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# **Research Paper**

# p63 expression confers significantly better survival outcomes in highrisk diffuse large B-cell lymphoma and demonstrates p53-like and p53-independent tumor suppressor function

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Abstract: The role of p53 family member, p63 in oncogenesis is the subject of controversy. Limited research has been done on the clinical implications of p63 expression in diffuse large B-cell lymphoma (DLBCL). In this study, we assessed p63 expression in *de novo* DLBCL samples (n=795) by immunohistochemistry with a pan-p63-monoclonal antibody and correlated it with other clinicopathologic factors and clinical outcomes. p63 expression was observed in 42.5% of DLBCL, did not correlate with p53 levels, but correlated with p21, MDM2, p16<sup>INK4A</sup>, Ki-67, Bcl-6, IRF4/MUM-1 and CD30 expression, *REL* gains, and *BCL6* translocation. p63 was an independent favorable prognostic factor in DLBCL, which was most significant in patients with International Prognostic Index (IPI) >2, and in activated-B-cell—like DLBCL patients with widetype *TP53*. The prognostic impact in germinal-center-B-cell—like DLBCL was not apparent, which was likely due to the association of p63 expression with high-risk IPI, and potential presence of  $\Delta$ Np63 isoform in *TP63* rearranged patients (a mere speculation). Gene expression profiling suggested that p63 has both overlapping and distinct functions compared with p53, and that p63 and mutated p53 antagonize each other. In summary, p63 has p53-like and p53-independent functions and favorable prognostic impact, however this protective effect can be abolished by *TP53* mutations.

# **INTRODUCTION**

TP63, a member of the TP53 gene family, encodes p63 with 2 types of isoforms: a form with the N-terminal transactivation (TA) domain (TAp63) and a truncated form without the N-terminus ( $\Delta Np63$ ). Both TAp63 and  $\Delta Np63$  have isoforms  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ , and  $\varepsilon$  owing to alternative splicing at the 3' end [1-5]. p63 shares structural and sequence homology with p53 and p73, the third member of the p53 family [1, 6]. Like p53, TAp63 has been implicated in cell cycle arrest and apoptosis in response to DNA damage, ectoderm development, maternal reproduction and metabolism, dependent or independent of p53-functions [1, 7-13]. For example, TAp63 can transactivate some wellknown p53 target genes including CDKNIA, BAX and MDM2 [1, 14]. Moreover, p53-dependent apoptosis in response to DNA damage required p63 and p73 in mouse developing brain and embryonic fibroblasts [7]. However, in a mouse model p63 and p73 did not contribute to p53 tumor suppression function in lymphoma development [15].  $\Delta Np63$ , on the other hand, interacts with p53, TAp63, and TAp73 in a dominant-negative fashion to inhibit their tumorsuppressive functions [3]. It is generally believed that TAp63, like p53, is a tumor suppressor, whereas  $\Delta Np63$ has a critical role in epidermal development and functions as an oncogene in a mouse model [16-19]. Furthermore, the  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ , and  $\varepsilon$  isoforms of TAp63 and  $\Delta Np63$  have differential functions [5, 14, 20-24].

In normal human tissues, p63 expression is tissuespecific and restricted to epithelial cells, certain subpopulations of basal cells, and occasionally cells in the germinal centers of lymph nodes [1, 25, 26]. Accordingly, in tumors structural disruption of *TP63* and aberrant p63 expression are commonly seen in squamous cell and transitional cell carcinomas, but are also observed in non-Hodgkin lymphomas, predominantly in In basal epithelial cells and squamous cell carcinomas, isoform, especially the  $\Delta Np63$  $\Delta Np63\alpha$ . is predominantly expressed, possibly due to the increased  $\Delta$ Np63 stability caused by the lack of the transactivation domain which is indispensable for proteasomedependent MDM2-independent degradation of p63 [24, 31]. In contrast, TAp63 is present mostly in epithelial lining cells at lower levels under normal physiological conditions, and in adenocarcinoma, thymoma and lymphoma cells; TAp63 accumulates in response to genotoxic stress [24, 26]. Although p63 expression has been shown in a few studies to indicate a poor prognosis in some carcinomas [32-34], its prognostic significance in DLBCL is unclear.

DLBCL is the most common type of non-Hodgkin lymphoma and can be divided into germinal center Bcell-like (GCB) and activated B-cell-like (ABC) subgroups by gene expression profiling [35]. Numerous genetic factors affecting the prognosis of DLBCL have been identified [36]. In our previous study, TP53 mutations were detected in approximately 20% of de novo DLBCL cases and conferred a worse prognosis among DLBCL patients treated with rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisolone (R-CHOP) [37]. Overexpression of mutated but not wild-type p53 (WT-p53) protein is also associated with a poor prognosis in DLBCL patients [38]. The dysregulation, expression, and clinical implications of p63 in DLBCL are less clear than those of p53; likewise, p63's role in tumorigenesis and its functional relationship with p53 are not well understood. p63, predominantly TAp63 (likely TAp63) and/or TAp63 $\gamma$ ) but not  $\Delta$ Np63 or p63 $\alpha$ , was found expressed in 15.1% to 52.5% of DLBCLs at higher levels than in normal lymphoid tissues [21, 25-27, 39]. Truncated p63 homologous to  $\Delta Np63$  due to TP63 gene rearrangements was also reported in 1.2%-5% of DLBCL, exclusive of GCB subtype [40, 41].

Conflicting results showing the effect of p63 expression on patients' prognosis have been reported [21, 27, 39, 42, 43], likely owing to small number of patients (fewer than 100) in each study, the use of different cutoffs for p63 positivity, the differential functions and complicated interactions of multiple p63 isoforms [23, 43].

To fill this knowledge gap, we studied the prognostic effects of p63 expression correlating with *TP53* status in a multicenter cohort of patients with well-characterized *de novo* DLBCL treated with R-CHOP. We found that p63 expression conferred better clinical outcomes in DLBCL which however could be compromised or abolished by the difference in International Prognostic Index (IPI) scores and/or the presence of *TP53* mutations. We further investigated p63-associated biology to understand possible underlying molecular mechanisms.

### RESULTS

### p63 expression in DLBCL

We observed nuclear expression of p63 at variable levels in tumor cells of 317 (61%) of 520 samples from patients in the training set and 180 (65%) of 275 samples from patients in the validation set. Representative immunohistochemical stains are shown in Fig 1A, B and the histograms of p63 expression by immunohistochemistry are shown in Fig 1C, D. The mean number of p63 positive tumor cells in the training set was 18%, which was significantly higher than that of WT-p53 (P=0.017) but significantly lower than that of mutated p53 (MUT-p53, P<0.0001, Fig 1E) (Supplemental Fig 1A, B) [37, 38], although the *TP63* mRNA levels were significantly lower than the *TP53* 



**Figure 1. p63 expression in diffuse large B-cell lymphoma (DLBCL) in comparison with p53 expression. (A-B)** Representative immunohistochemistry staining for p63 (10% and 95%) in DLBCL. **(C-D)** Histograms of p63 expression in the training and validation sets. **(E)** Comparisons between p63 and p53 expression levels in DLBCL. **(F)** Comparisons between *TP63* and *TP53* mRNA levels in DLBCL. **(G)** p63<sup>+</sup> DLBCL had significantly higher levels of *TP63* mRNA compared with p63<sup>-</sup> DLBCL. **(H)** p63<sup>+</sup> DLBCL had significantly higher *TP53* mRNA levels compared with p63<sup>-</sup> DLBCL. **(I)** Expression of p63 protein correlated with *TP63* mRNA levels. The *TP63* mRNA expression levels (Log2 values) were retrieved from the gene expression profiling data. The mean values of 3 probesets (1555581\_a\_at, 207382\_at, 209863\_s\_at) for each patient were used. The relative mRNA level refers to the difference between the *TP63* mRNA level for each patient and the mean *TP63* mRNA level for the entire cohort. **(J-K)** Comparisons of p63 protein and *TP63* mRNA expression levels between germinal center B-cell–like (GCB) and activated B-cell–like (ABC) subtypes of DLBCL patients. mRNA levels (*P*<0.0001, Fig 1F). p63 protein expression significantly correlated with *TP63* mRNA (Spearman rank correlation: r=0.596, *P*<0.0001).

Owing to the significantly lower level of p63 compared with MUT-p53 expression in DLBCL and the exclusion of potential false-positive cases, we used a cutoff value of 5% of tumor cells being p63-positive for p63 expression in DLBCL ( $p63^+$ : >5%). Using this cutoff, 221 patients (42.5%) in the training set, and 130 (47%) of 275 patients in the validation set had  $p63^+$  DLBCL. The  $p63^+$  group showed a significantly higher mean TP63 mRNA level compared with the p63<sup>-</sup> group (unpaired t test, P<0.0001, Fig 1G) and TP53 mRNA level (Fig 1H). Transcriptional activation appeared to be the most common mechanism for p63 expression in this study of DLBCL (Fig 1I). No significant difference in the expression levels of p63/TP63 was observed between the GCB and ABC subtypes of tumor samples, either at the protein (16.66% vs. 19.26%, P=0.21) or mRNA (P=0.90) levels (Fig 1J, K).

### Clinical and pathobiological features of p63<sup>+</sup> DLBCL

We compared the clinicopathologic features of patients with  $p63^+$  and  $p63^-$  DLBCL. The  $p63^+$  group more often had male (P=0.0056) and patients with small (< 5 cm) tumors (P=0.05) than did the p63<sup>-</sup> group. In addition, a higher proportion (41.9%) of p63<sup>+</sup> patients had an IPI score >2 compared with p63<sup>-</sup> patients (34.4%), but this difference was not significant (P=0.086); however, by unpaired t test, patients with IPI scores >2 showed significantly higher mean levels of p63 (P=0.05, Fig. 2A) and MUT-p53 (P=0.011, figure not shown) than did patients with IPI scores  $\leq 2$ . When DLBCL cases were stratified into the GCB and ABC subtypes, in GCB-DLBCL  $p63^+$  compared with  $p63^-$  patients was associated with IPI scores >2, small tumors, and possibly stage III/IV disease (P=0.06), whereas in ABC-DLBCL p63<sup>+</sup> patients had higher percentages of male gender and extranodal DLBCL (44% compared with the 31% in p63<sup>-</sup> ABC-DLBCL) (Table 1). In contrast, WT-p53 overexpression was more common in nodal DLBCL (data not shown).



**Figure 2. Correlations between p63 expression and other tumor associated factors.** (A) The group with high International Prognostic Index (IPI) scores had a significantly higher mean level of p63 expression. (B-D) p63 expression was associated with significantly higher levels of p21, MDM2, and p16-INK4a in both germinal center B-cell like (GCB) and activated B-cell like (ABC) subtypes of DLBCL patients. (E-F) p63 expression was associated with significantly higher levels Ki-67 and Bcl-6 in ABC-DLBCL. (G) p63 expression was associated with significantly higher levels of IRF4/MUM-1 in both GCB and ABC subtypes of DLBCL patients. (H) p63<sup>+</sup> ABC-DLBCL was associated with a trend toward higher c-Rel level. (I-J) The association of p63 expression with p21 and MDM2 is independent of p53 mutation status. (K-L) p53 mutation status did not impact the association with increased p16-INK4a, Bcl-6 and IRF4/MUM-1(figure not shown) levels.

### Table 1. Clinical characteristics of patients with *de novo* DLBCL in the training cohort

	<b>D</b> p63 <sup>+</sup>	pLBCL p63		<b>GCB-I</b> p63 <sup>+</sup>	p63 <sup>-</sup>		<b>ABC-D</b> p63 <sup>+</sup>	LBCL p63 <sup>-</sup>		
	N (%)	N (%)	Pl	N (%)	N (%)	P2	N (%)	N (%)	P3	P4
Patients	221	299		110	156		110	141	.59	
Age (yr) < 60 ≥ 60	92 (42) 129 (58)	133 (45) 166 (55)	.52	52 (47) 58 (53)	84 (54) 72 (46)	.29	39 (35) 71 (65)	47 (33) 94 (67)	.73	.075
Sex Female Male	76 (34) 145 (66)	139 (46) 160 (54)	.0056	41 (37) 69 (63)	68 (44) 88 (56)	.30	34 (31) 76 (69)	70 (50) 71 (50)	.0028	.32
Stage I-II III-IV	94 (44) 119 (56)	141(49) 148 (51)	.30	50 (48) 55 (52)	89 (59) 61 (41)	.06	44 (41) 63 (59)	51 (37) 86 (63)	.54	.34
B symptoms No Yes	136 (63) 79 (37)	183 (65) 97 (35)	.63	73 (67) 35 (32)	101 (70) 43 (30)	.67	62 (58) 44 (42)	81 (60) 53 (40)	.76	.17
<b>Serum LDH</b> Normal Elevated	79 (39) 124 (61)	107 (39) 169 (61)	.97	42 (42) 58 (58)	56 (39) 86 (61)	.70	37 (36) 65 (64)	51 (39) 81 (61)	.71	.40
No. of extranodal sites $0-1 \ge 2$	156 (75) 53 (25)	228 (79) 61 (21)	.27	79 (78) 22 (22)	121 (81) 28 (19)	.56	76 (71) 31 (29)	106 (77) 32 (23)	.30	.23
Performance status $0-1 \ge 2$	164 (85) 30 (15)	225 (83) 46 (17)	.66	78 (85) 14 (15)	119 (86) 19 (14)	.76	85 (84) 16 (16)	104 (79) 27 (21)	.35	.90
Size of largest tumor < 5cm ≥ 5cm	106 (65) 58 (35)	120 (55) 99 (45)	.05	56 (70) 24 (30)	62 (55) 50 (45)	.04	49 (59) 34 (41)	58 (54) 49 (46)	.51	.14
0-2 3-5	125 (58) 90 (42)	189 (66) 99 (34)	.086	65 (61) 41 (39)	111 (75) 38 (25)	.025	59 (55) 49 (45)	76 (56) 61 (44)	.90	.32
Therapy response CR PR SD PD	178 (81) 24 8 11	227 (76) 43 13 16	.21	87 (79) 13 6 4	118 (76) 19 7 12	.51	90 (82) 11 2 7	104 (74) 24 6 7	.13	.61
<b>Primary origin</b> Nodal Extranodal	131 (60) 88 (40)	193 (66) 99 (34)	.16	69 (64) 39 (36)	97 (64) 55 (36)	1.0	62 (56) 48 (44)	95 (69) 43 (31)	.048	.27
Ki-67 ≤ 70%	66 (30) 155 (70)	119 (40) 175 (60)	.0016	41 (37) 69 (63)	64 (42) 88 (58)	.45	24 (22) 86 (78)	55 (39) 86 (61)	.004	.018
TP53 mutations WT-TP53 MUT-TP53	154 (80) 40 (21)	206 (77) 60 (23)	.65	70 (74) 25 (26)	105 (76) 34 (24)	.76	83 (85) 15 (15)	100 (79) 26 (21)	.38	.059
MYC translocation No Yes	138 (89) 17 (11)	158 (88) 22 (12)	.86	62 (89) 8 (11)	73 (80) 18 (20)	.20	75 (89) 9 (11)	85 (95) 4 (5)	.15	1
BCL2 translocation No Yes	159 (84) 30 (16)	187 (81) 44 (19)	.44	68 (74) 24 (26)	74 (64) 41 (36)	.18	90 (94) 6 (6)	113 (97) 3 (3)	.31	.0002
BCL6 translocation No Yes	98 (60) 66 (40)	145 (74) 51 (26)	.0041	54 (69) 24 (31)	83 (78) 23 (22)	.16	43 (51) 42 (49)	61 (69) 28 (31)	.0016	.016
<b>REL gains</b> Normal Amplification/ polysomy	140 (86) 23 (14)	216 (92) 19 (8)	.068	62 (77) 18 (23)	118 (92) 10 (8)	.003	77 (94) 5 (6)	98 (92) 9 (8)	.036	.0001
REL amplification No Yes	156 (95) 8 (5)	227 (97) 8 (3)	.60	72 (91) 7 (9)	121 (95) 7 (5)	.40	83 (99) 1 (1)	106 (99) 1 (1)	1.0	.03
CD30 expression - +	175 (79) 46 (21)	259 (88) 34 (12)	.0048	86 (78) 24 (22)	133 (88) 19 (12)	.06	88 (80) 22 (20)	125 (89) 15 (11)	.049	.87
p53 expression < 20% ≥ 20%	116 (61) 74 (39)	172 (66) 87 (34)	.27	57 (61) 37 (39)	89 (66) 45 (34)	.40	59 (62) 37 (38)	83 (66) 42 (34)	.48	1.0

**Abbreviations**: DLBCL, diffuse large B-cell lymphoma; GCB, germinal center B-cell–like; ABC, activated B-cell–like; LDH, lactate dehydrogenase; IPI, International Prognostic Index; CR, complete remission; PR, partial response; SD, stable disease; PD, progressive disease.

**Note**: *P* values indicate the significance of differences in the positivity frequencies of listed parameters between 2 groups. *P1* values are for comparisons between overall  $p63^+$  and  $p63^-$  DLBCL patients; *P2* values are for comparisons between  $p63^+$  and  $p63^-$  GCB-DLBCL patients; *P3* values are for comparisons between  $p63^+$  and  $p63^-$  ABC-DLBCL patients; *P4* values are for comparisons between  $p63^+$  GCB-DLBCL and  $p63^+$  ABC-DLBCL patients. For therapy response, we calculated *P* values by comparing CR to other responses.

When correlating p63 expression with other genetic abnormalities and immunohistochemical biomarkers in DLBCL, we found that the p63<sup>+</sup> group had higher frequencies of *BCL6* translocation and CD30 positivity (21% compared with the 12% in p63<sup>-</sup> patients) (Table 1), as well as elevated expression levels of Bcl-6, IRF4/MUM-1, p21, MDM2, p16-INK4a, and Ki-67 (in ABC-DLBCL only); most of these associations were independent of *TP53* mutation status (Fig 2B-L). In addition, p63 expression was associated with *REL* gains (including amplification and polysomies) in both the GCB and ABC subsets. No significant differences in frequencies of *TP53* mutations, *MYC* or *BCL2* translocations, or the expression levels of p53, Myc, or Bcl-2, were observed between the p63<sup>+</sup> and p63<sup>-</sup> groups.

### p63 expression confers better clinical outcomes, more apparently in high-risk DLBCL and ABC-DLBCL

### Univariate survival analysis in the training set

With a median follow-up of 62 months, p63<sup>+</sup> DLBCL patients showed better progression-free survival (PFS, P=0.05) compared with p63<sup>-</sup> DLBCL patients (Fig 3a, b). When patients with low-risk (IPI score  $\leq 2$ ) and high-risk DLBCL (IPI score  $\geq 2$ ) were analyzed separately (Fig 3c, d), p63 expression showed prognostic significance only in the high-risk group and correlated with significantly better overall survival (OS) (P=0.006) and PFS (P=0.0043).



**Figure 3. Prognostic analysis of p63 expression in DLBCL.** (a-b) p63 expression correlated with significantly better progression-free survival (PFS) but not overall survival (OS) in DLBCL. (c-d) p63 expression correlated with significantly better overall survival in DLBCL patients with IPI scores > 2 but not in DLBCL patients with IPI scores  $\leq 2$ . (e-h) p63 expression correlated with significantly better PFS in ABC- but not GCB-DLBCL patients. (i-j) p63 expression was associated with trends towards better survival outcomes in GCB-DLBCL patients with IPI scores > 2. (k-l) p63 expression correlated with significantly better > 2.

		OS			PFS		
Variable	HR	95% CI	Р	HR	95% CI	Р	
Overall DI BCI							
IPI >2	3.08	2 21-4 38	< 0001	2 84	2 08-3 89	< 0001	
$n63^{+}$	62	45- 87	.006	66	48- 90	.009	
Female sex	.86	.62-1.20	.37	.92	.67-1.26	.60	
Tumor size ≥5 cm	1.30	.94-1.79	.11	1.26	.93-1.70	.14	
B symptoms present	1.32	.95-1.85	.10	1.24	.90-1.71	.18	
GCB-DLBCL							
IPI >2	4.00	2.36-6.79	<.0001	3.44	2.27-5.21	<.0001	
p63 <sup>+</sup>	.64	.4199	.045	.67	.42-1.09	.11	
Female sex	.94	.61-1.45	.78	1.00	.67-1.50	.99	
Tumor size ≥5 cm	1.53	.92-2.54	.10	1.46	.92-2.34	.11	
B symptoms present	1.08	.69-1.70	.74	1.21	.74-1.98	.44	
ABC-DLBCL							
IPI >2	2.35	1.61-3.43	<.0001	2.23	1.57-3.16	<.0001	
p63+	.56	.3883	.004	.58	.4083	.003	
Female sex	.77	.52-1.15	.20	.78	.54-1.12	.17	
Tumor size ≥5 cm	1.03	.58-1.56	.88	.99	.66-1.47	.94	
B symptoms present	1.06	.72-1.58	.76	1.14	.79-1.64	.49	
DLBCL with WT-TP53							
IPI >2	3.29	2.21-4.88	<.0001	3.21	2.18-4.72	<.0001	
p63 <sup>+</sup>	.61	.4091	.015	.63	.4392	.016	
p53 <sup>+</sup>	.97	.62-1.52	.90	.91	.60-1.40	.68	
Female sex	.91	.61-1.36	.65	.85	.57-1.26	.42	
Tumor size ≥5 cm	1.19	.81-176	.38	1.11	.76-1.62	.59	
B symptoms present	1.45	.97-2.17	.07	1.48	1.00-2.20	.049	
DLBCL with MUT-TP53							
IPI >2	2.43	1.17-5.05	.017	2.11	1.07-4.18	.032	
p63 <sup>+</sup>	.70	.34-1.44	.33	.72	.36-1.44	.36	
p53 <sup>+</sup>	3.16	1.17-8.52	.023	2.30	.97-5.45	.06	
Female sex	1.02	.50-2.11	.96	1.12	.57-2.20	.75	
Tumor size ≥5 cm	1.57	.77-3.20	.21	1.85	.95-3.63	.07	
B symptoms present	1.19	.54-2.60	.67	1.03	.49-2.17	.93	

### Table 2. Multivariate survival analysis

**Abbreviations**: DLBCL, diffuse large B-cell lymphoma; ABC, activated B-cell–like; OS, overall survival; PFS, progression-free survival; HR, hazard ratio; CI, confidence interval; IPI, International Prognostic Index.

When analyzed in GCB- and ABC-DLBCL subsets separately, patients with  $p63^+$  ABC-DLBCL showed significantly better PFS (*P*=0.0097) and a trend of better OS (*P*=0.08) compared with  $p63^-$  ABC-DLBCL patients. In contrast, in GCB-DLBCL, p63 expression did not show significant impact on OS or PFS (Fig 3e-h). Consistently, survival analysis based on *TP63* mRNA levels showed that *TP63* mRNA expression correlated with favorable OS and significantly better PFS in ABC-DLBCL patients only (*P*=0.06 and *P*=0.036 respectively, Supplemental Fig S1D-G).

Since the p63<sup>+</sup> GCB-DLBCL group had a higher portion of patients with high-risk DLBCL (Table 1) which may have confounded the analysis [44], patients with low-risk and high-risk DLBCL were analyzed separately. In GCB-DLBCL patients with IPI scores >2, p63<sup>+</sup> GCB-DLBCL patients showed trends of better OS and PFS (*P*=0.078 and *P*=0.057 respectively) (Fig 3i-j). Similarly, the prognostic impact of p63 expression in ABC-DLBCL patients was more apparent in those with IPI scores >2 (Fig 3k-l); For ABC-DLBCL patients with IPI scores  $\leq$  2, OS and PFS rates were higher for p63<sup>+</sup> patients but not the differences were not significant (P=0.48 and P=0.12 respectively).

### Multivariate survival analysis

We performed multivariate survival analysis for p63 expression adjustin g clinic al parameters including IPI score, sex, tumor size and B symptoms. p63 expression was found to be an independent prognostic factor for better OS in the overall DLBCL, GCB-DLBCL and ABC-DLBCL sets, and an independent prognostic factor for better PFS in the overall DLBCL and ABC-DLBCL sets but not in the GCB-DLBCL set (Table 2).

### Validation set

Similar to the training set, no significant difference was observed in p63 expression between the GCB and ABC subtypes (P=0.68). These similar prognostic impacts as in the training set were all significant with a  $\geq$ 5% cutoff value for p63 expression (P=0.02, P=0.047, and P=0.0007 for PFS in DLBCL, ABC-DLBCL and highrisk DLBCL respectively. Supplemental Fig S2). A multivariate survival analysis indicated that after adjusting clinical parameters, p63 expression  $\geq$ 5% was an independent favorable prognostic factor in overall DLBCL and ABC-DLBCL but not in GCB-DLBCL (data not shown).



**Figure 4. Relationship between** *TP63*/p63 and *TP53*/p53 expression in DLBCL. (A) Comparison of wild-type (WT) or mutated (MUT) p53 protein expression between p63<sup>+</sup> and p63<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL. (B) Comparison of *TP53* mRNA expression between p63<sup>+</sup> and p63<sup>-</sup> patients with *WT-TP53* or *MUT-TP53* and GCB/ABC DLBCL. (C) Comparison of p63 protein expression between *WT-TP53* and *MUT-TP53* DLBCL, and between p53<sup>+</sup> and p53<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL. (D) Comparison of *TP63* mRNA expression between *WT-TP53* and *MUT-TP53* DLBCL, and between p53<sup>+</sup> and p53<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL. (D) Comparison of *TP63* mRNA expression between *WT-TP53* and *MUT-TP53* DLBCL, and between p53<sup>+</sup> and p53<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL. (D) Comparison of *TP63* mRNA expression between *WT-TP53* and *MUT-TP53* DLBCL, and between p53<sup>+</sup> and p53<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL. (D) Comparison of *TP63* mRNA expression between *WT-TP53* and *MUT-TP53* DLBCL, and between p53<sup>+</sup> and p53<sup>-</sup> patients with DLBCL or GCB/ABC subtypes of DLBCL.

Relationships with *TP53* mutations and p53 expression

# Non-significant correlation with p53 expression and correlation with TP53 mRNA

By Spearman rank correlation *TP63* mRNA showed correlation with *TP53* mRNA levels in the overall DLBCL set (r=0.091, *P*=0.048) and *WT-TP53* subset (r=0.106, *P*=0.044) but not in the *MUT-TP53* subset. In contrast, p63 expression did not show significant correlation with overall p53 (r=0.071, *P*=0.132), WTp53 (r=0.08, *P*=0.135), or MUT-p53 (r=0.072, *P*=0.481). Using unpaired *t*-tests, p63 expression did not correlate with p53 levels (Supplemental Fig S1C), but was associated with elevated *TP53* mRNA levels (Fig 1H). Analysis in GCB/ABC DLBCL subsets with WT- p53 or MUT-p53 showed no significant correlations between p63 positivity and WT-p53/MUT-p53 expression levels (Table 1, Fig 4A, B). However, the WT-p53<sup>+</sup> ( $\geq$ 20% [38]) compared with the WT-p53<sup>-</sup> DLBCL group had a significantly higher mean level of p63 protein (Fig 4C(b)) but not *TP63* mRNA (Fig 4D).

### *Prognostic impact of p63 expression in the presence of WT-TP53 or MUT-TP53*

The clinicopathologic features of patients with  $p63^+$  or  $p63^-$  DLBCL with *WT-TP53* or *MUT-TP53* are shown in Table 3. p63 expression was associated with significantly better OS and PFS in patients with *WT-TP53* and IPI scores >2 (Fig 5A, B) and in ABC-DLBCL patients with *WT-TP53* (Fig 5C, D), and favorable trends in patients with MUT-p53<sup>+</sup> GCB-DLBCL (Fig 5G, H).



**Figure 5. Prognostic analysis for p63 expression in DLBCL patients with wild- type and mutated TP53.** (A-B) p63 expression correlated with significantly better survival outcomes in patients with high-risk (IPI > 2) DLBCL and *WT-TP53.* (C-D) p63 expression correlated with significantly better survival outcomes in patients with ABC-DLBCL and *WT-TP53.* (E) In patients with high-risk (IPI > 2) DLBCL and *MUT-TP53,* p63 expression did not correlate with survival outcomes although showed a trend toward better PFS. (F) p63 expression did not correlate with survival outcomes in patients with ABC-DLBCL and MUT-p53 overexpression. (G-H) p63 expression was associated with trends toward better survival outcomes in GCB-DLBCL patients with MUT-p53 overexpression (marginal *P* values).

		WT-TP53		MUT-T	P53	•
	p63 <sup>+</sup>	p63 <sup>-</sup>		p63 <sup>+</sup>	p63 <sup>-</sup>	
Characteristic	N (%)	N (%)	Р	N (%)	N (%)	Р
Patients	154	206		40	60	
Age (y)						
< 60	62 (40)	87 (42)	.75	16 (40)	25 (42)	1.0
$\geq 60$	92 (60)	119 (58)		24 (60)	35 (58)	
Gender						
Male	106 (69)	107 (52)	.0013	24 (60)	36 (60)	1.0
Female	48 (31)	99 (48)		16 (40)	24 (40)	
Stage	(2) $(12)$	07 (40)	21	17 (42)	29 (47)	(0
	62 (42) 84 (59)	97 (49)	.21	17 (43)	28 (47)	.68
III-1 V R symptoms	84 (38)	100 (31)		25 (38)	52 (55)	
No	100 (67)	196 (66)	92	23 (59)	38 (68)	37
Yes	50 (33)	65 (34)	.92	16 (41)	18 (32)	.57
LDH	00(00)			10(11)	10 (02)	
Normal	58 (41)	82 (44)	.66	12 (32)	19 (33)	.93
Elevated	82 (59)	105 (56)		25 (68)	38 (67)	
No. of extranodal sites	( )			× /	~ /	
0-1	105 (73)	155 (78)	.30	29 (74)	46 (78)	.68
$\geq 2$	38 (27)	43 (22)		10 (26)	13 (22)	
Performance status						
0-1	117 (87)	231 (85)	.62	30 (88)	50 (86)	.78
$\geq 2$	18 (13)	28 (15)		4 (12)	8 (14)	
Size of largest tumor						
< 5cm	81 (68)	90 (58)	.079	18 (58)	23 (48)	.38
$\geq$ 5 cm	38 (32)	66 (42)		13 (42)	25 (52)	
IPI risk group						
0-2	87 (58)	132 (67)	.10	19 (49)	38 (64)	.12
3-5	62 (42)	65 (33)		20 (51)	21 (36)	
Therapy response						
CR	126 (82)	163 (79)	.59	27 (68)	35 (58)	.40
PR	16	24		7	13	
SD	3	12		3	3	
PD V: 67	9	12		3	9	
KI-07	50 (22)	02 (41)	10	0 (22)	17 (20)	()
< /0%	50 (33)	83 (41)	.10	9 (22)	17 (29)	.64
$\geq 10\%$	104 (67)	119 (59)		31 (78)	42 (71)	
Primary origin						
Nodal	91 (40)	134 (66)	.21	25 (37)	39 (70)	.54
Extranodal	61 (60)	68 (66)		15 (63)	17 (30)	
DLBCL subtypes	70 (40)	105 (51)	24	25 ((2))	24 (57)	(0
GCB	/0 (46) 83 (54)	105 (51)	.34	25 (62)	34 (57)	.68
ABC	83 (34)	100 (49)		13 (38)	20 (43)	
BCL6 translocation	73 (63)	07 (70)	21	17(57)	35 (85)	007
+	43 (37)	41 (30)	.21	13(43)	6 (15)	.007
CD20	45 (57)	41 (50)		15(45)	0(15)	
- -	120 (78)	178 (89)	.0085	32 (80)	54 (92)	.13
+	34 (22)	23 (11)		8 (20)	5 (8.5)	
p53 expression	( -)			- ( - )		
< 20%	106 (70)	154 (77)	.18	10 (26)	18 (31)	.65
$\geq$ 20%	45 (30)	47 (23)		29 (74)	40 (69)	

Table 3.	Clinical characteristics of	DLBCL patients with	wild-type TP53 (WT	-TP53) or mutated	TP53 (MUT-TP53)
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**Abbreviations**: DLBCL, diffuse large B-cell lymphoma; R-CHOP, rituximab with cyclophosphamide, doxorubicin, vincristine, and prednisone; LDH, lactate dehydrogenase; IPI, international prognostic index; CR, complete remission; PR, partial response; SD, stable disease; PD, progressive disease; GCB, germinal center B-cell–like; ABC, activated B-cell–like.

	p63 <sup>+</sup> DLBCL FDR< 0.20		p63 <sup>+</sup> DLBCL FDR< 0.05, fo	with <i>MUT-TP53</i> old change > 1.68	p63 <sup>+</sup> ABC-DLBCL with <i>WT- TP53</i> FDR< 0.20	
Function categories	Upregulated	Downregulated	Upregula ted	Downregulated	Upregulated	Downregul ated
Signaling, immune response, inflammation	FLJ23834, TRAF1	SGPP1			FOXD1, PDE7A	GABRR2, MS4A2, COMMD5
Development, differentiation	SOX4, FOXC1			ZNF141, BACH2	SOX4, EPHA4	
Cell growth and proliferation, gene expression, metabolism	H2AFB1/2/3	MSI2, TBC1D1, ZNF652, TOR1AIP1, ZMYM2		STRBP, CDC2L5, DDX18, MS12, ZNF439, ZNF91, ZNF226, MTMR2	MEF2C, DCN, KDM2B, RPS15, NFYB, DDX3Y, FOXD1, UTY	CCND2
Apoptosis, cell death, DNA damage response	TP63, BCL2L1, ZAK, RFFL, ATG4B, MKL1, HIPK2	C13orf15	<i>TP63</i>		TP63, MKL1	
Protein folding, protein translocation, heat shock	PPIL6	HSF2, SEC62				
Transport, mobility, cell adhesion	KCNMA1, ATP2B1, KIF21A, ANKH, TRPM4	VAMP1		ITGB1, CXCR4	ECM2, RHOBTB3	
IncRNA and other unknown function	COBLL1, NCRNA00173	C17orf58, C8orf6		SETD5, SLMO2	DNAJC5B, TMEM57, ANUBL1, IQCK	C10orf53

#### Table 4. Genes differentially expressed between patients with p63<sup>+</sup> and p63<sup>-</sup> DLBCL

Abbreviations: DLBCL, diffuse large B-cell lymphoma; FDR, false discovery rate; IncRNA, long noncoding RNA.

#### Multivariate survival analysis

We further performed multivariate survival analysis including p63 expression, p53 overexpression, and clinical parameters in the WT-TP53 and MUT-TP53 subsets individually. In the WT-TP53 subset, p63 expression but not WT-p53 overexpression remained as an independent prognostic factor for better OS and PFS; in the MUT-TP53 subset, MUT-p53 overexpression but not p63 expression was an independent prognostic factor for poorer PFS (borderline P value for OS) (Table 2).

# Gene expression profiling signature of p63 expression

To gain insights into the potential molecular mechanisms underlying the prognostic observation, we performed a series of GEP analyses comparing  $p63^+$  and

p63<sup>-</sup> patients in the overall DLBCL group and various subsets stratified by GCB/ABC subtype, TP53 mutation and p53 overexpression status (Fig 6A-H, Supplemental Fig S3A-D). Counts of significant differentially expressed genes (DEGs) between compared groups with different false discovery rate (FDR) thresholds are listed in Supplemental Table S1. Largely, whether p63 expression was associated with distinct GEP signatures did not correlated with whether p63 showed apparent prognostic effects, and the GEP signature of p63 expression in the MUT-TP53 subset was much more prominent (Fig 6B, Table 4) than that in the WT-TP53 subset (7 genes only with a FDR threshold of 0.30, figure not shown). However, after dividing the WT-TP53 subset into GCB and ABC subtypes of DLBCL patients, p63 expression showed GEP signatures, more distinctive in ABC than in GCB (Fig 6C, Supplemental Fig S3A), which was opposite to the pattern for overall ABC and GCB (only few DEGs in ABC compared to the distinct GEP signature in GCB, Supplemental Table S1). The p63 GEP signatures in the *MUT-TP53* and *WT-TP53* subsets had both similarity (upregulated *ATP2A2* and downregulated *ZNF652*) and difference (three genes, *GABBR2*, *PDHA1* and *NFYB*, showed opposite up- or down-regulation). Reinforcing the idea that p63 GEP signatures are more highlighted in the absence of WT-p53 activities as shown in the *MUT-TP53* subset, we further found that in WT-p53<sup>-</sup> ABC-DLBCL but not in WT-p53<sup>+</sup> ABC-DLBCL, p63 expression was associated with significant DEGs (Supplemental Fig S3B, Supplemental Table S3).

To gain insights into the functional relationship between p53 and p63, we further analyzed the overlap and difference between the p53 [37, 38] and p63 GEP signatures. The results (Table 5) suggest p63 expression

had a WT-p53-like GEP signature either in the context of WT-TP53 (such as CTAG2, SOX4 and ELL2, accounting for approximately 21% of the DEGs between WT-TP53/p63<sup>+</sup> and WT-TP53/p63<sup>-</sup>) or MUT-TP53 (such as DSE, ATM, CDK13, CD47, ELF1, DYRK1A [45], PFDN4, and TMEM97, accounting for approximately 4% of the DEGs between MUT- $TP53/p63^+$  and  $MUT-TP53/p63^-$ ), yet remained some MUT-p53-like GEP signature mainly in the context of MUT-TP53 (such as CAMTA1 resembling the MUT-p53 GEP signature, and ABHD11, KCNN3, MART3, and *MRPL30* opposite to the WT-p53 GEP signature; accounting for approximately 1.4% of the DEGs between *MUT-TP53*/p63<sup>+</sup> and *MUT-TP53*/p63<sup>-</sup>). Moreover, only in the  $p63^+$  but not in the  $p63^-$  subset, expression of WT-p53 or MUT-p53 was associated with distinct GEP signatures (Fig 6G, H), which may suggest that p63 is important for p53 activities.



**Figure 6. Gene expression profiling analysis.** (A) Heatmap for comparison between  $p63^+$  and  $p63^-$  DLBCL patients (false discovery rate < 0.15). (B) Heatmap for comparison between  $p63^+$  and  $p63^-$  DLBCL patients with *MUT-TP53* (false discovery rate < 0.05, fold change >1.68). (C) Heatmap for comparison between  $p63^+$  versus  $p63^-$  patients with ABC-DLBCL and WT-TP53 (false discovery rate < 0.20). (D) Heatmap for comparison between  $p63^+$  versus  $p63^-$  patients with GCB-DLBCL (false discovery rate < 0.05).



**Figure 6. Gene expression profiling analysis.** (E) Heatmap for comparison between  $p63^+$  and  $p63^-$  DLBCL patients with MUT-p53 overexpression (false discovery rate < 0.15). (F) Heatmap for comparison between  $p63^+$  and  $p63^-$  DLBCL patients with no or low (< 20%) expression levels of MUT-p53 (false discovery rate < 0.10). (G) Heatmap for comparison between WT-p53<sup>+</sup> ( $\geq$  20%) and WT-p53<sup>-</sup> (< 20%) DLBCL patients with p63 expression (false discovery rate < 0.30). (H) Heatmap for comparison between MUT-p53<sup>+</sup> ( $\geq$  20%) and MUT-p53<sup>-</sup> (< 20%) DLBCL patients with p63 expression (false discovery rate < 0.20).

We also compared the p63 GEP signature with the MDM2 GEP signature [38], and found 21 DEGs were common between the GEP signatures of p63 and MDM2 expression, among which 16 DEGs were not shared by the p53 GEP signature (Table 5).

Although the p53 and p63 GEP signatures overlapped, majority of the DEGs were not shared. Nonetheless, a p53-like tumor suppressor role of p63 was suggested by the p63 GEP signatures, including downregulation of *CCND2* (in *WT-TP53*/p63<sup>+</sup> ABC-DLBCL), *CDC27* and *MYCT1* (in *WT-TP53*/p63<sup>+</sup> GCB-DLBCL), *CDC2L5/ CDK13* and *CXCR4* (in *MUT-TP53*/p63<sup>+</sup> DLBCL. *TP53* mutations were associated with increased CXCR4 levels especially in GCB- DLBCL as previously reported [46]), *ELF1* which encodes a transcription factor that activates *LYN* and *BLK* (in *MUT-TP53*/p63<sup>+</sup> GCB-DLBCL), *MYBL1* and *STRBP* which play roles in proliferation and growth (in *MUT-TP53*/p63<sup>+</sup> GCB-DLBCL), antiapoptotic *C9orf82* and *BCOR* (which encodes an interacting corepressor of BCL6 required for germinal center formation and may influence apoptosis) (in *MUT-TP53*/p63<sup>+</sup> ABC-DLBCL), as well as upregulation of *HIPK2* (which promotes apoptosis through the activation of p53/*TP53*) (in p63<sup>+</sup> DLBCL) and *WWOX* (which functions synergistically with p53/*TP53* to control genotoxic stress-induced cell death) (in *MUT-TP53*/p63<sup>+</sup> ABC-DLBCL) (Supplemental Fig S3C-D).

**Table 5** Lists of differentially expressed genes between p63<sup>+</sup> and p63<sup>-</sup> DLBCL that are also in the p53 signatures and MDM2 signatures

Commo	on genes shared	l by the p63 <sup>+</sup> and p53 <sup>+</sup> signatures			
	<i>WT-TP53</i> WT-p53 <sup>+</sup> vs	WT-p53 <sup>-</sup>	<i>MUT-TP53</i> MUT-p53 <sup>+</sup> vs WT-p53 <sup>-</sup> <i>MUT-TP53 vs WT-TP53</i>		
		Same		Same	
Up↑	DSE	↑ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> ↑ in MUT-p53 <sup>+</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>+</sup> /p63 <sup>-</sup>	BCAS1	$\uparrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	
	ELL2	↑ in <i>WT-TP53/</i> p63 <sup>+</sup> <i>vs WT-TP53/</i> p63 <sup>-</sup>			
	FDXR	↑ in WT-p53 <sup>-</sup> /p63 <sup>+</sup> ABC vs WT-p53 <sup>-</sup> /p63 <sup>-</sup> ABC			
	GRRP1	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>			
	HPGD	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>			
	PFDN4	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>			
	SOX4	<pre>↑ in p63+ vs p63<sup>-</sup> ↑ in WT-TP53/p63<sup>+</sup> vs WT-TP53/p63<sup>-</sup> ↑ in WT-p53<sup>-</sup>/p63<sup>+</sup> ABC vs WT-p53<sup>-</sup>/p63<sup>-</sup> ABC</pre>			
Down ↓	ATM	$\downarrow \text{ in } MUT\text{-}TP53/\text{p}63^+ vs MUT\text{-}TP53/\text{p}63^-$	CAMTA1	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT- TP53/</i> p63 <sup>-</sup> ↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> GCB vs <i>MUT- TP53/</i> p63 <sup>-</sup> GCB	
	C3orf63	$\downarrow$ in MUT-p53 <sup>+</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>+</sup> /p63 <sup>-</sup>			
	CCDC69	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB			
	<b>CD</b> 47	↓ in MUT-p53 <sup>+</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>+</sup> /p63 <sup>-</sup>			
	CDC2L5/C DK13	↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> ↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> GCB vs <i>MUT-TP53</i> /p63 <sup>-</sup> GCB			
	DCLRE1C	↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> ↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> GCB vs <i>MUT-TP53</i> /p63 <sup>-</sup> GCB			
	DYRK1A	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>			
	ELF1	$\downarrow$ in MUT-TP53/p63 <sup>+</sup> vs MUT-TP53/p63 <sup>-</sup> $\downarrow$ in MUT-TP53/p63 <sup>+</sup> GCB vs MUT-TP53/p63 <sup>-</sup> GCB $\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB			
	ESR2	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB			
	HCG18	1 in WT-TP53/p63 <sup>+</sup> vs WT-TP53/p63 <sup>-</sup>			
	HERC4	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> <i>vs MUT-TP53/</i> p63 <sup>-</sup> ↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> GCB <i>vs MUT-TP53/</i> p63 <sup>-</sup> GCB			
	ІТСН	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>			
	LOC645513	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>			
	ORC4L	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>			
	PPP1R2	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>			
	TBC1D1	↓ in p63+ vs p63 <sup>-</sup> ↓ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB			
	РХК	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB			
	TMCC1	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> ABC <i>vs MUT-TP53/</i> p63 <sup>-</sup> ABC			
	ZCCHC7	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>			
	ZNF221	$\downarrow$ in WT-p53 <sup>-</sup> /p63 <sup>+</sup> ABC vs WT-p53 <sup>-</sup> /p63 <sup>-</sup> ABC			
		Opposite		Opposite	
Up ↑	KCNN3	↓ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	CTAG2	↓ in WT-p53 <sup>-</sup> /p63 <sup>+</sup> ABC <i>vs</i> WT-p53 <sup>-</sup> /p63 <sup>-</sup> ABC	
	KIAA0564	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup> ↓ in MUT-p53 <sup>+</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>+</sup> /p63 <sup>-</sup>	TMEM97	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>	
	MATR3	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>	SLC16A1	↓ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	
	MRPL30	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>			
Down ↓	ABHD11	↑ in <i>MUT-TP53</i> /p63 <sup>+</sup> ABC vs <i>MUT-TP53</i> /p63 <sup>-</sup> ABC			

Common genes shared by the p63 <sup>+</sup> and MDM2 <sup>+</sup> signatures						
	WT-TP53 MDM2 <sup>+</sup> vs	MDM2 <sup>-</sup>	<i>MUT-TP53</i> MDM2 <sup>+</sup> vs MDM2 <sup>-</sup>			
		Same		Same		
Up↑	FAM83A	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> ABC vs <i>MUT-TP53/</i> p63 <sup>-</sup> ABC				
	FDXR	↑ in WT-p53 <sup>-</sup> /p63 <sup>+</sup> ABC <i>vs</i> WT-p53 <sup>-</sup> /p63 <sup>-</sup> ABC				
	MICAL2	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> GCB vs <i>MUT-TP53/</i> p63 <sup>-</sup> GCB				
	PCBP3	↑ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>				
	TCEB3	↑ in p63+ <i>vs</i> p63 <sup>-</sup>				
Down↓	ATM	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>				
	BPTF	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> $\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	ATG7	↓ in p63 <sup>+</sup> GCB <i>vs</i> p63 <sup>-</sup> GCB		
	BRWD1	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	ATP5C1	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT- TP53/</i> p63 <sup>-</sup> ↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> GCB vs <i>MUT- TP53/</i> p63 <sup>-</sup> GCB		
	CD22	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB	EIF2A	$\downarrow$ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>		
	DHX36	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>	PAK2	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-</i> <i>TP53/</i> p63 <sup>-</sup>		
	EIF2A	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>	PRICKLE4/ TOMM6	↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> ABC <i>vs MUT-TP53</i> /p63 <sup>-</sup> ABC		
	NKTR	↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> ↓ in MUT-p53 <sup>+</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>+</sup> /p63 <sup>-</sup> ↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> GCB vs <i>MUT-TP53</i> /p63 <sup>-</sup> GCB				
	RBM26	↓ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup> ↓ in MUT-p53 <sup>-</sup> /p63 <sup>+</sup> vs MUT-p53 <sup>-</sup> /p63 <sup>-</sup>				
	RPL34	↓ in <i>MUT-TP53/</i> p63 <sup>+</sup> vs <i>MUT-TP53/</i> p63 <sup>-</sup>				
	SLC35F5	$\downarrow$ in <i>MUT-TP53</i> /p63 <sup>+</sup> vs <i>MUT-TP53</i> /p63 <sup>-</sup>				
	<i>WT-TP53</i> N	MDM2 <sup>+</sup> vs MUT-TP53 MDM2 <sup>+</sup>				
		Same				
Down↓	LPP	$\downarrow$ in p63 <sup>+</sup> GCB vs p63 <sup>-</sup> GCB				

On the other hand, some DEGs promoting tumor cell survival were also shown in the comparison between overall  $p63^+$  and  $p63^-$  DLBCL patients, which may be due to the oncogenic function provided by MUT-p53 or p63 isoforms in the  $p63^+$  DLBCL subsets. For example, antiapoptotic *BCL2L1*, *RFFL* (which negatively regulates p53, CASP8 and CASP10 through proteasomal degradation), *ATG4B* (required for autophagy), and *MKL1* (which suppresses TNF-induced cell death by inhibiting caspase activation) were up-

regulated in  $p63^+$  DLBCL compared with  $p63^-$  DLBCL, whereas *C13orf15/RGCC* (in response to DNA damage) was downregulated in  $p63^+$  DLBCL patients (Table 4). Cytokine/receptor genes *IL17RC*, *IL4*, *IL411* and *IL8RB/CXCR2* which have been associated with poorer prognosis in cancers, were upregulated in *MUT-TP53/*p63<sup>+</sup> DLBCL (Supplemental Table S2); *MLL2* was upregulated in  $p63^+$  patients with ABC-DLBCL and *MUT-TP53* (Supplemental Fig S3D).



p63 (TAp63 mostly) expression has significant favorable impact on clinical outcomes of DLBCL, but the protective effect can be abolished by *TP53* mutations, or compounded by the presence of ΔNp63 in some GCB-DLBCLs.

**Figure 7**. A hypothetical model illustrating the regulation and roles of p53 and p63 in DLBCL lymphomagenesis and clinical outcomes suggested by our clinical and biological data.

### **DISCUSSION**

Abnormal p63 expression patterns instead of *TP63* mutations have been found to be important for tumorigenesis [5]. Little data are available with conflicting results regarding p63 expression and its prognostic role [27, 39, 43]. We found that p63 expression correlated with a superior survival in ABC-DLBCL with *WT-TP53* and in high-risk (IPI >2) DLBCL (regardless GCB or ABC), which is consistent with a previous study in high-intermediate and high risk DLBCL [27]. The association of p63 expression with high-risk IPI in GCB-DLBCL, and thus affecting its apparent prognostic effects in GCB and overall DLBCL, may contribute to the inconsistent findings from previous studies.

The prognostic effect of p63 expression suggests that p63 has a tumor suppressor role for DLBCL, although its protective effect can be antagonized or abolished by *TP53* mutations and high-risk DLBCL associated biology. In our cohort, p63 expression was associated with increased levels of IRF4/MUM-1, p21, MDM2, and p16-INK4a resembling that of WT-p53 yet independent of p53 mutation status. GEP analysis showed that compared to the prominent p63 GEP signature within the *MUT-TP53* subset, the comparison

between  $p63^+$  and  $p63^-$  patients with WT-TP53 had much fewer DEGs; DEGs were shown within the WTp53<sup>-</sup> but not WT-p53<sup>+</sup> ABC-DLBCL subset. These results may suggest that the tumor suppressor function of p63 may overlap with (and is probably weaker than) that of WT-p53, and when TP53 was mutated, p63 functions as a supplemental tumor suppressor alternative to WT-p53. However, MUT-p53 function remained or dominated p63 function in certain MUT-TP53 cases (Table 5), likely due to the significantly higher levels of MUT-p53 than p63 [47]. In addition to the GEP results as above, p63 expression correlated with MDM2 upregulation and BCL2 and MDM4 downregulation (P=0.0174, P=0.0487 and P=0.090 respectively) resembling WT-p53 expression GEP signature (although the FDRs for the comparison between  $p63^+$  and  $p63^-$  DLBCL were higher). In contrast, CDKN1A/p21, MCL1, B2M, and FYB showed great variation even opposite up/down regulation between the WT-p53<sup>+</sup> and the  $p63^+$  GEP signature. These phenomena may be explained by the remained MUT-p53-like function in the MUT-TP53/p63<sup>+</sup> cases, whereas TP63 mutations and expression of different p63 isoforms may not be significant factors as suggested by the previous studies [5, 25, 43] and our preliminary data of TP63 mutations in DLBCL (unpublished data).

These observations in DLBCL may support previous functional studies, which showed that TAp63 $\alpha$  and TAp63 $\gamma$  (but not  $\Delta$ Np63) could induce apoptosis at lesser levels than WT-p53 [48]; TAp63, and also TAp73, together with p53, may transactivate a group of common target genes in response to DNA damage, including damage resulting from exposure to doxorubicin, a component of R-CHOP; <sup>1</sup> TAp63 and MUT-p53 antagonize each other mainly in the regulation of metastasis and tumor dissemination [5]; p53 mutants may bind directly to p63 and inhibit the p63-mediated transcription of p53 target genes [49, 50]. Strategies to overcome MUT-p53 interaction with p63, decrease MUT-p53 levels and enhance p63 levels may have therapeutic value [47]. On the other hand, in mouse embryonic fibroblasts, p63 and p73 are required for p53-dependent apoptosis in response to DNA damage [7]. This may explain why our GEP comparisons between  $p53^+$  and  $p53^-$  DLBCL showed DEGs within the  $p63^+$  but not  $p63^-$  subset. Moreover, our data suggested that p63 act together with p53 in essential pathways yet also function some independently in many processes such as development, immune response and chemokinesis. Large variations between p63 signatures in the overall DLBCL patient population and in the GCB and ABC subsets may also imply a wide range of p63 activities. These characteristics of p63 function compared with p53, as well as association with high Ki-67 (consistent with previous studies [21, 43]) and high IPI may explain the limitation of p63's apparent prognostic effect in DLBCL.

It is also possible that the correlation between p63 expression and better survival outcomes may be also influenced by the escape from MDM2-mediated degradation. In our cohort, the p63's protective effects on patient survival were independent of MDM2 expression, yet GEP signatures were only shown in MDM2<sup>low</sup> but not in MDM2<sup>high</sup> subsets (data not shown), suggesting that MDM2 may suppress p63 function but the suppression is not significant to the p63's protective effect. Conversely, p63 may have confounded the MDM2's prognostic effect in DLBCL just as that of WT-p53 [38], suggested by the common genes shared by the MDM2 and p63 GEP signatures (Table 5). Previous studies have suggested that p63 degradation is independent of MDM2 [24, 31] and that MDM2 increases the protein level and transcriptional activity of p63 [51]. The MDM2 inhibitor p14ARF directly interacts with and impairs p63 transcriptional activity [52]. On the other hand, it has also been shown that MDM2 transports p63 out of nucleus and inhibits its transcription function [53].

Yang *et al.* speculated that p63 expression in cancer cells was due to *TP63* gene amplification by genomic

instability [3], and other researches showed that p63 expression was regulated via mRNA stability [4, 19]. TP63 rearrangements have been reported in 1.2-5% of DLBCL (exclusive of GCB subtype) and also in 5.8% of peripheral T-cell lymphomas, which resulted in a truncated p63 protein lacking the TA domain [40, 41]. Our data showed the associations of p63 expression with BCL6 (mapped to 3q27) translocations, which appears to suggest the possibility of concurrent translocation of TP63 gene (mapped to 3q27-28) due to chromosomal proximity in  $p63^+$  DLBCL subsets. In these cases it is possible that expressed p63 had oncogenic function like  $\Delta Np63$ , which may explain the oncogenic DEGs in the p63 GEP signatures, and the lack of p63's prognostic significance in GCB-DLBCL. In addition, genomic stress similar to that inducing p53 may also be the cause of p63 expression in subsets of  $p63^+$  DLBCL [54], since our data showed correlation between the WT-TP53 and TP63 mRNA levels, and both WT-p53 and p63 expression were associated with increased IRF4/MUM-1 and Ki-67 expression. Fig 7 illustrates these potential causes for p63 expression and possible relationships between p63 and WT-p53/MUTp53 function. Understanding the mechanisms regulating TP63 may lead to therapeutic strategies. In DLBCL cell lines, FOXP1, directly represses TP63 and cooperate with NF-kB signaling to promote lymphoma cell survival [42]. Consistently, our GEP data also suggest that molecules related to B-cell receptor signaling may be potential targets which suppresses p63 expression, as in GCB-DLBCL and MUT-TP53/p63<sup>+</sup> DLBCL, p63 expression was associated with downregulation of SYK and *ELF1* respectively (suggesting decreased B-cell receptor signaling).

In conclusion, we demonstrated the correlation of p63 expression and better survival outcomes in patients with high-risk DLBCL, ABC-DLBCL with *WT-TP53*, and biology associated with p63 expression supporting p63's tumor suppressor role in DLBCL. This study helps identify a subgroup of patients with better prognosis among patients who have ABC-DLBCL or high-risk DLBCL. Targeting p63 expression and function may be a novel therapeutic strategy for particular subgroups of DLBCL patients.

# **MATERIALS AND METHODS**

<u>Patients.</u> A total of 795 patients with *de novo* DLBCL from 20 medical centers treated with R-CHOP were studied, randomly divided into a training set (n=520) and a validation set (n=275). The diagnostic criteria, selection process, therapy, and treatment response have been described previously [37]. The study was approved as being of minimal or no risk or as exempt by the

institution review boards of all participating medical centers.

Immunohistochemistry. Tissue microarrays prepared from the diagnostic formalin-fixed, paraffin-embedded (FFPE) tissue blocks of all patients studied were stained with an anti-p63 antibody (4A4, Santa Cruz Biotechnology, Santa Cruz, CA) which can detect all p63 isoforms. Expression levels of p63 were determined by estimating the percentage of p63-positive tumor cells in the tissue array cores. X-tile software and receiver operating characteristic curve analysis by GraphPad Prism 6 Software were used to determine the percentage of p63-positive cells with maximal discriminatory power for the separation of DLBCL patients into 2 different prognostic groups. Evaluation of other biomarkers by immunohistochemistry was also performed on tissue microarrays using corresponding antibodies: p53 (DO-7, Dako, Carpinteria, CA), MDM2 (IF2, Calbiochem, Billerica, MA), p21 (Dako), Bcl-2 (Clone-124, Dako, Carpinteria, CA), Ki-67 (Dako), CD30 (clone BerH2, Dako), Bcl-6 (Dako), FOXP1 (Abcam), IRF4/MUM1 (Dako), CD10 (56C6, Vantana), c-Rel (Dako), and CXCR4 (Abcam, San Francisco, CA). Details of immunohistochemistry procedures and scoring processes have been described previously [38, 44, 55-58].

<u>TP53 and TP63 sequencing</u>, fluorescence in situ hybridization. Genomic DNA samples were extracted from FFPE tissues, and the *TP53* coding region and splice site sequence were determined for 460 patients in the training set using a p53 AmpliChip (Roche Molecular Systems, Pleasanton, CA) as described previously [37]. *TP63* coding region sequence was analyzed by Sanger sequencing method. *MYC*, *BCL2*, *BCL6*, and *REL* gene arrangements and copy number aberrations were detected by fluorescence in situ hybridization [56, 59, 60].

<u>Gene expression profiling.</u> Gene expression profiling was performed on Affymetrix GeneChips HG-U133 Plus Version 2.0 (Affymetrix, Santa Clara, CA) using total RNAs as described previously [37, 55]. The CEL files are deposited in the National Center for Biotechnology Information Gene Expression Omnibus repository (GSE#31312). The microarray data were quantified and normalized by the frozen robust multiarray analysis (RMA) algorithm. The differentially expressed genes were identified by using multiple *t*tests.

<u>Statistical analysis.</u> The clinical and pathologic features at the time of presentation were compared between various DLBCL subgroups by using the chi-square test and unpaired t test. Correlation between expression of different genes or proteins was evaluated by Spearman

rank correlation. Overall survival (OS) was calculated from the date of diagnosis to the date of last follow-up or death. Progression-free survival (PFS) was calculated from the date of diagnosis to the date of disease progression or death. OS and PFS curves of the various groups were analyzed by GraphPad Prism 6 software using the Kaplan-Meier method, and differences were compared with use of the log-rank (Cox-Mantel) test. Multivariate analysis was conducted by using the Cox proportional hazards regression model with the SPSS software version 19.0 (IBM, Armonk, NY). Any difference with a *P* value of < 0.05 was considered statistically significant.

# Authorship

Contribution: Z.Y.X-M, S.Z., and K.H.Y designed and conducted the research and performed the statistical analysis; Z.Y.X-M, S.Z., X.L., G.C.M., X.W., Y.X., C.V., A.T., S.M.M., K.D., A.C., A.O., Y.Z., G.B., K.L.R., E.D.H., W.W.L.C., J.H.K., J.H., M.P., A.J.M.F., X.Z., M.B.M., F.B., B.M.P., M.A.P., J.N.W., L.J.M., and K.H.Y. contributed vital new reagents, resources, technology, and analytical tools; Z.Y.X-M, S.Z., C.V., A.T., S.M.M., K.D., A.C., A.O., Y.Z., G.B., K.L.R., E.D.H., W.W.L.C., J.H.K., J.H., M.P., A.J.M.F., X.Z., M.B.M., F.B., B.M.P., M.A.P., J.N.W., and K.H.Y. collected clinical and follow-up data under approval by the Institutional Review Boards and the material transfer agreement; Z.Y.X-M, S.Z., L.J.M., and K.H.Y. wrote and edited the manuscript; and all authors contributed vital strategies, participated in discussions, and provided scientific input.

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#### **Conflict of interest statement**

KHY receives research support from Roche Molecular System, Gilead Sciences Pharmaceutical, Seattle Genetics, Dai Sanyo Pharmaceutical, Adaptive Biotechnology, Incyte Pharmaceutical, and HTG Molecular Diagnostics.

### **REFERENCES**

**1.** Su X, Chakravarti D and Flores ER. p63 steps into the limelight: crucial roles in the suppression of tumorigenesis and metastasis. Nat Rev Cancer. 2013; 13:136-143.

**2.** Yang A and McKeon F. P63 and P73: P53 mimics, menaces and more. Nat Rev Mol Cell Biol. 2000; 1:199-207.

**3.** Yang A, Kaghad M, Wang Y, Gillett E, Fleming MD, Dotsch V, Andrews NC, Caput D and McKeon F. p63, a p53 homolog at 3q27-29, encodes multiple products with transactivating, death-inducing, and dominant-negative activities. Molecular cell. 1998; 2:305-316.

**4.** Cho SJ, Jung YS and Chen X. Poly (C)-binding protein 1 regulates p63 expression through mRNA stability. PloS one. 2013; 8:e71724.

**5.** Pflaum J, Schlosser S and Muller M. p53 Family and Cellular Stress Responses in Cancer. Front Oncol. 2014; 4:285.

**6.** Levine AJ, Tomasini R, McKeon FD, Mak TW and Melino G. The p53 family: guardians of maternal reproduction. Nat Rev Mol Cell Biol. 2011; 12:259-265.

**7.** Flores ER, Tsai KY, Crowley D, Sengupta S, Yang A, McKeon F and Jacks T. p63 and p73 are required for p53-dependent apoptosis in response to DNA damage. Nature. 2002; 416:560-564.

**8.** Venkatanarayan A, Raulji P, Norton W, Chakravarti D, Coarfa C, Su X, Sandur SK, Ramirez MS, Lee J, Kingsley CV, Sananikone EF, Rajapakshe K, Naff K, et al. IAPP-driven metabolic re rogramming induces regression of p53-deficient tumours in vivo. Nature. 2015; 517:626-630.

**9.** Mills AA, Zheng B, Wang XJ, Vogel H, Roop DR and Bradley A. p63 is a p53 homologue required for limb and epidermal morphogenesis. Nature. 1999; 398:708-713.

**10.** Suh EK, Yang A, Kettenbach A, Bamberger C, Michaelis AH, Zhu Z, Elvin JA, Bronson RT, Crum CP and McKeon F. p63 protects the female germ line during meiotic arrest. Nature. 2006; 444:624-628.

**11.** Yang A, Schweitzer R, Sun D, Kaghad M, Walker N, Bronson RT, Tabin C, Sharpe A, Caput D, Crum C and McKeon F. p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. Nature. 1999; 398:714-718.

**12.** Sun R, Zhang Y, Lv Q, Liu B, Jin M, Zhang W, He Q, Deng M, Liu X, Li G, Li Y, Zhou G, Xie P, et al. Toll-like receptor 3 (TLR3) induces apoptosis via death receptors and mitochondria by upregulating the transactivating p63 isoform alpha (TAP63alpha). J Biol Chem. 2011; 286:15918-15928.

13. Levrero M, De Laurenzi V, Costanzo A, Gong J, Wang JY and Melino G. The p53/p63/p73 family of transcription factors: overlapping and distinct functions. J Cell Sci. 2000; 113:1661-1670.
14. Dohn M, Zhang S and Chen X. p63alpha and DeltaNp63alpha can induce cell cycle arrest and apoptosis and differentially regulate p53 target genes. Oncogene. 2001; 20:3193-3205.

**15.** Perez-Losada J, Wu D, DelRosario R, Balmain A and Mao JH. p63 and p73 do not contribute to p53-mediated lymphoma suppressor activity in vivo. Oncogene. 2005; 24:5521-5524.

**16.** Chakrabarti R, Wei Y, Hwang J, Hang X, Andres Blanco M, Choudhury A, Tiede B, Romano RA, DeCoste C, Mercatali L, Ibrahim T, Amadori D, Kannan N, et al. DeltaNp63 promotes stem cell activity in mammary gland development and basal-like breast cancer by enhancing Fzd7 expression and Wnt signalling. Nat Cell Biol. 2014; 16:1004-1015, 1001-1013.

**17.** Su X, Chakravarti D, Cho MS, Liu L, Gi YJ, Lin YL, Leung ML, El-Naggar A, Creighton CJ, Suraokar MB, Wistuba I and Flores ER. TAp63 suppresses metastasis through coordinate regulation of Dicer and miRNAs. Nature. 2010; 467:986-990.

**18.** Guo X, Keyes WM, Papazoglu C, Zuber J, Li W, Lowe SW, Vogel H and Mills AA. TAp63 induces senescence and suppresses tumorigenesis in vivo. Nat Cell Biol. 2009; 11:1451-1457.

**19.** Xu E, Zhang J, Zhang M, Jiang Y, Cho SJ and Chen X. RNAbinding protein RBM24 regulates p63 expression via mRNA stability. Mol Cancer Res. 2014; 12:359-369.

**20.** Truong AB, Kretz M, Ridky TW, Kimmel R and Khavari PA. p63 regulates proliferation and differentiation of developmentally mature keratinocytes. Genes Dev. 2006; 20:3185-3197.

**21.** Hedvat CV, Teruya-Feldstein J, Puig P, Capodieci P, Dudas M, Pica N, Qin J, Cordon-Cardo C and Di Como CJ. Expression of p63 in diffuse large B-cell lymphoma. Appl Immunohistochem Mol Morphol. 2005; 13:237-242.

**22.** Danilova N, Sakamoto KM and Lin S. p53 family in development. Mech Dev. 2008; 125:919-931.

**23.** Wolff S, Talos F, Palacios G, Beyer U, Dobbelstein M and Moll UM. The alpha/beta carboxy-terminal domains of p63 are required for skin and limb development. New insights from the Brdm2 mouse which is not a complete p63 knockout but expresses p63 gamma-like proteins. Cell Death Differ. 2009; 16:1108-1117.

**24.** Okada Y, Osada M, Kurata S, Sato S, Aisaki K, Kageyama Y, Kihara K, Ikawa Y and Katoh I. p53 gene family p51(p63)encoded, secondary transactivator p51B(TAp63alpha) occurs without forming an immunoprecipitable complex with MDM2, but responds to genotoxic stress by accumulation. Exp Cell Res. 2002; 276:194-200.

**25.** Di Como CJ, Urist MJ, Babayan I, Drobnjak M, Hedvat CV, Teruya-Feldstein J, Pohar K, Hoos A and Cordon-Cardo C. p63 expression profiles in human normal and tumor tissues. Clin Cancer Res. 2002; 8:494-501.

**26.** Nylander K, Vojtesek B, Nenutil R, Lindgren B, Roos G, Zhanxiang W, Sjostrom B, Dahlqvist A and Coates PJ. Differential expression of p63 isoforms in normal tissues and neoplastic cells. J Pathol. 2002; 198:417-427.

**27.** Hallack Neto AE, Siqueira SA, Dulley FL, Ruiz MA, Chamone DA and Pereira J. p63 protein expression in high risk diffuse large B-cell lymphoma. J Clin Pathol. 2009; 62:77-79.

**28.** Alexandrova EM and Moll UM. Role of p53 family members p73 and p63 in human hematological malignancies. Leuk Lymphoma. 2012; 53:2116-2129.

**29.** Aubry MC, Roden A, Murphy SJ, Vasmatzis G, Johnson SH, Harris FR, Halling G, Knudson RA, Ketterling RP and Feldman AL. Chromosomal rearrangements and copy number abnormalities of TP63 correlate with p63 protein expression in lung adenocarcinoma. Mod Pathol. 2015; 28:359-366.

**30.** Geddert H, Kiel S, Heep HJ, Gabbert HE and Sarbia M. The role of p63 and deltaNp63 (p40) protein expression and gene

amplification in esophageal carcinogenesis. Hum Pathol. 2003; 34:850-856.

**31.** Ying H, Chang DL, Zheng H, McKeon F and Xiao ZX. DNAbinding and transactivation activities are essential for TAp63 protein degradation. Mol Cell Biol. 2005; 25:6154-6164.

**32.** Lo Muzio L, Santarelli A, Caltabiano R, Rubini C, Pieramici T, Trevisiol L, Carinci F, Leonardi R, De Lillo A, Lanzafame S, Bufo P and Piattelli A. p63 overexpression associates with poor prognosis in head and neck squamous cell carcinoma. Hum Pathol. 2005; 36:187-194.

**33.** Stetsenko GY, Malekirad J, Paulson KG, Iyer JG, Thibodeau RM, Nagase K, Schmidt M, Storer BE, Argenyi ZB and Nghiem P. p63 expression in Merkel cell carcinoma predicts poorer survival yet may have limited clinical utility. Am J Clin Pathol. 2013; 140:838-844.

**34.** Takahashi Y, Noguchi T, Takeno S, Kimura Y, Okubo M and Kawahara K. Reduced expression of p63 has prognostic implications for patients with esophageal squamous cell carcinoma. Oncol Rep. 2006; 15:323-328.

**35.** Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, Boldrick JC, Sabet H, Tran T, Yu X, Powell JI, Yang L, Marti GE, et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. Nature. 2000; 403:503-511.

**36.** Testoni M, Zucca E, Young KH and Bertoni F. Genetic lesions in diffuse large B-cell lymphomas. Ann Oncol. 2015. 26:1069-1080

**37.** Xu-Monette ZY, Wu L, Visco C, Tai YC, Tzankov A, Liu WM, Montes-Moreno S, Dybkaer K, Chiu A, Orazi A, Zu Y, Bhagat G, Richards KL, et al. Mutational profile and prognostic significance of TP53 in diffuse large B-cell lymphoma patients treated with R-CHOP: report from an International DLBCL Rituximab-CHOP Consortium Program Study. Blood. 2012; 120:3986-3996.

**38.** Xu-Monette ZY, Moller MB, Tzankov A, Montes-Moreno S, Hu W, Manyam GC, Kristensen L, Fan L, Visco C, Dybkaer K, Chiu A, Tam W, Zu Y, et al. MDM2 phenotypic and genotypic profiling, respective to TP53 genetic status, in diffuse large B-cell lymphoma patients treated with rituximab-CHOP immunochemotherapy: a report from the International DLBCL Rituximab-CHOP Consortium Program. Blood. 2013; 122:2630-2640.

**39.** Park CK and Oh YH. Expression of p63 in reactive hyperplasias and malignant lymphomas. J Korean Med Sci. 2005; 20:752-758.

**40.** Vasmatzis G, Johnson SH, Knudson RA, Ketterling RP, Braggio E, Fonseca R, Viswanatha DS, Law ME, Kip NS, Ozsan N, Grebe SK, Frederick LA, Eckloff BW, et al. Genome-wide analysis reveals recurrent structural abnormalities of TP63 and other p53-related genes in peripheral T-cell lymphomas. Blood. 2012; 120:2280-2289.

**41.** Scott DW, Mungall KL, Ben-Neriah S, Rogic S, Morin RD, Slack GW, Tan KL, Chan FC, Lim RS, Connors JM, Marra MA, Mungall AJ, Steidl C, et al. TBL1XR1/TP63: a novel recurrent gene fusion in B-cell non-Hodgkin lymphoma. Blood. 2012; 119:4949-4952.

**42.** van Keimpema M, Gruneberg LJ, Mokry M, van Boxtel R, Koster J, Coffer PJ, Pals ST and Spaargaren M. FOXP1 directly represses transcription of proapoptotic genes and cooperates with NF-kappaB to promote survival of human B cells. Blood. 2014; 124:3431-3440.

**43.** Fukushima N, Satoh T, Sueoka N, Sato A, Ide M, Hisatomi T, Kuwahara N, Tomimasu R, Tsuneyoshi N, Funai N, Sano M, Tokunaga O and Sueoka E. Clinico-pathological characteristics of

p63 expression in B-cell lymphoma. Cancer Sci. 2006; 97:1050-1055.

**44.** Hu S, Xu-Monette ZY, Tzankov A, Green T, Wu L, Balasubramanyam A, Liu WM, Visco C, Li Y, Miranda RN, Montes-Moreno S, Dybkaer K, Chiu A, et al. MYC/BCL2 protein coexpression contributes to the inferior survival of activated B-cell subtype of diffuse large B-cell lymphoma and demonstrates high-risk gene expression signatures: a report from The International DLBCL Rituximab-CHOP Consortium Program. Blood. 2013; 121:4021-4031; quiz 4250.

**45.** Zhang Y, Liao JM, Zeng SX and Lu H. p53 downregulates Down syndrome-associated DYRK1A through miR-1246. EMBO reports. 2011; 12:811-817.

**46.** Chen J, Xu-Monette ZY, Deng L, Shen Q, Manyam GC, Martinez-Lopez A, Zhang L, Montes-Moreno S, Visco C, Tzankov A, Yin L, Dybkaer K, Chiu A, et al. Dysregulated CXCR4 expression promotes lymphoma cell survival and independently predicts disease progression in germinal center B-cell-like diffuse large B-cell lymphoma. Oncotarget. 2015; 6:5597-5614. DOI: 10.18632/oncotarget.3343.

**47.** Li Y and Prives C. Are interactions with p63 and p73 involved in mutant p53 gain of oncogenic function? Oncogene. 2007; 26:2220-2225.

**48.** Dietz S, Rother K, Bamberger C, Schmale H, Mossner J and Engeland K. Differential regulation of transcription and induction of programmed cell death by human p53-family members p63 and p73. FEBS Lett. 2002; 525:93-99.

**49.** Gaiddon C, Lokshin M, Ahn J, Zhang T and Prives C. A subset of tumor-derived mutant forms of p53 down-regulate p63 and p73 through a direct interaction with the p53 core domain. Mol Cell Biol. 2001; 21:1874-1887.

**50.** Strano S, Fontemaggi G, Costanzo A, Rizzo MG, Monti O, Baccarini A, Del Sal G, Levrero M, Sacchi A, Oren M and Blandino G. Physical interaction with human tumor-derived p53 mutants inhibits p63 activities. J Biol Chem. 2002; 277:18817-18826.

**51.** Calabro V, Mansueto G, Parisi T, Vivo M, Calogero RA and La Mantia G. The human MDM2 oncoprotein increases the transcriptional activity and the protein level of the p53 homolog p63. J Biol Chem. 2002; 277:2674-2681.

**52.** Calabro V, Mansueto G, Santoro R, Gentilella A, Pollice A, Ghioni P, Guerrini L and La Mantia G. Inhibition of p63 transcriptional activity by p14ARF: functional and physical link between human ARF tumor suppressor and a member of the p53 family. Mol Cell Bio. 2004; 24:8529-8540.

**53.** Kadakia M, Slader C and Berberich SJ. Regulation of p63 function by Mdm2 and MdmX. DNA Cell Biol. 2001; 20:321-330.

**54.** Gonfloni S, Di Tella L, Caldarola S, Cannata SM, Klinger FG, Di Bartolomeo C, Mattei M, Candi E, De Felici M, Melino G and Cesareni G. Inhibition of the c-Abl-TAp63 pathway protects mouse oocytes from chemotherapy-induced death. Nature medicine. 2009; 15:1179-1185.

**55.** Visco C, Li Y, Xu-Monette ZY, Miranda RN, Green TM, Li Y, Tzankov A, Wen W, Liu WM, Kahl BS, d'Amore ES, Montes-Moreno S, Dybkaer K, et al. Comprehensive gene expression profiling and immunohistochemical studies support application of immunophenotypic algorithm for molecular subtype classification in diffuse large B-cell lymphoma: a report from the International DLBCL Rituximab-CHOP Consortium Program Study. Leukemia. 2012; 26:2103-2113.

**56.** Tzankov A, Xu-Monette ZY, Gerhard M, Visco C, Dirnhofer S, Gisin N, Dybkaer K, Orazi A, Bhagat G, Richards KL, Hsi ED, Choi

WW, van Krieken JH, et al. Rearrangements of MYC gene facilitate risk stratification in diffuse large B-cell lymphoma patients treated with rituximab-CHOP. Mod Pathol. 2014; 27:958-971.

**57.** Ok CY, Chen J, Xu-Monette ZY, Tzankov A, Manyam GC, Li L, Visco C, Montes-Moreno S, Dybkaer K, Chiu A, Orazi A, Zu Y, Bhagat G, et al. Clinical Implications of Phosphorylated STAT3 Expression in De Novo Diffuse Large B-cell Lymphoma. Clin Cancer Res. 2014; 20:5113-5123.

**58.** Hu S, Xu-Monette ZY, Balasubramanyam A, Manyam GC, Visco C, Tzankov A, Liu WM, Miranda RN, Zhang L, Montes-Moreno S, Dybkaer K, Chiu A, Orazi A, et al. CD30 expression defines a novel subgroup of diffuse large B-cell lymphoma with favorable prognosis and distinct gene expression signature: a report from the International DLBCL Rituximab-CHOP Consortium Program Study. Blood. 2013; 121:2715-2724.

**59.** Visco C, Tzankov A, Xu-Monette ZY, Miranda RN, Tai YC, Li Y, Liu WM, d'Amore ES, Li Y, Montes-Moreno S, Dybkaer K, Chiu A, Orazi A, et al. Patients with diffuse large B-cell lymphoma of germinal center origin with BCL2 translocations have poor outcome, irrespective of MYC status: a report from an International DLBCL rituximab-CHOP Consortium Program Study. Haematologica. 2013; 98:255-263.

**60.** Xu-Monette ZY, Dabaja BS, Wang X, Tu M, Manyam GC, Tzankov A, Xia Y, Li Zhang, Visco C, Dybkaer K, Yin L, Chiu A, Orazi A, et al. Clinical features, tumor biology and prognosis associated with MYC rearrangement and overexpression in diffuse large B-cell lymphoma patients treated with rituximab-CHOP. Modern Pathology. 2015; 28:1555-1573.

### SUPPLEMENTAL DATA

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