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1	Original	article
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2 Understanding the legacy of widespread population translocations on the post-glacial genetic

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3 structure of the European beech, Fagus sylvatica L.
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5	M. J. Sjölund ^{1,2} , P. González-Díaz ¹ , J. J. Moreno-Villena ^{1,3} , and A. S. Jump ^{1,4}
6	¹ Biological and Environmental Sciences, Faculty of Natural Sciences, University of Stirling,
7	Stirling, FK9 4LA, UK. ² Current address: Science and Advice for Scottish Agriculture (SASA),
8	Roddinglaw Road, Edinburgh, EH12 9FJ, UK. ³ Current address: Department of Animal and
9	Plant Sciences, University of Sheffield, Western Bank, Sheffield, S10 2TN, UK. 4 Centre de
10	Recerca Ecològica i Aplicacions Forestals (CREAF), Campus UAB, Edifici C. E-08193, Bellaterra,
11	Barcelona, Spain.
12	
13	Corresponding author: M. J. Sjölund
14	Email: jennifer.sjolund@sasa.gsi.gov.uk
15	Address: Diagnostics, Wildlife and Molecular Biology (DWMB), Science and Advice for Scottish
16	Agriculture (SASA), Roddinglaw Road, Edinburgh, EH12 9FJ, UK.
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19	Natural colonisation signals persist despite beech forest translocation
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27 ABSTRACT

Aim Human impacts have shaped species ranges throughout the Holocene. The putative native range of beech, *Fagus sylvatica*, in Britain was obscured by its late post-glacial arrival and subsequent extensive management. We sought to differentiate the interacting effects of post-glacial colonization and anthropic impacts on the current genetic structure and diversity of beech by contrasting phylogeographic signals from putatively natural and translocated populations.

34 **Location** Samples were obtained from 42 sites throughout Great Britain.

Methods Chloroplast and nuclear microsatellite marker data were interpreted alongside palynological, historical and anecdotal evidence. Genetic structure was analysed using individual-based Bayesian assignment methods and colonization history was analysed using an approximate Bayesian computation framework.

39 Results Phylogeographic patterns suggested contemporary forests originated from putative 40 native south-eastern populations. High haplotypic diversity was found near the entry point of 41 beech into Britain. Cryptic signals of isolation-by-distance persisted in the putative native 42 range, together with higher levels of gene diversity in nuclear markers. Weak regional nuclear 43 genetic structure suggested high levels of contemporary gene flow throughout the country.

Main conclusions Genetic patterns driven by natural colonization persist despite widespread
 anthropic intervention. Forests in northerly regions were established from forests in the
 putative native range, diminishing the credibility of any present boundary between the native
 and non-native range of beech in Britain.

48 Key words Anthropogenic, Britain, colonization, *Fagus sylvatica*, gene flow, microsatellites,
49 phylogeography, post-glacial.

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52

53 INTRODUCTION

54 The migration of species during the Holocene (Taberlet et al., 1998) coincided with substantial human impact (Kalis & Merkt, 2003; Kalis et al., 2003) shaping species 55 56 contemporary ranges. Historic range limits have been examined from an ecological 57 perspective and post-glacial plant migrations from a phylogeographic perspective (Comes & Kadereit, 1998; Hewitt, 2000; Jump et al., 2010; Magri, 2010). However, many natural 58 59 systems have been under profound and persistent anthropic influence, which has shaped 60 species distributions and the genetic composition of their component populations (Bradshaw, 61 2004; Alessa & Iii, 2008; Schaberg et al., 2008). The influence of forest management, through 62 the selective removal of genotypes and the translocation of plant material, can impact 63 genetic diversity and its spatial distribution within populations and across regions (Savolainen 64 & Kärkkäinen, 1992; Bradshaw, 2004; Alessa & Iii, 2008; Schaberg et al., 2008). However, 65 despite widespread human influences, native forests might retain genetic signals from past 66 population distribution due to natural regeneration of local stock in combination with the 67 long life of individual trees (Petit & Hampe, 2006; Bradshaw, 2004), thereby allowing natural 68 ranges to be detected.

69

70 Given its many uses, including timber, fuel and fodder, the European beech, Fagus sylvatica 71 L., has experienced a long history of management (Nocentini, 2009; Read et al., 2010; 72 Packham et al., 2012) including wide-scale historical translocations throughout Great Britain. 73 Anecdotal, historical and palynological evidence suggest that the native range of beech was 74 limited to south-east England (Rackham, 1980; Pott, 2000; Packham et al., 2012). Despite this 75 commonly held view, this species has been classified as native throughout mainland Great 76 Britain, due to insufficient evidence available to accurately define the native range (Preston 77 et al., 2002). At the regional scale, the species range is believed to be under broad climatic 78 control (Huntley et al., 1989) and in Great Britain, beech has naturalised further north than its

79 presumed native range in the south-east.

80

81 Pollen records from Great Britain indicate that beech migrated into south-east England, with 82 its first establishment just before 3000 BP. Beech maintained a steady rate of spread of 83 100-200 m per year, whereas the majority of other tree species displayed a decrease in the 84 rate of spread. The relatively constant rate of spread suggests that beech had not reached its natural climatic limit by 1000 BP (Birks, 1989). Human intervention might have manipulated 85 86 the species' range before it reached its climatic range limit (Watt, 1931; Packham et al., 87 2012). Historical records suggests the existence of native populations that occur further north 88 than the predicted range as suggested by pollen evidence (Rackham, 1980). However, 89 confirmation of the species' presence in the pollen record does not directly indicate native 90 origin of forests that exist today. Beech therefore provides a valuable model for 91 understanding how human intervention through translocation of material might impact the 92 genetic structure and diversity of natural populations.

93

94 Studies that aim to determine the 'native status' of a species in a particular region have 95 hypothesised that introduced populations are genetically depauperate as they originate from 96 limited source propagules (Stone & Sunnucks, 1993; Fuentes-Utrilla et al., 2014). 97 Consequently, we sought to determine the interacting effects of past migration and anthropic 98 impacts on the current genetic structure and diversity of beech by assessing if a 99 phylogeographic signal of natural colonization can be identified in its putative native range. 100 We used a combination of conservative, maternally-inherited chloroplast markers (Reboud & 101 Zeyl, 1994; Magri et al., 2006) and highly variable nuclear markers to explore regional trends 102 in genetic variation in combination with historical and palynological data. Extensive sampling 103 was carried out in both the putative native and non-native range across Great Britain. We 104 hypothesised that; (1) patterns associated with natural colonization, such as spatial genetic

105 structure, should persist in the putative native range, and putative native sites will show high 106 genetic similarity; (2) patterns driven by natural colonization should be absent in the putative 107 non-native range due to extensive translocations of plant material by humans outside the 108 putative native range; (3) putative non-native sites should display lower levels of haplotypic 109 and genotypic diversity and greater inter-population divergence, compared to putative native 110 sites, due to the high levels of genetic drift associated with the translocation of a limited 111 number genotypes. We refer to sites as 'putative native' and 'putative non-native' as the a112 priori assigned origins of sites used in our study are purely for analytical purposes and should 113 not be taken as precedent.

114

115 MATERIALS AND METHODS

116 Study species

117 Fagus sylvatica L. is a broadleaved, monoecious, primarily outcrossing tree, with pollen 118 dispersed by wind and seeds dispersed by gravity and animals (Fig 1.; Wagner et al., 2010; 119 Packham et al., 2012). It covers approximately 14 million ha, forming the dominant forest 120 type in much of mainland Europe. With the exception of Great Britain, the distribution of beech is primarily climatically limited owing to the species' drought and late frost 121 122 susceptibility (Watt, 1923; Peterken & Mountford, 1996). Beech trees reach approximately 123 300 years of age, commencing flowering typically between 40 to 80 years (Firbas & Losert, 124 1949).

125

Palaeobotanical and genetic data indicate central and northern European populations were colonized from source populations in southern France, eastern Alps-Slovenia-Isteria and potentially Moravia and southern Bohemia. Residual populations in the Iberian, Italian and Balkan refugia are believed to have expanded relatively late and did not contribute significantly to the colonization of central and northern Europe (Magri *et al.*, 2006). Recent

evidence found in Denmark suggests that post-glacial colonization was aided by occasional long-distance dispersal events, leading to the establishment of beech and other temperate tree species ahead of their main colonization fronts (Overballe-Petersen *et al.*, 2012) and might have contributed significantly to the observed rates of spread of the species (Feurdean *et al.*, 2013).

136

137 Study sites

Forty-two populations were sampled across Great Britain (Fig. 2). Using a combination of historical records, palynological, and anecdotal evidence, study sites were designated *a priori* as stands of putative native or putative non-native origin (see Table S1.1 in Appendix S1 in Supporting Information). In each site, leaf samples were collected from 20 mature trees within a 10 ha area, preferentially sampling the oldest trees determined by using diameter at breast height (DBH) as a proxy for age. All samples were geo-referenced using a GARMIN 62s handheld GPS (GARMIN, Southampton, UK).

145

146 Molecular analysis

DNA was obtained from the silica gel dried leaf samples and isolated using the QIAGEN 147 148 DNeasy 96 Plant Kit (QIAGEN, Venlo, Netherlands). Out of 840 samples, 802 individuals were 149 successfully genotyped using four Chloroplast DNA (CpDNA) microsatellite markers (ccmp4, 150 ccmp7 (Weising & Gardner 1999), cmcs3, and cmcs12 (Sebastiani et al., 2004)). Chloroplast 151 markers were combined in one PCR multiplex, FSCplex, using 10ng of template DNA and the 152 QIAGEN Type-it Microsatellite PCR Kit with the following primer concentrations, ccmp4 at 0.5 153 μ M, ccmp7 at 0.5 μ M, cmcs3 at 3 μ M, and cmcs12 at 3 μ M. Annealing temperature was set to 154 55°C, with a total PCR reaction volume of 10µl. 837 individuals were successfully genotyped 155 using 13 nuclear microsatellite markers (fs1-03, fs1-15, fs3-04, fs4-46, fcm5 (Pastorelli et al., 156 2003), mfc7 (Tanaka et al., 1999), mfs11 (Vornam et al., 2004), sfc0007-2, sfc0018, sfc0036,

sfc1143, sfc1061, sfc1063 (Asuka *et al.*, 2004) processed in three multiplexes as detailed in
Sjölund & Jump (2015). A total of three chloroplast microsatellite loci were used, excluding
cmcs12 as it was monomorphic. Fragment analysis was performed on an ABI 3730 (Applied
Biosystems, Bleiswijk, Netherlands) and allele scoring on GENEMARKER 2.4.0 (SoftGenetics,
State College, PA, U.S.A).

162

163 Scoring errors and null alleles in nuclear loci were checked using MICRO-CHECKER (Van 164 Oosterhout et al., 2004). Analyses presented exclude fs4-46, fcm5, and fs1-15 due to null 165 allele presence and use a total of 10 nuclear microsatellite loci. Gametic disequilibrium was tested between pairs of nuclear loci using FSTAT 2.9.3.2 (Goudet, 1995), identifying 166 167 significant associations between loci by randomly associating genotypes at pairs of loci 1100 168 times, using a 5% nominal level after Bonferroni correction. The multilocus average error 169 rates were 0.0% for the 3 chloroplast loci and 0.4% for the 10 nuclear loci included in analysis. 170 The error rate per locus was calculated as the number of erroneously assigned loci over 45 171 repeated samples.

172

173 Measuring genetic diversity and structure using nuclear and chloroplast markers

174 Estimators of genetic diversity were prefixed with 'c' for chloroplast and 'n' for nuclear. 175 Multilocus estimates of haplotypic and genotypic diversity were mapped in ARCMAP 10 (ESRI 176 software) against the pollen isochrones from Birks' (1989) map for the rational limit of beech 177 pollen. Chloroplast haplotypic diversity was measured as the number of haplotypes (CH_N) and 178 the number of private haplotypes (cH_p) . Nuclear genetic diversity was measured as rarefied 179 allelic richness (nA_R) (Petit et al., 1998), gene diversity corrected for sample size (nH_S) (Nei, 180 1978), and the inbreeding coefficient (nF_{15}) with P-values derived from 10,000 permutations 181 of gene copies within individuals per site (Weir and Cockerham 1984), calculated in SPAGeDi 182 1.4b (Hardy & Vekemans, 2002), and rarefied private allelic richness (nA_{P}) calculated in ADZE 183 1.0 (Szpiech *et al.*, 2008). The minimum number of gene copies (*k*) used for rarefication 184 analysis of nA_R and nA_P is 38. To map nuclear genetic differentiation, for each site we 185 calculated the percentage of total sites that it was significantly differentiated from (i.e. 186 percentage of differentiated sites, *nDS* (%)), based on nF_{ST} values (Weir & Cockerham, 1984). 187 nF_{ST} values were obtained from pairwise tests of genetic differentiation not assuming 188 Hardy-Weinberg, with significances determined for a 5% nominal level after Bonferonni 189 correction in FSTAT 2.9.3.2 (Goudet, 1995).

190

191 To test for differences of nuclear-based measurements, nA_R , nH_S , nF_{IS} and nF_{ST} among a priori 192 determined groups of putative native and putative non-native sites, we performed 193 permutation tests using FSTAT 2.9.3.2 (Goudet, 1995). The difference between putative 194 native and putative non-native groups in the remaining estimators nA_P and cN_H were tested 195 using the non-parametric Mann-Whitney U test. To quantify the distribution of variation 196 displayed by the maps of nuclear genetic diversity and chloroplast haplotypes (considering 197 haplotype distances) we tested both marker sets in a hierarchical analysis of molecular 198 variance (AMOVA) performed in ARLEQUIN 3.5.1.2 (Excoffier & Lischer, 2010) with sites 199 grouped into potential stand origin, putative native or putative non-native.

200

201 Testing isolation-by-distance with nuclear markers

Subsets of sites with putative native and putative non-native origin were tested for isolation-by-distance following Rousset (1997) using nuclear markers in SPAGeDi 1.4b (Hardy & Vekemans, 2002) with significance of the slope of the correlation of the natural log of the linear spatial distance (Ln(Spatial Distance (km)) against $F_{ST}/(1-F_{ST})$ was computed after 10,000 permutations of sites among locations. IBD was analysed with and without the continental datasets to check whether a geographical cline in allelic frequencies was influencing clustering (Guillot *et al.*, 2009). To test for geographic gradients in genetic

diversity, we performed non-parametric corrected Spearman's Rank tests on all genetic
diversity estimators and all sites, including separate tests on subsets of putative native and
putative non-native sites.

212

213 Analysing population clusters

214 Individual-based Bayesian assignment methods were performed using data from nuclear loci 215 in STRUCTURE 2.3.4. (Pritchard et al., 2000) using the correlated allele frequency model 216 (Falush et al., 2007), the admixture ancestry model, and site location a priori (LOCPRIOR 217 option) to improve the detection of weak population structure (Hubisz et al., 2009). No stand 218 origin was included a priori in cluster analysis. To examine relationships between British 219 samples and putative colonization sources in continental Europe, we included nuclear 220 microsatellite data from a subset of 150 samples collected from native beech forests in 221 France, Germany and Italy as detailed in Sjölund & Jump (2015). Analysis without continental 222 samples revealed a similar structure in Britain to that found with continental samples, 223 therefore we present the data including continental samples to set the results in context. K 224 was set from 1 to 20, with 10 runs each. Runs consisted of 500,000 Markov chain Monte 225 Carlo (MCMC) iterations with a burn-in period of 100,000. To observe the consensus in 226 number of clusters in the data, we plotted the log probability of the data (LnP(D)), identifying 227 K where log likelihood values converged. Individual Q-matrices were computed in CLUMPP 228 1.1.2 (Jakobsson & Rosenberg, 2007), with graphics created in DISTRUCT 1.1 (Rosenberg, 229 2004). As results indicated cryptic genetic structure in the putative native range, we 230 performed a subsequent analysis on a subset of the 17 putative native sites (i.e. using the 231 putative stand origin a priori) under the same conditions as the main model and excluding 232 continental samples to test for further population sub-structure.

233

234 Demographic history

235 We used an approximate Bayesian computation (ABC) framework implemented in DIYABC 236 2.1.0 (Cornuet et al., 2010) to explore scenarios of demographic history that were likely to 237 have generated current regional genetic structure of beech in Britain according to the results 238 obtained from analyses of regional genetic structure, combined with palynological and 239 historical information. Populations were grouped regionally according to Birks' (1989) 240 isochrones, with British populations grouped into southern, central and northern 241 populations, and populations in Italy, France and Germany grouped into a continental 242 population (see Fig S1.1 in Appendix S1 in Supporting Information). Three scenarios were 243 tested (Fig. 6): Scenario 1 assumes natural colonization of beech in southern and central 244 populations from continental Europe, and northern populations originating from continental 245 European stock (i.e. beech is established in northern Britain by anthropogenic translocation 246 from the continent). Scenario 2 assumes natural colonization of southern and central 247 populations from continental Europe, and northern populations originating from southern 248 and central stock (i.e. northern populations are established from material derived from the 249 putative native region). Scenario 3 assumes natural colonization of southern and central 250 populations from continental Europe, and northern populations originating from an 251 admixture of southern, central and continental populations (i.e. some northern populations 252 are established from southern and central stock, the putative native region, and some from 253 continental stock).

254

Prior parameters for effective population sizes and timing of events were defined based on knowledge of beech colonization dynamics from palynological and historical information. Prior parameter distributions were uniform and bounded between $10-10^4$ for the effective population size of southern, central and northern Britain, and $10-10^5$ for continental Europe. Population divergence time priors ranged between 10-50 for t1, 25-100 for t2 and 25-200 for t3, with the additional setting t3>t2, and t2>t1. Priors for admixture rates ranged between

261 0.01 and 0.99. Nuclear microsatellite loci were assumed to follow a Generalized Stepwise 262 Mutation model and a uniform prior was assumed for the mean microsatellite mutation rate bounded between 10⁻³ and 10⁻⁴. 36 summary statistics were used for the ABC analysis. Three 263 264 single sample statistics were used (mean number of alleles, mean Nei's genetic diversity 265 index (Nei, 1987) and mean allele size variance), and three between-sample statistics (mean 266 allele size variance, F_{ST}, and mean index of classification (Rannala & Mountain, 1997; Pascual 267 et al., 2007). Following the methods outlined by Cornuet (2010), type I and type II errors were 268 estimated to evaluate the power of the model. The selected scenario was used to estimate 269 posterior distribution of demographic parameters.

270

271 **RESULTS**

272 Geographic trends in genetic diversity

273 Three variants were detected for chloroplast loci, ccmp4, ccmp7 and cmcs3. The number of 274 haplotypes (cH_N) within sites ranged from one to four, with a total of seven haplotypes 275 recorded (Table 1, Fig. 2; see Fig. S2.1 in Appendix S2 in Supporting Information). Haplotype 276 (A) was present in all sites and was the dominant haplotype within sites. Haplotype diversity 277 was highest in the putative native sites BLE ($cH_N = 5$) and LUL ($cH_N = 3$), the two most 278 south-easterly British sites. Six out of seven haplotypes (A to F) were represented collectively 279 in these two sites. Two private haplotypes, F and G, were present in sites BLE and DEV, 280 respectively, in single individuals. Significant genetic structuring was found in chloroplast and 281 nuclear markers between sites using AMOVA (Table 2), although no significant difference was 282 found between groups of putative native and putative non-native sites. The remaining 283 variation was present within individuals.

284

285 Multilocus estimates of genetic diversity were obtained for 10 nuclear loci, with an average 286 number of 13.3 alleles per locus, and a maximum of five to 30 alleles per locus (Table 1 and

287 Fig. 3). All nuclear loci were under gametic equilibrium. Rarefied allelic richness (nA_R) varied 288 from 4.58 to 7.19 with rarefied private allelic richness (nA_p) ranging from 0 to 0.156. Gene diversity estimates ranged from 0.606 to 0.750. Putative native site WYC and putative 289 non-native MAB displayed a significant homozygote excess (WYC nF_{IS} = 0.088, P < 0.05; MAB 290 nF_{IS} = 0.089, P < 0.05), whilst a heterozygote excess was found in sites BEE and CRA, both 291 292 putative non-native sites (BEE nF_{IS} = -0.152, P < 0.001; CRA nF_{IS} = -0.086, P < 0.05). The 293 percentage of significantly differentiated sites [nDS (%)] varied greatly, for example, site WYT 294 was significantly differentiated to 4.9% of sites, whilst site BEE was significantly differentiated 295 to 95.1% of all sites.

296

297 Differences between groups of putative native and non-native sites

Significantly higher levels of gene diversity (nH_s) were found in putative native sites, compared to putative non-native sites; H_s : putative native 0.708, putative non-native 0.690 (P< 0.05) (Table 1). The general trend of lower gene diversity in sites outside of the putative native range can be seen in the map for gene diversity in Fig. 3. No significant differences were found for other estimators; nA_R : putative native 6.225 v 6.252 putative non-native (P = 0.87), nA_P : 0.053 v 0.037 (U(40) = 245, Z = 0.84, P = 0.41), nF_{IS} : 0.022 v -0.005 (P = 0.07), nF_{ST} : 0.024 v 0.021 (P = 0.59), and cN_H : 1.5 v 1.4 (U(40) = 208, Z = -0.155, P = 0.89).

305

306 Isolation-by-distance

Significant isolation-by-distance was found in Britain amongst putative native sites (refer to Fig. 4C: slope = 0.0085, $R^2 = 0.10$, P < 0.05). IBD was not significant amongst putative nonnative sites, nor was it significant in all sites combined with and without the continental subset (refer to Fig. 4, A: British sites only (slope = 0.0011, $R^2 < 0.01$, P = 0.22), B: British and continental sites (slope = 0.0032, $R^2 = 0.03$, P = 0.06), D: putative non-native sites (slope = 0.0030, $R^2 = 0.02$, P = 0.09). Within the putative native sites, there was a significant reduction in haplotype number following an east to west gradient (rho = 0.70, P < 0.01; Fig. 2). This effect disappeared when all sites were analysed together (rho = 0.18, P = 0.241), and was not found in putative non-native sites alone (rho = -0.02, P = 0.90). Preliminary analysis revealed no significant correlations between nuclear genetic diversity estimators and geographic variables (i.e. latitude and longitude) overall sites and within subsets of putative native and putative non-native sites (see Table S2.1 in Appendix S2 in Supporting Information).

319

320 Detection of further regional structuring using nuclear markers

Mean log-likelihood values for each of the STRUCTURE runs on samples from Britain including the continental samples, gradually ceased to converge after K = 2, after which they began to plateau (see Fig. S3.1 in Appendix S3 in Supporting Information). Examination of Q-matrices indicated that K = 3 provided meaningful biological clusters that followed a regional distribution (Fig. 5). Although values of Ln P(D) for K = 3 varied more than K = 2, assignment of individuals to clusters were congruent between runs (see Fig. S3.2 in Appendix S3 in Supporting Information).

328

329 Several sites throughout Britain contained highly admixed individuals, whereas individuals 330 from continental sites displayed homogenous levels of admixture among individuals within 331 site. Organising the Q-matrix according to Birks' (1989) isochrones in approximate geographic 332 order revealed some consistency in cluster assignment between neighbouring sites (Fig. 5). 333 There appeared to be a geographic gradient in the continental clusters with the predominant 334 cluster being blue in ITA, to grey in FRA and GER, with the introduction of the red cluster in 335 GER. TAN appears to have the highest assignment to the blue cluster out of any other sites in 336 Britain. Individuals from Britain were largely assigned to the grey and red clusters. The red 337 cluster appeared to be associated with sites within the putative native range and the putative 338 non-native sites in the south-west (DEV, GOL, HEM, and BRI). Similar patterns were observed for K = 2 and K = 3 when analysing a subset of British samples alone and a subset of putative native sites, both revealing no further population sub-structuring (see Fig. S3.3 and Fig. S3.4 in Appendix S3 in Supporting Information).

342

343 Model-based inference using ABC analyses indicated that scenario 2 was supported with 344 maximum probability compared to around zero support for scenarios 1 and 3 (Fig. 6). This is 345 indicative of a gradual colonization of beech from continental Europe and expanding 346 northwards into and through Britain, with northern populations being derived from the 347 putative native range of beech in Britain. Confidence in scenario choice was high, with low 348 error rates (type 1 = 0.001-0.054, type 2 = 0-0.054). For scenario 2, median values for the 349 effective population size were 6830, 4920, 6270 and 25100 for N1 (northern Britain), N2 350 (central Britain), N3 (southern Britain) and N4 (Continental Europe). The median values for 351 divergence events corresponded to 16.9, 26.5 and 170 number of generations for t1, t2 and 352 t3.

353

354 **DISCUSSION**

Analysing genetic data according to palynological and historical evidence allowed us to tease apart the post-glacial history of beech in Britain despite the prolonged human impacts on this species. Patterns of past colonization dynamics persisted in putative native sites, while phylogeographic patterns and modelled demographic history support the hypothesis that throughout Britain, beech forests are largely derived from native stock originating from trees that colonized the island during the Holocene.

361

362 Native origins of beech in Britain

363 The distribution of chloroplast haplotypes and modelled demographic history strongly 364 support the colonization of northern beech populations from stock originating from the

365 putative native range of beech. The distribution of haplotypes in Britain matched the 366 expected phylogeographic signal of postglacial colonization, with the highest number of 367 haplotypes found in south-eastern sites, LUL and BLE (Table 1; Fig. 2) in close proximity to 368 the purported entry point of beech migration into Britain (Birks 1989). All the haplotypes, 369 except one (G), were represented in these sites, suggesting that the majority of Britain's 370 beech forests were colonized from native stock. Similar south to north patterns in haplotype 371 distribution have been found for other European tree taxa, including Alnus, Quercus and 372 Populus (King & Ferris et al., 1998; Petit et al., 2002a; Cottrell et al., 2005 along with genetic 373 clines indicative of gradual recolonization of Quercus robur and Quercus petraea in Ireland 374 (Kelleher et al., 2004). The loss of haplotype diversity occurring away from proposed entry 375 sites can be a result of founder effects induced by the progressive movement of the 376 migration front (Excoffier et al., 2009). The clinal pattern of haplotype diversity in F. sylvatica 377 in Britain is in contrast with Q. robur and Q. petraea, which displayed a highly clumped 378 distribution of haplotypes indicative of long-distance dispersal events (Cottrell et al., 2002a) 379 following the much earlier entry of these oaks into Britain (Birks 1989). There is little similar 380 evidence of long-distance dispersal events in F. sylvatica in Britain. site BLE harboured one of 381 the two private haplotypes found while haplotype (G) in site DEV may have arisen from a 382 single base mutation in haplotype A or a long-distance dispersal event (Overballe-Petersen et 383 al., 2012).

384

The high number of haplotypes ($cH_N = 7$) is in contrast to that found by Magri *et al.* (2006) who report one haplotype throughout Britain. This is likely a consequence of our larger sample size, the significant partitioning of haplotype variation between sites, and polymorphism in cmcs3. Restricting analysis to ccmp4 and ccmp7 used by Magri *et al.* (2006) reduces haplotype number to four (data not shown) and matches the regional trend seen with all three loci, with diversity restricted to south-eastern sites. The average levels of

391 rarefied allelic richness using nuclear markers in Britain $(nA_{R} = 6.25\pm0.08)$ were lower than 392 those reported in studies using some of the same microsatellite markers, approximately 393 ranging from 8.2 to 18.2 in other studies (Jump & Peñuelas, 2006; Buiteveld et al., 2007; 394 Sjölund & Jump, 2015). Sites in Britain also displayed lower levels of rarefied private allelic 395 richness ($nA_P = 0.044 \pm 0.007$) compared to sites in continental Europe sampled by Sjölund and 396 Jump (2015) where values for nA_{P} ranged between 1.51 and 2.36. Overall levels of gene 397 diversity were similar to those found in other studies ($nH_s = 0.696 \pm 0.004$) (Jump & Peñuelas, 398 2006; Buiteveld et al., 2007; Oddou-Muratorio et al., 2008; Sjölund & Jump, 2015).

399

400

401 Genetic variation between groups of different a priori stand origins

402 Isolation-by-distance occurs when the genetic differentiation between individuals or 403 populations increases with geographic distance (Wright, 1940). In plants, this is primarily a 404 consequence of restricted gene flow via seed or pollen (Loveless & Hamrick, 1984). Although 405 beech is assumed to show high levels of gene flow as a wind-pollinated tree, it displays 406 significant structuring at local (Chybicki et al., 2009; Jump et al., 2012; Piotti et al., 2013; 407 Sjölund & Jump, 2015) and regional scales (Jump & Peñuelas, 2006; de Lafontaine et al., 408 2013). In agreement with the significant genetic structuring found in natural populations of 409 beech, putative native sites displayed a weak but significant trend of IBD based on nuclear 410 loci, which was absent in putative non-native sites, with the addition of all sites obscuring the 411 IBD signal (Fig. 4).

412

Populations of beech in France with relatively recent colonization histories displayed stronger
IBD compared to southern refugial populations in France (de Lafontaine *et al.*, 2013). As
beech only arrived in Britain around 3000 BP (Birks, 1989), IBD in the putative native range is
likely driven by relatively recent colonization dynamics. In contrast, widespread

translocations are likely to have prevented the development of IBD between putative non-native populations, possibly due to the anthropic movement of plant material throughout the country. A similar result was found by Leonardi *et al.* (2012) in beech populations of different levels of fragmentation in central Italy. IBD in less fragmented nonmarginal populations was obscured when populations from marginal, fragmented populations where included. This finding was interpreted as a result of intense genetic drift in fragmented populations.

424

425 We found a significant decrease in gene diversity (nH_s) in putative non-native sites, 426 suggesting a reduction in genetic diversity due to founder effects, although the magnitude of 427 this difference may have been limited by high gene flow associated with wind pollination. 428 However, no significant difference was found for rarefied allelic richness between sites of 429 different origins. Allelic richness is expected to be more sensitive to reductions in effective 430 population sizes, as rare alleles, which do not contribute considerably to gene diversity, are 431 more likely to be lost first (Nei et al., 1975). Low levels of allelic richness throughout Britain 432 suggest a significant proportion was lost during post-glacial colonization, probably due to 433 founder effects. The lack of a pattern in allelic richness between putative native and putative 434 non-native sites may be due to a lack of sensitivity of the analysis arising from the 435 comparison of two already limited gene pools. In agreement with theoretical predictions, 436 gene diversity throughout Britain displayed similarly high levels to that found on the continent (Jump & Peñuelas, 2006; Buiteveld et al., 2007; Oddou-Muratorio et al., 2008; 437 438 Sjölund & Jump, 2015). Although putative native sites displayed the highest average private 439 allelic richness (nA_p) , this difference was not statistically significant.

440

441 Regional patterns of postglacial migration of beech into Britain

442 Within Britain, neighbouring sites displayed congruent levels of admixture with lower levels

443 of admixture in northern, putative non-native sites (Fig. 5). A gradient of admixture marked 444 the transition of continental regions to Britain, with continental sites assigned predominantly 445 to the blue and grey cluster. This is in contrast to that found in Quercus spp. (Petit et al., 446 2002b) and Populus nigra (Cottrell et al., 1997; Cottrell et al., 2002b; Cottrell et al., 2004), 447 where much longer periods of translocations have led to a decrease in genetic differentiation. 448 Individuals from the putative non-native site, TAN, may have arisen from translocation of 449 continental stock as it displayed the highest levels of assignment to the blue cluster in Britain, 450 similar to individuals in FRA. Individuals from the south of Britain displayed a relatively higher 451 probability of assignment to the red cluster, which was generally associated with the putative 452 native range, in addition to the south-west region, including putative non-native sites DEV, 453 GOL, HEM, and BRI. There appeared to be a transition between the assignment of individuals 454 to the red and grey cluster that occurred in proximity to the border for the 1000 BP 455 isochrone, with a tendency towards assigning individuals to the red cluster in regions pre-456 1000 BP. Only one putative native site, CWw, occurred outside of the border of the 1000 BP 457 limit and was predominantly assigned to the red cluster. However, CWw is only 15km away 458 from the 1000 BP isochrones.

459

460 Conclusion

461 Using multiple data sources, we were able to identify signals of natural colonization in a 462 forest system that has been heavily impacted by humans. Our results build upon existing 463 palynological and historical evidence to substantially advance our understanding of the 464 contemporary genetic impact of past population translocation. Chloroplast markers suggest 465 that the majority of trees sampled were derived from putative native stock either through 466 natural regeneration or translocation, whilst nuclear markers confirm the persistence of 467 signals of the natural colonization of beech in Britain in putative native sites. Although cryptic 468 genetic signals of population expansion remain in the putative native range of beech, we

469 caution against using this evidence as a means to classify stand origins, since gene flow 470 between neighbouring regions essentially blurs the borders of the native range. Warming 471 climate is increasing the productivity of this species in more northerly parts of its range while 472 decreasing growth and reproduction are predicted in the south. Given that our data suggest 473 native origin for most of Britain's populations, it is paramount that climate-induced range 474 shifts are considered in management plans and this species managed as a whole, irrespective 475 of regional boundaries.

476

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484 **REFERENCES**

- Alessa L. & Iii F.S.C. (2008) Anthropogenic biomes: a key contribution to earth-system science. *Trends in Biotechnology*, 23, 529–531.
- 487 Asuka Y., Tani N., Tsumura Y., & Tomaru N. (2004) Development and characterization of
- 488 microsatellite markers for *Fagus crenata* Blume. *Molecular Ecology Notes*, **4**, 101–103.
- Birks H.J.B. (1989) Holocene isochrone maps and patterns of tree-spreading in the British
 Isles. *Journal of Biogeography*, **16**, 503–540.
- 491 Bradshaw R.H.W. (2004) Past anthropogenic influence on European forests and some
- 492 possible genetic consequences. *Forest Ecology and Management*, **197**, 203–212.

- 493 Buiteveld J., Vendramin G.G., Leonardi S., Kamer K., & Geburek T. (2007) Genetic diversity
- 494 and differentiation in European beech (*Fagus sylvatica* L.) stands varying in

495 management history. *Forest Ecology and Management*, **247**, 98–106.

- 496 Chybicki I.J., Trojankiewicz M., Oleksa A., Dzialuk A., & Burczyk J. (2009) Isolation-by-distance
- 497 within naturally established populations of European beech (*Fagus sylvatica*). *Botany*,
 498 **87**, 791–798.
- Comes H.P. & Kadereit J.W. (1998) The effect of Quaternary climatic changes on plant
 distribution and evolution. *Trends in Plant Science*, **3**, 432–438.
- Cornuet J.M., Ravigné V., & Estoup A. (2010) Inference on population history and model
 checking using DNA sequence and microsatellite data with the software DIYABC (v1.0).
 BMC Bioinformatics, **11**, 401.
- Cottrell J.E., Forrest G.I., & White I.M.S. (1997) The use of RAPD analysis to study diversity in
 British black poplar (*Populus nigra* L. ssp. *betulifolia* (Pursch) W. Wettst. (Salicaceae)) in
 Great Britain. Watsonia, **21**, 305–312.
- 507 Cottrell J., Munro R.C., Tabbener H.E., Gillies A.C.M, Forrest G.I., Deans J.D., & Lowe A.J.

508 (2002a) Distribution of chloroplast DNA variation in British oak (*Quercus robur* and *Q*.

509 *petraea*): the influence of postglacial colonisation and human management. *Forest*

- 510 *Ecology and Management*, **156**, 181–195.
- 511 Cottrell J.E., Tabbener H.E., & Forrest G.I. (2002b) Distribution of variation in British black
- 512 poplar: role for human management. In: van Dam B.C., Bordács S. (Ed.), Genetic
- 513 Diversity in River Populations of European Black Poplar-implications for Riparian Eco-
- 514 system Management. Proceedings of an International Symposium, Szekza'rd, Hungary,
- 515 May 2001. Str. 73-84.

- 516 Cottrell J.E., Krystufek V., Tabbener H.E., Milner A.D., Connolly T., Sing L., Fluch S., Burg K.,
- 517 Lefévre F., Achard R., Bordács S., Gebhardt K., Vornam B., Smulders M.J.M., Vanden
- 518 Broeck A.H., Van Slycken J., Storme V., Boerjan W., Castiglione S., Fossati T., Alba N.,
- 519 Agundez D., Maestro C., Notivol E., Bovenschen J., & Van Dam B.C. (2005) Postglacial
- 520 migration of *Populus nigra* L.: lessons learnt from chloroplast DNA. *Forest Ecology and*
- 521 *Management*, **206**, 71-90.
- Excoffier L., Foll M., & Petit R.J. (2009) Genetic consequences of range expansions. *Annual Review of Ecology, Evolution, and Systematics*, 40, 481–501.

Excoffier L. & Lischer H.E.L. (2010) Arlequin suite ver 3.5: a new series of programs to perform
population genetics analyses under Linux and Windows. *Molecular Ecology Resources*,
10, 564–567.

- Falush D., Stephens M., & Pritchard J.K. (2007) Inference of population structure using
 multilocus genotype data: dominant markers and null alleles. *Molecular Ecology Notes*,
 7, 574–578.
- 530 Feurdean A., Bhagwat S.A., Willis K.J., Birks H.J.B., & Lischke H. (2013) Tree migration-rates:

531 narrowing the gap between inferred post-glacial rates and projected rates. *PLoS*532 *Genetics*, **8**, 1–7.

- Firbas F., Losert H. (1949) *Spät- und nacheiszeitliche Waldgeschichte Mitteleuropas närdlich der Alpen.* Fischer, Germany.
- Fuentes-Utrilla P., Venturas M., Hollingsworth P.M., Squirrell J., Collada C., Stone G.N., & Gil L.
 (2014) Extending glacial refugia for a European tree: genetic markers show that Iberian
 populations of white elm are native relicts and not introductions. *Heredity*, **112**, 105–
 113.

- Goudet J. (1995) FSTAT (version 1.2): A computer program to calculate F-statistics. *Journal of Heredity*, 86, 485–486.
- 541 Guillot G., Leblois R., Coulon A., & Frantz A.C. (2009) Statistical methods in spatial genetics.
 542 *Molecular Ecology*, 18, 4734–4756.
- Hardy O.J. & Vekemans X. (2002) Spagedi: a versatile computer program to analyse spatial
 genetic structure at the individual or population levels. *Molecular Ecology Notes*, 2,
 618–620.
- 546 Hewitt G.M. (2000) The genetic legacy of the Quaternary ice ages. *Nature*, **405**, 907–913.
- 547 Hubisz M.J., Falush D., Stephens M., & Pritchard J.K. (2009) Inferring weak population
- 548 structure with the assistance of sample group information. *Molecular Ecology*549 *Resources*, **9**, 1322–1332.
- 550 Huntley A.B., Bartlein P.J., Prentice I.C., Journal S., Nov N., Huntley B., Bartlein P.J., & Road S.
- 551 (1989) Climatic control of the distribution and abundance of Beech (*Fagus* L.) in Europe
- and North America. *Journal of Archaeological Science*, **16**, 551–560.
- Jakobsson M. & Rosenberg N.A. (2007) CLUMPP: a cluster matching and permutation
- program for dealing with label switching and multimodality in analysis of population
 structure. *Bioinformatics*, 23, 1801–1806.
- 556 Jump A.S., Cavin L., & Hunter P.D. (2010) Monitoring and managing responses to climate
- 557 change at the retreating range edge of forest trees. *Journal of Environmental*
- 558 *Monitoring*, **12**, 1791–1798.

- Jump A.S. & Peñuelas J. (2006) Genetic effects of chronic habitat fragmentation in a windpollinated tree. *Proceedings of the National Academy of Sciences of the United States of*America, 103, 8096–8100.
- 562 Jump A.S., Rico L., Coll M., & Peñuelas J. (2012) Wide variation in spatial genetic structure
- 563 between natural populations of the European beech (*Fagus sylvatica*) and its
- implications for SGS comparability. *Heredity*, **108**, 633–639.
- Kalis A.J. & Merkt J. (2003) Environmental changes during the Holocene climatic optimum in
 central Europe human impact and natural causes. Quaternary Science Reviews, 22, 33–
 79.
- Kelleher C.T., Hodkinson T.R., Kelly D.L., & Douglas G.C. (2004) Characterisation of chloroplast
 DNA haplotypes to reveal the provenance and genetic structure of oaks in Ireland. *Forest Ecology and Management*, **189**, 123–131.
- King, A.R., & Ferris, C. (1998). Chloroplast DNA phylogeography of *Alnus glutinosa* (L.) Gaertn. *Molecular ecology*, 7, 1151-1161.
- 573 De Lafontaine G., Ducousso A., Lefèvre S., Magnanou E., & Petit R.J. (2013) Stronger spatial
- 574 genetic structure in recolonized areas than in refugia in the European beech. *Molecular*575 *Ecology*, **22**, 4397–4412.
- Loveless M.D. & Hamrick J.L. (1984) Ecological determinants of genentic structure in plant
 populations. *Annual Review of Ecology and Systematics*, **15**, 65–95.
- 578 Magri D. (2010) Persistence of tree taxa in Europe and Quaternary climate changes.
- 579 *Quaternary International*, **219**, 145–151.

580	Magri D., Vendramin G.G., Comps B., Dupanloup I., Geburek T., Gömöry D.S., Litt T., Paule L.,
581	Roure J.M., & Tantau I. (2006) A new scenario for the Quaternary history of European
582	beech populations: palaeobotanical evidence and genetic consequences. New
583	Phytologist, 171 , 199–221.
584	Nei M. (1978) Estimation of average heterozygosity and genetic distance from a small
585	number of individuals. <i>Genetics</i> , 89 , 583–590.
586	Nei M. (1987) Molecular evolutionary genetics. Columbia University Press, USA.
587	Nocentini S. (2009) Structure and management of beech (Fagus sylvatica L.) forests in Italy.
588	iForest - Biogeosciences and Forestry, 2 , 105–113.
589	Oddou-Muratorio S., Vendramin G.G., Buiteveld J., & Fady B. (2008) Population estimators or

590 progeny tests: what is the best method to assess null allele frequencies at SSR loci?
591 *Conservation Genetics*, **10**, 1343–1347.

592 Van Oosterhout C., Hutchinson W.F., Wills D.P.M., & Shipley P. (2004) Micro-Checker:

593 software for identifying and correcting genotyping errors in microsatellite data.

- 594 *Molecular Ecology Notes*, **4**, 535–538.
- 595 Overballe-Petersen M. V., Nielsen a. B., Hannon G.E., Halsall K., & Bradshaw R.H. (2012) Long-
- 596 term forest dynamics at Gribskov, eastern Denmark with early-Holocene evidence for
- thermophilous broadleaved tree species. *The Holocene*, **23**, 243–254.
- Packham J.R., Thomas P.A., Atkinson M.D., & Degen T. (2012) Biological flora of the British
 Isles: *Fagus sylvatica*. *Journal of Ecology*, **100**, 1557–1608.

- 600 Pascual M., Chapuis M.P., Mestres F., Balanyà J., Huey R.B., Gilchrist G.W., Serra L., & Estoup
- 601 A. (2007) Introduction history of *Drosophila subobscura* in the New World: a
- 602 microsatellite-based survey using ABC methods. *Molecular Ecology*, **16**, 3069–3083.
- 603 Pastorelli R., Smulders M.J.M., Wastende V., Vosman B., Giannini R., Vettori C., & Vendramin
- 604 G.G. (2003) Characterization of microsatellite markers in *Fagus sylvatica* L . and *Fagus*
- 605 *orientalis* Lipsky. *Molecular Ecology Notes*, **96**, 76–78.
- Peterken G.F. & Mountford E.P. (1996) Effects of drought on beech in Lady Park Wood, an
 unmanaged mixed deciduous woodland. *Forestry*, 69, 125-136.
- Petit R.J., Mousadik A.E.L., & Pons O. (1998) Identifying populations for conservation on the
 basis of genetic markers. *Conservation Biology*, **12**, 844–855.
- 610 Petit J.R., Csaikl U.M., Bordács S., Burg K., Coart E., Cottrell J., Van Dam, B., Deans, J.D.,
- 611 Dumolin-Lapegues S., Fineschi S., Finkeldey R., Gillies A., Glaz I., Goicoechea P.G., Jensen
- J. S., König A. O., Iowe A.J., Madsen S.F., Matyás C., Munro R.C., Olalde, M., Pemonge
- 613 M.H., Popescu F., Slade D., Tabbener H., Taurchini D., De Vries, S.G.M., Ziegenhagen G.,
- 614 & Kremer A. (2002a) Chloroplast DNA variation in European white oaks. Phylogeography
- and patterns of diversity based on data from over 2600 populations. *Forest Ecology and*
- 616 *Management*, **156**, 5–26.
- 617 Petit J.R., Brewer S., Bordács S., Burg K., Cheddadi R., Coart E., Cottrell J., Csaikl U.M., Van
- 618 Dam B., Deans J.D., Espinel S., Fineschi S., Finkeldey R., Glaz I., Goicoechea P. G.,
- Jensen J.S., König A.O., Lowe A.J., Madsen S.F., Mátyás C., Munro R.C., Popescu F.,
- 620 Slade D., Tabbener H., De Vries, S.G.M., Ziegenhagen B., De Beaulieu J-L., & Kremer A.
- 621 (2002b) Identification of refugia and post-glacial colonisation routes of European

- white oaks based on chloroplast DNA and fossil pollen evidence. *Forest Ecology and Management*, **156**, 49-74.
- Petit, R. J., & Hampe, A. (2006). Some Evolutionary Consequences of Being a Tree. *Annual Review of Ecology, Evolution, and Systematics*, **37**, 187–214.
- 626 Piotti A., Leonardi S., Heuertz M., Buiteveld J., Geburek T., Gerber S., Kramer K., Vettori C., &
- 627 Vendramin G.G. (2013) Within-population genetic structure in beech (*Fagus sylvatica* L.)
- 628 stands characterized by different disturbance histories: does forest management
- 629 simplify population substructure? *PloS One*, **8**, e73391.
- 630 Pott R. (2000) Palaeoclimate and vegetation long-term vegetation dynamics in central
- 631 Europe with particular reference to beech. *Phytocoenologia*, **30**, 285–333.
- Preston C.D., Pearman D., Dines T. (2002) *New atlas of the British & Irish flora*. Oxford
 University Press, UK.
- 634 Pritchard J.K., Stephens M., & Donnelly P. (2000) Inference of population structure using
- 635 multilocus genotype data. *Genetics*, **155**, 945–59.
- Rannala B. & Mountain J. (1997) Detecting immigration by using multilocus genotypes.

637 Proceedings of the National Academy of Sciences of the United States of America, 94,
638 9197–9201.

- 639 Reboud X. & Zeyl C. (1994) Organelle inheritance in plants. *Heredity*, **72**, 132–140.
- 640 Rosenberg N.A. (2004) DISTRUCT: a program for the graphical display of population structure.
- 641 *Molecular Ecology Notes*, **4**, 137–138.

- 642 Savolainen O., Kärkkäinen K. (1992) Effect of forest management on gene pools. *New Forests*,
 643 6, 329–345.
- Schaberg P.G., DeHayes D.H., Hawley G.J., & Nijensohn S.E. (2008) Anthropogenic alterations
 of genetic diversity within tree populations: Implications for forest ecosystem resilience. *Forest Ecology and Management*, **256**, 855–862.
- 647 Sebastiani F., Carnevale S., & Vendramin G.G. (2004) A new set of mono- and dinucleotide
 648 chloroplast microsatellites in Fagaceae. *Molecular Ecology Notes*, 4, 259–261.
- 649 Sjölund M.J. & Jump A.S. (2015) Coppice management of forests impacts spatial genetic
- 650 structure but not genetic diversity in European beech (*Fagus sylvatica* L.). *Forest Ecology*
- 651 *and Management*, **336**, 65–71.
- 652 Stone G.N. & Sunnucks P. (1993) Genetic consequences of an invasion through a patchy

653 environment - the cynipid gallwasp Andricus. *Molecular Ecology*, **2**, 251–268.

- 654 Szpiech Z.A., Jakobsson M., & Rosenberg N.A. (2008) ADZE: a rarefaction approach for
- 655 counting alleles private to combinations of populations. *Bioinformatics*, **24**, 2498–2504.
- Taberlet P., Fumagalli L., Wust-Saucy A.G., & Cossons J.F. (1998) Comparative phylogeography
 and postglacial colonization routes in Europe. *Molecular Ecology*, 7, 453–464.
- Tanaka K., Tsumura Y., & Nakamura T. (1999) Development and polymorphism of
- 659 microsatellite markers for *Fagus crenata* and the closely related species , *F* . *japonica*.
- 660 Theoretical and Applied Climatology, **99**, 11–15.
- 661 Vornam B., Decarli N., & Gailing O. (2004) Spatial distribution of genetic variation in a natural

beech stand (Fagus sylvatica L.) based on microsatellite markers. Conservation Genetics,

5, 561–570.

664	Wagner S., Collet C., Madsen P., Nakashizuka T., Nyland R.D., & Sagheb-Talebi K. (2010) Beech
665	regeneration research: From ecological to silvicultural aspects. Forest Ecology and
666	Management, 259 , 2172–2182.

- 667 Watt A.S. (1923) On the ecology of British beechwoods with special reference to their
- regeneration. *Journal of Ecology*, **11**, 1–48.
- Watt A.S. (1931) Preliminary observations on Scottish beechwoods. Introduction and part I. *Journal of Ecology*, **19**, 137–157.
- 671 Weir B.S. & Cockerham C.C. (1984) Estimating F-statistics for population structure. *Evolution*,

672 **38**, 1358–1370.

- 673 Weising K. & Gardner R. C. (1999). A set of conserved PCR primers for the analysis of simple
- 674 sequence repeat polymorphisms in chloroplast genomes of dicotyledonous
- 675 angiosperms. *Genome*, **42**, 9-19.
- 676 Wright S. (1940) Breeding structure of populations in relation to speciation. *The American*
- 677 *Naturalist*, **74**, 232–248.

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680 Supporting Information

- 681 Additional Supporting Information may be found in the online version of this article:
- 682 Appendix S1 Stand origins for study sites
- 683 Appendix S2 Chloroplast and nuclear genetic data
- 684 Appendix S3 STRUCTURE analysis output
- 685

686 Data Accessibility

- 687 All microsatellite and GPS data for this study are available at DataSTORRE: Stirling Online
- 688 Repository for Research Data: http://hdl.handle.net/11667/90
- 689

690 Biosketch

- 691 Alistair Jump's research team has a strong focus on understanding biogeographic impacts of
- 692 past and present environmental changes from population genetics to demography and
- 693 remote sensing and how they interact with human interventions.
- 694 Author contributions: MJS and ASJ designed the research. MJS conducted field-based work.
- 695 MJS, JJM, and PGD conducted lab-based work. MJS and PGD conducted data analysis. ASJ
- 696 supervised the research project. MJS, ASJ, PGD, and JJM wrote the manuscript.
- 697 Editor: Jim Provan
- 698

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703 Tables

704	Table 1 Genetic diversity estimates obtained from nuclear (n) and chloroplast (c) markers
705	for beech in Great Britain. Data from chloroplast (c) markers include; cN , no. of samples; cH_N ,
706	no. of haplotypes, \dagger identifies private haplotypes (CH_P). Data from nuclear (n) markers
707	include; nN , no. of samples; nA_R , rarefied allelic richness; nA_P , rarefied private allelic
708	richness; nH_s , gene diversity; nF_{1s} , inbreeding coefficient; and, $nDS\%$, percentage of
709	significantly differentiated sites. Mean±SE is given per group (Putative Native and Non-native)
710	and overall for genetic diversity estimators. Significant P -values are indicated as * P < 0.05,
711	*** <i>P</i> < 0.001.
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	SITES	CIN	CHN	nN	nA _R	nA _P	nHs	nr _{is}	nDS (%)
Native		321	1.5±0.2	340	6.22±0.13	0.053±0.13	0.707±0.006*	0.022±0.010	46.1±6.4
	FEL	17	2	20	5.17	0.095	0.651	0.057	75.6
	BED	18	1	20	5.95	0.025	0.722	0.059	17.1
SEC 1		17	1	20	6.06	0.008	0.717	-0.040	56.1
	WYC	20	1	20	7.12	0.077 0.694		0.088*	9.8
	LAD 20 1 20 6.72 0.110 0.736		0.033	22.0					
	BUC	20	1	20	5.33	0.000	0.682	0.010	87.8
	CWe	20	1	20	6.64	0.146	0.750	0.055	85.4
	CWw	20	1	20	6.00	0.000	0.710	-0.037	85.4
	MON	19	1	20	6.71	0.156	0.716	-0.013	56.1
	GRE	20	1	20	5.98	0.004	0.712	0.038	26.8
	BUR	19	2	20	6.80	0.109	0.713	-0.003	22.0
:	SAV	16	1	20	6.09	0.001	0.687	0.041	43.9
	LUL	17	3	20	6.09	0.020	0.727	-0.054	29.3
	FRI	20	1	20	6.51	0.087	0.679	0.007	26.8
	BLE	18	4†	20	6.82	0.027	0.721	0.030	24.4
,	WEA	20 2 20 5.86 0.007 0.703		0.070	56.1				
	DEN	20	1	20	5.97	0.024	0.706	0.031	58.5
Non-nati	ive	481	1.4±0.1	497	6.25±0.11	0.037±0.008	0.690±0.006*	-0.006±0.010	32.9±4.5
	APP	19	2	19	6.68	0.065	0.682	0.004	14.6
	BAR	19	2	19	6.50	0.001	0.702	-0.012	17.1
	BEE	19	2	20	4.58	0.000	0.606	-0.152***	95.1
	BRI	20	1	20	6.21	0.080	0.663	-0.065	58.5
	CAR	18	1	19	5.92	0.000	0.685	0.057	26.8
	CLE	20	1	20	6.30	0.014	0.695	0.014	17.1
	CRA	20	1	20	5.29	0.000	0.706	-0.086*	75.6
	DEV	16	2†	20	6.30	0.038	0.691	-0.066	22.0
	DRU	20	1	20	6.44	0.042	0.692	-0.012	39.0
	DUN	19	2	20	7.16	0.103	0.684	0.005	24.4
	ECC	20	2	20	6.32	0.025	0.700	0.036	56.1
	GEL	20	1	20	6.32	0.013	0.700	0.014	17.1
	GOL	20	1	20	5.49	0.048	0.672	0.034	36.6
	HEM	20	1	20	5.86	0.001	0.672	-0.042	70.7
	KIN	20	1	20	5.87	0.013	0.675	-0.005	34.1
	MAB	20	1	20	6.14	0.011	0.701	0.089*	9.8
	PLO	17	2	20	6.04	0.013	0.693	0.019	24.4
:	STR	20	1	20	6.67	0.099	0.692	-0.012	12.2
	TAL	15	2	20	6.28	0.122	0.667	0.018	17.1
	TAN	20	1	20	7.19	0.077	0.747	0.065	51.2
	TON	20	1	20	6.23	0.000	0.680	0.023	24.4
	TWO	20	1	20	6.33	0.014	0.717	0.003	39.0
,	WAL	20	1	20	6.52	0.033	0.708	-0.069	7.3
	WYT	20	2	20	6.79	0.103	0.700	-0.005	4.9
,		-			6.04	0.000	0 722	0.010	26.0
	YEL	19	1	20	6.84	0.000	0.722	-0.012	26.8

774Table 2 Hierarchical analysis of molecular variance (AMOVA) for chloroplast and nuclear775markers for beech in Great Britain. The degrees of freedom (df), percentage of variation776explained by each level (Variation (%)), and the relevant F-statistic are presented with777significant P-values indicated as *** P < 0.001.</td>

779		Chloroplast			Nuclea		
780	Levels	df	Variation (%)	F-statistic	df	Variation (%)	F-statistic
781	Among groups	1	-0.22	<0.001	1	0.01	0.000
782	Among sites						
783	within groups	40	12.59	0.005***	40	2.25	0.022***
784	Within sites	763	87.63	0.032***	795	97.74	0.023***
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801 Figures

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- 803 Fig. 1 Beech in the putative non-native range in Scotland, Great Britain (site CRA, N57.5777
- 804 W4.1435). Photo courtesy of M. J. Sjölund.



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- 816 Fig. 2 Geographical variation of chloroplast haplotypic diversity of beech in Great Britain. A
- total of seven haplotypes are displayed against Birks' (1989) isochrones. Putative native sites
- 818 are outlined.

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Fig. 3 Geographical variation of estimators of beech nuclear genetic diversity in Great Britain. Estimators include rarefied allelic richness (nA_R), rarefied private allelic richness (nA_P), gene diversity (nH_s), and the percentage of significantly differentiated sites (nDS%). Putative native sites are outlined, with Birks' (1989) isochrones for *F. sylvatica* redrawn as broken lines.



Fig. 4 Comparison of isolation-by-distance analyses amongst beech populations in putative
native and putative non-native regions in Great Britain and continental Europe. Sites
included in the analysis are as follows; A) British sites only; B) British and continental sites; C)
putative native sites; D) putative non-native sites.



842 Fig. 5 Regional genetic structure of beech in Great Britain. Three clusters are shown in blue, 843 red, and grey. Each horizontal bar represents an individual with the proportions of its genetic 844 make-up assigned probabilistically to each of the three clusters. Sites are ordered on an 845 approximate geographical gradient by ordering sites following Birks' (1989) isochrones to 846 reflect the putative migration route of Beech into Britain. Continental samples are situated at 847 the bottom of the figure, with a general northward trend to the top of the graph. Stand 848 history of the sites are indicated on the right of the Q-matrix, with continental sites labelled 849 C, putative native sites N, and putative non-native sites left blank. Approximate borders of 850 the isochrones are indicated by a dashed line with years in BP, with site codes on the left.



Fig. 6 Comparison of modelled colonization scenarios of beech in Great Britain. There are four assumed populations of beech: South (SGB), Central (CGB) and North (NGB) populations of Great Britain, and continental Europe (EUR). All scenarios assume colonization from EUR towards SGB and CGB and either assumes NGB originates from EUR (scenario 1 - a); from natural colonization of SGB and CGB (scenario 2 - b); or from the admixture of both EUR and British CGB and SGB (scenario 3 - c). *t1*, *t2*, and *t3* correspond to the divergence times in generations.

