

Draft Genome Sequences of Two Benthic Cyanobacteria, *Oscillatoriales* USR 001 and *Nostoc* sp. MBR 210, Isolated from Tropical Freshwater Lakes

Shu Harn Te,^a Boon Fei Tan,^b Janelle R. Thompson,^b Karina Yew-Hoong Gin^{a,c}

NUS Environmental Research Institute, National University of Singapore, Singapore, Singapore^a; Centre for Environmental Sensing and Modelling, Singapore-MIT Alliance for Research and Technology, Singapore, Singapore^b; Department of Civil and Environmental Engineering, National University of Singapore, Singapore, Singapore^c

Genomes of two filamentous benthic cyanobacteria were obtained from cocultures obtained from two freshwater lakes. The cultures were obtained by first growing cyanobacterial trichome on solid medium, followed by subculturing in freshwater media. Subsequent shotgun sequencing, *de novo* assembly, and genomic binning yielded almost complete genomes of *Oscillatoriales* USR 001 and *Nostoc* sp. MBR 210.

Received 18 August 2016 Accepted 20 August 2016 Published 13 October 2016

Citation Te SH, Tan BF, Thompson JR, Gin KY-H. 2016. Draft genome sequences of two benthic cyanobacteria, *Oscillatoriales* USR 001 and *Nostoc* sp. MBR 210, isolated from tropical freshwater lakes. *Genome Announc* 4(5):e01115-16. doi:10.1128/genomeA.01115-16.

Copyright © 2016 Te et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](http://creativecommons.org/licenses/by/4.0/).

Address correspondence to Karina Yew-Hoong Gin, ceeginyh@nus.edu.sg.

Benthic cyanobacteria inhabit the bottom of a diverse range of bodies of water, including lakes, wetlands, estuaries, and oceans, forming benthic mats in these environments (1). Under favorable conditions, they can proliferate rapidly and synthesize undesirable secondary metabolites, including toxins and odors (2). However, they are less studied compared to planktonic cyanobacteria even though they are able to cause similar ecological and water quality impacts on affected waters (3). Two filamentous benthic cyanobacteria isolated from tropical freshwater lakes in Singapore were identified as *Oscillatoriales* USR 001 and *Nostoc* sp. MBR 210, based on morphological traits (4). Here, we present additional genomic information about these isolates, which is important for functional annotation, pathways analysis, comparative genomics and for better understanding of their roles in bloom formation.

The two filamentous cyanobacteria were acquired through an agar culturing method (5). Briefly, lake sediment samples were streaked across agar plates enriched with McBride *Listeria* agar (MLA) medium (6). The agar plates were incubated (25°C, light intensity: 25 $\mu\text{mol}/\text{m}^2\text{s}$) until green filaments appeared; then individual filaments were aseptically cut, transferred, and cultivated in sterile MLA media for 2 weeks. The genomic DNA extraction, Illumina HiSeq 2000 sequencing, and read quality controls were conducted following a method described previously (7). Subsequently, the two metagenomes were *de novo* assembled separately into scaffolds using CLC Genomics Workbench version 8. Contigs belonging to *Cyanobacteria* in each metagenome were separated from those of heterotrophic bacteria using MetaBAT (8), following which genome completeness and sequence contaminants were determined using CheckM (9), and the lack of a sequence contaminant was confirmed using a BLAST-based approach (10). The two genomes were annotated using RAST (11) and NCBI PGAP (http://www.ncbi.nlm.nih.gov/genome/annotation_prok).

Genomes of the two cyanobacteria have GC contents of 41%

and completeness of 99%, assessed using checkM by comparing 579 to 583 reference marker genes in 79 to 82 lineage-specific reference genomes. The draft genome for *Oscillatoriales* USR 001 comprises 5.9 Mbp contained in 96 scaffolds, while *Nostoc* sp. MBR 210 has a genome size of 6.9 Mbp contained in 36 scaffolds. The 16S rRNA of *Nostoc* sp. MBR 210 (1,482 bp) is 99% identical to that of *Nostoc piscinale* CENA21 (CP012036.1), whereas the 16S rRNA of *Oscillatoriales* USR 001 (1,492 bp) is 98% identical to three members of the family *Oscillatoriales*: *Kamptomena animale* (EF654087.1), *Phormidium animale* CCAP (HF678514.1), and *Oscillatoria lutea* (KM019965.1). Further comparison between the genome of *Oscillatoriales* USR 001 with all reference genomes of the three genera currently available in the NCBI and JGI IMG databases (12) revealed a two-way average nucleotide identity of <90%, precluding the classification of USR 001 to the genus level. As members of these two organisms are known to be potential toxin (microcystin and anatoxin) and odor (geosmin and 2-methylisoborneol) producers, we used antiSMASH version 3.0.5 (13) and BLASTp to search for potential gene- or gene cluster-encoding secondary metabolites, to which no toxin and off-flavor-producing gene (as above) was identified in both genomes. Both genomes carry genes for photosynthesis and CO₂ and nitrogen fixation (e.g., carboxysome and nitrogenase, respectively) and a limited number of genes encoding sugar utilization (e.g., beta-galactosidase in USR 001; alpha-mannosidase for mannose utilization in MBR 210), suggesting their potential roles as photoheterotrophic nitrogen fixers.

Accession number(s). These whole-genome shotgun projects have been deposited in DDBJ/ENA/GenBank under the accession numbers [MBRE00000000](https://www.ncbi.nlm.nih.gov/nuccore/MBRE00000000) (*Oscillatoriales* USR 001) and [MBRD00000000](https://www.ncbi.nlm.nih.gov/nuccore/MBRD00000000) (*Nostoc* sp. MBR 210). The versions described in this paper are the first versions, MBRE01000000 and MBRD01000000.

ACKNOWLEDGMENTS

We thank Boo Chek Yin for providing cultures and the Public Utilities Board of Singapore (PUB) for their collaboration in this project.

This research grant is supported by the Singapore National Research Foundation under its Environmental and Water Technologies Strategies Research Programme and administered by PUB. B.F.T. and J.R.T. were supported by the National Research Foundation of Singapore through the Singapore MIT Alliance for Research and Technology's (SMART) Center for Environmental Sensing and Modeling (CENSAM) research program.

FUNDING INFORMATION

This work, including the efforts of Shu Harn Te and Karina Yew-Hoong Gin, was funded by National Research Foundation Singapore (NRF) (1102-IRIS-14-02). This work, including the efforts of Boon Fei Tan and Janelle R. Thompson, was funded by Singapore-MIT Alliance for Research and Technology Centre (SMART).

REFERENCES

1. Carmichael WW. 2001. Health effects of toxin-producing Cyanobacteria: "the CyanoHABs." *Hum Ecol Risk Assess* 7:1393–1407. <http://dx.doi.org/10.1080/20018091095087>.
2. Zakaria M. 2016. Cyanobacterial toxins in water sources and their impacts on human health, p 120–149. In McKeown AE, George B (ed), *Impact of water pollution on human health and environmental sustainability*. IGI Global, Hershey, PA.
3. Catherine Q, Susanna W, Isidora E-S, Mark H, Aurélie V, Jean-François H. 2013. A review of current knowledge on toxic benthic freshwater cyanobacteria—ecology, toxin production and risk management. *Water Res* 47:5464–5479. <http://dx.doi.org/10.1016/j.watres.2013.06.042>.
4. Rippka R, Deruelles J, Waterbury JB, Herdman M, Stanier RY. 1979. Generic assignments, strain histories and properties of pure cultures of cyanobacteria. *Microbiology* 111:1–61. <http://dx.doi.org/10.1099/00221287-111-1-1>.
5. Anderson RA, Kawachi M. 2005. Traditional microalgae isolation techniques, p 83–100. In Anderson RA (ed), *Algal culturing techniques*. Elsevier Academic Press, New York.
6. Bolch CJS, Blackburn SI. 1996. Isolation and purification of Australian isolates of the toxic cyanobacterium *Microcystis aeruginosa* Kütz. *J Appl Phycol* 8:5–13. <http://dx.doi.org/10.1007/BF02186215>.
7. Mohamed Nor NH, Tan BF, Te SH, Thompson JR, Gin KY. 2016. Draft genome sequence of *Cylindrospermopsis* sp. strain CR12 extracted from the minimetagenome of a nonaxenic unialgal culture from a tropical freshwater lake. *Genome Announc* 4(1):e01726-15. <http://dx.doi.org/10.1128/genomeA.01726-15>.
8. Kang DD, Froula J, Egan R, Wang Z. 2015. MetaBAT, an efficient tool for accurately reconstructing single genomes from complex microbial communities. *PeerJ* 3:e1165. <http://dx.doi.org/10.7717/peerj.1165>.
9. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <http://dx.doi.org/10.1101/gr.186072.114>.
10. Tan B, Charchuk R, Li C, Nesbø C, Abu Laban N, Foght J. 2014. Draft genome sequence of uncultivated *Firmicutes* (*Peptococcaceae* SCADC) single cells sorted from methanogenic alkane-degrading cultures. *Genome Announc* 2(5):e00909-14. <http://dx.doi.org/10.1128/genomeA.00909-14>.
11. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). *Nucleic Acids Res* 42:D206–D214. <http://dx.doi.org/10.1093/nar/gkt1226>.
12. Markowitz VM, Chen IM, Palaniappan K, Chu K, Szeto E, Pillay M, Ratner A, Huang J, Woyke T, Huntemann M, Anderson I, Billis K, Varghese N, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG 4 version of the integrated microbial genomes comparative analysis system. *Nucleic Acids Res* 42:D560–D567. <http://dx.doi.org/10.1093/nar/gkt963>.
13. Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <http://dx.doi.org/10.1093/nar/gkv437>.