN	2.8 ± 0.1
5	styrene	CH ₂ Cl ₂	3.4 ± 0.07
6		CH ₃ CN	2.4 ± 0.1
7	ethylbenzene	CH ₂ Cl ₂	0.27 ± 0.03
8	thioanisole	CH ₂ Cl ₂	530 ± 12
9		CH ₃ CN	870 ± 8
10	<i>p</i> -fluorothioanisole	CH ₃ CN	1200 ± 40
11	<i>p</i> -chlorothioanisole	CH ₂ Cl ₂	120 ± 6
12		CH ₃ CN	550 ± 16
13	methyl <i>p</i> -tolyl sulfide	CH ₃ CN	2112 ± 60

Table 1. Kinetics of oxidation reactions by 3a^a

^aIn 2.0 mL of solvent with **3a** $(2.5 \times 10^{-5} \text{ M})$ generated *in situ* by oxidation of **2a** with *m*-CPBA (3 equiv.).

The second-order rate constants for sulfoxidations, epoxidations and hydroxylations follow their relative orders of reactivity, differing by several orders of magnitude. Notably, changing the concentration of 2a in CH₂Cl₂ affected the second-order rate constant for the same substrate, clearly indicating that the rate law is not actually first order in 3a. This effect was not observed in CH₃CN.

In general, epoxidations are faster in CH_2Cl_2 than in CH_3CN , while sulfoxidations follow the opposite pattern. Presumably, sulfoxidations proceed through a different mechanism in which positive charge accumulates on the sulfur.

For comparison, the reactivity of $Mn^{IV}(TMP)O$ (4c) towards OAT was also studied under single-turnover, pseudo-first-order reaction conditions. In particular, *p*substituted styrenes were examined. These second-order rate constants are reported in Table 2. 4c is significantly more reactive towards OAT than 3a owing to the dianionic nature of the porphyrin ligand. Corrole ligands are trianionic, and consequentially, their high-valent metal complexes are inherently more stable than high-valent metal-oxo porphyrins.

Entry	Substrate	$k_2 (\times 10^3 \mathrm{M}^{-1}\mathrm{s}^{-1})$
1		820 ± 9
2	CI	680 ± 14
3	F	1100 ± 20
4	Me	1900 ± 50
5	MeO	5200 ± 70

Table 2. Rate constants of OAT to *p*-substituted styrenes by 4c^a

^aIn 2.0 mL CH₃CN with 4c (1.8×10^{-5} M) formed *in situ* by oxidation of 2c by PhI(OAc)₂ (10 equiv.).

3.2.4 Hammett Correlation Studies

To more thoroughly probe the OAT mechanism, a series of *p*-substituted thioanisole substrates were oxidized with **3a**. Table 3 shows the second-order rate constants obtained from these substrates. A Hammett plot describes the relationship between the rate constants of reactions with substituted aromatic substrates and a substituent constant σ^+ . The substituent constant combines several factors, including electronegativity and electron density, to essentially quantify the electron donating or withdrawing nature of a substituent. Larger values indicate more electron-withdrawing substituents. When the ratio of the rate constants associated with the substituted and unsubstituted substrate is plotted versus σ^+ , the resulting linear plot quantifies how significantly the identity of a substituent can affect the rate of a reaction. The slope of the Hammett plot, or reaction constant, ρ , is specific to the type of reaction. The slope of ρ indicates whether electron-donating or electron-withdrawing substituents increase the rate of reaction, while the magnitude of ρ corresponds to the magnitude of the effect. The Hammett plot of substituted thioanisoles with **3a** in CH₃CN is shown in Figure 26.

Entry	Substrate	$k_2 (\times 10^3 \mathrm{M}^{-1}\mathrm{s}^{-1})$
Entry	Substrate	$K_2 (\times 10 \text{ WLS})$
1	S S	870 ± 8
2	CI	550 ± 16
3	F	1200 ± 40
4	Me	2100 ± 60
5	MeO	6700 ± 80

Table 3. Rate constants of OAT to *p*-substituted thioanisoles by $3a^{a}$

^aIn 2.0 mL CH₃CN with **3a** (2.5×10^{-5} M) formed *in situ* by oxidation of **2a** with *m*-

CPBA (3 equiv.).

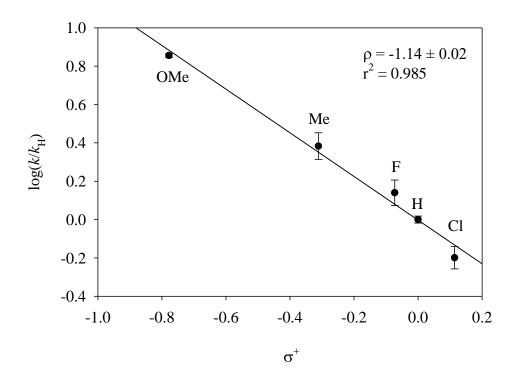


Figure 26. The Hammett plot for oxidations of *p*-substituted thioanisoles by 3a in CH₃CN.

The slope of $\rho = -1.14$ is similar to previously-reported values for these reactions under both chemical and photochemical conditions.^{35, 53} Given that the slope of ρ for this plot is negative, the reaction rate is positively affected by electron-donating substituents. This is discernible from Table 3, and it follows the relative reactivities of these substrates towards sulfoxidation. The magnitude of ρ , however, indicates that the reaction is significantly affected by the nature of the substituents; it describes the selectivity of **3a** as an OAT catalyst. The linearity of the plot implies an electrophilic mechanism, strongly suggesting that the active oxidant in CH₃CN is **3a**. Notably, the fastest-reacting of these substrates exclusively reformed **2a**, even in CH₂Cl₂.

For comparison, a Hammett plot was also constructed from the reactions of **4c** as shown in Table 2. This plot is shown in Figure 27. Although OAT by **4c** is significantly

faster than OAT by **3a**, the effect of the substituents is similar in magnitude and identical in sign; electron donating substituents significantly increase the rate.

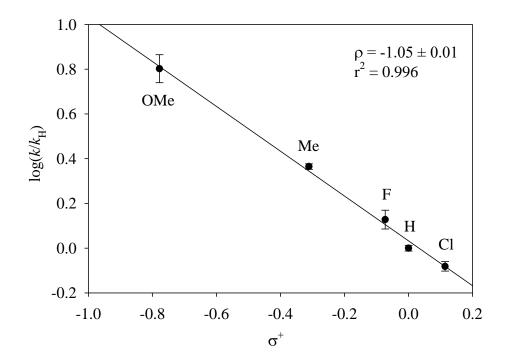
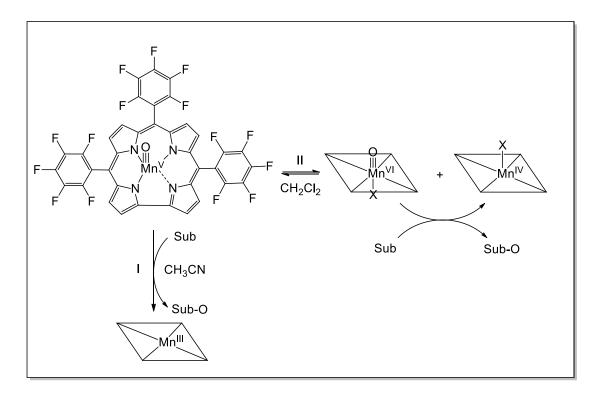


Figure 27. The Hammett plot for epoxidations of *p*-substituted styrenes by 4c in CH₃CN.

3.3 Mechanistic Considerations

The significant solvent effect present in this system supports the previously reported two-pathway mechanism of oxidation.⁵³ This mechanism suggests that the system can either undergo disproportionation or participate in direct oxidation. The disproportionation pathway converts two equivalents of **3a** to one equivalent each of a Mn^{IV}(TPFC)X and a Mn^{VI}(TPFC)O, where the manganese(VI) species acts as the active oxidant, and in performing the oxidation, is reduced to the manganese(IV) species. In the direct oxidation pathway, however, **3a** is the active species and directly performs the

oxidation on the substrate, simultaneously being reduced back to **2a**. Both pathways are shown in Scheme 9.



Scheme 9. The proposed two-pathway mechanism of oxidation by **3a**. Pathway I is the direct oxidation, while II is the disproportionation pathway.

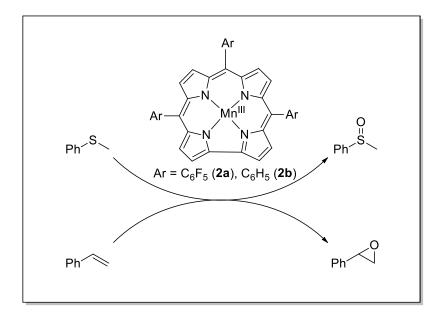
The products of the decay of **3a** in CH₃CN and CH₂Cl₂ support the direct oxidation and disproportionation pathways, respectively; reactions in CH₃CN result in regeneration of **2a**, while reactions in CH₂Cl₂ result in accumulation of the manganese(IV) species. Moreover, highly concentrated solutions of **3a** in CH₂Cl₂ decomposed faster into the manganese(IV) species than less concentrated solutions. Ideally, the concentration of **3a** should not affect its rate of decay; the effect implies that **3a** disproportionates, contributing to the accumulation of the manganese(IV) species. Essentially, the rate law is not first order in **3a**. This further supports the suggestion of a disproportionation pathway; Newcomb et al. proved using the steady-state

approximation that disproportionation would result in a fractional order for 3a.⁶⁵ This multiple-pathway mechanism is also supported by the report of a similar solvent effect under photochemical conditions. Kwong et al. reported a two-pathway, solvent-dependent mechanism of oxidation by photochemically-generated 3a.³⁵

4. CATALYTIC OXIDATIONS BY MANGANESE CORROLES

4.1 Introduction

Iodobenzene diacetate [PhI(OAc)₂] was chosen as the sacrificial oxidant for catalytic oxidations (Scheme 10). PhI(OAc)₂ is milder than commonly used oxidants such as *m*-CPBA, *tert*-butyl-hydroperoxide (TBHP) or iodosylbenzene (PhIO), each of which can bleach the catalyst, resulting in low catalytic efficiency. Additionally, these strong oxidants can directly oxidize the substrates, artificially increasing yields of oxidized products. Particularly, *m*-CPBA performs nearly quantitative epoxidation of alkenes. Under single-turnover, pseudo-first-order conditions, as outlined in the previous kinetic studies, the use of *m*-CPBA is not a concern because of the comparatively high concentrations of substrate. However, under catalytic conditions, the amount of sacrificial oxidant needed to propogate the catalytic cycle is significantly larger; in turn, the use of an oxidant that reacts with the substrate becomes less feasible. PhI(OAc)₂ is commercially available, and it is significantly safer to handle than stronger oxidants. In this section, the utility of PhI(OAc)₂ was further explored for the catalytic oxidation of alkenes and sulfides by manganese corrole complexes.



Scheme 10. Catalytic sulfoxidations and epoxidations by manganese(III) corroles.

4.2 Screening Studies

4.2.1 Solvent Effect

In an effort to determine optimal conditions for catalysis, screening studies were first performed. Given that a significant solvent effect on the OAT mechanism has been established in the previous section, the solvent effect was reexamined under catalytic conditions. The epoxidation of *cis*-cyclooctene was chosen as the probe for screening studies because of the relatively clean product, typically resulting in the epoxide only. Allylic oxidation to the corresponding alcohol or over-oxidation to the ketone often interferes with the epoxidation of cyclohexene. Likewise, sulfides can be overoxidized to form sulfones as opposed to the desirable sulfoxides. Table 4 shows the conversion of *cis*-cyclooctene to *cis*-cyclooctene oxide in CH₂Cl₂, CH₃CN and CH₃OH by **2a**.

Entry	Solvent	Time (h)	Conversion (%) ^c
1	dichloromethane	3	10
2		5	11
3	methanol	3	10
4		5	13
5 ^b		3	28
6	acetonitrile	3	11
7		5	13
8 ^b		3	33

Table 4. Solvent effect on the epoxidation of *cis*-cyclooctene by $2a^a$

^aUnless otherwise noted, all reactions took place in 0.5 mL of solvent at 23 ± 2 °C with 0.2 mol% of **2a**, 0.2 mmol *cis*-cyclooctene and 0.3 mmol PhI(OAc)₂.

 ${}^{b}40 \pm 2 {}^{\circ}C.$

^cDetermined by GC-MS analysis of the crude reaction mixture.

The reaction in dichloromethane was examined at 3 h, and *cis*-cyclooctene oxide was obtained as the only identifiable product at ca. 10% conversion (Table 4, entry 1). Slightly higher conversions were obtained after 5 h of reaction (entry 2). The slowing of the reaction over the course of the last 2 h indicates that the catalyst was bleached during the catalytic cycle. This is supported by observations of the color change of the solutions; the bright red color of the dichloromethane and acetonitrile solutions began to fade after about 3 h of reaction. Highest conversions at room temperature were seen in acetonitrile, while dichloromethane exhibited the lowest conversions (entry 2, entry 7).

All reactions benefitted from higher reaction temperature, and conversions of up to 33% were reached when heated to 40 °C (entry 8).

Notably, **2a** is catalytically active in methanol, resulting in conversions of 28% with heating (entry 5. However, **3a** was unable to be formed during the single-turnover kinetic studies. Methanol, which can form hydrogen bonds, may strongly coordinate to the metal, potentially preventing the binding of an oxygen source, which is crucial for the formation of a high-valent metal-oxo species. However, this is unlikely given the significant catalytic activity in this solvent; a high-valent metal-oxo species is proposed as the major oxidizing intermediate in the catalytic cycles of metallocorrole- and metalloporphyrin-catalyzed oxidations. Alternatively, the high-valent metal-oxo species could be so reactive in this solvent that it is reacted faster than it is formed, preventing the detection of this intermediate. The latter explanation is more reasonable because it accounts for the considerable catalytic activity in the solvent. Because CH₃CN exhibited the highest conversions, it was chosen for all remaining catalytic reactions with **2a**. For comparison, the solvent effect on the catalytic epoxidation of *cis*-cyclooctene by **2b** was also examined. Similar results were obtained and are shown in Table 5.

Entry	Solvent	Time (h)	Conversion (%) ^c
1	dichloromethane	3	6
2		5	8
3	methanol	3	7
4		5	10
5 ^b		3	18
6	acetonitrile	3	7
7		5	11
8 ^b		3	22

Table 5. Solvent effect on the epoxidation of *cis*-cyclooctene by 2b^a

^aUnless otherwise noted, all reactions took place in 0.5 mL of solvent at 23 ± 2 °C with 0.2 mol% **2b**, 0.2 mmol *cis*-cyclooctene and 0.3 mmol PhI(OAc)₂.

 $^{b}40 \pm 2 \ ^{\circ}C$

^cDetermined by GC-MS analysis of the crude reaction mixture.

The general trends in reactivity are similar to those of 2a. The reaction in dichloromethane was examined at 3 h, and *cis*-cyclooctene oxide was obtained as the only identifiable product with ca. 6% conversion (Table 5, entry 1). As with 2a, the reaction slowed over the course of the last 2 h, giving only an 8% conversion after 5 h total reaction time (entry 2), indicating catalyst bleaching. All reactions benefitted from heating, and in acetonitrile, conversions of 22% were reached with heating (entry 8). Acetonitrile shows the highest conversions of about 11% with no heating (entry 7) compared to dichloromethane's 8% under the same conditions (entry 2). Again, there

was catalytic activity in methanol, and ca. 10% conversion was obtained at room temperature (entry 4).

Interestingly, **2b** yielded lower conversions in all solvents. This is likely related to the electron-deficient nature of the pentafluorophenyl groups on **2a**, resulting in a more reactive high-valent manganese(V)-oxo species in comparison to **2b**. Additionally, fluorination of the corrole ligand in **2a** can increase its stability against catalyst bleaching under catalytic conditions.

4.2.2 Water Effect

In several metalloporphyrins and a reported iron(III) corrole with PhI(OAc)₂, catalytic efficiency is increased with the addition of small amounts of water. According to Chen et al., this effect is likely due to the hydrolysis of PhI(OAc)₂ into the stronger oxidant PhIO.⁶⁶ This explanation is also supported by the decrease in selectivity of the reported iron(III) corrole catalyst upon addition of water. Interestingly, catalytic activity for the reported iron(III) corrole and for reported metalloporphyrins began to decrease upon larger addition of water. In essence, these catalysts exhibited maximum conversions with small amounts of added water, and the conversion subsequently decreased as more water was added. This effect is most likely due to the highly polar nature of water; it can coordinate with the metal center and prevent the formation of critical high-valent metal-oxo species. The epoxidation of *cis*-cyclooctene by **2a** with added water was examined, and the results are present in Table 6.

Entry	Water (µL)	Conversion (%) ^b
1	0	18
2	5	16
3	10	15
4	20	14
5	40	11

Table 6. Water effect on the epoxidation of *cis*-cyclooctene by $2a^{a}$

^aAll reactions took place in 0.5 mL CH₃CN at 23 ± 2 °C with 0.2 mol% **2a**, 0.2 mmol *cis*-cyclooctene and 0.3 mmol PhI(OAc)₂ over 24 h.

^bDetermined by GC-MS analysis of the crude reaction mixture.

Of note, 2a expresses no positive water effect. Even the miniscule addition of 5 μ L negatively affects conversions (Table 6, entry 2). This may indicate that water coordinates more strongly to manganese(III) corroles than to iron(III) corroles, possibly reducing the formation rate of the high-valent manganese-oxo intermediate. For comparison, the epoxidation of *cis*-cyclooctene by **2b** was also tested under these conditions, and results are present in Table 7.

Entry	Water (µL)	Conversion (%)
1	0	14
2	5	12
3	10	11
4	20	9
5	40	6

Table 7. Water effect on the epoxidation of *cis*-cyclooctene by 2b^a

^aAll reactions took place in 0.5 mL CH₃CN at 23 ± 2 °C with 0.2 mol% **2b**, 0.2 mmol *cis*-cyclooctene and 0.3 mmol PhI(OAc)₂ over 24 h.

^bDetermined by GC-MS analysis of the crude reaction mixture.

2b exhibits essentially the same water effect as 2a, indicating that the lack of a positive water effect is likely unrelated to the corrole ligand itself, and instead more relevant to the manganese(III) center. This supports the assertion that the unique water effect is caused by strong coordination between the manganese center and water, preventing the formation of the manganese(V)-oxo species and thus slowing down the catalytic cycle.

4.3 Catalytic Oxidations of Sulfides to Sulfoxides

Catalytic sulfoxidations by 2a were investigated based on the results of the screening studies. Table 8 shows the results of sulfoxidations of *p*-substituted thioanisoles.

Entry	Substrate	Products	Conversion	Product
			(%) ^b	Ratio ^c
1	S_		49	67:33
2	CI		53	62:38
3	F		47	57:43
4	Me	Me Me	57	54:46
5	MeO	MeO MeO	59	51:49

Table 8. Catalytic sulfoxidation of *p*-substituted thioanisoles by 2a^a

^aAll reactions took place in 0.5 mL CH₃CN at 23 ± 2 °C with 0.2 mol% **2b**, 0.2 mmol substrate and 0.3 mmol PhI(OAc)₂ over 5 h.

^bDetermined by GC-MS analysis of the crude reaction mixture.

^cRatio of sulfoxide:sulfone products.

The sulfoxidation of thioanisole resulted in only 49% conversion with 2:1 formation of the sulfoxide over the sulfone (Table 8, entry 1). The incomplete conversion of substrate is due to catalyst bleaching. As expected based on the Hammett correlation studies, catalytic conversions increased with more electron-rich substrates.

The *p*-methoxythioanisole had conversions of 59% (entry 5), while selectivity dropped to a nearly even mix of the two products.

4.4 Catalytic Epoxidation of Alkenes

Catalytic epoxidations of p-substituted styrenes by 2a were investigated under the optimal conditions from the screening studies. These data are shown in Table 9.

Entry	Substrate	Product	Conversion (%) ^b	Product Ratio ^c
1			28	68:11:21
2	CI	CI	.o 29	57:11:32
3	F	F	o 31	54:11:35
4	Me	Me	.o 36	52:17:31
5	MeO	MeO	 O 39 	52:48 ^d

Table 9. Catalytic epoxidation of p-substituted styrenes by $2a^a$

^aAll reactions took place in 0.5 mL CH₃CN at 23 ± 2 °C with 0.2 mol% **2b**, 0.2 mmol substrate and 0.3 mmol PhI(OAc)₂ over 5 h.

^bDetermined by GC-MS analysis of the crude reaction mixtures.

^cRatio of epoxide:benzaldehyde:phenylacetaldehyde products

^dRatio of epoxide:benzyl alcohol products

The oxidation of styrene over 5 h yielded a mixture of products, including the epoxide, benzaldehyde and phenylacetaldehyde with 68% selectivity of epoxide formation (Table 9, entry 1). As with sulfoxidation, there is a clear trend wherein electron-donating *p*-substituents increase the conversions, and the *p*-fluorostyrene substrate resulted in higher conversions than the unsubstituted styrene at 31 and 28%, respectively (entry 1, entry 3). Selectivity decreased as reactivity increased, with the most reactive *p*-methoxystyrene reaching conversions of 39%, but yielding only 52% epoxide product. For comparison, **2c** was used to perform the same epoxidations under similar conditions established for manganese porphyrin catalysis.¹⁸ Results are shown in Table 10.

After 5 h of reaction in literature-established conditions for manganese porphyrin catalysis, near quantitative conversion was reached for even the unsubstituted styrene (Table 10, entry 1). The selectivity, however, is diminished to only 55% epoxide. The selectivity decreases with more electron-donating substituents, where the p-fluorostyrene substrate resulted in only 49% epoxide and significant amounts of the benzaldehyde and phenylacetaldehyde overoxidized products (entry 3). Additionally, the most reactive p-methoxystyrene produced three distinct overoxidized products in addition to the desired epoxide, including a 1° and 2° benzyl alcohol and a ketone. Although the metalloporphyrin catalyst was more reactive, giving near quantitative conversions, the selectivity is diminished compared to the less reactive metallocorrole catalyst.

Entry	Substrate	Products	Conversion (%) ^b	Product Ratio ^c
1			O 99 √	55:12:33
2	CI	CI	< <mark>0</mark> 99	51:17:32
3	F	F	\leq ⁰ 100	49:16:35
4	Me	Me	< <mark>○</mark> 100	48:17:35
5	MeO	MeO	100	44:12:10:33 ^d

Table 10.	Catalytic e	poxidation of	<i>p</i> -substituted	styrenes b	y 2c ^a
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^aAll reactions took place in 0.5 mL CH₃CN at 23 ± 2 °C with 0.2 mol% **2c**, 0.2 mmol

substrate, 5 μ L H₂O and 0.3 mmol PhI(OAc)₂ over 5 h.

^bDetermined by GC-MS analysis of the crude reaction mixture.

^cRatio of epoxide:benzaldehyde:phenylacetaldehyde products

^dRatio of epoxide:1° benzyl alcohol:ketone:2° benzyl alcohol products

5. CONCLUSIONS

In this work, manganese porphyrin and corrole systems (2) were synthesized as biomimetic models of Cytochrome P450 enzymes and subsequently characterized by UV-vis absorbance and ¹H-NMR. The role of high-valent manganeseoxo species in OAT by these systems was examined, and the catalytic abilities of these complexes towards epoxidation and sulfoxidation with PhI(OAc)₂ as an oxygen source were explored.

Manganese(V)-oxo corroles (3) and a manganese(IV)-oxo porphyrin (4) were generated from oxidation of the corresponding manganese(III) precursors (2) using mchloroperoxybenzoic acid (m-CPBA) as an oxygen source. Under single-turnover conditions, **3** complexes showed appreciable reactivity towards sulfoxidation, epoxidation and hydroxylation. In the tris(pentafluorophenyl)corrole system (3a), results from spectral studies indicate that the active oxidizing intermediate is solventdependent. In acetonitrile, the active oxidizing intermediate is most likely the manganese(V)-oxo species (3a). However, in dichloromethane, the active oxidant is suspected to be а putative manganese(VI)-oxo species generated from disproportionation of the manganese(V)-oxo species. This significant solvent effect is not present in other reported manganese corrole or porphyrin systems.

The catalytic activity of tris(pentafluorophenyl)corrolato manganese(III) (**2a**) towards thioanisoles and styrenes was examined with iodobenzene diacetate [PhI(OAc)₂] as a mild oxygen source. This complex was then compared to triphenylcorrolato manganese(III) (**2b**) and tetramesitylporphyrinato manganese(III) (**2c**). **2a** exhibited higher conversions than **2b**, most likely because of the relative

stability of **3b** compared to **3a**. In contrast, **2c** was much more reactive than **2a**, resulting in much higher conversions, but much less selectivity. Previously reported manganese porphyrin and iron corrole systems showed an accelerating effect upon addition of small amounts of water, attributed to the hydrolysis of the oxygen source to the more reactive iodosylbenzene (PhIO). In contrast to these systems, **2a** and **2b** did not have a positive water effect, but instead had decreased conversions with any added water. This is attributed to the strong coordination between manganese(III) and water, preventing the oxygen source from coordinating with the metal center. This coordination is crucial to generate the active metal-oxo intermediate.

The use of metallocorrole catalysts for selective oxidations is widely applicable. Most notably, selective sulfoxidation is relevant in the synthesis of many pharamceuticals, including proton pump inhibitors. In the literature, **3a** has been generated under photochemical conditions and used to perform OAT under environmentally-friendly conditions.³⁵ Currently, our lab is exploring the photochemical generation of other high-valent metallocorroles and metalloporphyrins for environmentally-friendly oxidations.

REFERENCES

 Sheldon, R. A., *Metalloporphyrins in Catalytic Oxidations*. Marcel Dekker: New York, 1994.

2. Ortiz de Montellano, P. R., *Cytochrome P450 Structure, Mechanism, and Biochemistry*. 3rd ed.; Kluwer Academic/Plenum: New York, 2005.

Brink, G. T., Green, Catalytic Oxidation of Alcohols in Water. *Science* 2000, 287 (5458), 1636-1639.

 Punniyamurthy, T.; Velusamy, S.; Iqbal, J., Recent Advances in Transition Metal Catalyzed Oxidation of Organic Substrates with Molecular Oxygen. *Chem.Rev.* 2005, *105* (6), 2329-2363.

5. Poulos, T. L.; Finzel, B. C.; Howard, A. J., Crystal Structure of Substrate-Free *Pseudomonas putida* Cytochrome P450. *Biochemistry* **1986**, *25*, 5314-5322.

Nelson, D. R.; Kamataki, T.; Waxman, D. J.; Guengerich, F. P.; Estabrook, R.
 W.; Feyereisen, R.; Gonzalez, F. J.; Coon, M. J.; Gunsalus, I. C.; Gotoh, O., The P450
 Superfamily: Update on New Sequences, Gene Mapping, Accession Numbers, Early
 Trivial Names of Enzymes, and Nomenclature. *DNA Cell Biol.* 1993, *12* (1), 1-51.

7. Wrighton, S. A.; Stevens, J. C., The Human Hepatic Cytochromes P450 Involved in Drug Metabolism. *Crit. Rev. Toxicol.* **1992**, *22* (1), 1-21.

8. Hasemann, C. A.; Kurumbail, R. G.; Boddupalli, S. S.; Peterson, J. A.; Deisenhofer, J., Structure and Function of Cytochromes P450: A Comparitive Analysis of Three Crystal Structures. *Structure* **1995**, *3* (1), 41-62.

Sono, M.; Roach, M. P.; Coulter, E. D.; Dawson, J. H., Heme-Containing
 Oxygenases. *Chem. Rev.* 1996, 96 (7), 2841-2887.

10. Denisov, I. G.; Makris, T. M.; Sligar, S. G.; Schlichting, I., Structure and chemistry of cytochrome P 450. *Chem. Rev.* **2005**, *105* (6), 2253-2277.

11. Poulos, T. L.; Finzel, B. C.; Howard, A. J., High-resolution Crystal Structure of Cytochrome P450_{cam}. *J. Mol. Biol.* **1987**, *195*, 687-700.

Marques, S.; Ramos, J. L., Transcriptional Control of the *Pseudomonas putida* TOL plasmid catabolic pathways. *Mol. Microbiol.* **1993**, *9* (5), 923-929.

Meunier, B., *Metal-Oxo and Metal-Peroxo Species in Catalytic Oxidations*.
 Springer-Verlag: Berlin, 2000.

Meunier, B.; Visser, S. I.; Shaik, S., Mechanism of Oxidation Reactions
 Catalyzed by Cytochrome P450 Enzymes. *Chem. Rev.* 2004, *104*, 3947-3980.

15. Che, C.-M.; Huang, J.-S., Metalloporphyrin-based oxidation systems: from bioimetic reactions to application in organic synthesis. *Chem. Commun.* **2009**, 3996-4015.

16. Meunier, B., Metalloporphyrins as Versatile Catalysts for Oxidation Reactions and Oxidative DNA Cleavage. *Chem. Rev.* **1992**, *92* (6), 1411-1456.

Rittle, J.; Green, M. T., Cytochrome P450 Compound I: Capture,
 Characterization, and C-H Bond Activation Kinetics. *Science* 2010, *330*, 933-937.

18. Kwong, K. W.; Chen, T. H.; Luo, W.; Jeddi, H.; Zhang, R., A biomimetic oxidation catalyzed by manganese(III) porphyrins and iodobenzene diacetate: Synthetic and mechanistic investigations. *Inorg. Chim. Acta* **2015**, *430*, 176-183.

19. Nam, W., High-Valent Iron(IV)-Oxo Complexes of Heme and Non-Heme Ligands in Oxygenation Reactions. *Acc. Chem. Res.* **2007**, *40*, 522-531.

59

20. Che, C.-M.; Yu, W.-Y., Ruthenium-oxo and -tosylimido porphyrin complexes for epoxidation and aziridination of alkenes. *Pure and Applied Chemistry* **1999**, *71* (2), 281-288.

Gross, Z.; Ini, S., Asymmetric Catalysis by a Chiral Ruthenium Porphyrin:
 Epoxidation, Hydroxylation, and Partial Kinetic Resolution of Hydrocarbons. *Org. Lett.* 1999, *1* (13), 2077-2080.

22. Groves, J. T., Reactivity and Mechanisms of Metalloporphyrin-Catalyzed Oxidations. *J. Porph. Phthal.* **2000**, *4*, 350-352.

23. Groves, J. T.; Haushalter, R. C.; Nakamura, M.; Nemo, T. E.; Evans, B. J., High-valent iron-porphyrin complexes related to peroxidase and cytochrome P-450. *J. Am. Chem. Soc.* **1981**, *103* (10), 2884-6.

24. Groves, J. T. K., William J., Jr.; Haushalter, Robert C., Hydrocarbon oxidations with oxometalloporphinates. Isolation and reactions of a (porphinato)manganese(V) complex. *J. Am. Chem. Soc.* **1980**, *102* (20), 6375-7.

25. Groves, J. T.; Stern, M. K., Olefin epoxidation by manganese (IV) porphyrins: evidence for two reaction pathways. *J. Am. Chem. Soc.* **1987**, *109* (12), 3812-14.

26. Leeladee , P.; Goldberg, D. P., Epoxidations Catalyzed by Manganese(V) Oxo and Imido Complexes: Role of the Oxidant–Mn–Oxo (Imido) Intermediate. *Inorg. Chem.* **2010**, *49*, 3083-3085.

27. Bougher, C. J.; Liu, S.; Hicks, S. D.; Abu-Omar, M. M., Valence Tautomerization of High-Valent Manganese(V)-Oxo Corrole Induced by Protonation of the Oxo Ligand. *J. Am. Chem. Soc.* **2015**, *137*, 14481–14487.

60

28. Leeladee, P.; Baglia, R. A.; Prokop, K. A.; Latifi, R.; de Visser, S. P.; Goldberg,
D. P., Valence Tautomerism in a High-Valent Manganese-Oxo Porphyrinoid Complex
Induced by a Lewis Acid. *J. Am. Chem. Soc.* 2012, *134* (25), 10397-10400.

29. Zaragoza, J. P.; Baglia, R. A.; Siegler, M. A.; Goldberg, D. P., Strong Inhibition of O-Atom Transfer Reactivity for Mn^{IV}(O)(pi-Radical-Cation)(Lewis Acid) versus Mn^V(O) Porphyrinoid Complexes. *J. Am. Chem. Soc.* **2015**, *137* (20), 6531-6540.

Baglia, R. A.; Prokop-Prigge, K. A.; Neu, H. M.; Siegler, M. A.; Goldberg, D.
P., Mn(V)(O) versus Cr(V)(O) Porphyrinoid Complexes: Structural Characterization and Implications for Basicity Controlling H-Atom Abstraction. *J. Am. Chem. Soc.* 2015, *137* (34), 10874-10877.

Groves, J. T.; Nemo, T. E.; Myers, R. S., Hydroxylation and Epoxidation
 Catalyzed by Iron-Porphine Complexes. Oxygen Transfer from Iodosylbenzene. *J. Am. Chem. Soc.* 1979, *101*, 1032-1033.

32. Nam, W.; Kim, I.; Lim, M. H.; Choi, H. J.; Lee, J. S.; Jang, H. G., Isolation of an oxomanganese(V) porphyrin intermediate in the reaction of a manganese(III) porphyrin complex and H2O2 in aqueous solution. *Chem. Eur. J.* **2002**, *8* (9), 2067-2071.

33. Collman, J. P.; Chien, A. S.; Eberspacher, T. A.; Brauman, J. I., Multiple Active Oxidants in Cytochrome P-450 Model Oxidations. *J. Am. Chem. Soc.* **2000**, *122* (45), 11098-11100.

34. In, J. H.; Park, S. E.; Song, R.; Nam, W., Iodobenzene diacetate as an efficient terminal oxidant in iron(III) porphyrin complex-catalyzed oxygenation reactions. *Inorg. Chim. Acta* **2003**, *343*, 373-376.

35. Kwong, K. W.; Lee, N. F.; Ranburger, D.; Malone, J.; Zhang, R., Visible Light-Induced Formation of Corrole-Manganese(V)-Oxo Complexes: Observation of Multiple Oxidation Pathways. *J. Inorg. Biochem.* **2016**, *163*, 39-44.

36. Suslick, K. S.; Watson, R. A., The photochemistry of chromium, manganese, and iron porphyrin complexes. *New J. Chem.* **1992**, *16* (5), 633-42.

37. Hoshino, M. A., S.; Yamaji, M.; Hama, Y., Laser photolysis studies of nitric oxide adducts of cobalt(II) porphyrins. Photoinduced denitrosylation at the temperature range 160-300 K. *J. Phys. Chem.* **1986**, *90*, 2109-2111.

38. Groves, J. T.; Quinn, R., Aerobic epoxidation of olefins with ruthenium porphyrin catalysts. *J. Am. Chem. Soc.* **1985**, *107* (20), 5790-2.

39. Hodgkin, D. C.; Pickworth, J.; Robertson, J. H.; Trueblood, K. N.; Prosen, R. J.;
White, J. G., Structure of Vitamin B₁₂: The Crystal Structure of the Hexacarboxylic
Acid Derived from B₁₂ and the Molecular Structure of the Vitamin. *Nature* 1955, *176* (325-328).

40. Visser, S. P.; Ogliaro, F.; Gross, Z.; Shaik, S., What is the Difference between the Manganese Porphyrin and Corrole Analogues of Cytochrome P450's Compound I? *Chem. Eur. J.* **2001**, *7*, 4954-4960.

41. Johnson, A. W.; Kay, I. T., Corroles. Part I. Synthesis. *J. Chem. Soc.* **1965**, *306*, 1620-1629.

42. Gross, Z.; Galili, N.; Simkhovich, L.; Saltsman, I.; Botoshansky, M.; Blaeser, D.;
Boese, R.; Goldberg, I., Solvent-Free Condensation of Pyrrole and
Pentafluorobenzaldehyde: A Novel Synthetic Pathway to Corrole and
Oligopyrromethenes. *Org. Lett.* **1999**, *1* (4), 599-602.

43. Agadjanian, H.; Weaver, J. J.; Mahammed, A.; Rentsendorj, A.; Bass, S.; Kim,
J.; Dmochowski, I. J.; Margalit, R.; Gray, H. B.; Gross, Z.; Medina-Kauwe, L. K.,
Specific Delivery of Corroles to Cells via Noncovalent Conjugates with Viral Proteins. *Pharm. Res.* 2006, *23*, 367-377.

44. Agadjanian, H.; Ma, J.; Rentsendorj, A.; Valluripalli, V.; Hwang, J. Y.;
Mahammed, A.; Farkas, D. L.; Gray, H. B.; Gross, Z.; Medina-Kauwe, L. K., Tumor
Detection and Elimination by a Targeted Gallium Corrole. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106* (15), 6105-6110.

45. Teo, R. D.; Gray, H. B.; Lim, P.; Termini, J.; Domeshek, E.; Gross, Z., A Cytotoxic and Cytostatic Gold(III) Corrole. *Chem. Commun.* **2014**, *50*, 13789-13792.

Hwang, J. Y.; Lubow, J.; Chu, D.; Ma, J.; Agadjanian, H.; Sims, J.; Gray, H. B.;
Gross, Z.; Farkas, D. L.; Medina-Kauwe, L. K., A Mechanistic Study of TumorTargeted Corrole Toxicity. *Mol. Pharm.* 2011, 8 (6), 2233-2243.

47. Lim, P.; Mahammed, A.; Okun, Z.; Saltsman, I.; Gross, Z.; Gray, H. B.; Termini,
J., Differential Cytostatic and Cytotoxic Action of Metallocorroles against Human
Cancer Cells: Potential Platforms for Anticancer Drug Development. *Chem. Res. Toxicol.* 2012, 25, 400-409.

48. Sims, J. D.; Hwang, J. Y.; Wagner, S.; Alonso-Valenteen, F.; Hanson, C.;
Taguiam, J. M.; Polo, R.; Harutyunyan, I.; Karapetyan, G.; Sorasaenee, K.; Ibrahim, A.;
Marban, E.; Moats, R.; Gray, H. B.; Gross, Z.; Medina-Kauwe, L. K., A Corrole
Nanobiologic Elicits Tissue-Activated MRI Contrast Enhancement and Tumor-Targeted
Toxicity. *J. Control. Release* 2015, *217*, 92-101.

49. Haber, A.; Gross, Z., Catalytic Antioxidant Therapy by Metallodrugs: Lessons from Metallocorroles. *Chem. Commun.* **2015**, *51* (27), 5812-5827.

50. Okun, Z.; Gross, Z., Fine Tuning the Reactivity of Corrole-Based Catalytic Antioxidants. *J. Inorg. Chem.* **2012**, *51* (15), 8083-8090.

51. Okun, Z.; Kupershmidt, L.; Amit, T.; Mandel, S.; Bar-Am, O.; Youdim, M. B.
H.; Gross, Z., Manganese Corroles Prevent Intracellular Nitration and Subsequent Death of Insulin-Producing Cells. *ACS Chem. Biol.* 2009, *4*, 910-914.

52. Gross, Z.; Golubkov, G.; Simkhovich, L., Epoxidation catalysis by a manganese corrole and isolation of an oxomanganese(v) corrole. *Angew. Chem. Int. Ed. Engl.* **2000**, *39* (22), 4045-4047.

53. Kumar, A.; Goldberg, I.; Botoshansky, M.; Buchman, Y.; Z., G., Oxygen Atom Transfer Reactions from Isolated (Oxo)manganese(V) Corroles to Sulfides. *J. Am. Chem. Soc.* **2010**, *132*, 15233-15245.

54. Caron, S.; Dugger, R. W.; Ruggeri, S. G.; Ragan, J. A.; Ripin, D. H. B., Large-Scale Oxidations in the Pharmaceutical Industry. *Chem. Rev.* **2006**, *106*, 2943-2989.

55. Baeckvall, J. E., *Modern oxidation methods*. Wiley-VCH Verlag: Weinheim,2004.

Wojaczynska, E.; Wojacznski, J., Enantioselective Synthesis of Sulfoxides:
 2000-2009. *Chem. Rev.* 2010, *110*, 4303-4356.

57. Hosseinpoor, F.; Golchubian, H., Mn(III)-Catalyzed Oxidation of Sulfides to Sulfoxides with Hydrogen Peroxide. *Tetrahedron Lett.* **2006**, *47* (29), 5195-5197.

58. Kaczorowska, K.; Kolarska, Z.; Mitka, K.; Kowalski, P., Oxidation of Sulfides to Sulfoxides. Part 2: Oxidation by Hydrogen Peroxide. *Tetrahedron* **2005**, *61* (35), 8315-8327.

59. Surendra, K.; Krishnaveni, N. S.; Kumar, V. P.; Sridhar, R.; Rao, K. R.,

Selective and Efficient Oxidation of Sulfides to Sulfoxides with *N*-Bromosuccinimide in the Presence of Cyclodextrinin Water. *Tetrahedron Lett.* **2005**, *46* (27), 4581-4583.

60. Gross, Z.; Gray, H. B., Oxidations catalyzed by metallocorroles. *Adv. Syn. Catal.*2004, *346* (2+3), 165-170.

61. Gross, Z.; Galili, N.; Saltsman, I., The First Direct Synthesis of Corroles from Pyrrole. *Angew. Chem. Int. Ed.* **1999**, *38*, 1427-1429.

62. Koszarna, B.; Gryko, D. T., Efficient synthesis of meso-subsitutted corroles in a H2O-MeOH mixture. *J. Org. Chem.* **2006**, *71*, 3707-3717.

63. Lindsey, J.; Wagner, R. D., Investigation of the synthesis of ortho-substituted tetraphenylporphyrins. *J. Org. Chem.* **1989**, *54*, 828-836.

64. Bard, A. J.; Faulkner, L. R., *Electrochemical Methods: Fundamentals and Applications*. 2nd Ed. ed.; Wiley: New York, 2001.

65. Zhang, R.; Harischandra, D. N.; Newcomb, M., Laser flash photolysis generation and kinetic studies of corrole-manganese(V)-oxo intermediates. *Chem. Eur. J.* 2005, *11* (19), 5713-5720.

66. Chen, T.-H.; Kwong, K.-W.; Carver, A.; Luo, W. L.; Zhang, R., Enhanced iron(III) corrole-catalyzed oxidations with iodobenzene diacetate: Synthetic and mechanistic investigations. *Appl. Catal. A* **2015**, *497*, 121-126.

CURRICULUM VITAE

Honors and Awards

American Institute of Chemists Outstanding Graduate Student Award	Spring 2017
WKU Analytical Chemistry Award	Spring 2016
Faculty–Undergraduate Student Engagement (FUSE) Grant	Fall 2015
C.P. McNally Undergraduate Scholarship Award	Fall 2015

Presentations

- Jeddi, H.; Zhang, R. (oral presentation, 1st place). "Kinetic and Catalytic Studies of Oxidations by an Electron-Deficient Manganese(V)-oxo Corrole." 102nd Annual Meeting of the Kentucky Academy of Science, 2016, Louisville, KY.
- Jeddi, H.; Zhang, R. (poster). "Kinetic and Mechanistic Studies of a Manganese(V)-oxo Corrole." University of Kentucky Undergraduate Research in Chemistry Regional Poster Competition, 2016, Lexington, KY.
- Jeddi, H.; Zhang, R. (oral presentation). "Kinetic Studies of Oxygen Atom Transfer Reactions by an Oxo-manganese(v) Corrole." 46th Annual WKU Student Research Conference, 2016, Bowling Green, KY.
- Jeddi, H.; Luo, W.; Zhang, R. (poster). "Kinetic and Competition studies of Oxygen Atom Transfer Reactions with a Corrole-manganese(V)-oxo Species." 251st Annual National Meeting of the American Chemical Society, Spring 2016, San Diego, CA.
- **Jeddi, H.**; Luo, W.; Zhang, R. (poster, 1st place). "Kinetic and Catalytic Studies of a Corrole-manganese Species." 101st Annual Meeting of the Kentucky Academy of Science, 2015, Highland Heights, KY.
- Jeddi, H.; Luo, W.; Zhang, R. (poster). "Synthesis and Kinetic Studies of a Manganese(V)-oxo Corrole." 67th Annual Southeastern Regional Meeting of the American Chemical Society, 2015, Memphis, TN.

Publications

- Lee, N.F.; Malone, J.; **Jeddi, H.**; Kwong, K.W.; Zhang, R. Visible-Light Photolysis of Corrole-Manganese(IV) Nitrates to Generate Corrole-Manganese(V)-Oxo Complexes. *Inorg. Chem. Commun.* In press.
- Kwong, K. W.; Chen, T.-H.; Luo, W.; Jeddi, H.; Zhang, R. A Biomimetic Oxidation Catalyzed by Manganese(III) Porphyrins and Iodobenzene Diacetate. *Inorg. Chim. Acta.* 2015, *430*, 176–183.

Ar	Aryl
Al ₂ O ₃	Alumina
$BF_3 \cdot OEt_2$	Boron trifluoride diethyl etherate
CYP450s	Cytochrome P450s
DMF	N,N-Dimethylformamide
DDQ	2,3-Dichloro-5,6-dicyanobenzoquinone
FID	Flame ionization detector
GC	Gas chromatography
H ₂ TMP	meso-Tetramesitylporphyrin
H ₃ TPC	Triphenylcorrole
H ₃ TPFC	Tris(pentafluorophenyl)corrole
H_2O_2	Hydrogen peroxide
k _{obs}	Observed pseudo-first-order rate constant
k_2	Second order rate constant
LFP	Laser flash photolysis
Mn ^{III} (TMP)Cl	Tetramesitylporphyrinato manganese(III) chloride
Mn ^{III} (TPC)	Triphenylcorrolato manganese(III)
Mn ^{III} (TPFC)	Tris(pentafluorophenyl)corrolato manganese(III)
MS	Mass spectrometry
<i>m</i> -CPBA	meta-Chloroperoxybenzoic acid
NADH	Nicotinamide adenine dinucleotide
NMR	Nuclear magnetic resonance

PhIO	Iodosylbenzene
PhI(OAc) ₂	Iodobenzene diacetate
ppm	Parts per million
Sub	Substrate
TBHP	tert-Butylhydroperoxide
TON	Turnover number
TMS	Tetramethylsilane
UV-vis	UV-visible